

VETERİNER HEKİMLER DERNEĞİ DERGİSİ

JOURNAL OF THE TURKISH VETERINARY MEDICAL SOCIETY

ISSN: 0377-6395 e-ISSN: 2651-4214 Dergi ana sayfası: Journal homepage:

http://dergipark.org.tr/vetheder

DOI: 10.33188/vetheder.599455 Olgu Sunumu / Case Report

Anisakis spp. infection in Atlantic mackerel (Scomber scombrus, Linnaeus 1758) from the Sea of Marmara

Gökben ÖZBAKIŞ BECERIKLISOY 1, a*, Ceren AŞTI 1, b, Bahadır GÖNENÇ 1, c

¹ Ankara University Faculty of Veterinary Medicine, Department of Parasitology, 06110, Ankara, Turkey ORCID: 0000-0002-3354-9150 °; 0000-0002-8424-2343 b; 0000-0003-0410-7326 °

MAKALE BİLGİSİ:

ARTICLE INFORMATION:

Geliş / Received: 31 Temmuz 2019 31 July 2019

Kabul / Accepted: 23 Ağustos 2019 23 August 2019

Keywords: Anisakidae Anisakis spp. L3 stages

Anahtar Sözcükler: Anisakidae Anisakis spp. L3 dönemi

ABSTRACT:

This study was aimed to determine the parasite infection of fish samples. Ten Atlantic mackerel (*Scomber scombrus*), which were suspected with parasite infection, were sent to the laboratory by the fisherman at the Sea of Marmara, Turkey. The skin, fins, eyes, buccal cavity, and gills were examined regarding ectoparasites, while internal organs were investigated concerning endoparasites. Seven of 10 fish were infected with endoparasites. A total of 27 nematode larvae were detected from the abdominal cavity of fish. The detected nematode larvae were rinsed in 0.9 % isotonic saline and preserved in glycerine-alcohol until examined. After that the anterior and posterior end of samples were cleared in lactophenol. The larvae had a prominent boring tooth at the anterior end. The body features of the larvae were measured (body width, oesophagus length, ventriculus length and mucron length) on a light microscope (4x-10x-40x). Morphological analysis showed that all samples were identified as *Anisakis* Type I larvae (L3). It is known that anisakiasis is one of the important fish-borne zoonotic diseases. Also, this nematode species had been previously reported in Turkish water from different fish species. The present case contributes valuable information about one of the intermediate hosts of *Anisakis* spp. in Turkey.

Marmara denizi Atlantik uskumrularındaki (Scomber scombrus, Linnaeus 1758) Anisakis spp. enfeksiyonu

ÖZET:

Bu çalışmada balık örneklerinde paraziter enfeksiyonların belirlenmesi amaçlandı. Marmara denizi balıkçıları tarafından paraziter enfeksiyon şüpheli on adet Atlantik uskumrusu (*Scombrus scombrus*) laboratuvara gönderildi. Balıklara ait deri, yüzgeçler, gözler, ağız boşluğu ve solungaçlar ektoparazitler yönünden incelenirken, iç organların da endoparazitler yönünden araştırmaları yapıldı. Gönderilen on adet balığın yedisinin enfekte olduğu saptandı. Balıkların karın boşluğunda toplam 27 nematod larvası tespit edildi. Saptanan nematod larvaları % 0.9'luk izotonik tuzlu su ile yıkandı ve gliserinalkol içinde saklandı. Sonrasında, örneklerin anterior ve posterior uçları laktofenolde şeffaflandırıldı. Larvaların ön kısmında belirgin bir kesici diş (boring tooth) görüldü. Larvaların vücut özellikleri (boy uzunluğu, özefagus uzunluğu, ventrikül ve mucron uzunluğu) ışık mikroskopu altında ölçüldü (x-4-x10-x40). Morfolojik analizler sonucu tüm örneklerin Anisakis Tip I larvaları (L3) olduğu saptandı. Anisakiasis balık-kaynaklı (fish-borne) zoonotik hastalıkların en önemlilerinden biri olarak bilinmektedir. Ayrıca, bu nematod türlerinin daha önce Türk sularında farklı balık türlerinde de bulunduğu bildirilmiştir. Bu çalışma ile, *Anisakis* spp.'nin Türkiye'deki ara konaklarından biri hakkında değerli bilgi elde edilecektir.

