






The Effects of Drinking Water Supplemented with Essential Oils on Performance, Egg Quality and Egg Yolk Fatty Acid Composition in Laying Hens

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Abstract

This study was performed to examine the effect of different concentrations of an essential oil mixture (EOM) added to drinking water on growth performance, internal and external egg quality, and egg yolk fatty acid composition in laying hens. A total of 240 Brown Nick laying hens aged 20 weeks were used in this study. These hens were divided into one control and three treatment groups whose drinking water was supplemented with 0, 0.1, 0.2, and 0.3 ml/L of Oregofarm EOM. Oregofarm is a commercially available product containing peppermint, oregano, and anise oil. Each treatment group was further categorized into 10 subgroups, each consisting of 6 laying hens. The hens were fed basal rations for 16 weeks. For data collection and analysis, a 16-week period was divided into three time periods of 5 weeks. The weights of 25- to 30- and 35-week eggs were recorded weekly. In the groups whose drinking water was supplemented with the EOM, there was a significant improvement in egg production (EP) ($p < 0.001$), weekly egg weights

during the entire study period ($p < 0.001$), and feed conversion ratio ($p < 0.05$). Furthermore, egg shell thickness and yolk height were significantly increased ($p < 0.05$) at week 35. The Haugh unit was also significantly improved ($p < 0.05$) at weeks 25 and 30. However, feed intake and body weight were not affected by the treatment ($p > 0.05$). The levels of saturated fatty acids were decreased, whereas the levels of polyunsaturated fatty acids increased with EOM supplementation, although the differences were not statistically significant ($p > 0.05$). Therefore, based on the beneficial effects of supplementing drinking water with different concentrations of EOM on egg weight, egg production, feed conversion ratio, and egg yolk fatty acid compositions, it is suggested that EOM can be safely added at concentrations of up to 0.3 ml/L (i.e., the levels used in this study) to drinking water for laying hens.

Keywords: Egg quality, essential oils, laying hens, performance, yolk fatty acids

Introduction

Due to cross and multiple resistance issues, many antibiotic growth promoters in poultry nutrition are restricted by the EU. Consequently, researchers have been exploring the use of phyto-genic feed additives (PFA) such as plants, plant extracts, essential oils (EOs) and individual or combined active EOs ingredients (Bozkurt et al., 2014). Essential oils are a mixture of fragile and volatile compounds - often referred to as aromatic - by plant origin and plant rooting. Hence, the chemical composition and concentra-

tion of active plant compounds vary greatly dependent on their source (Hippenstiel et al., 2011). Mint (*Mentha piperita*) is a member of the Labiatae family and is probably of East Asian origin. Such herbal medicines are widely used and are believed to be particularly useful in combating the immune system and secondary infections (Akbari et al., 2015). As an aromatic plant growing mostly in the Mediterranean region, Oregano (*Origanum vulgare* L.) EO can be used as a phyto-genic additive. The medicinal plant has been used as an anion for digestive, antiparasitic, antibacterial and antifungal stimulant effects (Ertaş et al., 2005).

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In some studies, the successful use of EOs as alternative growth promoters has already been proven and has started to play an important role in poultry nutrition. Phytogetic additives found in a wide variety of plants, spices and derivatives have a beneficial effect on the quality of animal products and animal health and are safe for use in the food industry (Ertaş et al., 2005). It has been reported that thyme EOs have various biological activities *in vitro* and *in vivo*, together with antimicrobial, antioxidant and antifungal effects (Espina et al., 2015; Mooyottu et al., 2014). Essential oils using herbs and spices might increase the acceptability of feed because of their flavorful characteristics and therefore, could advance feed intake by being added to poultry diets (Williams and Losa, 2001). Similarly, some studies showed positive effects on performance traits such as egg production rate, egg weight and egg mass output (Aydın et al., 2008; Bozkurt et al., 2012b), however some experiments showed that varying levels of dietary EOM did not significantly affect the performance, damaged eggs and eggshell weight (Olgun and Yıldız, 2014; Özek et al., 2011). There are limited studies reporting the effects of herbs and EOs on egg quality parameters of laying hens with contradictory results (Bölükbaşı et al., 2008 and 2010).

