

INTERNATIONAL ENGINEERING, SCIENCE AND EDUCATION GROUP	Middle East Journal of Science (2019) 5(1):86-93 Published online June, 2019( <u>http://dergipark.org.tr/mejs</u> ) doi: 10.23884/mejs.2019.5.1.09 e-ISSN 2618-6136 Received: April 18, 2019 Accepted: June 19, 2019 Submission Type: Research Article
--	--

# THE EFFECTS OF PRETREATMENT FACTORS ON SEED GERMINATION AND SEEDLING GROWTH OF ANISE (*PIMPINELLA ANISUM L.*)

 Mohamad Hesam Shahrajabian<sup>1,2,3\*</sup>, Mehdi Khoshkharam<sup>3</sup>, Wenli Sun<sup>1,2</sup>, Qi Cheng<sup>1,2</sup>

 ORCID:0000-0002-8638-1312
 ORCID:0000-0002-9301-801X
 ORCID:0000-0002-1705-2996
 ORCID:0000-0003-1269-6386

<sup>1</sup>Biotechnology Research Institute, Chinese Academy of Agricultural Sciences, Beijing 100081, China <sup>2</sup>Nitrogen Fixation Laboratory, Qi Institute, Building C4, No.555 Chuangye, Jiaxing 314000,

Zhejiang, China

<sup>3</sup>Department of Agronomy and Plant Breeding, Faculty of Agriculture, Islamic Azad University, Isfahan (Khorasgan) Branch, Isfahan, Iran

 $* Corresponding \ author: hesamshahrajabian@gmail.com$ 

Abstract:Seed dormancy is one of the major problems in agricultural studies, especially for medicinal plants. Anise (Pimpinella anisum L.)is an important economic medicinal plant with dormant seeds and distributed only in its natural habitats. An experiment was conducted as a Factorial layout within a completely randomized design with four replications to evaluate the effects of some pretreatment factors on primary growth and germination characteristics of anise. Pre-chilling treatments were 0, 15, 30 and 45 days treatments and hormone treatments were  $GA_3$  (Gibberellic BA (benzyladenine), kinetin (Kinetinnetin),  $GA_3+BA$ ,  $GA_3+kinetin$  BA+kinetin, Acid), GA<sub>3</sub>+BA+kinetin, KNO<sub>3</sub>, H<sub>2</sub>SO<sub>4</sub> and distilled water as a control treatment.Prechilling treatment effects on coleoptile and radicle length, seedling length, germination percentage, mean time for germination, germination rate and seed vigor index showed significant differences (p<0.01) among them. Similarly, different hormone treatments also had significantly different influence on coleoptile and radicle length, seedling length, germination percentage, mean time germination, germination rate and seed vigor index. The highest germination percentage and germination rate was related to the usage of BA+ kinetin. The highest values for radicle length and uniformity of seed germination were achieved in BA and kinetin, respectively. Moreover, application of  $GA_3+BA+kinetin$  had given the highest seed vigor index. It seems that application of exogenous  $GA_3+KINETIN$  and BA+kinetinconcentration, which is provided mostly by chilling treatment, is the most effective factor for breaking the seed dormancy. On the basis of the results, usage of 45 days moist prechilling accompanied with application of  $GA_3$ +kinetin and BA+kinetin in Esfahan cultivar was appropriate.

