



Synthesis of Medicinal Leech (*Hirudo Verbana*) Saliva-Incorporated Hybrid Nanoflowers and Evaluations of Their Biological Applications

Erdal YILMAZ ¹ Fatih Doğan KOCA ¹ Ayşe DEMİRBAŞ ² İzzet Burçin SATICIOĞLU ³ Behzat ÇİMEN ⁴ İsmail ÖÇSOY ⁴

¹ Erciyes Üniversitesi, Veteriner Fakültesi, Su Ürünleri ve Hastalıkları Anabilim Dalı, Kayseri/Türkiye

² Erciyes Üniversitesi, Su Ürünleri Fakültesi, Su Ürünleri Avlama ve İşleme Teknolojisi Bölümü, İşleme Teknolojisi Anabilim Dalı, Rize/Türkiye

³ Uludağ Üniversitesi, Veteriner Fakültesi, Su Ürünleri Hastalıkları Anabilim Dalı, Bursa/Türkiye

⁴ Erciyes Üniversitesi, Eczacılık Fakültesi, Temel Eczacılık Bilimleri, Kayseri/Türkiye

◆ Geliş Tarihi/Received: 05.05.2025

◆ Kabul Tarihi/Accepted: 15.05.2025

◆ Yayın Tarihi/Published: 30.06.2025

Bu makaleye atıfta bulunmak için/To cite this article:

Yılmaz E, Koca FD, Demirbaş A, Satıcıoğlu İB, Çimen B, Öçsoy İ. Synthesis of Medicinal Leech (*Hirudo Verbana*) Saliva-Incorporated Hybrid Nanoflowers and Evaluations of Their Biological Applications. Bozok Vet Sci (2025) 6, (1):***.

Abstract: In this study, medicinal leech (*Hirudo verbana*) saliva-incorporated nanoflower (LS-NF) was synthesized and evaluated its peroxidase mimic and antimicrobial activities at the first time. Herein, we reported the formation of medicinal leech saliva (LS)-incorporated organic-inorganic nanoflower (LS-NF) with peroxidase mimic and antimicrobial activities. The medicinal leech saliva was used as an organic component and copper ion (Cu²⁺) acted as an inorganic part in the synthesis of LS-NF. Leech saliva was obtained from medicinal leeches *Hirudo verbana*, contains various amino acids, and it can react with Cu²⁺ ions in phosphate buffer through its accessible amine groups to form flower-shaped structures. These LS-NFs showed quite uniform and monodispersed morphology. They were systematically characterized with SEM, FTIR, and XRD for evaluation of morphology, bond vibrations, and crystal structure, respectively. The LS-NF acted as a Fenton reagent in the presence of hydrogen peroxide (H₂O₂), then we investigated their peroxidase-like and antimicrobial activities through the Fenton reaction. It was concluded that LS-NFs can be used in various scientific and industrial applications, and also as an effective antimicrobial agent.

Keywords: Medicinal leech saliva, Hybrid nanoflower, Fenton reaction, Peroxidase-like activity, Antimicrobial activities

Tıbbi Sülük Salyası İçeren Çiçek Şekli Hibrit Nanoyapıların Sentezlenmesi Ve Biyolojik Uygulamalarda Değerlendirilmesi

Özet: Bu çalışmada tıbbi sülük (*Hirudo verbana*) salyası kullanılarak çiçek şekilli nano yapılar sentezlenmiş ve ilk defa peroksidaz ve antimikrobiyal aktiviteleri değerlendirilmiştir. Araştırmada, tıbbi sülük salyası (*H. verbana*) kullanarak organik ve inorganik çiçek şekilli nanoyapıların (LS-NF) peroksidaz ve antimikrobiyal etkilere sahip olduğu bildirilmiştir. Çiçek şekilli nano yapıların sentesinde sülük salyası organik bileşen olarak kullanılırken, bakır iyonları (Cu²⁺) ise inorganik bileşen olarak rol almıştır. Birçok amino asit içeren tıbbi sülük salyası fosfat tampon içindeki Cu²⁺ iyonlarının amin grupları üzerinden çiçek şekilli nano yapılar oluşturmaktadır. Bu çiçek şekilli nano yapılar oldukça düzgün ve homojen bir dağılım sergiledi. Sentezlenen bu nano yapılar morfolojik, bağ titreşimleri ve kristal yapıları bakımından SEM, FTIR ve XDR analizleri ile sistematik bir şekilde karakterize edildi. Çiçek şekilli nano yapılar (LS-NF) hidrojen peroksit (H₂O₂) varlığında bir Fenton reaktifi olarak davranmış olup, bunlar peroksidaz benzeri ve antibiyotik aktiviteler bakımından incelendi. Çalışmada, LS-NF'lerin etkili bir antimikrobiyal ajan olmalarının yanı sıra birçok bilimsel ve endüstriyel uygulamada kullanılabileceği sonucuna varılmıştır.

