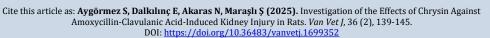


### Van Veterinary Journal

https://dergipark.org.tr/tr/pub/vanveti





ISSN: 2149-3359 Original Article e-ISSN: 2149-8644

# Investigation of the Effects of Chrysin Against Amoxycillin-Clavulanic Acid-Induced Kidney Injury in Rats

Serpil AYGÖRMEZ<sup>1,\*</sup> Elif DALKILINÇ<sup>2</sup> Nurhan AKARAS<sup>3</sup> Şaban MARAŞLI<sup>1</sup>

- <sup>1</sup> Kafkas University, Faculty of Veterinary Medicine, Department of Biochemistry, 36100, Kars, Türkiye <sup>2</sup> Ataturk University, Faculty of Veterinary Medicine, Department of Biochemistry, 25010, Erzurum, Türkiye <sup>3</sup> Aksaray University, Faculty of Medicine, Department of Histology and Embryology, 68100, Aksaray, Türkiye
  - Received: 14.05.2025 Accepted: 22.07.2025

#### ABSTRACT

The amoxycillin-clavulanic acid (ACA) used in the treatment of various bacterial infections often causes druginduced tissue damage, but the mechanism of this damage has not yet been fully elucidated. Chrysin (CHR) is a natural flavonoid with various pharmacological properties as well as antioxidant and anti-inflammatory properties. In this study, the protective effect of CHR against ACA-induced kidney damage, which is frequently used in human and animal health, was investigated. Twenty-eight female rats were divided into four groups as control, CHR, ACA and ACA+CHR. ACA (30 mg/kg) and CHR (50 mg/kg) were administered orally once a day for seven days. Renal function, oxidative stress and inflammation parameters were analyzed to determine renal tissue damage. Histopathologic analysis was also performed to detect tissue damage and structural changes. According to the data obtained from these analyses, ACA increased urea and creatinine levels in kidney tissue. ACA administration also increased malondialdehyde (MDA) and decreased glutathione peroxidase (GPx), superoxide dismutase (SOD), catalase (CAT) activities and glutathione (GSH). Nuclear factor kappa B (NF- $\kappa$ B), tumor necrosis factor-alpha (TNF- $\alpha$ ) and interleukin 1 $\beta$  (IL-1 $\beta$ ) expression levels were found to increase. Administration of CHR together with ACA decreased urea, creatinine, MDA, NF-κB, TNF-α, IL-1β levels and increased GSH level and GPx, SOD, CAT activities. When the findings were evaluated together, it was determined that ACA caused renal damage by increasing renal function levels, oxidative stress and inflammation, while supportive treatment of CHR reduced renal damage by bringing these parameters closer to normal.

Keywords: Amoxycillin-Clavulanic Acid, Chrysin, Oxidative stress, Inflammation, Kidney.

### öz Sıçanlarda Amoksisilin/Klavulanik Asit ile Oluşturulan Böbrek Hasarına Karşı Krisin'in Etkilerinin Araştırılması

Çeşitli bakteriyel enfeksiyonların tedavisinde kullanılan amoksisilin-klavulanik asit (ACA), çoğunlukla ilaç kaynaklı doku hasarına neden olmaktadır fakat bu hasarın mekanizması henüz tam olarak aydınlatılmamıştır. Krisin (CHR), antioksidan ve antiinflamatuar gibi özelliklerinin yanı sıra çeşitli farmakolojik özellikleri bulunan doğal bir flavonoiddir. Bu çalışmada, insan ve hayvan sağlığında sıklıkla kullanılan ACA'nın neden olduğu böbrek hasarına karşı CHR'nin koruyucu etkisi araştırıldı. Yirmi sekiz adet dişi sıçan kontrol, CHR, ACA ve ACA+CHR olmak üzere dört grup ayrıldı. ACA (30 mg/kg) ve CHR (50 mg/kg) yedi gün boyunca günde bir kez oral yoldan uygulandı. Böbrek dokusunda hasarı belirlemek için böbrek fonksiyonu, oksidatif stres ve inflamasyon parametreleri analiz edildi. Doku hasarı ve yapısal değişiklikleri tespit etmek için histopatolojik analiz yapıldı. Bu analizler sonucunda elde edilen verilere göre ACA, böbrek dokusunda üre ve kreatin düzeylerini artırdı. ACA uygulamasının aynı zamanda malondialdehit (MDA)'i arttırdığı, glutatyon peroksidaz (GPx), süperoksit dismutaz (SOD), katalaz (CAT) aktivitelerini ile glutatyon (GSH)'u azalttığı bulundu. Nuclear factor kappa B (NF-κB), tumor necrosis factor-alpha (TNF-α) ve interleukin 1β (IL-1β) ekspresyon düzevlerini artırdığı tespit edildi. CHR'nin ACA ile birlikte uygulanmasının üre, kreatin, MDA, NF-κB, TNF-α, IL-1β düzeylerini azalttığı, GSH düzeyi ile GPx, SOD, CAT aktivitelerini artırdığı belirlendi. Elde edilen bulgular birlikte değerlendirildiğinde, ACA'nın böbrek fonksiyon düzeylerini, oksidatif stresi ve inflamasyonu artırarak böbrek hasarına neden olduğu, CHR'nin destekleyici tedavisinin ise bu parametreleri normale yakınlaştırarak böbrekte hasarı azalttığı tespit edildi.