Keywords: Seed dormancy, Seed germination, Seedling growth, Anise



### 1.Introduction

Anise (Pimpinella anisum L.) is a flowering plant in the family Apiaceae native to the eastern Mediterranean region, west and Southwest Asia [1-2-3]. Anise is also famous in traditional Chinese medicine (TCM), Ayurveda and Unani medicine. Anise has been used for different purposes in the traditional medicine system of Iran [4]. Seed priming treatments have been employed to accelerate germination, seedling growth and yield in most seeds under normal and stress conditions [5-6-7-8-9-10-11]. Seed germination can be controlled by many factors like natural germination and growth inhibitors [12-13-14-15]. These are the derivatives of gibberellic acid (GA<sub>3</sub>), abscisic acid (ABA), cinnamic acid, kinetinnetin (KİNETİN), bonzyladenin (BA), coumarin, jasmonic and etc. The variation in seed dormancy and seedling emergence are controlled by environmental conditions. The origin of research into Gibberellins can be traced to Japanese plant pathologists who were investigating the causes of the bakane (foolish seedling) disease that seriously lowered to the yield of rice crop in Japan, Taiwan, and some other Asian countries [16]. Gibberellic acid is a plant growth hormone that has an important role in seed germination [16]. It has been reported that the stimulating effects of GA3 on seed germination are not similar in all crop species [17]. GA3 has been also reported to promote growth in cotton, rice and in some halophytes under saline conditions [18-19]. Tsygankova et al. [20] confirmed specific auxin-like, cytokinetinnin-like and minor gibberellins-like effect of synthetic heterocyclic compounds on cell division, cell proliferation, cell elongation and cell differentiation that are the basic processes of plant growth and development. Gibberellins is known to eliminate the chilling requirements of peach and apple seed and increased their germination [21]. Primed with gibberelin improve quality of seeds and germination [22]. Liopa-Tsakalidi et al. [23] also suggest that germination and seedling growth of 11 species responded differently to different levels of GA<sub>3</sub>. Fernandez et al. [24] revealed that cold stratification has a direct influence on the production of gibberellins (Gas) in seeds of Arabidopsis thaliana. Exogenously applied GA overcomes seed dormancy in several species [25]. Hormone priming increased antioxidant enzyme activity and decrease the amount of reactive oxygen space. Sharifi and Pouresmael [26] concluded that only cold treatments such as gibberellic acid, cytokinetinnin, potassium nitrate, washing and light treatments are not useful. It has been reported that GA is effective in breaking seed dormancy in snowberry [27]. Nkomo and Kambizi [28] noted that prechilling followed by exposure to a temperature higher than 30°C encourages the germination of C. Olitorious seeds. Rouhi et al. [29] concluded that applying 500 ppm concentration of GA<sub>3</sub> and KNO<sub>3</sub> resulted in higher germination in waterlily dormant seeds. Plant hormones are used in breaking seed dormancy [30]. Cytokinins and auxin are the most common plant growth regulators used in in vitro culture of plant tissues [31-32]. Cytokinins constitute a major class of plant growth regulator that is involved in a wide range of physiological processes [33]. Cytokinins have a stimulatory or an inhibitory role in different development processes, such as control of apical dominance in the shoot, root growth and branching, leaf senescence, and chloroplast development [34]. In spite of the fact that Anise is an important and expensive medicinal and spice plant, not enough information is available on the effects of moist pre-chilling and application of hormones on different cultivars of it. So, the aim of this study is to survey the certain effects of different treatments to stimulate seed germination and seedling growth of Anise.



### 2.Materials and methods

The study evaluates the influence of some pretreatments on growth and germination characteristics of anise (*Pimpinella anisum L.*), an experiment was conducted as Factorial layout within completely randomized design with four replications at Research laboratory of Mojgan Agricultural Company, Mahmood Abad, Isfahan, Iran. Pre-chilling treatments were 0, 15, 30 and 45 days treatments and hormone treatments were GA<sub>3</sub> (Gibberellic Acid), BA (benzyladenine), kinetin, GA<sub>3</sub>+BA, GA<sub>3</sub>+ kinetin, BA+ kinetin, GA<sub>3</sub>+BA+ kinetin, KNO<sub>3</sub>, H<sub>2</sub>SO<sub>4</sub> and distilled water as a control treatment. First, seeds were surface sterilized in 1.5% (w/v) sodium hypochlorite solution for 15 minutes and then rinsed three times with sterile distilled water. For each treatment, 4 Petri dishes were used and 30 seeds were put into each of them, then, each Petri dish was covered with 10 mm of each specific treatment. In the first trial, seeds were chilled for 15, 30 and 45 days, and after that, seeds were soaked and treated with 10 hormone treatments. In the second experiment, the seeds were treated without pre-chilling treatments. In the thirds experiment, seeds treatments were done with polyethylene glycol. Equation number one and number two were used to calculate germination percentage and germination rate, respectively.