Anahtar Kelimeler: Tıbbi sülük salyası, Hibrit nanoçiçek, Fenton reaksiyonu, Peroksidaze benzeri aktivite, Antimikrobiyal aktiviteler

1. Introduction

The discovery of protein-inorganic hybrid nanoflower (NFs) formation with enhanced stability and catalytic activities was reported for the first time by Zare and co-workers (1). The enzyme incorporated-NFs have received great attention from researchers owing to their unique biological activities, and they have been in use for a variety of applications, including biosensor fabrication, catalysis, dye decolorization, and free-radical polymerization (2-11). Wang et al. reported how the allosteric effect and morphology influence enzymatic activities of α -amylase@CaHPO₄ NFs. In terms of

mechanism, α -amylase was first activated by binding Ca²⁺ into the allosteric site in inactive α -amylase, then amylase@CaHPO₄ NFs showing great activities and stabilities were formed with self-assembly strategy in phosphate buffer (2). Yang et al. demonstrated how to use acetylcholinesterase-based NF as an electrochemical sensor for the detection of dichlorvos via square-wave voltammetry (11). In a further study, Dadi et al. prepared horseradish peroxidase NFs@carbon nanotube (HRP-NF@CNTs) as novel hybrid nanocatalysts with in situ and post-modification procedures. While both HRP-NF@CNT showed an increase in catalytic activities, the HRP-NF@CNT prepared by post-

modification exhibited outstanding exhibited greatly enhanced peroxidase activity and stability owing to benefiting from both the HRP-NF and synergistic effect between CNT and Cu₃(PO₄)₂ crystals (6).

In addition to that, the development of dual enzyme combined NFs opened up a new avenue for multiple functions and cascade-type reactions (12-14). For instance; while Gul et al. synthesized dual enzyme-based NFs consisting of HRP and laccase for effective and simultaneous degradation of a cationic dye and an anionic azo dye, Zhu et al. developed a combination of glucose oxidase (GOx) and HRP enzymes in the NFs for quantitative detection of glucose. Simply, glucose is catalyzed by GOx to produce hydrogen peroxide (H₂O₂), then HRP used the H₂O₂ to oxidize a colorless 3,3',5,5'-Tetramethylbenzidine (TMB) into a blue product (13, 14).

Alternative to enzyme-based NFs, researchers have developed non-enzyme-based NFs and investigate their peroxidase mimic activities in the presence of H₂O₂ through mechanism of Fenton reaction (15-22). The NFs can be formed from organic or biological molecules containing functional groups (amine, carboxyl or thiol and etc). For instance, We et al. used natural amino acids (AA) as organic components to form AA-based NFs with an intrinsic peroxidase-mimic activity (22). Aslan et al. reported ethylenediaminetetraacetic acid (EDTA) based NFs and utilized them as novel irrigation solutions for biofilms formed by *Enterococcus faecalis* and *Candida albicans* (19). To extend this concept, whole plant extracts were employed as organic parts for the synthesis of plant extract-based NFs (23-27). These plant extract-based NFs have been used for various scientific and industrial applications, including the fabrication of novel antimicrobial, catalytic, and dye-removal agents. For instance, Demirbas et al. synthesized NFs using umbilical decussate extract and investigated their peroxidase mimic, antimicrobial, and dye degradation properties against guaiacol, bacteria (*Aeromonas hydrophila*, *Aeromonas sobria*, *Escherichia coli*, *Salmonella enterica*, and *Staphylococcus aureus*) and malachite green, respectively (28).

Medicinal leeches, also known as "blood-sucking leeches" (such as *Hirudo verbana*), secrete saliva that occurs naturally in the salivary glands and has anticoagulant properties (29). It has also been reported in the literature that medicinal leeches secrete saliva containing approximately 60 different proteins (30). In this research, we synthesized leech saliva-incorporated nanoflower (LS-NF) and evaluated its peroxidase mimic and antimicrobial activities at the first time. The various characterization methods were used to elucidate the formation of the LS-NFs. All biological activities of the LS-NF were observed by the addition of H₂O₂, which relied on the Fenton reaction mechanism.

