

Black Sea Journal of Agriculture Open Access Journal e-ISSN: 2618 - 6578



**Research Article** 

Volume 1 - Issue 2: 29-33 / April 2018 (Cilt 1 - Sayı 2: 29-33 / Nisan 2018)

### EFFECT OF *FUSARIUM CULMORUM* SPORE SUSPENSION ON MORTALITY OF ROOT LESION NEMATODES IN VITRO CONDITIONS

## Fatma Gül GÖZE ÖZDEMİR<sup>1</sup>\*, Şerife Evrim ARICI<sup>1</sup>, Bülent YAŞAR<sup>1</sup>, Halil İbrahim ELEKCİOĞLU<sup>2</sup>

<sup>1</sup>Süleyman Demirel University, Agricultural Faculty, Department of Plant Pathology, 32260, Isparta, Turkey

<sup>2</sup>Çukurova University, Agricultural Faculty, Department of Plant Protection, 01330, Adana, Turkey

*Submission:* January 26, 2018; *Published:* April 01, 2018 *(Gönderi:* 26 Ocak 2018; *Yayınlanma:* 01 Nisan 2018)

#### Abstract

Three different *Fusarium culmorum* isolates (B4, ISP, Fc5) were used in the study and the effects of these spore suspension of isolates were investigated on the root lesion nematodes *Pratylenchus thornei*, *P. neglectus* and *P. penetrans* in vitro. As a result of evaluating the percent mortality rate, ISP isolate had the highest mortality effect on root nematodes and more effective on *P. thornei*. The lowest mortality effect of isolate was Fc5. However, Fc5 effect on *P. neglectus* was higher than ISP and B4 isolates. Percentage of mortality rate of B4 isolate were found 16.2 on *P. penetrans*, 13.1 on *P. thornei* and 5.5 on *P. neglectus* and showed the lowest effect on *P. neglectus*. It was found that *F.*culmorum spore suspension could suppress root lesion nematodes at low levels in vitro and changed depending on the isolates.

**Keywords:** *Fusarium culmorum*, Root lesion nematode, Spore suspension, Mortality

\*Corresponding author: Süleyman Demirel University, Agricultural Faculty, Department of Plant Pathology, 32260, Isparta, Turkey E mail: fatmagoze@sdu.edu.tr (F.G. GÖZE ÖZDEMİR)

#### 1. Introduction

Root lesion nematodes (*Pratylenchus* spp.) and soil-borne pathogen *Fusarium* spp. are organisms that cause severe damage to grains. Root lesion nematode is one of the most destructive of the three plants parasitic nematode species and is common worldwide. They migrate through root epidermal and cortical cells and cause lesions in the root as a result of feeding (Agrios, 1997). However, they help to induce many soil-borne diseases (Lasserre et al., 1994; Taheri et al., 1994). One of the most economically important genera of phytopathogenic fungi is *Fusarium* 

spp. due to wide host range, presence different geographic region and environmental conditions around the world and produce a large number of toxins (van der Lee et al., 2015). *Fusarium culmorum* (W.G. Smith) Saccardo\_infects roots and cause dry rot that the bottom of the plant appears to be brown. When infected plants are exposed to water stress, the damage increases and can be seen white spike before ripening (Burgess et al., 2001; Smiley et al., 2004). The main mycotoxins of the *F. culmorum* species are trichothecenes, zearalenone, fusarin C, butenolid (Wagacha and Muthomi, 2007). Trichothecenes have immunosuppressive effect even at

low concentrations and they inhibit eukaryotic protein synthesis by binding ribosomes (Gutleb et al., 2002).