How to cite this article: Beceriklisoy ÖG, Aşti C, Gönenç B: Anisakis spp. infection in Atlantic mackerel (Scomber scombrus, Linnaeus 1758) from the Sea of Marmara. Veteriner Hekimler Dernegi Dergisi, 91(1): 80-85, 2020, DOI: 10.33188/vetheder.599455

^{*} Sorumlu yazar e-posta adresi / Corresponding author e-mail address: ozbakis@ankara.edu.tr

1. Introduction

Anisakis spp. (Dujardin, 1845) is found in the digestive tract of marine mammals (cetaceans) and fish-eating birds. The parasite occurs in the Atlantic ocean, the Pacific ocean and Mediterranean sea in a wide range of marine mammals species (12). Although, there are few studies on parasites of sea mammals in Turkey (8, 27, 33), Anisakis spp. were detected in 6 striped dolphins (Stenella coeruleoalba) on the Eastern Mediterranean coast of Turkey (8).

The larvae of *Anisakis* spp. infect the aquatic invertebrates (cephalopods) and non-mammalian vertebrates (fish species) as an intermediate host or paratenic host. The crustaceans are the first intermediate hosts of anisakids (21, 22). According to Berland (10) morphological features of *Anisakis* spp. larvae can be distinguished as Type I and Type II. Anisakis Type I includes; *Anisakis simplex* complex (*A. simplex* sensu stricto (s.s), *A. pegreffii*, *A. simplex* C), *A. typica*, *A. ziphidarum* and *A. nascetti*, besides Type II includes; *A. paggiae*, *A. physeteris* and *A. brevispiculata* (21, 22, 24). Inside of them, *A. simplex* s.s., *A. pegreffii* from Type I and *A. physeteris* from Type II are also reported in humans (6, 23). Humans can be infected by ingesting raw or undercooked fish or squids as a paratenic hosts. It is one of the important pathogens associated with human and animal health. The third stage of larvae can penetrate the mucosa of the gastrointestinal tract, producing acute or chronic abdominal syndromes (abdominal pain, vomiting, diarrhea and mild fever) in humans. Also in some cases, serious allergic reactions such as urticaria, angioedema and anaphylaxis can occur (7, 29).

The *Anisakis* spp. larvae were detected from throughout the world involving Africa, Australia, Oceania, Europe, North America and South America in a wide range of fish species (1). Besides, larvae of *Anisakis* spp. had been reported in Turkish coastal waters from different researchers and fish species (Table 1).

Table 1: Data on the distribution of *Anisakis* spp. larvae in Turkish coastal waters *Tablo 1:* Türkiye kıyı sularında Anisakis spp. larvalarının dağılımına ilişkin veriler

References	Fish species			Diagnostic methods			
		Origine	Number of examined fish (n)	Morphological	Molecular	Number of infected fish (n/ %)	Number of detected larvae (n)
(3)	Red mullet (Mullus surmuletus)	AS	142	A. simplex	-	4/ 2.8	7
(4)	Chup mackerel (Scomber japonicus)	AS	110	A. simplex	-	40/ -	163
(4)		M	122			9/ -	14
(19)	Atlantic mackerel (Scomber scombrus)	SM	20	A. simplex	-	2/ 10.0	21
(5)	Conger eel (Conger conger L.)	AS	26	A. simplex	-	4 / 15.38	14
(32)	Horse Mackerels (Trachurus trachurus)	BS	-	-	A. pegreffii A. simplex	-	6
(2)	11 different species	AS	235	A. simplex	-	33/ -	160+>100
(20)	Merlangius merlangus	BS	3	-	A. simplex	3/ -	10
(30)	Trachurus mediterraneus	BS	56	A. pegreffii	- A. pegreffii	1/2	11
(25)	10 different species	AS	70	Anisakis Type I	A. typica Hybrid	33/47.1	569
	7 1:66	М	50	4 Li- T I	genotypes	22/46.0	107
(2.4)	7 different species	M	50	Anisakis Type I	A. pegreffii	23/46.0	197
(34)	John Dory (Zeus faber)	M BS	1	Anisakis Type I	A. pegreffii	1/ -	15
(26)	23 different species	AS M	475	Anisakis Type I	-	-	102