2. Materials and Methods

2.1. Synthesis of Leech Saliva-Incorporated Nanoflower

The medicinal leech saliva used in this study was supplied by a local company called Cansuyu Sülük (Kayseri, Türkiye), which produces *Hirudo verbana* species. The LS-NFs were synthesized by the modification of reported work (22). Briefly, a 120 mM CuSO₄ solution was freshly prepared as a stock solution, then a certain volume (the final concentration of Cu²⁺ is 0.8 Mm) of stock CuSO₄ solution was added into 10 mM phosphate buffered saline (PBS) at pH: 7.4. Leech saliva solution (the final concentration is 0.02 mg/mL) was added into above mixture, then resulting mixture was vigorously vortexed for 30 sn to provide homogeneous interaction between saliva molecules and Cu²⁺ ions. The final mixture was left undisturbed for 3 days of incubation. After incubation, the blue precipitate at the bottom reaction vial can be considered as an indication for the formation of the LS-NFs. The final product, LS-NFs, was washed with deionized water by centrifugation to remove unreacted of metal ions and saliva. The collected LS-NFs were dried in an oven at 60 C. The powder LS-NF was stored for further use.

2.2. Catalytic Activity of Leech Saliva- Incorporated Nanoflower

Briefly, LS-NFs (3 mg) were added to the solution containing H₂O₂ (22.5 mM), guaiacol (45 mM), and PBS (pH 6.8) (23). The reaction volume was manipulated to adjust the final concentrations of H₂O₂, and guaiacol to 7.5 mM and 15 mM, respectively. The oxidation of guaiacol (2-methoxyphenol) to colored 3,3-dimethoxy-4,4-diphenylquinone was recorded by measuring absorbance values at 470 nm by using a UV-Vis spectrophotometer.

2.3. Antimicrobial Activity of Leech Saliva-Incorporated Nanoflower

In terms of standard microorganisms, *Escherichia coli* (E. coli) ATCC 35218 (Gram-negative) and *Staphylococcus aureus* (S. aureus) ATCC 25923 (Gram-positive) used as bacterial strains, and *Candida albicans* (C. albicans) ATCC 10231 used as a fungus (obtained from the Erciyes University Pharmaceutical Microbiology Laboratory collection). Antimicrobial activities of free leech saliva, LS-NF with H₂O₂, LS-NF, and H₂O₂ were evaluated based on Clinical Laboratory Standards Institute (CLSI) guidelines via broth microdilution method.

In the antibacterial activity procedure, the bacterial cells were cultured in Mueller Hinton broth and incubated at 37 °C for 14 hrs, and their concentration was arranged to 0.5 McFarland. Each antimicrobial agent was mixed with bacterial and fungus strains, and each mixture was incubated at 37 °C for 18–24 hrs for bacteria and 48–72 hrs for fungi. All experiments were carried out in triplicate.

3. Results and Discussions

3.1. Characterization of Leech Saliva-Incorporated Nanoflower

In a typical NF synthesis procedure, Cu^{2+} ions react with phosphate ions (PO_4^{3-}) in PBS to form primary copper phosphate complexes ($\text{Cu}_3(\text{PO}_4)_2$). Enzymes or proteins preferentially bind with Cu^{2+} through their primary amine groups in $\text{Cu}_3(\text{PO}_4)_2$ complexes to create protein- $\text{Cu}_3(\text{PO}_4)_2$ nanocrystals as nuclei. During the incubation process, protein- $\text{Cu}_3(\text{PO}_4)_2$ nanocrystals formed large petals, and each petal was combined with each other to form a flower-shaped structure.

The leech saliva has many polypeptides containing packs of amino acids, so it has various available amine groups to form NFs. The LS-NFs synthesized (in PBS pH 7.4) at room temperature (RT: 25°C) have quite spherical and uniform morphology due to various available amine groups in leech saliva. The LS-NFs are well-monodispersed with diameters of $\sim 3\ \mu\text{m}$ size (Figure 1A).

The boundaries of petals in blooming structures of LS-NF were clearly observed in a magnified SEM image (Figure 1B).