Anahtar Kelimeler: Amoksisilin/Klavulanik Asit, Krisin, Oksidatif stres, İnflamasyon, Böbrek.

 $\odot$ 

#### INTRODUCTION

Kidneys play a role in essential functions in the organism, such as detoxification of toxic substances and excretion of some drugs. Therefore, they can be exposed to many drugs or harmful substances, including antibiotics, through blood flow (Abouzed et al. 2021; Daoudi et al. 2025). Antibiotics, which are widely used in the treatment of bacterial infections, may cause kidney damage by causing the formation of free oxygen radicals due to their side effects (Badr et al. 2025; Daoudi et al. 2025). Although the exact cause of Amoxycillin-clavulanic acid (ACA)-induced tissue damage has not yet been proven, it is thought that oxidative stress plays an important role in the pathogenesis of drug-induced tissue damage and its induction is associated with tissue damage (El-Hosseiny et al. 2016; Jamshidi and Negintaji 2021). When the endogenous antioxidant system is depleted in the organism, reactive oxygen species (ROS)-mediated oxidative stress develops and tissue damage occurs, causing the onset of adverse outcomes (El-Emam et al. 2023). Interest in plant-derived compounds that can be used in the treatment of many diseases with their antiinflammatory and antioxidant properties is increasing in research (Kankılıç et al. 2024a).

Flavonoids, which are effective in various health problems and abundant in fruits and vegetables to prevent diseases caused by oxidative stress, have been reported to be effective against multidrug resistance with their antioxidant effect (Temel et al. 2021; Öztürk et al. 2025). Chrysin (5,7-dihydroxyflavone) (CHR), found in honey, many plants and propolis, is a natural flavonoid used for therapeutic purposes (Aksu et al. 2018; Küçükler et al. 2022). It has been stated that CHR, which has no side effects, has many pharmacological effects as well as antioxidant and anti-inflammatory effects (Çelik et al. 2020; Akaras et al. 2023a). Thanks to these properties, it is effective in reducing or preventing many tissue damages (Şimşek et al. 2023; Varışlı et al. 2023).

In the present study, we aimed to investigate the effects of CHR on kidney damage caused by ACA, which is frequently used in human and animal health treatment.

#### **MATERIAL AND METHODS**

### Chemicals

ACA (Amoclavin®-BID, Tablet, 1000 mg, Tekirdağ) and CHR (Sigma, Cas No: 480-40-0, 97% purity) were commercially available.

#### **Groups and Experimental Procedures**

In this study, 28 female *Sprague Dawley* rats (220-250 g) were obtained from Atatürk University Experimental Research and Application Center (ATADEM) (Erzurum/Turkey). Ethical approval was obtained from Atatürk University Animal Experiments Local Ethics Committee (Approval No: 2025/01/06, Date: 30.01.2025). Rats were housed in cages in an environment with a temperature of 24-25 °C and 12 h dark-light cycle. The rats were randomly divided into four groups (n=7).

**Control:** Saline was given orally for seven days.

Chrysin (CHR): 50 mg/kg CHR was given orally for seven days (Kankılıç et al. 2024a).

**Amoxycillin-Clavulanic Acid (ACA):** 30 mg/kg ACA was given orally for seven days (Mohammed et al. 2024).

**Amoxycillin-Clavulanic** Acid+Chrysin (ACA+CHR): Animals were given ACA (30 mg/kg) orally, followed by CHR (50 mg/kg) half an hour later. Treatment continued for seven days.

One day after the last ACA and CHR administration, blood samples were collected from the jugular vein under mild sevoflurane (Sevorane®; Queenborough, UK) anesthesia, followed by kidney tissues. Blood samples were collected in 5 mL vacuum gel tubes for urea and creatinine analyses. Serum was obtained by centrifugation at 3000 rpm for 10 minutes. One kidney tissue and serum sample was stored at -80 °C for biochemical analyses, while the other kidney tissue sample was stored in 10% formaldehyde solution for histological examination.

#### **Kidney Function Analyses**

Serum creatinine, urea levels were analyzed using commercial kits (Diasys Diagnostic Systems, Istanbul, Turkey) to evaluate renal function.

#### **MDA and GSH Analyzes**

Total protein content of kidney tissue was determined according to the method of Lowry et al. (1951). Kidney tissue, malondialdehyde (MDA) and glutathione (GSH) levels were homogenized with 1.15% potassium chloride (KCl) and the supernatant was obtained by centrifugation. The absorbance at 532 nm of the color formed by thiobarbituric acid reaction was measured to determine the level of MDA in kidney tissue (Placer et al. 1966). GSH level was analyzed according to the method of Sedlak and Lindsay (1968).