Due to increased dietary fat and increased coronary heart disease, egg yolk fatty acid composition is very important for consumers (Simopoulos and Salem, 1992). Methods of feeding animals can change the fatty acid composition of the eggs (Yi et al., 2014). Essential oils or mixtures added to diets have a positive effect on lipid metabolism (Acamovic and Brooker, 2005). Contrary to this statement, Ding et al. (2017) reported that there was no significant effect on the fatty acid composition of egg yolk in the experimental groups where Enviva EO additions were made and not made. Up to now, although the effects on the performance parameters of essential oil addition to poultry diets have been studied, there have very few reports on the effects of essential oils on egg yolk fatty acid composition. The study was designed to define the effects of essential oil mixture (EOM) (peppermint, oregano, and anise EOs) added to drinking water on growth performance, egg quality and egg yolk fatty acid compositions in laying hens.

Materials and Methods

Animals and experimental design

This study has been reviewed and approved by an ethical or advisory board of Animal Ethics Committee at Kafkas University (KAU-HADYEK/2016-032) (Kars, Turkey). A total of 240 *Brown-Nick* laying hens aged 20 weeks were used in the study. Drinking water was supplemented with EOs (Oregofarm EO, Farmavet International, Feed and Water Additives Specialist, Manisa, Turkey) at concentrations of 0.1, 0.2, and 0.3 mL/L respectively and were provided for 16 weeks (20 to 35 weeks). The EOM components are presented in Table 1. In the experiment, 4 groups of chickens were divided into 1 control group and 3 experimental groups. Ten sub-groups were created for each trial group and each sub group contained 6 laying chick-

ens. In the study, the hens were fed with laying hen rations containing 16% HP and 2750 kcal / kg ME. The basal ration was prepared using maize and soybean meal. The ingredient and nutrient levels of the basal diet as presented in Table 2 met the NRC recommendations (1994).

The rations were applied to all groups as isocaloric and isonitrogenic mixes. Hens were sheltered in a hen house where light-

Table 1. The active componenets of the essential oil mixture

	Components (mg/kg)
Thymol (thyme oil)	2000
B-phellanderene (thymeoil)	1300
Limonene (thyme oil+pepermint oil)	3525
B-pinene (thyme oil+pepermit oil)	1977
Linalool (oreganum oil)	3645
Carcacrol (oreganum oil)	8910
Anethole (anise oil)	10712
Menthole (pepermint oil)	6375

Table 2. Composition and nutrient level of basal diet

Ingredient composition	Basal Diet, g/kg ⁻¹
Corn	620
Soybean meal	205
Wheat bran	61.8
Oil	10
Limestone	85
Dicalcium phosphate	12.9
Salt	2.5
Vitamin mineral premix ^a	2.5
DL-Methionine	0.3
Calculated nutrient levels	
Metabolisable energy ^b (kcal kg ⁻¹)	2751
Crude protein	160
Dry matter	904
Ether extract	29.3
Crude ash	125
Crude fibre	28.4
Calcium	35.07
Total phosphorus	5.91
Available phosphorus	6.7
Starch	422.7
Sugar	33.4

^aSupplied per kilogram of diet: 15372000 IU vitamin A, 6.28 mg vitamin E, 0.64 mg vitamin K3, 37.36 mg Mn, 89 mg Fe, 25 mg Zn, 8.76 mg Cu, 0.03 mg Co, 0.05 mg MgI, 0.91 mg Se, 2400.000 IU D₃

^bMetabolisable energy content of diets was estimated according to the equation of Carpenter and Clegg.

ing, temperature and ventilation were controlled. The temperature was maintained at approximately 22-24°C. The house had controlled lighting (16L: 8D). The experiment was completed in 16 weeks in the layer chicken unit of the Research and Application Farm at Kafkas University. All hens were supplied with diet and water *ad-libitum*. 15 liter water tanks were prepared for each trial group and connected via nipples. The daily water consumption of the animals was estimated. Fresh drinking water and EOM were added to the water tanks daily. The animals were constantly monitored and kept well hydrated throughout the experiment. The nutrient composition of the basal diet was determined according to the AOAC (1995).