The Cu^{2+} ions are considered as the cornerstone of the LS-NFs because they form primary nanocrystals $\text{Cu}_3(\text{PO}_4)_2$ to form LS-NFs. However, no flower-shaped structure was formed without Cu^{2+} ions. When the Cu^{2+} ions are removed from pre-synthesized LS-NFs, the LS-NFs collapse, and no more flower-shaped morphology is seen. The presence of Cu metal in LS-NF was monitored by Energy dispersive X-ray (EDX) as shown in Figure 1C.

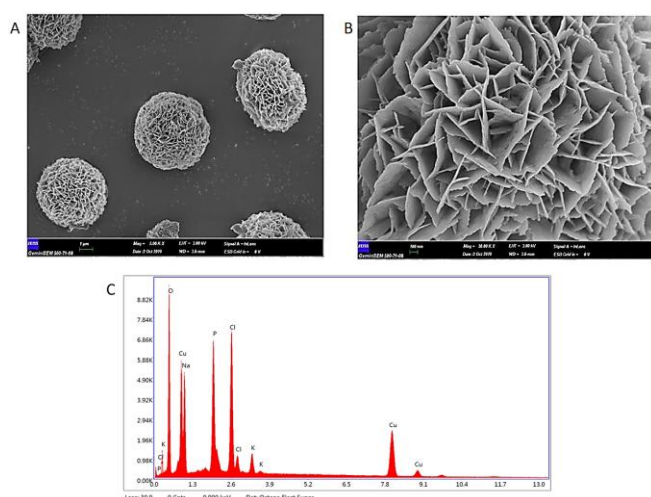


Figure 1. A) SEM image of LS-NFs with 5.00 KX magnification, B) Magnified SEM image of LS-NFs with 30.00 KX magnification. C) The Cu analysis of the LS-NF in the EDX spectrum.

The characteristic stretching and bending vibrations of leech saliva in LS-NFs were analyzed by FTIR analysis, as shown on the FTIR spectrum in Figure 2A.

The amine groups ($-\text{NH}_2$) gave the bending and stretching vibrations were recorded at $1622\ \text{cm}^{-1}$. Additionally, the peak that appeared at $1139\ \text{cm}^{-1}$ was attributed to the stretching of the C-N bond in NH_2 . The stretching vibration of hydroxyl bond ($-\text{OH}$) at $3289\ \text{cm}^{-1}$. The characteristic stretching vibration peaks of PO_4^{3-} (P-O and P=O vibrations in $\text{Cu}_3(\text{PO}_4)_2$ primary crystals were seen at $1097\ \text{cm}^{-1}$ and $557\ \text{cm}^{-1}$. All characteristic stretching and bending peaks can be indications of LS-NF formation.

In addition to that, the crystal structure of LS-NF was elucidated by XRD analysis, as given in Figure 2B.

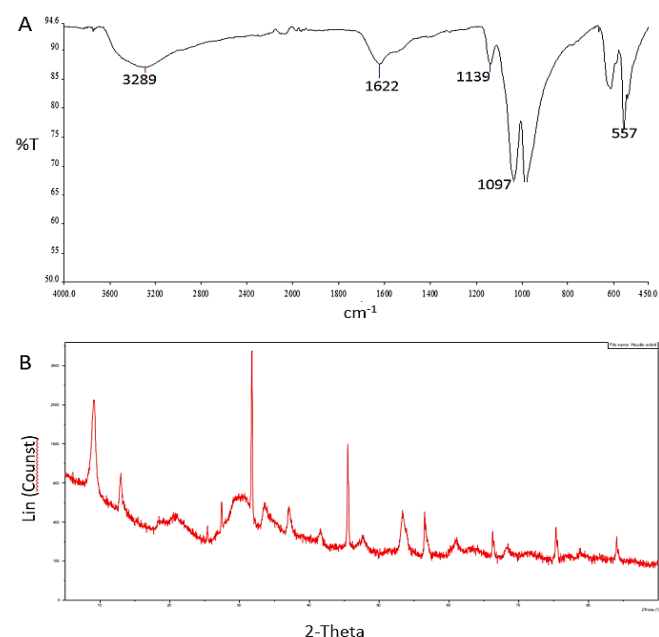


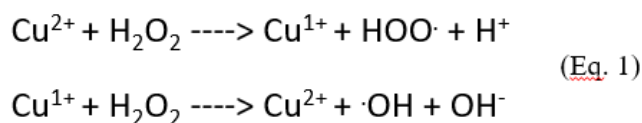
Figure 2. A) FTIR spectrum of LS-NF and B) XRD patterns of LS-NF.