Germination percentage= (Number of germinated seed /total number of seed)  $\times$  100 (1)

1) 
$$GR = \Sigma \frac{N}{\Sigma(n \times g)}$$
 (2)

Where N is the number of germinated seed on growth day and g is the number of germination seeds Analysis of variance (ANOVA) was used to determine the significant differences. Uniformity of seed germination and mean time for seed germination (MTG) was evaluated by equation number 3 and 4. Seed vigor index was calculated by equation number 5.

Uniformity of seed germination=
$$\frac{1}{\frac{\sum (D - \overline{D})^2 \times N}{\sum N}}$$
(3)  
MTG =  $\frac{\Sigma(nd)}{\Sigma n}$  (4)

n: The number of germinated seed in the specific day.

d: The number of days from the beginning of germination.

 $\sum n$ : The total number of germinated seed.

Seed vigor index: Germination percentage  $\times$  mean of seedling length (mm)(both coleoptile and radicle) / 100 (5)

The means were separated by Duncan's Multiple Range Test  $(p \le 0.05)$ . All statistical analysis was performed with the SAS computer statistical software.



#### 3. Results and discussion

Prechilling had a significant impact on coleoptile length, radicle length, seedling length, germination percentage, mean time for germination, germination rate and seed vigor index was evaluated. The highest coleoptile length was related to 45 days chilling (2.45 mm) which had significant differences with other treatments. Although, the higher value of radical length was obtained for 45 days (0.8485 mm) chilling, its difference with 30 days chilling was not significant. The minimum coleoptiles (0.7536 mm) and radical length (0.6819 mm) was related to control treatment. The higher values of seedling length (2.98 mm), and germination percentage (70.02%) was obtained for 45 days of chilling followed by chilling for 30 days, 15 days and 0 days (control treatment). There were significant differences in seedling length and germination percentage between 45 days of chilling and other treatments. Control treatment had obtained the highest mean time for germination (11.18) which had significant differences with 30 days and 45 days pre-chilling, although, its difference with control treatment was not significant. The maximum value for uniformity of germination rate (5.87%), and seed vigor index (2.244) was achieved in 45 days pre-chilling which had significant differences with other treatments. Both the germination rate and seed vigor index was increased significantly from control treatment to 45 days pre-chilling. The maximum and the minimum uniformity of seed germination was related to control treatment (0.2165), and 15 days pre-chilling (0.1075), which had no significant differences with each other (Table 1). Gupta et al. [30] reported that prechilling treatment also improved seed germination in Isabgol.

The highest coleoptiles length (1.663 mm), and seedling length (2.574 mm) was related to the application of GA<sub>3</sub>+ kinetin and the minimum one was observed in KNO<sub>3</sub>. Patel and Mankad [16] concluded that low concentrations of GA<sub>3</sub> influence all developmental and physiological processes in plants. The maximum and the minimum radical length was related to the application of BA (1.7232 mm), and  $KNO_3$  (0.5286 mm), respectively, which had significant differences with each other. Benzyladenine (BA) at a high concentration was also shown to be effective in shoot regeneration in P. vulgaris [35]. Application of BA + kinetin had obtained the highest value for germination percentage (66.33%) and germination rate (4.126%), which had significant differences with other treatments. Sawan et al. [36] demonstrated that kinetinnetin application improved seed viability and seedling vigor as shown by lengths of the hypocotyls, radical and the entire seedling, as well as seedling fresh weight. Gibberellic acid is also known to play an essential role in seed germination, stem elongation and flower development [37]. The maximum and the minimum mean time for germination was achieved in usage of BA + kinetin (66.33), and KNO<sub>3</sub> (25.45), respectively. Narra et al. [38] also found that the seedling under the GA<sub>3</sub> influence showed enhanced germination, seedling elongation and dry weight accumulation on Trachyspermum ammi. Although the higher value for uniformity of seed germination was related to kinetin (0.1523), followed by other treatments, there were no significant differences between treatments. application of  $ga_3+ba+$  kinetin had obtained the highest seed vigor index (1.545), and the minimum one was related to the application of distilled water (0.4076) (Table 1). The efficacy of BA in inducing multiple shoots was also demonstrated in chickpea [39], mungbean [40] and pigeonpea [41]. Gupta et al. [30] concluded that GA has shown promising effect in breaking seed dormancy with accelerated seed germination (speed of germination, vigor index) and seedling growth (seedling dry weight).