Antagonistic relationships between plant parasitic nematodes and soil-borne pathogens were identified and known that many fungi produce nematicidal compounds (Chen et al., 2000; Meyer et al., 2000; Köpcke et al., 2001). It was reported that the presence of Fusarium oxysporum f. sp. *lentis* in the soil were reduced in population density of sedentary endoparasite nematode Meloidogyne incognita (Fazal et al., 1994). To developing and egg laying of Meloidogyne spp. and Heterodera spp. species, nutrients from giant cells or syncytia is necessary. If Fusarium spp. were colonized and consumed nutrients, the female nematode would have died without laying the eggs (Nordmeyer and Sikora, 1983). In addition, the presence of nutritional deficiencies reduced the number of females (Triantaphyllou, 1960). Metabolites produced by some pathogenic fungi were suppressed the egg-laying of the nematode larvae (Vaishnav et al., 1985; Ciancio et al., 1988). Colletotrichum atramentarium reduces cyst formation of Globodera rostochienensis with inhibits larval development, reduces the number of eggs and increased male ratio (Powell, 1971).

While *Fusarium* species are commonly found in soil environments, there must be wide opportunities for these pathogens and their secondary metabolites to interact with plant parasitic nematodes. There are limited studies on the effects of toxins and enzymes of *Fusarium* species on mortality of nematodes (Ciancio, 1995) and no study has been conducted on the effect of *F. culmorum* toxins and enzymes on root lesion nematodes. In order to understand the interactions of *F. culmorum* and root lesion nematodes (*Pratylenchus* spp.), it has been firstly considered necessary to study *in vitro* conditions. In this study, the mortality effect of *F.culmorum* against root lesion nematodes was investigated.

#### 2. Materials and Methods

This study was conducted with Pratylenchus penetrans (Cobb, 1917), P. thornei Sher et Allen and P. neglectus (Rensch) Filipjev Schuurmanns & Stekhoven root lesion nematodes. nematodes distribution These and molecular identification made by Söğüt and Devran, (2011). Fusarium culmorum isolated from Adana (B4), Isparta (ISP) and Ankara, Turkey (Fc5/ provided Suleyman Demirel University, Department of Agricultural Biotechnology) that had highest pathogenicity were used in the study (Arıcı, 2006; Arıcı et al., 2013).

#### 2.1. Preparation of nematode inoculum

The carrot disc method was used in the mass production of root lesion nematodes (Zuckerman et al., 1985). After 2 months, carrot cultures were transferred to 12 cm diameter petri dishes and cut into pieces. Sterile purified water was placed on it and steep for 4-6 hours for nematodes passing nematodes into water. Then, the nematodes were passed through a 38 and 20  $\mu$ m sieve and transfered into a centrifuge tube. The larvae + adult densities were counted under light microscope and taken into tubes and made ready for inoculation.

#### 2.2. Preparation of *Fusarium culmorum* inoculum

*Fusarium culmorum* isolates (B4, ISP, Fc5) were cultured at 24  $\pm$  1°C on PDA (Potato Dextrose Agar) for 10 days. Then, mycelia were scraped with 5 ml of sterile water and filtering through miracloth into sterile tubes. Spore concentration was determined using a haemocytometer and adjusted to 2.5x10<sup>3</sup> spores/ml (Arici, 2006).

# 2.3. Effect of *Fusarium culmorum* Spore Suspension on mortality of Root Lesion Nematodes In Vitro Conditions

*Fusarium culmorum* isolates (B4, ISP, Fc5) and three root lesion nematodes (*P. penetrans, P. thornei, P. neglectus*) were used in the experiment. 200 root lesions nematode larvae + adult and  $2.5 \times 10^3$  spores / mL fungus spores were added simultaneously in the  $35 \times 10$  mm petri dish (Hassan et al., 2012). The dishes were covered with parafilm and they were cultured at  $25^{\circ}C\pm1$  for 3 days. Only nematode-treated petri dishes were taken as controls (C). The treatments were arranged in a randomized complete block design with 3 replicates. After 3 days, live-dead individuals were counted, and percentage deaths were calculated (Liu et al., 2011).

