Original Article

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Effect of NRAMP1 gene polymorphism on levels of (TNF-α1 and IL-1β) cytokines in cutaneous Leishmaniasis patients in Iraq

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Abstract

Background: Cutaneous leishmaniasis (CL) is vector-borne disease. and endemics in most regions of Iraq especially with poor populations. "Natural resistance-associated macrophage protein 1 (NRAMP1)" gene play an essential role in susceptibility to CL and disease pathology, NRAMP1 influences a production and activation of pro-inflammatory cytokines like IL-1 β and TNF- α . Proand anti-inflammatory cytokines play an essential role in susceptibility/resistance and the immunopathogenesis of Leishmania infection. These cytokines are crucial factors in the initiation and enhances of protective immunity against Leishmania infection. This study aimed to determine effect of polymorphism in NRAMP1 genes on cytokines secretion and their effect in susceptibility to CL infection.

Materials and Methods: Samples of blood were collected from patients (n: 60) with CL and apparently healthy controls (n: 32). Polymorphism of Nramp1 (D543N) detected by PCR-RFLP technique in patients and control groups while (TNF- α and IL-1 β) cytokine concentrations detected by ELISA technique using a quantitative sandwich enzyme immunoassay technique.

Results: Results indicate to effect of *Nramp1* gene polymorphism on levels of (IL-1 β and TNF- α) cytokines and this a clearly recorded in present study were A allele is associated with lower levels of (TNF- α and IL-1 β) in patients and control groups compression to that absorbed in allele G with statically significant (p \leq 0.05).

Conclusions: Cytokines (IL-1 β and TNF- α) plays an essential role in the resolution of CL infection, were its concentration in patients serum of all age groups were significant increase in comparison to that observed in their control groups. In polymorphisms of *Nramp1* (*D543N*) gene, were A allele is associated with lower levels of (IL-1 β and TNF- α) compression to that absorbed in allele G, and this decreased production may be associated with susceptibility and proliferation of parasites in the macrophage.

Key words: Cutaneous leishmaniasis, NRAMP1polymorphisms, TNF- α and IL-1 β , Cytokine

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Introduction

Cutaneous leishmaniasis (CL) is a parasitic disease transmitted by sand flies, and caused by obligate intra-macrophage protozoa, characteristic by a spectrum of cutaneous, mucocutaneous and visceral diseases that depend largely on the species of the parasite involved and host immune response (1, 2). CL is the most common form of leishmaniasis, with about 1.5 million cases every year, and about 50 to 70% of all cases in the world (2, 3). "Natural resistance-associated macrophage protein 1 (NRAMP1)" gene is a member of the solute carrier family 11 (proton-coupled divalent metal ion transporter), member (A1) SLC11A1 (4, 5). "Homologs of natural resistance-associated macrophage protein (NRAMP) or solute carrier 11 (SLC11), conserved in eukaryotes and bacteria, form a family of proton-coupled transporters that maintain divalent metal (Me²⁺, including Mn²⁺, Fe²⁺, Co²⁺, and Cd²⁺) homeostasis (6, 7). There are two Nramp genes that are associated with diseases in vertebrates, an Nramp1 (also called SLC11A1), in humans, Nramp1 gene is located in the chromosome region 2q35, containing 16 exons (8). NRAMP1 is protein transport divalent metal ions through the phagosomal membrane and might be an essential factor for resistance to some microbial infections. Nramp1 gene plays an essential role in activation of the macrophage pathway. It has many effects on macrophages function as regulation of the CXC chemokine KC, synthesis and activation of pro-inflammatory cytokines as human "Tumor Necrosis Factor-alpha" and human Interleukin-1 beta (7, 9). During an intracellular infection, NRAMP1 protein transports essential elements (Mn²⁺, Fe²⁺, Co²⁺) vital for the survival of the parasite, from the phagolysosome into the cytosol and hence starving and restricting their growth (10). Pro-inflammatory cytokine (TNF-α) primary produce by mononuclear phagocyte, fibroblast, B and T cell, macrophages participate in production of TNF- α , T cell induce macrophages to produce nitric oxide (NO), which cause control or killing parasites, TNF- α that secretion by macrophages also mediate in secretion of nitric oxide as well as activation of macrophages and parasite killing (11). IL-1\(\beta\) is primarily produced by several cells include the monocytes, mononuclear endothelial, keratinocytes, astrocytes, synovial cells, glial cells, osteoblasts, neutrophils, and numerous other cells, there are variant agent like endotoxins, microorganisms, antigens as well as other cytokines which mediate stimulation of IL-1\beta production, which contributes to the immunopathology effects observed in cutaneous leishmaniasis patients (12, 13).