Performance and egg quality parameters

In order to determine the changes in body weight, the body weights of the chickens were recorded at the beginning and end of the study. Egg production and weight and FI were registered weekly during the experiment. Egg production and FCR were calculated for each period. The magnitude of production was adjusted according to mortality rates, which were recorded daily. To determine egg quality parameters, in every 5 week period, 20 eggs were randomly selected per treatment (2 eggs per replicate). These were then evaluated in terms of eggshell weight, albumen and yolk weight, yolk height, eggshell thickness and HU. The eggs were weighed with a special instrument (TP-2000A-0.01 g, İstanbul, Turkey). The eggs were broken on a glass table. The eggshell thickness was measured at three different sections (upper-middle and ends) using a micrometer (Standardgage-200mm-8IN, Asia). The height of the yolk was measured with a tripod micrometer (Mitutoya-20mm, Kawasaki, Japan). Albumen quality was measured in terms of the HU calculated from the weight of the egg and the height of the albumen. While calculating HU values for each egg, the following formula was used:

$$HU = 100 \log (H - 1.7W^{0.37} + 7.6)$$

(H is the observed height of the albumen (mm) and W is the weight of the egg (g)).

Fatty acid profile of egg yolk

During the final week of fatty acids analysis, the oil extraction was carried out on 10 egg samples from each group, according to the method laid out by the AOAC (1995), and then processed with methyl esters with Boron Trifluoride. The fatty acid methyl esters were condensed under nitrogen gas and then analyzed in GC-MS (HP 6890/5972). An Agilent HP88 100X250 micro-Mx250 mm column was used in the analysis. The initial temperature of the column was 120°C, while the final temperature was 230°C. The injector and detector temperature was set at 250°C. The injection speed was set at 50:1 and helium was used as a carrier gas.

Statistical analysis

The statistical analysis and the significance of the mean values between the groups were determined by the analysis of vari-

ance. The multiple range test was used to determine differences between the trial groups. The statistical analyses were performed with Statistical Package for the Social Sciences 16.0 (SPSS Inc.; Chicago, IL, USA) (Dawson and Trap, 2001).

Results and Discussion

Laying hen performance and egg production

Initial and final BW, FI, FCR and egg production data for the layer hens fed drinking water containing EOM is presented in Table 3. Performance analysis showed that there were no effects on body weight and feed intake in the experimental group. During the experimental period, egg production and FCR improved significantly ($p < 0.05$) with the supplementation of 0.2 and 0.3 mL/L EOM. Studies reported beneficial effects on laying performance with EOM supplementation in layer's diet (Bozkurt et al., 2012b; Özek et al., 2011). Radwan et al. (2008) reported that FI and BW were not significantly affected by dietary herbs. No differences in FI were reported in hens when the EO was supplemented to basal diets (Bozkurt et al., 2012a and 2012c; Çabuk et al., 2014). In contrast, Abdo et al. (2010) found that adding green tea EO in the ratio significantly reduced feed intake. Similarly, Bölükbaşı et al. (2008) found that EO supplementation in layer diets lowered FI and FCR was improved when the birds were fed EOM in their diet. Certain feeding experiments with layer hens didn't find any differences in FCR when the oregano EO was added to diets (Florou-Paneri et al., 2005). Similarly, Bozkurt et al. (2012a; 2012c) revealed that dietary inclusion of EO did not improve FCR in white layer hens. However, some beneficial effects on FCR using EO of inclusion level 24 mg/kg feed have been reported for the brown layer strain under hot environmental conditions (Çabuk et al., 2006). Some studies confirmed the positive effects of herbs and their respective EO on the BW of hens. Çabuk et al. (2006) reported that the BW of layer hens fed diets of an EO mixture increased compared to that of the control group over a 20-week period. Similarly, in another study, significant improvements were determined in the BW of hens when they were fed different levels of garlic (Khan et al., 2007). However, no effect was reported on BWG in response to dietary supplementation with tea leaves or green tea EO (Abdo et al., 2010), or dietary oregano EO (Florou-Paneri et al., 2005). Bozkurt et al. (2009) observed that adding an EO mixture to diet had no effect on egg production and weight. Similarly, Bölükbaşı et al. (2010) observed no effect on egg production but there was an increase in egg weight when diets were supplemented with thyme, sage or rosemary over a period of 12 weeks. However, Xianjiing He et al. (2017) found that added oregano EOs in layer diets significantly influenced egg production and average egg weight.