**Table 1.** Mean comparison for coleoptile length (mm), radicle length (mm), seedling length (mm), germination percentage (%), meantime for germination, germination rate (%), uniformity of seed germination and seed vigor index.

germination and seed vigor index.									
Treatment	Coleoptile	Radicle	Seedling	Germination	Meantime	Germination	Uniformity	seed	
	length	length	length	percentage	for	rate	of seed	vigor	
	-	-			germination		germination	index	
Prechilling (day)	_								
0	0.7536d	0.6819bc	1.463c	14.22d	11.18a	0.3626d	0.2165a	0.3287d	
15	1.000c	0.5625c	1.554c	35.28c	11.00a	1.489c	0.1075a	0.6328c	
30	1.340b	0.7635ab	2.133b	53.74b	7.81b	3.356b	0.1336a	1.239b	
45	2.145a	0.8485a	2.986a	70.02a	5.52c	5.87a	0.1352a	2.244a	
Hormone									
GA3	1.352bc	0.8112abc	2.185b	37.91d	10.44a	2.533d	0.1471b	1.056ab	
BA	1.428abc	1.7232bcd	2.143bc	61.22b	9.862a	3.837b	0.1066b	1.365ab	
KİNETİN	1.536ab	0.8687ab	2.385ab	47.33c	9.575a	3.199b	0.1523b	1.351ab	
GA3+BA	1.345bc	0.6835cd	2.036bc	52.26b	10.61a	3.007bc	0.1474b	1.263bc	
GA3+KİNETİN	1.663a	0.9123a	2.574a	40.23cd	10.66a	2.673cd	0.1132b	1.484ab	
BA+KİNETİN	1.425abc	0.7348be	2.176bc	66.33a	9.74a	4.126a	0.1035b	1.464ab	
GA3+BA+KİNETİN	1.563abc	0.8140abc	2.356ab	54.00b	10.28a	3.179b	0.1202b	1.545a	
KNO3	0.7537d	0.5286e	1.343d	25.45e	6.40b	1.855e	0.1148b	0.4661de	
H <sub>2</sub> SO <sub>4</sub>	1.256c	0.5573de	1.915c	25.69e	6.26b	1.632e	0.1242b	0.6842d	
Distilled water	0.8223d	0.5647de	1.465d	25.68e	5.48b	1.987e	0.1247b	0.4076e	

Common letters within each column do not differ significantly.