Based on the XRD spectrum, almost all diffraction peaks of $\text{Cu}_3(\text{PO}_4)_2$ in the LS-NFs were consistent with the standard of the JCPDS card (00-022-0548). The distinct and clear diffraction peaks in the LS-NFs were observed in XRD spectrum. The diffraction peaks positions at around $\sim 8.5^\circ$, $\sim 13^\circ$, $\sim 21^\circ$, $\sim 27^\circ$, $\sim 32^\circ$, $\sim 33^\circ$, $\sim 45^\circ$, $\sim 48^\circ$, $\sim 53^\circ$, $\sim 57^\circ$ and $\sim 68^\circ$ in the 2-theta plane. This XRD pattern of LS-NF revealed that its crystal structure is overlapped with the crystal pattern of $\text{Cu}_3(\text{PO}_4)_2 \cdot 3\text{H}_2\text{O}$ (JCPDS card (00-022-0548)).

3.2. Catalytic Activity of Leech Saliva-Incorporated Nanoflower

Peroxidase-like activities of LS-NF were tested by oxidation of guaiacol. The LS-NF exhibited catalytic activity via a Fenton-like reaction mechanism in the presence of H_2O_2 . The Fenton reaction relies on the formation of Cu^{2+} and Cu^{1+} ions in the LS-NF shown in equation 1 (Eq. 1). Briefly, Cu^{2+}

ions in $\text{Cu}_3(\text{PO}_4)_2$ of the-NF were reduced to Cu^{1+} by H_2O_2 , then Cu^{1+} reacted with H_2O_2 to re-create Cu^{2+} ions and to form reactive hydroxyl radical (ROS). This continuous cascade reaction produces much ROS, which causes oxidation of guaiacol used as a standard substrate.



The peroxidase-mimic activity of the LS-NF used as a Fenton reagent (20 $\mu\text{g}/\text{ml}$) was examined towards guaiacol. The LS-NF rapidly and efficiently oxidized guaiacol to convert into an oxidized product called “3,3-dimethoxy-4,4-diphenoquinone”. The formation of the product was monitored by measurement of its absorbance peak at 470 nm via Uv-Vis spectrophotometer (Figure 3).

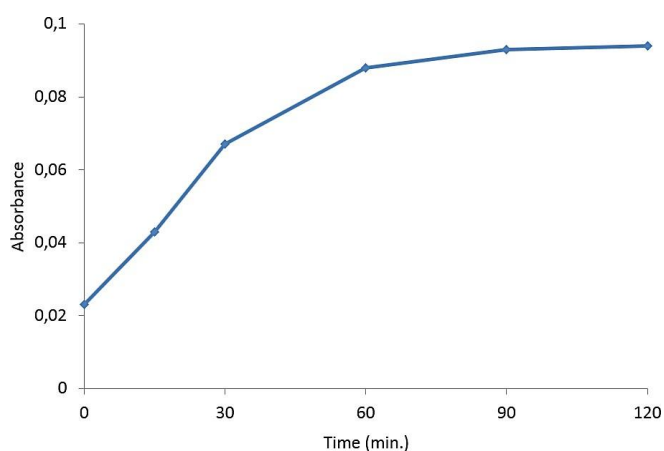


Figure 3. Peroxidase-mimic activity of LS-NF via Fenton reaction.

3.3. Antimicrobial Activity of Leech Saliva-Incorporated Nanoflower

We tested antimicrobial activities of LS-NF with and without H_2O_2 , and antimicrobial activities of the free leech saliva and H_2O_2 were used for comparison towards *E. coli*, *S. aureus*, and *C. albicans* (Figure 4).

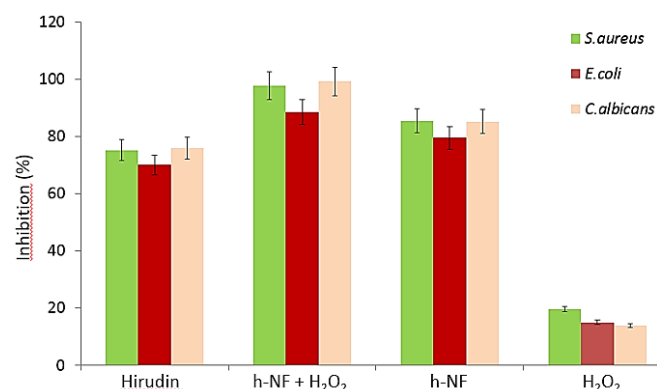


Figure 4 Antimicrobial activities of free leech saliva, LS-NF with H_2O_2 , LS-NF, and H_2O_2 .