#### 2.4. Statistical analysis

SPSS (version 16.0) program was used for all statistical analysis. One-way ANOVA was conducted to demonstrate variability among treatment means and were separated using Duncan's studentised range test.

#### 3. Results and Discussions

The highest number of living *P. penetrans* individuals was found in the Fc5 *F. culmorum* isolate. The lowest number of living *P. penetrans* individuals determined ISP and B4 isolates, but the differences among them were statistically insignificant ( $p \ge 0.05$ ). On the controry, the number of dead individuals were found to be highest in ISP and B4 isolates and no statistically significant difference was found between them ( $p \ge 0.05$ ). The ISP and B4 isolates had a mortality percentage of 18.6 and 16.2, respectively. The ISP isolate was thought to have a higher mortality than B4. The nematicidal effect of ISP isolate on *P. penetrans* was determinated at low mortality (Table 1).

on r rutylenchus penetruns					
<i>F.</i>	Live	Dead	Percentage		
culmorum	individuals	individuals	of		
isolate			mortality		
K	192.6±1.45 <sup>c*</sup>	$7.3 \pm 1.45^{a}$	$3.6 \pm 0.72^{a}$		
B4	167.6±1.45ª	32.3±1.45 <sup>c</sup>	16.2±0.72 <sup>c</sup>		
ISP	162.6±1.45ª	37.3±1.45 <sup>c</sup>	18.6±0.72 <sup>c</sup>		
Fc5	183.6±1.85 <sup>b</sup>	16.3±1.85 <sup>b</sup>	8.2±0.92b		
a h c Different letters in the same column indicate significant					

**Table 1.** Effect of Fusarium culmorum Spore Suspensionon Pratylenchus penetrans

 $^{\rm a,\ b,\ c}$  Different letters in the same column indicate significant difference (p < 0.05).

**Table 3.** Effect of Fusarium culmorum Spore Suspensionon Pratylenchus thornei

<i>F.</i>	Live	Dead	Percentage		
culmorum	individuals	individuals	of		
isolate			mortality		
К	$180.6 \pm 1.76^{bc^*}$	$19.3 \pm 1.76^{ab}$	$9.6\pm0.8^{ab}$		
B4	$173.6 \pm 6.38^{b}$	$26.3 \pm 6.38^{b}$	13.1±3.19 <sup>b</sup>		
ISP	$147.3 \pm 5.04^{a}$	52.6±5.04 <sup>c</sup>	26.3±2.5°		
Fc5	190.3±2.60 <sup>c</sup>	$9.6 \pm 2.6^{a}$	4.8±1.3 <sup>a</sup>		

 $^{a,\ b,\ c}$  Different letters in the same column indicate significant difference (p < 0.05).

*Pratylenchus neglectus* live individuals were found that the highest in the B4 isolate (189.0), the lowest in the ISP (160.0) and the differences ( $p \le 0.05$ ) among them were statistically significant (Table 2). The number of dead *P. neglectus* individuals were 5.0 in C. The highest mortality between *P. neglectus* (20.0) and *P. penetrans* (18.6) was determined in the application of ISP isolate. The mortality rate of *P. neglectus* was higher than *P. penetrans*, and was found to be at least 5.5 in B4 isolate the mortality effect of Fc5 isolate was higher in *P.neglectus* (12.8) than *P. penetrans* (8.2), whereas B4 isolate was lower in *P.neglectus* (5.5) than *P. penetrans* (16.2). All mortalities rates were found higher than the control group (Table 1 and 2).

**Table 2.** Effect of Fusarium culmorum Spore Suspensionon Pratylenchus neglectus

	0		
<i>F.</i>	Live	Dead	Percentage
culmorum	individuals	individuals	of
isolate			mortality
К	195.0±1,15 <sup>c*</sup>	$5.0 \pm 1.15^{a}$	2.3±0.72 <sup>a</sup>
B4	189.0±2,08 <sup>b</sup>	$11.0 \pm 2.08^{b}$	$5.5 \pm 1.04^{b}$
ISP	160.0±1,15ª	$40.0 \pm 1.15^{d}$	20.0±0.6 <sup>d</sup>
Fc5	174.3±2,3 <sup>b</sup>	25.6±2.3 <sup>c</sup>	12.8±1.16 <sup>c</sup>

<sup>a, b, c, d</sup> Different letters in the same column indicate significant difference (p < 0.05).