Material and methods

Subjects and study design

A total 32 apparently healthy people and 60 patients with CL were included in this study during the period between February / 2017 to April/ 2017 in the out-patients clinic of the dermatology department in Al-Hussein Teaching hospital and specialized center of sensitivity in Al-Muthanna Province in Iraq. Cases diagnosed clinically by a special dermatologist as cutaneous leishmaniasis and confirmed as CL patients based on clinical symptoms and parasitological parameters (14).

Nramp1 (D543N) Typing

Genomic DNA from blood samples was extracted by using Geneaid DNA extraction kit (Whole Blood), according to the manufacturers' instructions. Polymerase chain reaction was used to amplify a 244 bp fragment. The forward primer was 5-ACT-AAGAAA-GAC-CCG-AGG-C-3 and the reverse primer was 5'-GGG-GCA-CGT-TGG-TGTTTA-C-3. The annealing temperature used was 58°C. Then REFLP-PCR master mix did according to instructions of the company (Biolabs/U.K). The PCR products were digested with Ava II restriction endonuclease. After that, REFLP-PCR product was analyzed by electrophoresis (2.5%) agarose gel, there is three genotypes observed; GG, GA, and AA with band size 126/79/39 pb, 205/ 126/79/39 pb, and 205/39 pb respectively.

Determination of cytokines

Three milliliters of blood in the plain tube (serum tube), then the blood samples were centrifuged at (4700 rpm for 5 min) to obtain blood serum then frozen at -20 °C until the time of test. Serum TNF-alpha and IL1-beta cytokines levels were identified by ELISA technique using a quantitative sandwich enzyme immunoassay technique (EASIA kits for TNF- α and IL-1 β by PeproTech Company/Germany). All tests were done according to company's instruction. The results calculated by ELISA reader (optical density at 405nm immediately) and applied on a standard curve in order to sort out the cytokines concentration.

Statistical analysis

Statistical analysis was conducted by using SPSS version 23. Determine the statistical differences among different groups and associations between allelic and genotypes of Nramp1 gene was performed by using the Pearson Chi-square (x2) test and mean cytokine concentration were compared between groups using t-test (15). The probability of ($p \le 0.05$) was considered to be statistically significant.

Results

Distribution of *Nramp1* (D543N) polymorphism was detected by PCR-RFLP technique, at this locus there is three genotypes; GG, GA and AA with band sizes 126/79/39 pb, 205/ 126/79/39 pb and 205/39 pb respectively. Allele GG was 44 (73.30%) in patients and 18 (56.25%) in control with (p=0.096), Allele GA was 14 (23.3%) in patients and 8 (25%) in control with (p=0.858), and Allele AA was 2 (3.3%) in patients and 6 (18.75%) in control with (p=0.012), in other hand Allele G was (85%) in in patients and (68.75%) in control with (p=0.01), and Allele A was (15%) in patients and (31.25%) in control with (p=0.01).

Figure 1 shows the mean TNF- α interleukin was significant increase in patients group in comparison to control subjects (2.698+0.122ng/ml) versus (0.414+0.015 ng/ml) respectively (p=0.000).

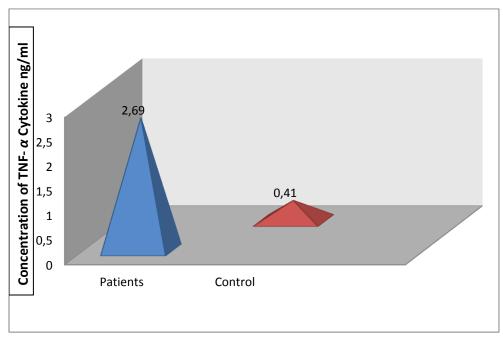


Figure 1. Comparison of mean serum TNF- α cytokine between patient and control groups.

Figure 2 shows the mean IL-1 β interleukin was significant increase in patients group in comparison to control subjects (0.814+ 0.054 ng/ml) versus (0.482+0.020 ng/ml) respectively (p=0.000).