The leech saliva in free from displayed kind of effective antimicrobial activities with ~70% inhibition for all three microorganisms. While almost ~85%, ~97%, and ~99% of *E. coli*, *S. aureus*, and *C. albicans* were respectively killed by the LS-NF in the presence of H_2O_2 , only LS-NF without H_2O_2 caused ~78%, ~82%, and ~82% inactivation for *E. coli*, *S. aureus* and *C. albicans*, respectively. In addition to that, the H_2O_2 solution killed around 10% of each bacterial and fungal cell.

We revealed that free leech saliva showed over mild-level antimicrobial activities; however, saliva in NF form exhibited highly effective antimicrobial activities, especially in the presence of H_2O_2 , which can be attributed to morphology and ROS production capability of the LS-NFs. The ROS produced by the LS-NF oxidizes the membrane of each cell, and cell membrane rigidity is decomposed, then eventual cell death is observed.

In conclusion, we have examined formation of medicinal leech (*Hirudo verbana*) saliva-incorporated organic-inorganic nanoflower (LS-NF) with peroxidase mimic and antimicrobial activities through the Fenton reaction. We showed that amino acid rich- leech saliva produced quiet uniform, compact and monodisperse NFs at the first time. The LS-NFs acted as Fenton reagent in the presence of H_2O_2 and generate ROS radicals which are in charge of intrinsic peroxidase-like and antimicrobial activities. We concluded that LS-NFs can be implemented in various scientific and industrial applications, and also as an effective antimicrobial agent.

Acknowledgements

This work was supported by the Erciyes University Scientific Research Projects Coordination Office, Project Number: FCD-2018-8410.

Declarations

Conflict of interest

The authors declare no competing interests.

Ethical approval

Not applicable

Consent to participate

Non applicable

Consent to publish

All authors consent to the publication of this manuscript

Reference

- Ge J, Lei J, Zare RN. Protein-inorganic hybrid nanoflowers. *Nature nanotechnology* 2012; 7: 428-432, 2012. doi: 10.1038/nnano.2012.80.