*Pratylenchus thornei* live individuals were found the highest in the Fc5 (190.3) and the lowest in ISP (147.3) and the differences among them were statistically significant (p < 0.05). The lowest mortality percentage of *P. thornei* was found in the Fc5 isolate and was lower than the control. Death effect on B4 isolate was lower than ISP, higher than Fc5. Percentage mortalities rate of *P.thornei* were 9.6, on control, 13,1 on B4 and 26.3 on ISP and 4.8 on Fc5 (Table 3).

ISP was found the most effective isolate on three-root lesion nematode (Table 1, 2 and 3). Percentage mortalities rate of ISP were 18.6 on *P. penetrans*, 20.0 on *P. neglectus* and 26.3 on *P. thornei* (Table 1, 2 and 3). Fc5 isolate had the lowest mortality effect among the other isolates (Table 1, 2 and 3). *F. culmorum* isolates did not have the same effect on root lesion nematodes. It suggested that each species of root lesion nematodes could have the morphological and physiological structure.

In the study, it was showed that F. culmorum spore suspension could suppress root lesion nematodes in vitro, even at low levels, and suppressing depending on the lesion nematode species and isolate of *F.culmorum*. Spore suspension of F.culmorum should have a nematicidal effects because of secretion of metabolites, enzymes or mycotoxins (Ciancio, 1995). It has been reported that some Fusarium species was secreted toxic compounds against plant parasitic nematodes and these compounds were affected egg laying, viability and larval mobility (Nitao et al., 1999). El-Borai et al. (2002a) reported that Tylenchus semipenetrans eggs were inhibited by mycelial growth of Phythopthora nicotianae and F. solani in vitro. Metabolites produced by some pathogenic fungi have been suppressed the laying of nematode larvae (Vaishnav et al., 1985, Ciancio et al., 1988).

The results from our study indicated that there may be an antagonistic relationship between F. culmorum and root lesion nematodes (Pratylenchus spp.). Similar results have been found by some investigators to have antagonistic relationships between different nematodes and fungi. It was reported that *T. semipenetrans* population density was increased in citrus plants, while P. nicotinae infection was decreasing (Graham and Duncan, 1997; El-Borai et al., 2002b). In addition, it was determinated negative interaction between Helicotylenchus multicinctus and F. oxysporum f.sp. cubense (Poornima et al., 2007). Sankaralingam and McGawley (1994) reported that there was antagonistic interaction between Rhizoctonia solani and Rotylenchulus reniformis in cotton plant and when the intensity of *R. solani* increases, population density of nematodes decreases significantly. Some researchers were reported that Nematode and fungal species, population density, plant species, variety, line and soil structure was important between nematode and fungal interactions (Riedel et al., 1985; Khan and Husain, 1989; Bowers et al., 1996; Back et al., 2002; Mohiddin and Rahman-Han, 2014). For these reasons, more detailed studies are needed to determinate the association between *F. culmorum* and root lesion nematodes. In addition, the same studies are planned with culture filtrates of *F. culmorum*. The work will be supported by pots and field studies. This planned study will contribute to a better understanding of the antagonistic relationship.