GA<sub>3</sub>= Gibberellic Acid

KİNETİN= Kinetinnetin

BA= Benzyladenine

## 4. Conclusion

Seed germination is a complex physiological process that responds to environmental signals such as light, water, and other factors. Also, Seed germination is very important to know the germination pattern of a plant, especially the medicinal plants. Prechilling treatment effects on coleoptile and radicle length, seedling length, germination percentage, mean time for germination, germination rate and seed vigor index were significant. Different hormone treatments had a significant influence on coleoptile and radicle length, seedling length, germination percentage, mean time germination, germination rate, and seed vigor index. Prechilling treatment for 45 days had obtained the highest coleoptile and radicle length, seedling length, germination percentage, germination rate, and seed vigor index. While control treatment had obtained the maximum mean time for germination and uniformity of seed germination. Application of GA3+kinetin had obtained the highest coleoptile length, seedling length, and mean time for germination. The highest germination percentage and germination rate was related to the usage of BA+kinetin. The higher values for radicle length and uniformity of seed germination were achieved in the application of BA and kinetin, respectively. Moreover, application of  $GA_3+BA+kinetin$  had resulted in the highest seed vigor index. All in all, in conclusion, it was shown that GA<sub>3</sub>, kinetin, and BA had greatly enhanced the germination parameters in terms of germination percentage, seedling elongation and other characteristics.

## References

[1] Kadan, S., Rayan, M., Rayan, A., "Anticancer activity of Anise (*Pimpinella anisum L.*) seed extract", *The Open Nutraceuticals Journal*, 6, 1-5, 2013.



[2] Saxena, S.N., Verma, M., Kakani, R.K., Rathore, S.S., Saxena, R., Sharma, L.K., "Analysis of medicinally important compounds and antioxidant properties of Anise (*Pimpinella anisum*) seed extract and shoot callus", *International J. Seed Spices*, 4(1), 55-62, 2014.

[3] Ibrahim Doa<sup>•</sup> an Anwar., "Medicinal benefits of Anise seeds (*Pimpinella Anisum*) and *Thymus Vulgaris* in a sample of healthy volunteers". *Int. J. Res. Ayurveda Pharm.*, 8(3), 91-95, 2017.

[4] Shojaii, A., Abdollahi Fard, M., "Review of pharmacological properties and chemical constituents of *Pimpinella anisum*". *ISRN Pharmaceutics*. Article ID 510795, 8 pages, 2012.

[5] Ashraf, M., Foolad, M.R., "Pre-sowing seed treatments shotgun approach to improve germination, plant growth and crop yield under saline and non-saline conditions", *Advances in Agronomy.*, 88, 223-271, 2005.

[6] Shahrajabian, M.H., Soleymani, A., Naranjani, L., "Grain yield and forage characteristics of forage sorghum under different plant densities and nitrogen levels in second cropping after barley in Isfahan, Iran", *Research on Crops.*, 12(1), 68-78, 2011.

[7] Shahrajabian, M.H., Xue, X., Soleymani, A., Ogbaji, P.O., Hu, Y., "Evaluation of physiological indices of winter wheat under different irrigation treatments using weighing lysimeter",*International Journal of Farming and Allied Sciences*, 2(24), 1192-1197, 2013.

[8] Shahrajabian, M.H., Wenli, S., Qi, C., "The power of natural Chinese medicine, ginger root in an organic life", *Middle-East Journal of Scientific Research.*, 27(1), 64-71, 2019.

[9] Mahdavi, B., "Effects of priming treatments on germination and seedling growth of Anise (*Pimpinella anisum L.*)", *Agriculture Science Development.*, 5(3), 28-32, 2016.

[10] Soleymani, A., Khoshkharam, M., Shahrajabian, M.H., "Germination rate and initial growth of silage corn grown under various fertility systems", *Research on Crops.*, 13(3), 1035-1038, 2012.

[11] Soleymani, A., Shahrajabian, M.H., Khoshkharam, M., "The impact of barley residue management and tillage on forage maize", *Romanian Agricultural Research.*, 33, 161-167, 2016.

[12] Bhardwaj, A.K., Kapoor, S., Naryal, A., Bhardwaj, P., Warghat, A.R., Kumar, B., Chaurasia, O.P., "Effect of various dormancy breaking treatments on seed germination, seedling growth and seed vigour of medicinal plants", *Tropical Plant Research.*, 3(3), 508-516, 2016.

[13] Shahrajabian, M.H., Soleymani, A., Khoshkharam, M., "Influence of green manuring from different cover crops and farm yard manures on quantitative and qualitative characteristics of forage corn in low input farming", *Research on Crop Ecophysiology*, 12(2), 62-68, 2017.