2. Wang LB, Wang YC, He R. A new nanobiocatalytic system based on allosteric effect with dramatically enhanced enzymatic performance. *Journal of American Chemical Society* 2013; 135(4): 1272-1275. doi:10.1021/ja3120136.
3. Ozaydin G, Mirioglu M, Dadi S. Investigation of the free-radical polymerization of vinyl monomers using horseradish peroxidase (HRP) nanoflowers. *Polymer Bulletin* 2025; <https://doi.org/10.1007/s00289-025-05664-z>.
4. Ozaydin G, Mirioglu M, Kaplan N: Horseradish peroxidase (HRP) nanoflowers-mediated polymerization of vinyl monomers. *Journal of Polymer Research* 2024; 31: 363, 2024. doi:10.1007/s10965-024-04217-8.
5. Gokturk E, Ocsoy İ, Turac E, Sahmetlioglu E: Horseradish peroxidase-based hybrid nanoflowers with enhanced catalytical activities for polymerization reactions of phenol derivatives. *Polymers For Advanced Technologies* 2020; 31:2371–2377. doi: 10.1002/pat.4956.
6. Dadi S, Temur N, Gul OT, Yilmaz V, Ocsoy I. In Situ Synthesis of Horseradish Peroxidase Nanoflower@Carbon Nanotube Hybrid Nanobiocatalysts with Greatly Enhanced Catalytic Activity. *Langmuir* 2023b; 39(13): 4819-4828. doi:10.1021/acs.langmuir.3c00260.
7. Dadi S, Ocsoy I. Role of pretty nanoflowers as novel versatile analytical tools for sensing in biomedical and bioanalytical applications. *Smart Medicine* 2024; 3(1): e20230040. doi:10.1002/SMMD.20230040.
8. Tran TD, Kim MI. Organic-inorganic hybrid nanoflowers as potent materials for biosensing and biocatalytic applications. *BioChip Journal* 2018; 12(4):268–279. doi: 10.1007/s13206-018-2409-7.
9. Cheon HJ, Adhikari MD, Chung M, Tran TD, Kim J, Kim MI. Magnetic Nanoparticles-Embedded Enzyme-Inorganic Hybrid Nanoflowers with Enhanced Peroxidase-Like Activity and Substrate Channeling for Glucose Biosensing. *Advanced Healthcare Materials* 2019; 8(9): e1801507. doi:10.1002/adhm.201801507.
10. Wang Q, Li X, Ren Y. Rapid and precise treatment selection for antimicrobial-resistant infection enabled by a nano-dilution SlipChip. *Biosensors and Bioelectronics* 2025; 271: 117084. doi:10.1016/j.bios.2024.117084.
11. Yang L, Zhang X, Li M, Qu L, Liu Z. Acetylcholinesterase-Cu₃(PO₄)₂ hybrid nanoflowers for electrochemical detection of dichlorvos using square-wave voltammetry. *Analytical Methods* 2022; 14(39): 3911-3920. doi:10.1039/d2ay01014c.
12. Gul OT, Ocsoy I: Co-Enzymes based nanoflowers incorporated-magnetic carbon nanotubes: A new generation nanocatalyst for superior removal of cationic and anionic dyes with great repeated use. *Environmental Technology and Innovation* 2021a; 24: 101992. doi: 10.1016/j.eti.2021.101992.
13. Gul OT, Ocsoy I. Preparation of magnetic horseradish peroxidase-laccase nanoflower for rapid and efficient dye degradation with dual mechanism and cyclic use. *Materials Letters* 2021b; 303: 130501. doi: 10.1016/j.matlet.2021.130501.
14. Zhu X, Huang J, Liu J, Zhang H, Jiang J, Yu R. A dual enzyme-inorganic hybrid nanoflower incorporated microfluidic paper-based analytic device (μPAD) biosensor for sensitive visualized detection of glucose. *Nanoscale* 2017; 9 (17): 5658-5663. doi:10.1039/c7nr00958e.
15. Koca FD, Demirezen Yilmaz D, Ertas Onmaz N, Ocsoy I. Peroxidase-like activity and antimicrobial properties of curcumin-inorganic hybrid nanostructure. *Saudi Journal of Biological Sciences* 2020; 27(10): 2574-2579. doi:10.1016/j.sjbs.2020.05.025.
16. Yilmaz SG, Demirbas A, Karaagac Z, Dadi S, Celik C et al. Synthesis of taurine-Cu₃(PO₄)₂ hybrid nanoflower and their peroxidase-mimic and antimicrobial properties. *Journal of Biotechnology* 2022; 343: 96-101. doi:10.1016/j.jbiotec.2021.11.009.
17. Dadi S, Celik C, Mandal AK, Ocsoy I. Dopamine and norepinephrine assistant-synthesized nanoflowers immobilized membrane with peroxidase mimic activity for efficient detection of model substrates. *Applied Nanoscience* 2021; 11: 117–125. doi:10.1007/s13204-020-01577-7.
18. Dadi S, Cardoso MH, Mandal AK, Franco OL, Ildiz N, et al. Natural Molecule-Incorporated Magnetic Organic-Inorganic Nanoflower: Investigation of Its Dual Fenton Reaction-Dependent Enzyme-Like Catalytic Activities with Cyclic Use. *ChemistrySelect* 2023a; 8: e202300404. doi: 10.1002/slct.202300404.
19. Aslan T, Dadi Ş, Kafdag O, Temur N, Ildiz N et al. Rational design of EDTA-incorporated nanoflowers as novel and effective endodontic disinfection against biofilms. *Odontology* 2024; 112(2): 444-452. doi:10.1007/s10266-023-00857-2.
20. Dadi S, Celik C, Ocsoy I: Gallic acid nanoflower immobilized membrane with peroxidase-like activity for m-cresol detection. *Science Reports* 2020; 10(1): 16765. doi:10.1038/s41598-020-73778-7.
21. Altinkaynak C, Ildiz N, Baldemir A, Ozdemir N, Yilmaz V, et al. Synthesis of organic-inorganic hybrid nanoflowers using *Trigonella foenum-graecum* seed extract and investigation of their antimicrobial activity. *Derim* 2019; 36 (2): 159-167. doi:10.16882/derim.2019.549151.
22. Wu ZF, Wang Z, Zhang Y, Ma YL, He CY, et al. Amino acids-incorporated nanoflowers with an intrinsic peroxidase-like activity. *Science Reports* 2016; 6: 22412. doi: 10.1038/srep22412.
23. Ildiz N, Baldemir A, Altinkaynak C, Özdemir N, Yilmaz V, et al. Self assembled snowball-like hybrid nanostructures comprising *Viburnum opulus* L. extract and metal ions for antimicrobial and catalytic applications. *Enzyme and Microbial Technology* 2017;102: 60-66. doi:10.1016/j.enzmitec.2017.04.003.
24. Baldemir A, Köse NB, Ildiz N, Ilgun S, Yusufbeyoglu S, et al. Synthesis and characterization of green tea (*Camellia sinensis* (L.) Kuntze) extract and its major components-based nanoflowers: a new strategy to enhance antimicrobial activity *RSC Advances* 2017; 7: 44303-44308. doi: 10.1039/C7RA07618E.
25. Baldemir Kılıç A, Ildiz N, Yusufbeyoglu S, Ocsoy I. Nanoflower synthesis formed at different pH based on *Crocus sativus* L. (Crocus stigma, saffron) extract and its major components: a new approach for enhancing antioxidant, antimicrobial and catalytic activities. *Inorganic and Nano-Metal Chemistry* 2023; 55(1): 18–28. doi: 10.1080/24701556.2023.2240757.
26. Kalaycı B, Kaplan N, Mirioglu M, Dadi Ş, Ocsoy İ, et al. Investigation of Peroxidase-Like Activity of Flower-Shaped Nanobiocatalyst from *Viburnum Opulus* L. Extract on the Polymerization Reactions. *Jotcsa* 2024;11(3):1321-1328. doi: 10.18596/jotcsa.1451444.