Egg quality parameters

The effects of Oregofarm EO on egg weight and internal and external egg quality are listed in Table 4 and 5. Supplementation of Oregofarm EO significantly increased the egg weight ($p < 0.05$) weekly. In all periods, the egg weight was effected significantly

Table 3. The effects of adding EOM on the performance (mean±standard error) to layer hens' drinking water

	Essential oil mixed supplementation (ml/L)				p
	0	0.1	0.2	0.3	
Initial body weight, g	1734±17.63	1734±14.44	1771±16.55	1782±15.1	0.62
Final body weight, g	1736±62.91	1661±22.52	1742±21.99	1771±20.44	0.14
Feed intake, g day ⁻¹ per bird	102.24±1.52	99.94±1.26	103.83±1.61	104.13±1.59	0.191
Egg production, %	90.88±0.44 ^b	89.74±0.48 ^b	92.41±0.41 ^a	92.90±0.40 ^a	<0.001
Feed conversion ratio, kg feed per kg egg	2.01±0.04 ^a	1.97±0.03 ^{ab}	1.91±0.03 ^b	1.90±0.02 ^b	0.038

^{a,b}indicated the difference within a row was significant (p<0.05)

Table 4. The effects on the weekly egg weights of adding EOMs to layer hens' drinking water (g)

Weeks	Essential oil mixed supplementation (mL/L)				p
	0	0.1	0.2	0.3	
1-wk	55.00±0.59 (n:55)	56.1±0.55 (n:53)	56.36±0.64 (n:58)	56.39±0.71 (n:59)	0.36
2-wk	55.66±0.55 ^b (n:56)	55.81±0.45 ^b (n:56)	57.48±0.55 ^a (n:52)	57.81±0.63 ^a (n:55)	0.007
3-wk	56.8±0.64 (n:52)	57.86±0.49 (n:54)	58.47±0.64 (n:57)	58.38±0.46 (n:55)	0.15
4-wk	56.7±0.05 (n:59)	57.07±0.45 (n:57)	58.12±0.57 (n:55)	58.25±0.59 (n:57)	0.103
5-wk	57.16±0.59 ^b (n:58)	56.89±0.43 ^b (n:56)	58.98±0.60 ^a (n:59)	58.40±0.52 ^{ab} (n:57)	0.019
6-wk	57.17±0.52 ^b (n:58)	57.51±0.50 ^b (n:59)	58.17±0.61 ^{ab} (n:57)	59.18±0.52 ^a (n:57)	0.047
7-wk	56.90±0.59 ^c (n:48)	57.56±0.47 ^{bc} (n:52)	59.62±0.65 ^a (n:51)	58.98±0.52 ^{ab} (n:54)	0.002
8-wk	56.68±0.66 ^b (n:55)	56.93±0.56 ^b (n:58)	59.60±0.66 ^a (n:58)	59.33±0.58 ^a (n:56)	0.001
9-wk	56.38±0.59 ^b (n:48)	56.35±0.57 ^b (n:49)	57.65±0.58 ^{ab} (n:50)	58.53±0.58 ^a (n:53)	0.019
10-wk	57.44±0.70 ^b (n:51)	56.81±0.64 ^{ab} (n:46)	58.74±0.57 ^a (n:55)	59.13±0.61 ^a (n:55)	0.037
11-wk	57.50±0.52 ^b (n:47)	58.92±0.61 ^{ab} (n:49)	59.77±0.63 ^a (n:51)	60.30±0.54 ^a (n:57)	0.005
12-wk	57.75±0.67 ^c (n:44)	58.68±0.57 ^{bc} (n:51)	60.98±0.64 ^a (n:53)	60.39±0.59 ^{ab} (n:53)	0.001
13-wk	57.61±0.72 ^b (n:47)	57.06±0.57 ^b (n:47)	60.25±0.52 ^a (n:55)	60.18±0.54 ^a (n:56)	0.000
14-wk	57.87±0.73 ^b (n:57)	58.77±0.69 ^{ab} (n:50)	60.42±0.59 ^a (n:55)	59.80±0.50 ^a (n:56)	0.024
15-wk	58.24±0.75 ^b (n:51)	57.58±0.54 ^b (n:54)	59.27±0.57 ^{ab} (n:56)	60.57±0.60 ^a (n:56)	0.003
16-wk	57.55±0.69 ^b (n:54)	57.81±0.63 ^b (n:55)	60.37±0.69 ^a (n:56)	60.27±0.59 ^a (n:56)	0.001
All Periods	56.67±0.16 ^b (n:840)	57.34±0.14 ^b (n:846)	59.17±0.16 ^a (n:878)	59.11±0.15 ^a (n:892)	<0.001