[14] Shahrajabian, M.H., Wenli, S., Qi, C., "A review of Goji berry (*Lycium barbarum*) in traditional Chinese medicine as a promising organic superfood and superfruit in the modern industry", *Academia Journal of Medicinal Plants*, 6(12), 437-445, 2018.

[15] Soleymani, A., Shahrajabian, M.H., "Changes in germination and seedling growth of different cultivars of cumin to drought stress", Cercetari *Agronomice in Moldova*, 1(173), 91-100, 2018.

[16] Patel, R.G., Mankad, A.U., "Effect of gibberellins on seed germination of *Tithonia rotundifolia* Blake", *International Journal of InnovativeResearch in Science, Engineering and Technology*, 3(3), 10680-10684, 2014.

[17] Bell, D.T., Rokich, D.P., Mcchesney, C.J., Plummer, J.A., "Effects of temperature, light and gibberellic acid on the germination of seeds of 43 species native to Western Australia", *J. Vegetat. Sci.*, 6, 797-806, 1995.

[18] Zhao, K.F., Li, M.L., Liu, J.Y., 1986. "Reduction by GA<sub>3</sub> of NaCl-induced inhibition of growth and development in Suaeda ussuriensis", *Austr. J. Plant Physiol.*, 13, 547-551, 1986.



[19] Lin, C.C., Kao, C.H., "NaCl stress in rice seedlings, starch mobilization and the influence of gibberellic acid on seedling growth", *Bot. Bull. Acad. Sin.*, 36, 169-173, 1995.

[20] Tsygankova, V., Andrusevich, Y., Shtompel, O., Myroljubov, O., Hurenko, A., Solomyanny, R., Mrug, G., Frasinyuk, M., Shablyn, O., Brovarets, V., "Study of Auxin, cytokinetin and gibberellinslike activity of heterocyclic compounds derivatives of pyrimidine, pyridine, pyrazole and isoflavones", *European Journal of Biotechnology and Bioscience*, 4(12), 29-44, 2016.

[21] El-Barghathi, M.F., El-Bakkosh, A., "Effect of some mechanical and chemical pre-treatments on seed germination and seedling growth of *Quercus coccifera* (Kemes Oaks)", Jerash Private University, 2005.

[22] Shekari, F., Abbasi, A., Mustafavi, S.H., "Effect of gibberellic acid, salicylic acid and paclobutrazol on oxidative stress in wheat seed under accelerated ageing", *Crop Research*. (1, 2 & 3), 25-32, 2015.

[23] Liopa-Tsakalidi, A., Zakynthinos, G., Varzakas, T., Xynias, I.N., "Effect of NaCl and GA<sub>3</sub> on seed germination and seedling growth of eleven medicinal and aromatic crops", *Journal of Medicinal Plants Research*, 5(17), 4065-4073, 2011.

[24] Fernandez, H., Perez, C., Revilla, M.A., Perez-Gar-cia, F., "The level of GA<sub>3</sub> and GA<sub>20</sub> may be associated with dormancy release in *Onopordum nervosum* seeds", *Plant Growth Regulation*. 38(2), 141-143, 2002.

[25] Hassan, M.A., Fardous, Z., "Seed germination, pollination and phenology of *Gloriosa superb L*. (Liliaceae)", *Bangladesh Journal of Plant Taxonomy*, 10(1), 95-97, 2003.

[26] Sharifi, M., Pouresmael, M., "Breaking seed dormancy in *Bunium persicum* by stratification and chemical substances", *Asian J. Plant Sci.*, 5(4), 695-699, 2006.

[27] Rosner, L.S., Harrington, J.T., Dreesen, D.R., Murray, I., "Effect of gibberellic acid and standard seed treatments on mountain snowberry germination", *Native Plants Journal*, 3(2), 155-162, 2002.