#### References

- Agrios GN. 1997. Plant Pathology, 4th edition. Academic Press, USA. 635 pp.
- Arıcı ŞE. 2006. In Vıtro Selection For Resistans To Head Blight (Fusarium Spp.) Via Somaclonal Variation in Wheat (Triticum aestivum L.) Phd Thesis, University Of Çukurova Institute Of Natural and Applied Sciences Department of Plant, Adana.
- Arıcı ŞE, Arap Ü, Yatağan FB. 2013. Identification of Soilborne Fungal Dieases on Wheat Field in Province of Isparta and Burdur. Suleyman Demirel University J Nat Appl Sci, 17(2): 26-30.
- Back MA, Haydock PPJ, Jenkinson P. 2002. Disease complexes involving plant parasitic nematodes and soilborne pathogens. Plant Pathol, 51: 683-697.
- Bowers JH, Nameth ST, Riedel RM, Rowe RC. 1996. Infection and colonisation of potato roots by *Verticillium dahliae* as affected by *Pratylenchus penetrans* and *P. crenatus*. Phytopathol, 86: 614–621.
- Burgess LW, Backhouse D, Summerell BA, Swan LJ. 2001. Crown rot of wheat. In: Summerell BA, Leslie JF, Backhouse D, Bryden WL, Burgess LW (eds.): Fusarium, pp. 271-294. American Phytopathological Society Press.
- Chen, S.Y., Dickson, D.W., Mitchell, D.J., 2000. Viability of *Heterodera glycines* exposed to fungal filltrates. J Nematol, 32(2): 190-197.
- Ciancio A, Logrieco A, Lamberti F, Bottalicao A. 1988. Nematicidal effects of some Fusarium toxins. Nematol Medit, 16: 137-138.
- Ciancio, A., 1995. Observations on the nematicidal properties of some mycotoxins. Fund Appl Nematol, 18: 451-454.
- Cobb, N.A., 1917. A new parasitic nema found infesting cotton and potatoes. J Agri Res, 11: 27-33.
- El-Borai FE, Duncan LW. Graham JH. 2002a. Infection of citrus roots by *Tylenchulus semipenetrans* reduces root infection by *Phytophthora nicotianae*. J Nematol, 34(4): 384–389.
- El-Borai KF, Duncan LW, Graham JH. 2002b. Eggs of *Tylenchulus* semipenetrans inhibit growth of *Phytophthora nicotianae* and *Fusarium solani* in vitro. J Nematol, 34: 267–272.
- Fazal M, Khan MI, Raza MMA, Siddiqui ZA. 1994. Interaction between *Meloidogyne incognita* and *Fusarium oxysporum* f. sp. *lentis* on lentil. Nematol Medit, 22: 185-187.
- Gutleb AC, Morrison E, Murk AJ. 2002. Cytotoxicity Assay for mycotoxins produced by Fusarium strains. J Envir Toxic Pharm, 11:309-320.
- Graham JH, Duncan LW. 1997. Suppression of *Phytophthora nicotianae* in citrus roots by the citrus nematode (*Tylenchulus semipenetrans*). Phytopathol, 87:35.