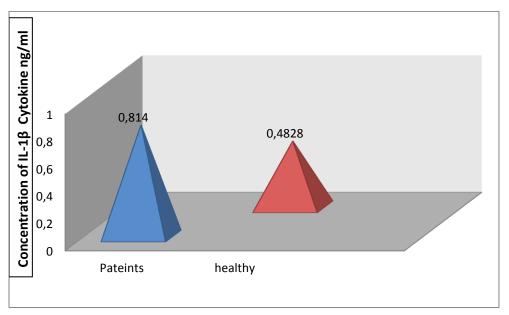


Figure 2. Comparison of concentration mean of IL-1 β cytokine between patient and control groups.

Figure 3 shows the mean TNF- α cytokine concentration according to genotype in *Nramp1* gene. It was found that the mean of TNF- α cytokine level decreased in (*D543N*) A allele in patients groups (2.527±0.104 ng/ml) compression with G allele (2.761±0.162 ng/ml) in patient group (p≤0.05), also it decreased in (*D543N*) A allele

in control groups $(0.402\pm0.0262 \text{ ng/ml})$ compression with observed in G allele $(0.430\pm0.019 \text{ 4ng/ml})$ in control group (p \leq 0.05).

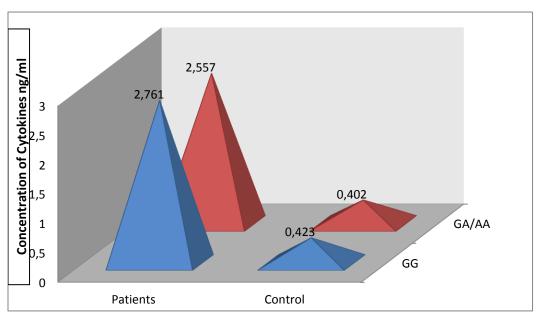


Figure 3. Correlation between Nramp1 (D543N) genotype and serum TNF- α in patient and control groups.

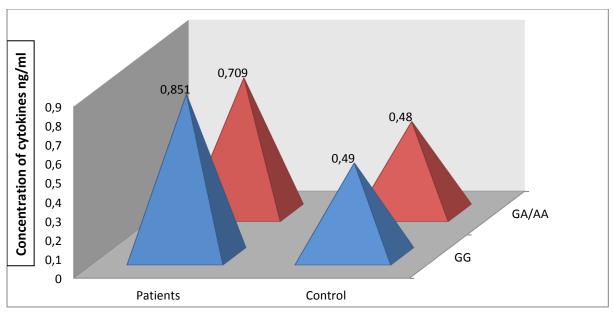


Figure 4. Correlation between Nramp1 (D543N) genotype and serum IL-1 β in patients and control groups.

Figure 4 shows the mean IL-1 β cytokine concentration according to genotype in *Nramp1* gene. It was found that the mean of IL-1 β cytokine level decreased in (D543N) A allele in patients groups (0.709±0.64 ng/ml) compression with G allele (0.852±0.069 ng/ml) in patient group (p≤0.05), also it decreased in (*D543N*) A allele

in control groups $(0.468\pm0.028 \text{ ng/ml})$ compression with observed in G allele $(0.490\pm0.030 \text{ ng/ml})$ in control group $(p\le0.05)$.

Discussion

Present data revealed that TNF- α concentration was a significant increase in patients group in comparison to control group, Figure 1, "this increased expression in cytokine levels might be due to an increase of cellular activation or a relative increase in the number of cytokine-producing cells". The current study finding was agreement with other studies in in Iraq (16) and Turkey (17), the reason in higher concentration of TNF- α in serum of CL patients may be due to responsive to treatment with sodium stibogluconate (pentostam), suggestion generally by that pentostam induce cytokines to activate macrophages (18).

Through present study were found that, the mean concentration of total IL-1 β in all CL patients were significant increase in comparison to that observed in their control groups, figure 2, the current result finding was agreement with other studies in Iraq (16) and Turkey (17) were found serum levels of (IL-1 β) was significant increase in patients group in comparison to control group, as well as (19) which found that IL-1 β concentration in patients infected with Leishmania donovan more than control groups, this result may be due to stimulation essential immunological component in response to pentavalent antimonial (20,21). While, some studies refer to that cytokine secretion important in process of healing following treatment with pentostam (22,23).