#### **Antioxidant Enzyme Analysis**

Kidney tissue was homogenized with 1.15% KCl for superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT) activities and the supernatant was obtained by centrifugation. SOD activity was analyzed according to Sun et al. (1988), CAT activity according to Aebi (1984), and GPx activity according to Lawrence and Burk (1976).

## RNA extraction and real-time polymerase chain reaction (RT-PCR) analysis

According to the manufacturer's guidelines, total RNA was extracted from kidey tissue using QIAzol Lysis Reagent (Qiagen, Germany). The concentration and purity of the RNA samples were evaluated using a NanoDrop® spectrophotometer (BioTek Epoch). cDNA was synthesized from 2  $\mu g$  of total RNA using the Qiagen High-Capacity cDNA Kit (Thermoscientific). RT-PCR was conducted using the Power SYBR Green Master Mix PCR kit (Qiagen) on the Rotor-Gene Q 5plex HRM platform (Qiagen, Germany). The mRNA levels of NF- $\kappa$ B, TNF- $\alpha$ , and IL-1 $\beta$  in the kidney tissues were analyzed in triplicate using gene-specific primers (Table 1).  $\beta$ -actin was used as reference gene. The relative gene expression levels were determined from the Ct value and calculated using the  $2^{-\Delta\Delta Ct}$  method Livak ve Schmittgen, (2001).

#### Histopathological Analysis

Kidney tissue obtained at the end of the experiment were kept in 10% formalin solution for 48 hours for fixative purposes. After fixation, the tissues were washed under tap water overnight and dehydrated in increasing levels of ethanol. They were then clarified with xylene and embedded in paraffin wax. From the paraffin blocks obtained, 5  $\mu$ m sections were taken using a microtome. Kidney tissues were stained with hematoxylin and eosin (H&E) for light microscopy (Olympus Cx43; Japan).

The resulting slides were evaluated using a blinded method and semi-quantitative scoring was performed. The criteria included inflammatory cell infiltration, vascular congestion and glomerular atrophy. The scoring was as follows: 0 = no lesions, 1 = mild lesions (<30%), 2 = moderate lesions (30-50%) and 3 = severe lesions (>50%).

#### **Statistical Analysis**

The data obtained at the end of the study were statistically analyzed using SPSS 26.0 software. Data were presented as mean ± standard error (SEM). Tukey post hoc tests and one-way analysis of variance (ANOVA) were applied for multiple comparisons. Statistical significance was determined at p<0.05 level.

**Table 1.** Primer sequences of genes analyzed in RT-PCR. (RT-PCR: Real-time polymerase chain reaction, NF- $\kappa$ B: Nuclear factor kappa B, TNF- $\alpha$ : Tumor necrosis factor-alpha, IL-1 $\beta$ : İnterleukin 1 $\beta$ ).

Gene	<b>Accession Number</b>	Primers	Product Size (bp)
β-actin	NM_031144.3	F: GGAGATTACTGCCCTGGCTCCTAGC R: GGCCGGACTCATCGTACTCCTGCTT	155
NF-κB	NM_001415012.1	F: CAGCACTCCTTATCAACCACC R: CTCCTGAGCGTTGACTTCTG	125
TNF-α	NM_012675.3	F: ATGGGCTCCCTCTCATCAGT R: GCTTGGTGGTTTGCTACGAC	106
IL-1β	NM_031512.2	F: AGCTCTCCACCTCAATGGAC R: TTGTTTGGGATCCACACTCTCC	187

#### **RESULTS**

#### Effects of ACA and CHR on Markers of Renal Function

The effects of ACA and CHR on the kidney tissue were examined and the findings are given in Figure 1. Urea (Figure 1A) and creatinine (Figure 1B) levels were evaluated to determine the markers of renal function in kidney. According to the data obtained, urea, creatinine levels in the ACA group increased compared to the control and CHR groups (p<0.05) and ACA+CHR treatment was found to be effective in bringing these markers closer to normal levels.

## Effects of ACA and CHR on Oxidative Stress Levels in Kidney Tissue

MDA (Figure 2A), GSH (Figure 2B) levels and SOD (Figure 2C), CAT (Figure 2D) and GPx (Figure 2E) activities were evaluated to determine oxidative stress in kidney tissue. It was found that MDA level increased in ACA group compared to control and CHR groups (p<0.05), while GSH level and SOD, GPx, CAT activities decreased (p<0.05). It was determined that CHR supportive treatment with ACA decreased MDA level and increased CAT, SOD, GPx activities and GSH and strengthened the antioxidant defense system in kidney tissue.

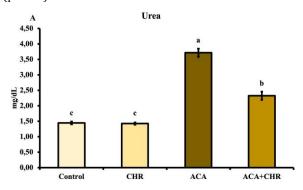
## Effects of ACA and CHR on NF- $\kappa B,\ TNF-\alpha$ and IL-1 $\beta$ Levels in Kidney Tissue

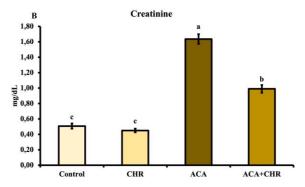
The effects of ACA on the kidney of rats were examined and the findings are shown in Figure 3. TNF- $\alpha$  (Figure 3A), IL-1 $\beta$  (Figure 3B) and NF- $\kappa$ B (Figure 3C) levels were evaluated to determine the level of inflammation in kidney tissue. According to the data obtained, TNF- $\alpha$ , NF- $\kappa$ B, IL-1 $\beta$  levels increased in ACA group compared to control and CHR groups (p<0.05) and ACA+CHR treatment was found to be effective in bringing these markers closer to normal levels.