27. Heidari N, Tarahhomi A, van der Lee A. Structural and Molecular Packing study of Three New Amidophosphoric Acid Esters and Assessment of Their Inhibiting Activity Against SARS-CoV-2 by Molecular Docking. *ChemistrySelect* 2022; 7 (29): e202201504. doi:10.1002/slct.202201504.
28. Demirbas A, Karsli B, Dadi S, Arabaci N, Koca FD, Halici MG, et al. Formation of Umbilicaria decussata (Antarctic and Turkey) Extracts Based Nanoflowers with Their Peroxidase Mimic, Dye Degradation and Antimicrobial Properties. *Chemical Biodiversity* 2023; 20(8): e202300090. doi:10.1002/cbdv.202300090.
29. Haycraft JB. On the action of a secretion obtained from the medicinal leech on the coagulation of the blood. *Proc R Soc Lond.*; 36: 478-87, 1883.
30. Baskova IP, Zavalova LL, Basanova AV, Moshkovskii SA, Zgoda VG. Protein profiling of the medicinal leech salivary gland secretion by proteomic analytical methods. *Biochemistry* 2004; 69(7):770-775. doi:10.1023/b:biry.0000040202.21965.2a.
31. Minuti A, Ahmed S, Trevisi E, Piccioli-Cappelli F, Bertoni G, Jahan N, et al. Experimental acute rumen acidosis in sheep: consequences on clinical, rumen, and gastrointestinal permeability conditions and blood chemistry. *Journal of Animal Science*, 2014; 92(9): 3966-3977.
32. Moeser AJ, Klok CV, Ryan KA, Wooten JG, Little D, Cook VL, et al. Stress signaling pathways activated by weaning mediate intestinal dysfunction in the pig. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 2007; 292(1): G173-G181. Kvidera SK, Horst EA, Abuajamieh M, Mayorga EJ, Fernandez MV, Baumgard LH. Glucose requirements of an activated immune system in lactating Holstein cows. *Journal of Dairy Science*, 2017; 100(3): 2360-2374.
33. Wood KM, Palmer SI, Steele MA, Metcalf JA, Penner GB. The influence of age and weaning on permeability of the gastrointestinal tract in Holstein bull calves. *Journal of Dairy Science*, 2015; 98(10): 7226-7237.
34. Zhang S, Albornoz RI, Aschenbach JR, Barreda DR, Penner GB. Short-term feed restriction impairs the absorptive function of the reticulo-rumen and total tract barrier function in beef cattle. *Journal of Animal Science*, 2013; 91(4): 1685-1695.
35. Abuajamieh M, Kvidera SK, Fernandez MV, Nayeri A, Upah NC, Nolan EA, et al. Inflammatory biomarkers are associated with ketosis in periparturient Holstein cows. *Research in Veterinary Science*, 2016; 109: 81-85.
36. Araujo G, Yunta C, Terré M, Mereu A, Ipharraguerre I, Bach A. Intestinal permeability and incidence of diarrhea in newborn calves. *Journal of Dairy Science*, 2015; 98(10): 7309-7317.