In a resting macrophage, Nramp1 gene encoded to protein which assembled into the membrane of late endosome, were phagocytosis it is relocated to the membrane of phagosome (24,25). NRAMP1 protein transport divalent metal ions through the phagosomal membrane and might be an essential factor for resistance to some microbial infections, NRAMP1 induce a variant types of antimicrobial responses of a macrophage, including induction of nitric oxide intermediates and radical oxygen, synthesis and activation of various pro-inflammatory cytokines such as (TNF- α and IL-1 β) (7,26).

However, when mutations occur in the *Nramp1* gene result in a non-functional or unstable protein and then leading to an increased proliferation of parasites in the macrophage might be the reason by deficient antimicrobial responses that confer by NRAMP1 protein (27-29). In current study were result showed that allele A was able to induce less TNF- α secretion in patients and control groups compression with allele G, Figure 3, in another hand when mean serum IL-1 β was studied in relation to allele A, there was decrease in mean serum IL-1 β compression to that absorbed in allele G in patients and control group, figure 4, were unstable NRAMP1protein because mutation results in less expression in (TNF- α and IL-1 β) and resulting in an increased susceptibility and proliferation of parasites in the macrophage, this result agreement with other studies on CL (27, 28).

Conclusion

Cytokines (IL-1 β and TNF- α) plays an essential role in the resolution of CL infection, were its concentration in patients serum of all age groups were significant

increase in comparison to that observed in their control groups. In polymorphisms of Nramp1 (D543N) gene, were A allele is associated with lower levels of (IL-1 β and TNF- α) compression to that absorbed in allele G, and this decreased production may be associated with susceptibility and proliferation of parasites in the macrophage.

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Informed Consent: Yes

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References

- **1.** Ahmed S, Colmenares M, Soong L, Goldsmith-Pestana K, Munstermann L, Molina R, et al. Intradermal infection model for pathogenesis and vaccine studies of murine visceral leishmaniasis. Infection and immunity. 2003;71(1):401-410.
- **2.** Blum J, Desjeux P, Schwartz E, Beck B, Hatz C. Treatment of cutaneous leishmaniasis among travellers. Journal of Antimicrobial Chemotherapy. 2004;53(2):158-166.
- **3.** Tripathi P, Singh V, Naik S. Immune response to leishmania: paradox rather than paradigm. FEMS Immunology & Medical Microbiology. 2007;51(2):229-242.
- **4.** Vidal S, Tremblay ML, Govoni G, Gauthier S, Sebastiani G, Malo D, et al. The Ity/Lsh/Bcg locus: natural resistance to infection with intracellular parasites is abrogated by disruption of the Nramp1 gene. Journal of Experimental Medicine. 1995;182(3):655-666.
- **5.** Mulero V, Searle S, Jenefer M, Brock JH. Solute carrier 11a1 (Slc11a1; formerly Nramp1) regulates metabolism and release of iron acquired by phagocytic, but not transferrin-receptor-mediated, iron uptake. Biochemical Journal. 2002;363(1):89-94.
- **6.** Mackenzie B, Hediger MA. SLC11 family of H+-coupled metal-ion transporters NRAMP1 and DMT1. Pflügers Archiv. 2004;447(5):571-579.
- **7.** Courville P, Chaloupka R, Cellier M. Recent progress in structure–function analyses of Nramp proton-dependent metal-ion transporters This paper is one of a selection of papers published in this Special Issue, entitled CSBMCB—Membrane Proteins in Health and Disease. Biochemistry and Cell Biology. 2006;84(6):960-978.
- **8.** Taype C, Castro J, Accinelli R, Herrera-Velit P, Shaw M, Espinoza J. Association between SLC11A1 polymorphisms and susceptibility to different clinical forms of tuberculosis in the Peruvian population. Infection, Genetics and Evolution. 2006;6(5):361-367.
- **9.** Blackwell JM, Searle S, Goswami T, Miller EN. Understanding the multiple functions of Nramp1. Microbes and infection. 2000;2(3):317-321.
- **10.** Ganz T. Iron in innate immunity: starve the invaders. Current opinion in immunology. 2009;21(1):63-67.
- **11.** Belosevic M, Davis C, Meltzer M, Nacy C. Regulation of activated macrophage antimicrobial activities. Identification of lymphokines that cooperate with IFN-gamma for induction of resistance to infection. J Immunol. 1988;141(3):890-896.
- **12.** Awasthi A, Mathur RK, Saha B. Immune response to Leishmania infection. Indian Journal of Medical Research. 2004;119(6):238.
- **13.** Santos D, Campos TM, Saldanha M, Oliveira SC, Nascimento M, Zamboni DS, et al. IL-1β production by intermediate monocytes is associated with immunopathology in cutaneous leishmaniasis. Journal of Investigative Dermatology 2018;138(5):1107–1115.
- **14.** Da-Cruz AM, Bittar R, Mattos M, Oliveira-Neto MP, Nogueira R, Pinho-Ribeiro V, et al. T-cell-mediated immune responses in patients with cutaneous or mucosal leishmaniasis: long-