BS: Black Sea, SM: Sea of Marmara, AS: Aegean Sea, M: Mediterranean Sea

Atlantic mackerel (Scomber scombrus, Linnaeus 1758) is represented by the Scombridae family of the Actinopterygii class. It is one of the widely distributed migratory pelagic fish species and lives in North-Atlantic

82 Vet Hekim Der Derg 91(1): 80-85, 2020

including Mediterranean (18). It has been assessed as the least concern (LC) in The Red List of Threatened Species since 2011 by The IUCN (15). In Turkey, Atlantic mackerel were reported in the Black Sea, Sea of Marmara, Aegean Sea, and the Mediterranean Sea (11). The 46.6-728.2 tons of Mackerel caught in-between years of 2009 and 2018 from these waters (31).

The aim of the present case was to investigate the presence of the Anisakid larvae in Atlantic mackerel from Sea of Marmara, Turkey.

2. Case Story

Ten Atlantic mackerel were sent by the fishermen in the Sea of Marmara, Turkey. Fish were stored individually, and transported with ice-box to the Ankara University, Faculty of Veterinary Medicine, Parasitology laboratory. Fish were investigated with parasitological examination. The skin, fins, eyes, buccal cavity and gills were examined in terms of ectoparasites, while internal organs were investigated regarding endoparasites. The nematode larvae were observed on 7 of 10 fish (3.85±3.38; min: 1, max: 10). Total of 27 nematode specimens were collected from the abdominal cavity. The samples were rinsed in isotonic solution 0.9 % and fixed in glycerine-alcohol after that anterior and the posterior end of the larvae were clarified in lactophenol. The samples were evaluated under the light microscope (x4-x10-x40) and the body features were measured (body width, length of oesophagus, ventriculus, and mucron) (Table 2). Morphological identification was conducted according to Berland (10). The larvae were white and cylindrical shape. The larvae had a prominent boring tooth, excretory pore opened ventrally at the anterior end and the distinct mucron was located at the caudal end. The morphological analysis confirmed that all examined samples were *Anisakis* Type I larvae (Figure 1).

Table 2: The comparative body measurements of *Anisakis* Type I larvae (mm)

Tablo 2: Anisakis Type I larvalarının karşılaştırmalı vücut ölçümleri (mm)

	Present study	A. simplex (28)	A. pegreffii (34)	A. pegreffi x A.simplex (ss) (17)	A. typica (16)
Total body length	9.12- 25.36	12.75-29.94	12.80-24.65	9.0-17.5	21.05-24.97
Length of oesophagus	0.95-1.79	1.18-2.58	0.98-1.87	0.56-1.75	1.50-1.70
Ventriculus length	0.39-0.74	0.90-1.50	0.53-0.74	0.20-0.40	0.89-0.94
Mucron length	0.01-0.02	0.02-0.03	0.01-0.02	0.01-0.03	0.005-0.010

3. Discussion and Conclusion

Anisakiss is a fish-borne zoonotic parasitic disease and were detected throughout the world in humans (13). Anisakis Type I larvae, including A. simplex sensu stricto and A. pegreffi have been reported as agents of human anisakiasis (9, 14). It is well-known, Anisakis larvae reported surviving in various fish species (13). The larvae of A. simplex and A. pegreffi had been reported in Turkish coastal waters from different researchers (25, 26, 32). Anisakis spp. larvae were detected in Aegean (25) and the Sea of Marmara (19) from Atlantic mackerel as previously reported. In the present case, the higher rate of infected Atlantic mackerel (7/10) might be assessed as a potential intermediate host of anisakiasis for Turkish waters.