^{a,b}indicated the difference within a row was significant (p<0.05)

by essential oil mixes (p<0.05). Bozkurt et al. (2012b) declared that EOM added to layer hen diet significantly enhanced egg production rate and egg weight. Other studies showed no significant change in egg production and egg weight when hens were fed a diet supplemented with EOM (Bozkurt et al., 2012b; Florou-Paneri et al., 2005; Özek et al., 2011). Similarly, Bozkurt et al. (2009) discovered that the addition of EOM at 24 to 48 mg/kg to diet had no effect on the production and weight of eggs for broiler breeders in chickens between the ages of 26 and 46 weeks. This variability in results may be related to the dose of EOs, different application methods, components of EOs or their utilization in different types of poultry.

Eggshell weight increased at week 25 week (p<0.05). At the 30th and 35th weeks of the trial, eggshell weight gradually in-

creased with EOM supplementation, but the difference was not significant (p>0.05). Supplements in drinking water had no effect on albumen and yolk weight (p>0.05) in early period hens at 25 weeks. Eggshell thickness was significantly improved at all periods measured (p<0.05). Adding EOM (0.1, 0.2 or 0.3 mL/L) to drinking water influenced HU at the age of 25 and 30 weeks (p<0.05) The highest HU was obtained in the eggs of layer hens fed drinking water containing 0.3 mL/L EOM. In some studies, it has been observed that the use of different forms of EOs or EOM leads to significant improvements in egg shell weight (Bozkurt et al., 2012b and 2012c). In contrast, Akbari et al. (2015) reported that feeding peppermint and thyme EO had no effect on shell weight in layer hens. In our study, the supplementation of EO significantly increased the eggshell thickness in all weeks (p<0.05). Sim-

Table 5. The effects on the internal and external egg quality (mean± standard error)¹ of adding EOMs to layer hens' drinking water

Item	Essential oil mixed supplementation (mL/L)				p
	0	0.1	0.2	0.3	
Unit of egg weight					
25 wk	57.18±1.17 ^b	58.13±0.91 ^{ab}	58.32±0.99 ^{ab}	60.32±0.83 ^a	0.15
30 wk	57.46±0.74	57.59±0.68	57.85±0.75	58.08±1.03	0.94
35 wk	59.66±0.95	59.39±0.90	61.64±0.65	61.02±0.79	0.10
Eggshell weight (g)					
25 wk	5.95±0.12 ^b	5.97±0.12 ^b	6.03±0.096 ^b	6.37±0.13 ^a	0.04
30 wk	6.03±0.12	6.04±0.12	6.35±0.09	6.21±0.11	0.12
35 wk	5.89±0.09	6.09±0.08	6.10±0.12	5.95±0.12	0.36
Albumen weight (g)					
25 wk	37.09±0.97	38.48±0.79	37.97±0.69	39.49±0.62	0.19
30 wk	37.91±0.77	36.89±0.84	36.08±0.84	36.14±0.97	0.82
35 wk	38.68±0.93 ^a	38.43±0.89 ^a	40.36±0.78 ^a	40.36±0.75 ^a	0.05
Yolk weight (g)					
25 wk	14.14±0.29	13.68±0.17	14.31±0.31	14.46±0.26	0.19
30 wk	14.48±0.27 ^b	14.66±0.25 ^b	15.42±0.22 ^a	15.72±0.30 ^a	0.003
35 wk	15.09±0.22 ^a	14.86±0.23 ^b	15.18±0.19 ^a	14.71±0.19 ^b	0.05
Yolk height (mm)					
25 wk	28.94±0.70 ^a	28.96±1.16 ^a	26.12±0.35 ^b	26.20±0.41 ^b	0.004
30 wk	26.96±0.27	26.65±0.26	27.08±0.24	26.74±0.29	0.65
35 wk	27.87±0.33 ^b	29.03±0.27 ^a	29.11±0.28 ^a	28.57±0.25 ^{ab}	0.01
Eggshell thickness (mm)					
25 wk	0.32±0.04 ^a	0.28±0.05 ^b	0.29±0.04 ^b	0.31±0.05 ^a	0.035
30 wk	0.33±0.05 ^a	0.33±0.04 ^a	0.31±0.05 ^b	0.32±0.05 ^{ab}	0.001
35 wk	0.31±0.03 ^b	0.33±0.03 ^a	0.33±0.08 ^a	0.34±0.06 ^a	0.008
Haugh unit					
25 wk	78.35±1.22 ^{ab}	75.99±0.89 ^b	78.72±1.04 ^{ab}	80.95±1.25 ^a	0.024
30 wk	79.26±0.28 ^b	79.15±0.37 ^b	79.86±0.31 ^{ab}	80.62±0.52 ^a	0.03
35 wk	79.69±0.33	80.31±0.32	80.51±0.58	80.80±0.64	0.43