#### **Histopathologic Results**

Hematoxylin and eosin (H&E) staining results of kidney tissue are shown in Figure 4. No histologic changes were observed in the control and CHR treated groups. In the ACA-treated group, atrophic glomeruli and enlarged bowman spaces were observed. In the tubules, there were degenerative and pyknotic changes in epithelial cells.

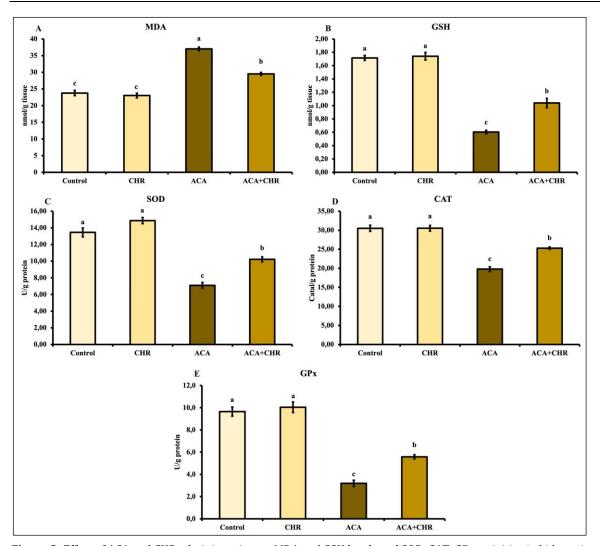
Vascular congestion, hemorrhage and increased inflammatory cells were also observed in interstitial areas. When the images of rats treated with CHR simultaneously with ACA were examined, there were more regular renal bodies and tubules. There was less vascular congestion and inflammatory cells in the interstitial area. According to the histopathologic score results, inflammatory cell infiltration, vascular congestion and glomerular atrophy were significantly increased in the ACA group compared to the control group, whereas they were significantly decreased in the group treated with ACA with CHR (p<0.05).





**Figure 1:** Effect of ACA and CHR administration on urea and creatinine levels in kidney tissue.

Each group values are given as mean  $\pm$  SEM. Different letters in the columns (a-b-c) indicate the difference in the groups (p<0.05). (ACA: Amoxycillin-Clavulanic Acid, CHR: Chrysin).



**Figure 2:** Effect of ACA and CHR administration on MDA and GSH levels and SOD, CAT, GPx activities in kidney tissue. Each group values are given as mean ± SEM. Different letters in the columns (a-b-c) indicate the difference in the groups (p<0.05). (ACA: Amoxycillin-Clavulanic Acid, CHR: Chrysin, MDA: Malondialdehyde, SOD: Superoxide dismutase, CAT: Catalase, GPx: Glutathione peroxidase, GSH: Glutathione).

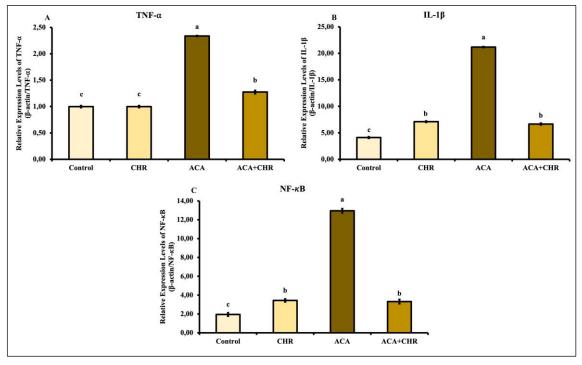


Figure 3: Effect of ACA and CHR administration on NF- $\kappa$ B, TNF- $\alpha$  and IL-1 $\beta$  levels in kidney tissue.

Each group values are given as mean  $\pm$  SEM. Different letters in the columns (a-b-c) indicate the difference in the groups (p<0.05). (ACA: Amoxycillin-Clavulanic Acid, CHR: Chrysin, TNF- $\alpha$ : Tumor necrosis factor-alpha, IL-1 $\beta$ : Interleukin 1 $\beta$ , NF- $\kappa$ B: Nuclear factor kappa B).

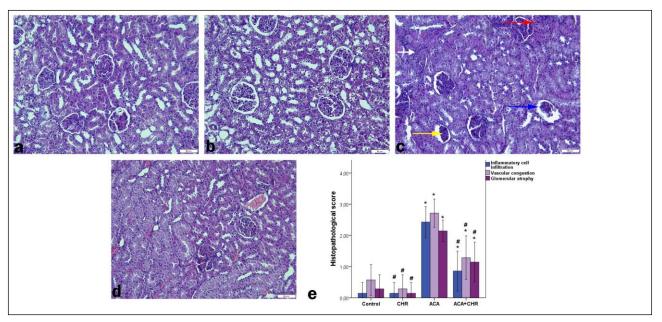


Figure 4. Histopathologic changes in kidney tissues of ACA and CHR treated rats (H&E staining, 200x).