Despite the extensity of this nematoda in Turkish coast, there is no reported case in human from Turkey. Because of symptoms of human anisakiasis are usually not specific and the disease is thought to be frequently misdiagnosed and underdiagnosed (9).

Vet Hekim Der Derg 91(1): 80-85, 2020

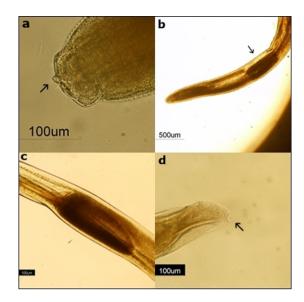


Figure 1: Anisakis Type I larvae (a) anterior end, larval tooth (arrow) (x40), (b) anterior end, ventriculus (arrow) (x4) (c) ventriculus (x10), (d) caudal end, mucron (arrow) (x20)

Şekil 1: Anisakis Tip I larva (a) ön nihayet, larval diş (ok ucu) (x40), (b) ön nihayet, ventriculus (ok ucu) (x4) (c) ventriculus (x10), (d) arka nihayet, mukron (ok ucu) (x20)

According to Berland (10), larval morphological features including the absence of a ventricular appendage and an intestinal caecum are useful for the distinction between several anisakid genera. *Anisakis Type I or Type II* larvae can be identified based on ventriculus length and the presence of a tail spine (or mucron), however, these measurements insufficient for species identifications. For this purpose, the latest studies were conducted via molecular approaches. Although our preservation methods of samples were useful for PCR protocol, we couldn't obtain sufficient amount of DNA. Therefore, the identification of larvae samples could not be performed via PCR.

Although five hundred twelve marine fish species were previously identified from the Turkish coasts (11), the *Anisakis* spp. larvae were detected in only 23 fish species in Turkey so far (2, 25). Due to the rich variety of Turkish marine fish species, further studies are required. Every new data is essential to understand the epidemiology of anisakiasis.

Acknowledgments

This case was presented on the 13th International Symposium on Fisheries and Aquatic Sciences, 21-23 November 2018, Ankara, Turkey.

References

- 1. Aibinu IE, Smooker PM, Lopata AL (2019): Anisakis nematodes in fish and shellfish-from infection to allergies. Int J Parasitol Parasites Wildl, 9, 384-393.
- **2. Akmırza A** (2013): *Gökçeada kıyı sularındaki balıkların parazitik nematodları*. Turkiye Parazitol Derg, **37**, 199-202.
- **3. Akmirza A** (2000): *Gökçeada civarında avlanan tekir (Mullus surmuletus L.) balığının metazoan parazitleri.* Vet Fak Derg (İstanbul), **26(1)**, 129-140.
- **4. Akmirza A** (2003): Distribution of parasites fauna of Chup mackerel in Aegean and Mediterranean Sea. Turkish J Marine Science, **9(3)**, 187-195.
- **5. Akmirza A** (2012): *Metazoan parasite fauna of Conger eel (Conger conger L.) near Gökçeada, northeasten Aegean Sea, Turkey.* Kafkas Univ Vet Fak Derg, **18(5)**, 845-848.

6. Arizono N, Yamada M, Tegoshi T, Yoshikawa M (2012): Anisakis simplex sensu stricto and Anisakis pegreffii: biological characteristics and pathogenetic potential in human anisakiasis. Foodborne Pathog Dis, 9(6), 517-521.