^{a,b}indicated the difference within a row was significant (p<0.05)

¹Means of 20 eggs per treatment

ilarly, Bozkurt et al. (2012b) reported that supplementation of EOM increased egg shell weight, egg shell thickness and shell breaking strength.

Adding oregano EO (50 or 100 mg kg⁻¹) to the diet had no effect on HU at the age of 32 weeks (Florou-Paneri et al., 2005). Similarly, Çabuk et al. (2006) observed that adding EOMs or antibiotics to the diet did not significantly affect the egg quality characteristics HU, shell weight or yolk weight in laying quails. Ding et al. (2017) reported that Enviva EO supplementation in layer hens diet had no significant effect on albumen height and HU. However, Xianjing et al. (2017) observed that the use of oregano EO supplementation had no effect on eggshell ratio but found that it did significantly affect yolk ratio, egg shape index and HU.

Fatty acid compositions

The mean value of the yolk fatty acids percentage in different treatment groups is shown in Table 6. When compared to the control diet, the proportion of DHA (C22:6 n-3) in the egg yolk was significantly decreased in the treatment groups (p<0.05). Palmitoleik acid (16:1) increased in drinking water containing EOM when compared to the control group (p<0.05). There was no significant effect on total saturated fatty acids and omega 3 and 6 fatty acid in the experimental groups. To our knowledge, there have been few reports about the effects of EO on the fatty acid composition of egg yolk. In the present study, there were significant differences in the treatment groups in a linolenic acid, linolenic acid, palmitoleic acid, arachidonic acid, lignoceric acid and DHA (p<0.05). However, Ding et al. (2017) found that EO in

Table 6. The effects on egg yolk fatty acid composition (mean± standard error) of adding EOMs to layer hens' drinking water