Control (a) and CHR (b) group showing normal architecture of renal tissues, ACA (c) group showing atrophic glomeruli (yellow arrow), enlarged Bowman's spaces (blue arrow), inflammatory cell infiltration (white arrow), vascular congestion (red arrow), ACA+CHR (d) group showing histologic structure similar to control group and histopathologic score (e) graph.

#### **DISCUSSION AND CONCLUSION**

ACA, a broad-spectrum antibiotic, is used in the treatment of bacterial infections and is known to cause tissue damage. This damage is thought to be related to the formation of oxidative stress as a result of increased ROS production and activation of damage pathways such as inflammation triggered by oxidative stress. Therefore, in the present study, we investigated the effects of CHR, which has no side effects, on ACA-induced kidney damage.

Kidney function has an important role in ensuring the overall homeostasis of the organism (Bencheikh et al. 2021). Urea and creatinine, which are important indicators of the structural integrity and function of the kidneys, are the end products of metabolism and are eliminated from the body through the kidneys (Shehata et al. 2022, Gur and Kandemir 2023). In this study, it was found that ACA administration to rats increased urea and creatinine levels in renal tissue and caused damage to the renal tissue as a result. This suggests that the change in pharmacokinetic effect of ACA, along with the changes in urea and creatinine function parameters, may cause damage. In previous studies, it was stated that antibiotic administration caused damage by increasing creatinine and urea levels in kidney tissue as a result of decreased glomerular filtration rate (Babaeenezhad et al. 2024, Abukhalil et al. 2025). Coadministration of CHR, which has no side effects, with ACA was found to reduce the increased creatinine and urea levels in kidney tissue. These results suggest that CHR improves renal function directly or indirectly through its antioxidant effect. Studies show that CHR reduces kidney damage as a result of increasing antioxidant activities of kidney tissue against different toxic agents (Kucukler et al. 2021, Simsek et al. 2023).

Oxidative stress occurs as a result of excessive ROS production in cells and tissues (Jamshidi and Negintaji 2021, Küçükler et al. 2024a). MDA, the most important indicator of oxidative stress, is the end product of polyunsaturated fatty acids (Aydin et al. 2009, Akarsu et al. 2023). SOD, which converts superoxide into hydrogen

peroxide (H<sub>2</sub>O<sub>2</sub>), CAT, which catalyzes oxygen and water, and GPx, which neutralizes it, are antioxidant enzymes of oxidative stress (Akaras et al. 2023b, Gur et al. 2023). GSH neutralizes free radicals (Cakmak et al. 2023, Küçükler et al. 2024b). Oxidative stress increases as a result of decreased antioxidant activity in the organism and increased lipid peroxidation (Kankılıç et al. 2024b, Şimşek et al. 2024, Tuncer et al. 2023). In the present study, it was determined that MDA increased in ACA-induced damage in kidney tissue in rats, oxidative stress developed as a result of decreased GSH and SOD, CAT, GPx enzyme activities, and as a result, tissue damage was caused. The results suggest that ACA causes oxidative stress by producing free radicals and ROS at the cellular level. In different studies on the subject, it has been stated that antibiotics cause kidney damage and oxidative stress due to ROS production produced by drug metabolites (Elgendy et al. 2024, Ekinci Akdemir et al. 2025). Co-administration of CHR with ACA was found to regulate oxidative stress markers in kidney tissue. CHR, a natural flavonoid, is thought to protect kidney tissue by improving antioxidant status and reducing free radical damage. Studies have shown that CHR has the ability to reduce lipid peroxidation and increase antioxidant activities against different toxic agents and prevent kidney tissue damage (Şimşek et al. 2023, Kankılıç et al. 2024a).

Inflammation is significantly triggered by oxidative stress (Yıldız et al. 2022, Yılmaz et al. 2024a). Activation of NF- $\kappa$ B stimulates proinflammatory cytokines (Ileriturk et al. 2022, Yardim et al. 2022, Yılmaz et al. 2024b). Molecules involved in the initiation of acute phase reactions are proinflammatory cytokines such as IL-1 $\beta$ , TNF- $\alpha$  (Akaras et al. 2024, Akarsu et al. 2024). In the present study, it was found that ACA administration increased the mRNA expression levels of NF- $\kappa$ B, TNF- $\alpha$  and IL-1 $\beta$  cytokines, which play an important role in inflammation in the kidney tissue of rats, and as a result, caused damage to the kidney tissue. The results suggest that ACA causes oxidative stress and inflammation by increasing NF- $\kappa$ B and other inflammation parameters. It has been reported that antibiotic administration increases proinflammatory

cytokines in kidney tissue, resulting in kidney damage and dysfunction (Hassanein et al. 2021, Wu et al. 2021, Ayusso et al. 2024). In the present study, co-administration of CHR, which has no side effects, with ACA was found to alleviate inflammatory damage by showing anti-inflammatory effect by reducing the increased IL-1 $\beta$ , NF- $\kappa$ B, TNF- $\alpha$  mRNA expression levels in kidney. These results support that CHR has anti-inflammatory effects by reducing proinflammatory cytokine expression. Studies have reported the therapeutic potential of CHR in reducing inflammation and alleviating histopathological changes (Şimşek et al. 2023, Kankılıç et al. 2024a).