[28] Nkomo, M., Kambizi, L., "Germination studies on *Corchorus olitorius L*. (Tiliaceae) (Jew's Mallow); a wild leafy vegetable for possible domestication in the Eastern Cape, South Africa", *Aspects of Applied Biology*, 96, 55-59, 2010.

[29] Rouhi, H.R., Tavakkol Afshari, R., Shakarami, K., "Seed treatments to overcome dormancy of waterlily tulip (*Tulipa kaufmanniana Regel.*)", *Australian Journal of Crop Science*, 4(9), 718-721, 2010.

[30] Gupta, A., Parihar, S.S., Choudhary, V.K., Naseem, M., Maiti, R.K., "Germination, dormancy and its removal in Isabgol (*Plantago ovata Forsk*)", *Int. J. Agric. Environ & Biotech.*, 1(3), 117-124, 2008.

[31] Eudes, F., Acharya, S., Laroche, A., Selinger, L.B., Cheng, K.J., "Novel method to induce direct somatic embryogenesis, secondary embryogenesis and regeneration of fertile green cereal plants", *Plant Cell, Tissue and Organ Culture*, 73, 147-157, 2003.

[32] Abu-Romman, S.M., Al-Hadid, K.A., Arabiyyat, A.R., "Kinetin is the most effective cytokinin on shoot multiplication from cucumber", *Journal of Agricultural Sciences*, 7(10), 159-165, 2015.

[33] Davies, P.J., "Plant hormones: Their nature, occurrence and function. In P. J. Davies (Ed.), Plant Hormones: Physiology, Biochemistry and Molecular Biology (pp. 1-12)", Kluwer Academic Publishers, Dordrecht. 1995. <u>http://dx.doi.org/10.1007/978-94-011-0473-9\_1</u>



[34] Mok, M.C., "Cytokinins and plant development- An overview. In D. W. S. Mok & M. C. Mok (Eds.), Cytokinnis-Chemistry, Activity, and Function (Chapter 12, pp. 155-166)", Boca Raton: CRC Press. 1994.

[35] Malik, K.A., Saxena, P.K., "Thidiazuron induces high-frequency shoot regeneration in intact seedlings of pea (*Pisum sativum*), chickpea (*Cicer arientinum*) and lentil (*Lens culinaris*)", *Aust J Plant Physiol.*, 19, 731-740, 1992.

[36] Sawan, Z.M., Mohamed, A.A., Sakr, R.A., Tarrad, A.M., "Effect of kinetin concentration and methods of application on seed germination, yield components, yield and fiber properties of the Egyptian cotton (*Gossypium barbadense*)", *Environmental and Experimental Botany*, 44, 59-68, 2000.

[37] Sharma, R.K., Sharma, S., Sharma, S.S., "Seed germination of some medicinal plants of Lahaul and Spiti cold desert (Himachal Pradesh): implications for conservation and cultivation", *Current Science*. 90(8), 1113-1118, 2006.

[38] Narra, H., Mamidala, P., Mehta, P.M., "Gibberellic acid and cycloheximide influenced the growth and biochemical constituents of a medicinally important plant- *trachyspermum ammi* (*I*.) *Sprague*", *Current Trends in Biotechnology and Pharmacy*, 4(1), 596-603., 2010.

[39] Polisetty, R., Paul, V., Deveshwar, J.J., Khetarpal, S., Suresh, K., Chandra, R., "Multiple shoot induction by benzyladenine and complete plant regeneration from seed explants of chickpea (*Cicer arietinum L.*)", *Plant Cell Reports*, 16, 565-571, 1997.

[40] Gulati, A., Jaiwal, P.K., "Plant regeneration from cotyledonary node explants of mungbean (*Vigna radiate L. Wilczek*)",*Plant Cell Rep.*, 13, 523-527, 1994.

[41] Prakash, N.S., Pental, D., Sarin, N.B., "Regeneration of pigeonpea (*Cajanus cajan*) from cotyledonary node via multiple shoot formation", *Plant Cell Rep.* 13, 623-627, 1994.