- 7. Audicana L, Audicana MT, Fernandez de Corres L, Kennedy MW (1997): Cooking and freezing may not protect against allergenic reactions to ingested Anisakis simplex antigens in humans. Vet Rec, 140(9), 235.
- **8.** Aytemiz I, Dede A, Danyer E, Tonay AM (2012): Morphological identification of parasites found in the stomach contents of bycaught striped dolphins (Stenella coeruleoalba) from Turkish Eastern Mediterranean Sea coast. J Black Sea/Mediterr, **18**, 238-245.
- 9. Bao M, Pierce GJ, Strachan NJC, Pascual S (2019): Human health, legislative and socioeconomic issues caused by the fish-borne zoonotic parasites Anisakis: challenges in risk assessment. Trends Food Sci Tech, 86, 298-310.
- **10.** Berland B (1961): Nematodes from some Norwegian marine fishes. Sarsia, **2**, 1-50.
- 11. Bilecenoğlu M, Kaya M, Cihangir B, Çiçek E (2014): An updated checklist of marine fishes of Turkey. Turk J Zool, 38, 901-929.
- 12. Bilska-Zajac E, Różycki M, Chmurzyńska E, Karamon J, Sroka J, Kochanowski M, Kusyk P, Cencek T (2015): Parasites of Anisakidae family— geographical distribution and threat to human health. J Agr Sci Tech-Iran, 5, 146-152.
- **13. Buchmann K, Mehrdana F** (2016): Effects of anisakid nematodes Anisakis simplex (s.l.) Pseudoterranova decipiens (s.l.) and Contracaecum osculatum (s.l.) on fish and consumer health. FAWPAR, **4**, 13-22.
- 14. Cipriani P, Sbaraglia GL, Palomba M, Giulietti L, Bellisario B, Bušelić I, Mladineoc I, Cheleschid R, Nascetti G, Mattiucci S (2018): Anisakis pegreffii (Nematoda: Anisakidae) in European anchovy Engraulis encrasicolus from the Mediterranean Sea: Fishing ground as a predictor of parasite distribution. Fish Res, 202, 59-68.
- 15. Collette B, Boustany A, Carpenter KE, Di Natale A, Fox W, Graves J, Juan Jorda M, Kada O, Nelson R, Oxenford H (2011): Scomber scombrus. The IUCN Red List of Threatened Species 2011: e.T170354A6764313. Erişim adresi: http://dx.doi.org/10.2305/IUCN.UK.2011-2.RLTS.T170354A6764313.en. Erişim tarihi: 28.07.2019
- 16. Da Fonseca MCG, Knoff M, Felizardo NN, Di Azevedo MIN, Torres EJL, Gomes DC, Iñiguez AM, De São Clemente SC (2016): Integrative taxonomy of Anisakidae and Raphidascarididae (Nematoda) in Paralichthys patagonicus and Xystreurys rasile (Pisces: Teleostei) from Brazil. Int J Food Microbiol, 235, 113-124.
- 17. Eissa AE, Showehdi ML, Ismail MM, El-Naas AS, Mhara, AAA, Abolghait SK (2018): Identification and prevalence of Anisakis pegreffii and A. pegreffii x A. Simplex (ss) hybrid genotype larvae in Atlantic horse mackerel (Trachurus trachurus) from some North African Mediterranean coasts. Egypt J Aquat Res, 44(1), 21-27.
- **18. Fish Base** (2019): *Scomber scombrus* Linnaeus 1758, Atlantic mackerel. Erişim adresi: https://www.fishbase.se/Summary/SpeciesSummary.php?ID=118&AT=atlantic+mackerel Erişim tarihi: 27.07.2019
- 19. Keser R, Bray RA, Oğuz MC, Çelen S, Erdoğan S, Doğutürk S, Aklanoğlu G, Martı B (2007): Helminth parasites of digestive tract of some teleost fish caught in the Dardanelles at Çanakkale, Turkey. Helminthologia, 44 (4), 217-221.
- **20. Keskin E, Genc E, Unal EM** (2013): *Identification of Anisakis simplex using DNA barcoding*. Aquaculture Europe 2013-Trondheim, Norway.
- **21. Mattiucci S, Nascetti G (2006):** *Molecular systematics, phylogeny and ecology of anisakid nematodes of the genus Anisakis Dujardin, 1845: an update.* Parasite, **13(2)**, 99-113.
- **22.** Mattiuci S, Nascetti G (2008): Advances and trends in the molecular systematics of anisakid nematodes, with implications for their evolutionary ecology and host—parasite co-evolutionary processes. Adv Parasitol, **66**, 47-148.