In conclusion, ACA interfered with the damage pathways in renal tissue and caused alterations in markers of oxidative stress, renal function, inflammation, and tissue structural and functional changes. CHR, a natural flavonoid, showed anti-inflammatory and antioxidant effects on ACA-induced kidney injury. These effects suggest that CHR may be considered as a potential therapeutic agent due to its ability to protect the structural integrity of kidney tissue.

#### **CONFLICTS OF INTEREST**

The authors report no conflicts of interest.

#### **AUTHOR CONTRIBUTIONS**

Idea / Concept: SA, ED, NA, ŞM Supervision / Consultancy: SA, ED, NA, ŞM Data Collection and / or Processing: SA, ED, NA Analysis and / or Interpretation: SA, ED, NA, ŞM Writing the Article: SA, ED, NA

Critical Review: SA, ED, NA, ŞM

#### REFERENCES

- **Abouzed TK, Sherif EAE, Barakat MES (2021).** Assessment of gentamicin and cisplatin-induced kidney damage mediated via necrotic and apoptosis genes in albino rats. *BMC Vet Res*, 17 (1), 350.
- Abukhalil MH, Al-Alami Z, Altaie HAA et al. (2025). Galangin prevents gentamicin-induced nephrotoxicity by modulating oxidative damage, inflammation and apoptosis in rats. Naunyn Schmiedebergs Arch Pharmacol, 398 (4), 3717-3729.
- Aebi H (1984). [13] Catalase in vitro. Methods Enzymol. 105, 121-126.
- Akaras N, Ileriturk M, Gur C et al. (2023a). The protective effects of chrysin on cadmium-induced pulmonary toxicity; a multi-biomarker approach. Environ Sci Pollut Res Int, 30 (38), 89479-89494.
- Akaras N, Gur C, Kucukler S, Kandemir FM (2023b). Zingerone reduces sodium arsenite-induced nephrotoxicity by regulating oxidative stress, inflammation, apoptosis and histopathological changes. *Chem Biol Interact*, 374, 110410.
- Akaras N, Kucukler S, Gur C, Ileriturk M, Kandemir FM (2024). Sinapic acid protects against lead acetate-induced lung toxicity by reducing oxidative stress, apoptosis, inflammation, and endoplasmic reticulum stress damage. *Environ Toxicol*, 39 (7), 3820-3832.
- Akarsu SA, Gür C, İleritürk M et al. (2023). Effect of syringic acid on oxidative stress, autophagy, apoptosis, inflammation pathways against testicular damage induced by lead acetate. J Trace Elem Med Biol, 80, 127315
- Akarsu SA, Gür C, Küçükler S (2024). Protective Effects of Syringic Acid Against Oxidative Damage, Apoptosis, Autophagy, Inflammation, Testicular Histopathologic Disorders, and Impaired Sperm Quality in the Testicular Tissue of Rats Induced by Mercuric Chloride. Environ Toxicol, 39 (10), 4803-4814.
- **Aksu EH, Kandemir FM, Küçükler S, Mahamadu A (2018).** Improvement in colistin-induced reproductive damage, apoptosis, and autophagy in testes via reducing oxidative stress by chrysin. *J Biochem Mol Toxicol*, 32 (11), e22201.
- Aydin M, Cevik A, Kandemir FM, Yuksel M, Apaydin AM (2009).

  Evaluation of hormonal change, biochemical parameters, and histopathological status of uterus in rats exposed to 50-Hz electromagnetic field. *Toxicol Ind Health*, 25 (3), 153-158.