Fatty acid (%)	Essential oil mixed supplementation (mL/L)				p
	0	0.1	0.2	0.3	
C14:0	0.28±0.016 ^b	0.29±0.015 ^{ab}	0.34±0.015 ^a	0.34±0.024 ^a	0.022
C14:1	0.005±0.003 ^b	0.008±0.004 ^b	0.016±0.005 ^b	0.033±0.009 ^a	0.004
C15:0	0.009±0.005 ^b	0.020±0.006 ^b	0.02±0.006 ^b	0.036±0.002 ^a	0.009
C16:0	24.40±0.20 ^b	23.52±0.42 ^b	24.72±0.21 ^a	24.90±0.34 ^a	0.014
C16:1	2.67±0.09 ^c	2.86±0.16 ^b	3.23±0.14 ^{ab}	3.48±0.21 ^a	0.033
C17:0	0.21±0.011 ^b	0.21±0.013 ^b	0.21±0.012 ^b	0.28±0.019 ^a	0.005
C17:1	0.21±0.05	0.18±0.03	0.29±0.04	0.28±0.053	0.272
C18:0	9.57±0.20 ^a	9.42±0.35 ^a	8.50±0.38 ^b	7.82±0.30 ^b	0.001
C18:1	41.15±0.71	41.35±0.49	41.32±0.38	41.16±0.70	0.991
C18:2 n6	16.96±0.78	17.64±0.41	17.42±0.44	17.93±0.62	0.696
C18:3 n6	0.50±0.035	0.49±0.052	0.48±0.030	0.48±0.031	0.973
C18:3 n3	0.74±0.058 ^b	0.94±0.059 ^a	0.90±0.041 ^a	0.96±0.044 ^a	0.021
C20:0	0.17±0.029	0.16±0.043	0.13±0.025	0.10 ± 0.017	0.393
C20:1	0.25±0.029	0.30±0.034	0.24±0.018	0.27±0.017	0.420
C20:2 n6	0.22±0.029	0.26±0.040	0.18±0.022	0.19±0.015	0.209
C20:3 n3	0.20±0.033	0.18±0.029	0.19±0.018	0.17±0.018	0.869
C20:4 n6	1.68±0.058 ^a	1.45±0.11 ^{ab}	1.23±0.14 ^{bc}	1.07±0.13 ^c	0.004
C22:6 n3	0.61±0.03 ^a	0.51±0.05 ^{ab}	0.44±0.06 ^b	0.38±0.05 ^b	0.014
C24:0	0.16±0.018 ^{ab}	0.21±0.03 ^a	0.11±0.014 ^b	0.12±0.014 ^b	0.015
ΣSFA	34.81±0.33	33.84±0.32	34.06±0.43	33.60±0.51	0.187
ΣMUFA	44.27±0.75	44.69±0.48	45.10±0.46	45.22±0.78	0.710
ΣPUFA	20.91±0.86	21.47±0.51	20.84±0.38	21.18±0.73	0.897
ΣUFA	65.19±0.33	66.16±0.32	65.94±0.43	66.40±0.51	0.187
ΣPUFA/ΣSFA	0.60±0.028	0.64±0.018	0.61±0.016	0.63±0.026	0.695
ΣUFA/ΣSFA	1.88±0.028	1.96±0.027	1.98±0.048	1.94±0.018	0.198
n6	19.37±0.81	19.84±0.46	19.31±0.36	19.68±0.67	0.911
n3	1.54±0.062	1.63±0.083	1.53±0.049	1.50±0.088	0.608
n6/n3	12.60±0.34	12.36±0.49	12.72±0.48	13.29±0.51	0.549
NUTRVALUE	2.08±0.036 ^b	2.17±0.055 ^a	2.02±0.028 ^b	1.97±0.043 ^b	0.014
ATHERNGINX	0.17±0.003 ^a	0.16±0.005 ^a	0.15±0.006 ^{ab}	0.14±0.006 ^b	0.008
THRINDEX	0.60±0.008	0.58±0.007	0.58±0.010	0.57±0.012	0.191

n=10

^{ab} indicated the difference within a row was significant (p<0.05)

ΣSFA= total saturated fatty acids; ΣMUFA= total monounsaturated fatty acids; ΣPUFA= total polyunsaturated fatty acids; n-6: omega-6 fatty acids; n-3: omega-3 fatty acids

layer hen diet had no effect on the fatty acid composition of egg yolk of layer hens. The addition of EOM to the drinking water of layer hens had no effect on the n-6/n-3 ratio in the egg (p>0.05). Bölükbaşı et al. (2010) reported that bergamot oil supplementation in layer diets significantly increased the proportion of DHA and n-3 in the egg yolk. While the addition of essential oil to drinking water reduced the SFA concentration, it increased the MUFA concentration, numerically. In agreement with the present study, Ding et al. (2017) reported that SFA concentrations were decreased but PUFA and MUFA concentrations were increased

with dietary EO. Bölükbaşı et al. (2008) showed that SFA and PUFA concentrations in leg and breast tissues in broilers were decreased, whereas, MUFA concentrations were increased. To our knowledge, the studies on the addition of essential oils to drinking water are limited. Therefore, further work is needed in order to give more detailed information on this topic in layer hens.

Conclusion

As a result of supplementing peppermint, oregano, and anise essential oils in the amounts of 0.2 mL/L and 0.3 mL/L to drink-

ing water, there were beneficial effects on performance and egg quality without adverse effects on other parameters.

Ethics Committee Approval: All experimental protocols adhered to and approved by the guidelines of Animal Ethics Committee of Kafkas University (KAU-HADYEK/2016-032) (Kars, Turkey).

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Conflict of Interest: The authors have no conflicts of interest to declare.

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