Vet Hekim Der Derg 91(1): 80-85, 2020 **85**

23. Molina-Fernández D, Adroher FJ, Benítez R (2018): A scanning electron microscopy study of Anisakis physeteris molecularly identified: from third stage larvae from fish to fourth stage larvae obtained in vitro. Parasitol Res, 117, 2095–2103.

- 24. Orecchia P, Paggi L, Mattiucci S, Nascetti G, Smith JW, Bullini L (1986): Electrophoretic identification of larvae and adults of Anisakis (Ascaridida: Anisakidae). J Helminthol, 60, 331-339.
- 25. Pekmezci GZ, Onuk EE, Bolukbas CS, Yardimci B, Gurler AT, Acici M, Umur S (2014): Molecular identification of Anisakis species (Nematoda: Anisakidae) from marine fishes collected in Turkish waters. Vet Parasitol, 201, 82–94.
- **26.** Pekmezci GZ, Yardimci B (2019): On the occurrence and molecular identification of Contracaecum larvae (Nematoda: Anisakidae) in Mugil cephalus from Turkish waters. Parasitol Res, **118(5)**, 1393-1402.
- 27. Pekmezci GZ, Yardimci B, Gurler AT, Bolubas CS, Acici M, Umur S (2013): Survey on the presence of nematodes and associated with pathology in marine mammals from Turkish waters. Kafkas Univ Vet Fak Derg, 19(6), 1035-1038.
- **28.** Quiazon KMA, Yoshinaga T, Ogawa K, Yukami R (2008): Morphological differences between larvae and in vitro-cultured adults of Anisakis simplex (sensu stricto) and Anisakis pegreffii (Nematoda: Anisakidae). Parasitol Int, **57(4)**, 483-489.
- 29. Şimşek E, Yılmaz E (2016): Anisakiasis ve halk sağlığı. Turkiye Klinikleri J Food Hyg Technol-Special Topics, 202(3), 38-44.
- **30. Tepe Y, Oğuz MC** (2013): Nematode and acanthocephalan parasites of marine fish of the eastern Black Sea coasts of Turkey. Turk J Zool, **37**, 753-760.
- **31. Tüik** (2019): Su ürünleri istatistikleri: Deniz ürünleri: Avlanan deniz balıkları miktarı. Erişim adresi: http://www.tuik.gov.tr/PreTablo.do?alt id=1005 Erişim tarihi: 31.07.2019
- **32.** Utuk AE, Piskin FC, Balkaya I (2012): Molecular detection of Anisakis pegreffii in Horse mackerels (Trachurus trachurus) sold for human consumption in Erzurum province of Turkey. Kafkas Univ Vet Fak Derg, 18(2), 303-307.
- **33.** Veryeri NG (2012): Postmortem examinations of stranded dolphins found on the Black Sea coast near Ordu, Turkey (Mammalia: Cetacea). Zool Middle East, **55(1)**, 129-132.
- **34.** Yardımcı B, Pekmezci GZ, Onuk EE (2014): Pathology and molecular identification of Anisakis pegreffii (Nematoda: Anisakidae) infection in the John Dory, Zeus faber (Linnaeus, 1758) caught in Mediterranean Sea. Ankara Univ Vet Fak Derg, **61**, 233-236.