- **Ayusso LL, Girol AP, Souza HR et al. (2024).** The anti-inflammatory properties of green tea extract protect against gentamicin-induced kidney injury. *Biomed Pharmacother*, 179, 117267.
- Babaeenezhad E, Dezfoulian O, Sarabi MM, Ahmadvand H (2024). Monoterpene linalool restrains gentamicin-mediated acute kidney injury in rats by subsiding oxidative stress, apoptosis, and the NF- $\kappa$ B/iNOS/TNF- $\alpha$ /IL-1 $\beta$  pathway and regulating TGF- $\beta$ . Naunyn Schmiedebergs Arch Pharmacol, 397 (8), 5701-5714.
- Badr NS, Talaat A, El-Saidy SA, Ghoneim AZA (2025). Revealing Sarcophyton extract's alleviating potential against gentamicin-induced renal and testicular toxicity in rat model. *JOBAZ*, 86, 9.
- Bencheikh N, Bouhrim M, Kharchoufa L et al. (2021). The Nephroprotective Effect of Zizyphus lotus L. (Desf.) Fruits in a Gentamicin-Induced Acute Kidney Injury Model in Rats: A Biochemical and Histopathological Investigation. *Molecules*, 26 (16), 4806.
- Cakmak F, Kucukler S, Gur C et al. (2023). Morin provides therapeutic effect by attenuating oxidative stress, inflammation, endoplasmic reticulum stress, autophagy, apoptosis, and oxidative DNA damage in testicular toxicity caused by ifosfamide in rats. *Iran J Basic Med Sci*, 26 (10), 1227-1236.
- Çelik H, Kucukler S, Çomaklı S et al. (2020). Neuroprotective effect of chrysin on isoniazid-induced neurotoxicity via suppression of oxidative stress, inflammation and apoptosis in rats. Neurotoxicology, 81, 197-208.
- Daoudi NE, Marghich M, Aziz M et al. (2025). Comparative analysis of nephroprotective effects of roasted and unroasted argan seed oils against gentamicin-induced nephrotoxicity in *Wistar* rats. *S Afr J Bot*, 177, 100-108.
- Ekinci Akdemir FN, Yildirim S, Kandemir FM (2025). Protective Effects of Baicalein and Bergenin Against Gentamicin-Induced Hepatic and Renal Injuries in Rats: An Immunohistochemical and Biochemical Study. Basic Clin Pharmacol Toxicol, 136 (1), e14121.
- Elgendy SA, Soliman MM, Shukry M et al. (2024). Screening impacts of Tilmicosin-induced hepatic and renal toxicity in rats: protection by Rhodiola rosea extract through the involvement of oxidative stress, antioxidants, and inflammatory cytokines biomarkers. *Naunyn Schmiedebergs Arch Pharmacol*, 397 (10), 7623-7637.
- El-Emam MMA, Mostafa M, Farag AA et al. (2023). The Potential Effects of Quercetin-Loaded Nanoliposomes on Amoxicillin/Clavulanate-Induced Hepatic Damage: Targeting the SIRT1/Nrf2/NF-κB Signaling Pathway and Microbiota Modulation. *Antioxidants (Basel)*, 12 (8), 1487.
- El-Hosseiny LS, Alqurashy NN, Sheweita SA (2016). Oxidative Stress Alleviation by Sage Essential Oil in Co-amoxiclav induced Hepatotoxicity in Rats. Int J Biomed Sci, 12 (2), 71-78.
- Gur C, Kandemir FM (2023). Molecular and biochemical investigation of the protective effects of rutin against liver and kidney toxicity caused by malathion administration in a rat model. *Environ Toxicol*, 38 (3), 555-565.
- Gur C, Akarsu SA, Akaras N, Tuncer SC, Kandemir FM (2023). Carvacrol reduces abnormal and dead sperm counts by attenuating sodium arsenite-induced oxidative stress, inflammation, apoptosis, and autophagy in the testicular tissues of rats. *Environ Toxicol*, 38 (6), 1265-1276.
- Hassanein EHM, Ali FEM, Kozman MR, El-Ghafar OAMA (2021).

  Umbelliferone attenuates gentamicin-induced renal toxicity by suppression of TLR-4/NF-κB-p65/NLRP-3 and JAK1/STAT-3 signaling pathways. Environ Sci Pollut Res Int, 28 (9), 11558-11571.
- Ileriturk M, Kandemir O, Kandemir FM (2022). Evaluation of protective effects of quercetin against cypermethrin-induced lung toxicity in rats via oxidative stress, inflammation, apoptosis, autophagy, and endoplasmic reticulum stress pathway. Environ Toxicol, 37 (11), 2639-2650.
- Jamshidi HR, Negintaji S (2021). Effects of Thymol on Co-amoxiclav-Induced Hepatotoxicity in Rats. IJML, 8 (1), 44-54.
- Kankılıç NA, Şimşek H, Akaras N et al. (2024a). The ameliorative effects of chrysin on bortezomib-induced nephrotoxicity in rats: Reduces oxidative stress, endoplasmic reticulum stress, inflammation damage, apoptotic and autophagic death. Food Chem Toxicol, 190, 114791.
- Kankılıç NA, Şimşek H, Akaras N, et al. (2024b). Protective effects of naringin on colistin-induced damage in rat testicular tissue: Modulating the levels of Nrf-2/HO-1, AKT-2/FOXO1A, Bax/Bcl2/Caspase-3, and Beclin-1/LC3A/LC3B signaling pathways. *J Biochem Mol Toxicol*, 38 (2), e23643.
- **Kucukler S, Benzer F, Yildirim S et al. (2021).** Protective Effects of Chrysin Against Oxidative Stress and Inflammation Induced by Lead Acetate in Rat Kidneys: a Biochemical and Histopathological Approach. *Biol Trace Elem Res*, 199 (4), 1501-1514.
- Küçükler S, Kandemir FM, Yıldırım S (2022). Protective effect of chrysin on indomethacin induced gastric ulcer in rats: role of multi-pathway regulation. *Biotech Histochem*, 97 (7), 490-503.