term evaluation after therapy. Clinical and diagnostic laboratory immunology. 2002;9(2):251-256.

- **15.** Team RC. R Foundation for Statistical Computing; Vienna, Austria: 2014. R: A language and environment for statistical computing. 2015:2013.
- **16.** Al-Aubaidi IK. Serum cytokine production in patients with cutaneous leishmaniasis before and after treatment. Iraqi journal of medical sciences. 2011;9(1):55-60.
- **17.** Kocyigit A, Gur S, Gurel MS, Bulut V, Ulukanligil M. Antimonial therapy induces circulating proinflammatory cytokines in patients with cutaneous leishmaniasis. Infection and immunity. 2002;70(12):6589-6591.
- **18.** Roberts WL, Rainey PM. Antileishmanial activity of sodium stibogluconate fractions. Antimicrobial Agents and Chemotherapy. 1993;37(9):1842-1846.
- **19.** Sodhi A, Pai K, Singh RK, Singh SM. Activation of human NK cells and monocytes with cisplatin in vitro. International journal of immunopharmacology. 1990;12(8):893-898.
- **20.** Beebe AM, Mauze S, Schork NJ, Coffman RL. Serial backcross mapping of multiple loci associated with resistance to Leishmania major in mice. Immunity. 1997;6(5):551-557.
- **21.** García M, Monzote L, Montalvo AM, Scull R. Screening of medicinal plants against Leishmania amazonensis. Pharmaceutical biology. 2010;48(9):1053-1058.
- **22.** Murray H, Oca M, Granger A, Schreiber R. Requirement for T cells and effect of lymphokines in successful chemotherapy for an intracellular infection. Experimental visceral leishmaniasis. Journal of Clinical Investigation. 1989;83(4):1253.
- **23.** Rais S, Perianin A, Lenoir M, Sadak A, Rivollet D, Paul M, et al. Sodium stibogluconate (Pentostam) potentiates oxidant production in murine visceral leishmaniasis and in human blood. Antimicrobial agents and chemotherapy. 2000;44(9):2406-2410.
- **24.** Canonne-Hergaux F, Calafat J, Richer E, Cellier M, Grinstein S, Borregaard N, et al. Expression and subcellular localization of NRAMP1 in human neutrophil granules. Blood. 2002;100(1):268-275.
- **25.** Kim J, Lee S, Lee S, Sin C, Shim J, In K, et al. *NRAMP1* genetic polymorphisms as a risk factor of tuberculous pleurisy. The International Journal of Tuberculosis and Lung Disease. 2003;7(4):370-375.
- **26.** Blackwell JM, Searle S, Mohamed H, White JK. Divalent cation transport and susceptibility to infectious and autoimmune disease: continuation of the Ity/Lsh/Bcg/Nramp1/Slc11a1 gene story. Immunology letters. 2003;85(2):197-203.
- **27.** Fritsche G, Nairz M, Werner ER, Barton HC, Weiss G. Nramp1-functionality increases iNOS expression via repression of IL-10 formation. European journal of immunology. 2008;38(11):3060-3067.
- **28.** Nugraha J, Anggraini R. NRAMP1 polymorphism and susceptibility to lung tuberculosis in Surabaya, Indonesia. Southeast Asian Journal of Tropical Medicine and Public Health. 2011;42(2):338.
- **29.** Wessling-Resnick M. Nramp1 and other transporters involved in metal withholding during infection. Journal of Biological Chemistry. 2015;290(31):18984-18990.



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