



Comment on : 'Distribution, phenotypic characterisation and antibiogram of bacterial species from hospital environment in Nigeria: **Public health implications'**

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Sir.

I read with interest the article by Onuoha et al. entitled 'Distribution, phenotypic characterization and antibiogram of bacterial species from hospital environment in Nigeria: Public health implications'. The authors performed antimicrobial susceptibility testing for the bacteria that were isolated from various surfaces of a federal teaching hospital. It is clear that the authors made a great effort for the work. However, I have some concerns about the method that used for the determination of vancomycin susceptibility of Staphylococcus aureus isolates and some of the antimicrobial agents that were chosen for the antimicrobial susceptibility testing for the isolates of Salmonella spp. and Shigella spp.

The authors used disk diffusion method for the detection of vancomycin resistance among S. aureus isolates. According to current guidelines, the disk diffusion test with vancomycin is unreliable and should not be used for the detection of vancomycin resistance in S. aureus. Clinical and Laboratory standards Institute (CLSI) and European Committee on Antimicrobial Susceptibility Testing (EUCAST) recommend vancomycin MIC testing to determine the susceptibility of staphylococci to vancomycin (1, 2). I must indicate that the percentage of S. aureus isolates that were found to be resistant to vancomycin is extremely high (87.5%) in the study of Onuoha et al. Isolates of vancomycin-resistant S. aureus (VRSA) are rarely described worldwide (3, 4). Vancomycin MIC testing should be performed for all S. aureus strains in this study to reliably determine the vancomycin susceptibility profile of the isolates.

The authors also performed antimicrobial susceptibility testing for Salmonella spp. and Shigella spp. isolates. According to CLSI guidelines, first and second generation cephalosporins, cephamycins and aminoglycosides may appear active in vitro, but these agents are clinically ineffective and shouldn't be reported as susceptible (1).

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Journal of Immunology and Clinical Microbiology © 2019

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The authors included cephalothin (a first-generation cephalosporin), cefuroxime (a secondgeneration cephalosporin), streptomycin and gentamicin (aminoglycosides) among the antimicrobial agents tested against the isolates of *Salmonella* spp. and *Shigella* spp. They reported various susceptibility rates for cefuroxime, gentamicin and streptomycin for *Salmonella* spp. isolates. Also, 95.2% and 4.8% of *Shigella* spp. isolates were reported to be susceptible to gentamicin and streptomycin, respectively. It would be better not to use first and second generation cephalosporins, cephamycins and aminoglycosides for antimicrobial susceptibility testing of *Salmonella* spp. and *Shigella* spp. in order to prevent misunderstandings.

Ethics Committee Approval: NA Informed Consent: NA Peer-review: Externally peer-reviewed. Conflict of Interest: No conflict of interest was declared by the author. Financial Disclosure: The author declared that this study has received no financial support.

References

1. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Susceptibility Testing: Twenty-eighth Informational Supplement M100-S28, Wayne, PA (USA), 2018.

2. EUCAST. Breakpoint Tables for Interpretation of MICs and Zone Diameters, Version 9.0 http://www.eucast.org/clinical_breakpoints/ (March 2019, date last accessed).

3. Mirza HC. Glycopeptide resistance in S. aureus. In: Enany S, Alexander LE, eds. The rise of virulence and antibiotic resistance in Staphylococcus aureus. Croatia: InTech; 2017. p. 43-59.

4. Gardete S, Tomasz A. Mechanisms of vancomycin resistance in Staphylococcus aureus. J Clin Invest. 2014; 124(7): 2836-40.



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