- Küçükler S, Çelik O, Özdemir S et al. (2024a). Effects of rutin against deltamethrin-induced testicular toxicity in rats: Biochemical, molecular, and pathological studies. Food Chem Toxicol, 186, 114562.
- Küçükler S, Caglayan C, Özdemir S, Çomaklı S, Kandemir FM (2024b). Hesperidin counteracts chlorpyrifos-induced neurotoxicity by regulating oxidative stress, inflammation, and apoptosis in rats. *Metab Brain Dis*, 39 (4), 509-522.
- Lawrence RA, Burk RF (1976). Glutathione peroxidase activity in selenium-deficient rat liver. Biochem Biophys Res Commun, 71 (4), 952-958
- **Livak KJ, Schmittgen TD (2001).** Analysis of relative gene expression data using real-time quantitative PCR and the  $2-\Delta\Delta$ CT method. *Methods.* 25, 402-408.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951). Protein measurement with the Folin phenol reagent. *J Biol Chem*, 193 (1), 265-275.
- Mohammed H, Galal AT, Rahman MA, Hosny YB (2024). The Possible Ameliorative Effect of Vitamin C Against Amoxicillin-Clavulanic Acid Toxicity in the Liver of Adult Male Albino Rats. SMJ, 28 (2), 64-73.
- Öztürk AB, Şimşek H, Akaras N, Kandemir FM (2025). Chrysin Counteracts Sodium Hydroxide-Induced Alkali Esophageal Burn by Regulating Beclin-1/H0-1/NQ01, PERK/IRE1-α/ATF-6, Oxidative Stress, Inflammation, Apoptosis Signaling Pathways and Ki-67, EGF Expressions in Rats. Bratisl Med I,
- Placer ZA, Cushman LL, Johnson BC (1966). Estimation of product of lipid peroxidation (malonyl dialdehyde) in biochemical systems. *Anal Biochem*, 16 (2), 359-364.
- **Sedlak J, Lindsay RH (1968).** Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal Biochem*, 25, 192-205.
- Shehata AM, Salem FMS, El-Saied EM et al. (2022). Evaluation of the Ameliorative Effect of Zinc Nanoparticles against Silver Nanoparticle-Induced Toxicity in Liver and Kidney of Rats. Biol Trace Elem Res, 200 (3), 1201-1211.
- Sun Y, Oberleyi LW, Li Y (1988). A simple method for clinical assay of superoxide dismutase. Clin Chem, 34 (3), 497-500.
- Şimşek H, Akaras N, Gür C, Küçükler S, Kandemir FM (2023). Beneficial effects of Chrysin on Cadmium-induced nephrotoxicity in rats:

- Modulating the levels of Nrf2/HO-1, RAGE/NLRP3, and Caspase-3/Bax/Bcl-2 signaling pathways. *Gene*, 875, 147502.
- Şimşek H, Gür C, Küçükler S et al. (2024). Carvacrol Reduces Mercuric Chloride-Induced Testicular Toxicity by Regulating Oxidative Stress, Inflammation, Apoptosis, Autophagy, and Histopathological Changes. *Biol Trace Elem Res*, 202 (10), 4605-4617.
- Temel Y, Çağlayan C, Ahmed BM, Kandemir FM, Çiftci M (2021). The effects of chrysin and naringin on cyclophosphamide-induced erythrocyte damage in rats: biochemical evaluation of some enzyme activities in vivo and in vitro. Naunyn Schmiedebergs Arch Pharmacol, 394 (4), 645-654.
- Tuncer SÇ, Akarsu SA, Küçükler S, Gür C, Kandemir FM (2023). Effects of sinapic acid on lead acetate-induced oxidative stress, apoptosis and inflammation in testicular tissue. *Environ Toxicol*, 38 (11), 2656-2667.
- Varişli B, Caglayan C, Kandemir FM et al. (2023). Chrysin mitigates diclofenac-induced hepatotoxicity by modulating oxidative stress, apoptosis, autophagy and endoplasmic reticulum stress in rats. Mol Biol Rep, 50 (1), 433-442.
- Yardim A, Gur C, Selim Comakli S et al. (2022). Investigation of the effects of berberine on bortezomib-induced sciatic nerve and spinal cord damage in rats through pathways involved in oxidative stress and neuro-inflammation. *Neurotoxicology*, 89, 127-139.
- Yıldız MO, Çelik H, Caglayan C et al. (2022). Neuromodulatory effects of hesperidin against sodium fluoride-induced neurotoxicity in rats: Involvement of neuroinflammation, endoplasmic reticulum stress, apoptosis and autophagy. Neurotoxicology. 90, 197-204.
- Yilmaz S, Gur C, Kucukler S, Akaras N, Kandemir FM (2024a). Zingerone attenuates sciatic nerve damage caused by sodium arsenite by inhibiting NF-κB, caspase-3, and ATF-6/CHOP pathways and activating the Akt2/FOXO1 pathway. *Iranian J Basic Med Sci*, 27 (4), 485-491.
- Yılmaz S, Küçükler S, Şimşek H, Aygörmez S, Kandemir FM (2024b). Naringin protects against colistin-induced sciatic nerve damage by reducing oxidative stress, apoptosis and inflammation damage. *JECM*, 41(1), 53-59.
- Wu O, Li W, Zhao J et al. (2021). Apigenin ameliorates doxorubicininduced renal injury via inhibition of oxidative stress and inflammation. Biomed Pharmacother, 137, 111308.