- Hassan Gh A, Kh Al-Assas T, Al-Fadil A. 2012. Interactions between *Heterodera avenae* and *Fusarium culmorum* on yield components of wheat, nematode reproduction and crown rot severity. Nematropica, 42: 260-266.
- Khan TA, Husain SI. 1989. Relative resistance of six cowpea cultivars as affected by the concomitance of two nematodes and a fungus. Nematol Medit, 17: 39–41.
- Köpcke B, Wolf D, Anke H, Sterner O. 2001. New natural products with nematicidal activity from fungi. *British Mycological Society International Symposium*, Bioactive Fungal Metabolites – Impact and Exploitation, UW Swansea, UK, 22-27.
- Lasserre F, Rivoal R, Cook R. 1994. Interactions between *Heterodera avenae* and *Pratylenchus neglectus* on wheat. J Nematol, 26: 336-344.
- Liu JH, Wang L, Qiu JY, Jiang Ll, Yan JY, LiuT, Liu WC, Duan YX. 2011. Nematicidal activity of *Gymnoascus reesii* against *Meloidogyne incognita*. African J Microb Res, 5: 2715-2719.
- Meyer SLF, Massoud SI, Chitwood DJ, Roberts DP. 2000. Evaluation of *Trichoderma virens* and *Burkholderia cepacia* for antagonistic activity against root-knot nematode, *Meloidogyne incognita*. Nematology, 2: 871-879.
- Mohiddin FA, Rahman Khan M. 2014. Root-knot nematode: Ignored soil borne plant pathogen causing root diseases of chickpea. European J Biotech Biosci, 2(1): 04-10.
- Nitao JK, Meyer SLF, Chitwood DJ. 1999. In-vitro assays of *Meloidogyne incognita* and *Heterodera glycines* for detection of nematode-antagonistic fungal compounds. J Nematol, 31: 172–183.
- Nitao JK, Meyer SLF, Schmidt WF, Fettinger JC, Chitwood DJ. 2001. Nematode-antagonistic trichothecenesfrom *Fusarium equiseti*. J Chem Ecol, 27: 859-869.
- Nordmeyer D, Sikora RA. 1983. Studies on the interaction between *Heterodera daverti, Fusarium avenaceum* and *F. oxysporum* on *Trifolium subterraneum*. Revue de Nematol, 6: 193–198.
- Powell NT. 1971. Interaction between nematode and fungi in disease complexes. Annual Rev Phytopathol, 9: 253–274.
- Poornima K, Angappan K, Kannan R, Kumar N, Kavino M, Balamohan TN. 2007. Interactions of nematodes with the fungal panama wilt disease of banana and its management. Nematol Medit, 35: 35-39.
- Riedel RM, Rowe RC, Martin MJ. 1985. Differential interactions of *Pratylenchus crenatus, Pratylenchus penetrans* and *Pratylenchus scribneri* with *Verticillium dahliae* in potato early dying disease. Phytopathol, 75: 419–422.
- Sankaralingam A, McGawley EC. 1994. Interrelationships of Rotylenchulus reniformis with *Rhizoctonia solani* on cotton. J Nematol, 26: 475–485.
- Smiley RW, Gourlie JA, Whittaker RG, Easley SA, Kidwell KK. 2004. Economic impact of Hessian fly (Diptera: Cecidomyiidae) on spring wheat in Oregon and additive yield losses with *Fusarium crown* root and lesion nematode. J Econ Entomol, 97: 397-408.
- Sögüt MA, Devran Z. 2011. Distribution and Molecular Identification of Root Lesion Nematodes in Temperate Fruit Orchards of Turkey. Nematropica, 41: 91-99.
- Taheri A, Hollamby GJ, Vanstone VA. 1994. Interaction between root lesion nematode, Pratylenchus neglectus (Rensch 1924) Chitwood and Oteifa 1952, and root rotting fungi of wheat. New Zealand J Crop Horticultural Sci, 22: 181-185.
- Triantaphyllou AC. 1960. Sex determination in *Meloidogyne incognita* Chitwood, 1949 and intersexuality in *M. javanica* (Treub, 1885) Chitwood, 1949. Annals Institut Phytopathol, 3: 12-31.

- Vander Lee T, Zhang H, vanDiepeningen A, Waalwijk C. 2015. Biogeography of *Fusarium graminearum* species complex and chemotypes: A review. Food Addit Contam Part A Chem Anal Control Expo Risk Assess, 32: 453–460.
- Vaishnav MU, Patel HR, Dhruj IU. 1985. Effect of culture filtrates of *Aspergillus* spp. on *Meloidogyne arenaria*. Indian J Nematol, 15: 116-117.
- Wagacha JM, Muthomi JW. 2007. *Fusarium culmorum*: Infection process, mechanisms of mycotoxin production and their role in pathogenesis in wheat. Crop Protect, 26: 877–885
- Zuckerman BM, Mai WF, Harrison MB. 1985. Plant Nematology Laboratory Manual. The University of Massachusetts Agricultural Experiment Station Amherst, Massachusetts 01003, pp: 212.