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EXAMINATION OF POLLEN MORPHOLOGY OF SOME AETHIONEMA (BRASSICACEAE), FROM TURKEY

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ABSTRACT. Pollen morphology of 23 species of taxonomically difficult genus Aethionema W.T.Aiton was examined by light microscopy (LM) and scanning electron microscopy (SEM). Pollen grains have showed significant variation in aperture number and type. Generally pollen is tricolpate, but tricolpate-syncolpate, tricolpate-syncolpate-tetracolpate, syncolpatetetracolpate, or merely syncolpate apertures also occur and make a heteromorphic assemblage. Pollen shape is prolate, prolate-spheroidal, subprolate, spheroidal or suboblate, and pollen ranges from 9.4 to 29 m (P), 6.2 to 23.9 m (E). Five main pollen types were identified based on the sculpturing of the exine. The pollen shape, size, surface ornamentation, muri and lumina shape and size are found as important and useful features for distiguishing of the taxa.

1. INTRODUCTION

The genus Aethionema W.T.Aiton (Brassicaceae) has 75 species in the worldwide, of which 31 species are endemic to Turkey [1,2]. Turkey is gene-center of the genus and maintains 51 taxa [3,4,5,6]. Aethionema is a taxonomically difficult genus and few macromorphological characters are available for species delimitation. Life span (annual or perennial) and fruit morphology are very importance at the species level [3]. Brassicaceae contains nectary plant taxa, and these are very important for honey bees [7].

Many researchers have emphasized the importance of pollen morphology for the Brassicaceae family [8,9,10,11,12]. İnceoğlu and Karamustafa (1977) was studied the pollen morphology of *Aethionema arabicum* (L.) Andrz. Ex DC and *A. armenum* Boiss. with light microscopy [13]. Atçeken (2016) studied pollen morphology of four taxa (*A. karamanicum* Ertugrul & Beyazoglu, *A. cordatum* Boiss., *A. arabicum* and

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A. armenum) of genus [14]. Karaismailoğlu (2017) examined pollen morphology of eleven Aethionema taxa (A. froedinii Rech. Fil., A. arabicum, A. eunomioides (Boiss.) Bornm., A. fimbriatum Boiss., A. speciosum Boiss ssp. speciosum, A. speciosum ssp. compactum, A. saxatile, A. oppositifolium (Pers.) Hedge, A. iberideum (Boiss.) Boiss., A. armenum and A. grandiflorum Boiss. & Hohen.) distributed in Turkey [15].

Our objective was to examine the pollen morphology of 23 Aethionema species and determine the contributions of pollen morphology to taxonomy of the genus.

2. Material & Method

All pollen samples were taken from herbarium specimens in the GAZI collection. The voucher specimens are listed in the Appendix.

The pollen grains were studied by light microscopy (LM) and also by scanning electron microscopy (SEM) in order to investigate the exine sculpturing in detail. Pollen for LM was prepared by the methods of Wodehouse (1935) [16]. The LM studies were performed using a Nikon E-600 microscope. The following parameters were measured: polar (P) and equatorial (E) axis, colpus length (Clg) and width (Clt) and exine and intine thickness. For most specimens, the measurements of P (polar axis) and E (equatorial axis) represent the mean of 50 or more pollen grains, and of some 10 measurements for exine and intine thickness. The LM photographs were taken with a Nikon FDX–35 camera connected to a Nikon E-600 photomicroscope. For SEM, the air-dried pollen were covered with gold during exaporation prior to examination on the screen of a jeol electron microscope. Terminology follows that of Faegri-Iversen [17] and Brochmann [8]. The Simpson and Roe graphical test was used for graphical calculations [18]. The order of the species in the list adopted to Davis [19]. All samples have been deposited by Adıgüzel, N. in the Gazi University Herbarium. Specimens examined were given in Table 1.

A. lepidioides (Hub-Mor.)	B6 Sivas
A. carneum (Banks & Sol.) B.Fedtsch.	C7 Şanlıurfa
A. syriacum (Boiss.) Bornm.	C7 Şanlıurfa
A. heterocarpum J.Gay	C5 Adana
A. virgatum (Boiss) Hedge	Iran Zanjan
A. lycium I.A.Andersson et al.	C3 Antalya
A. stylosum DC.	C4 İçel
A. trinervium (DC.) Boiss.	B9 Van, Iran Tehran
A. spicatum Post	B5 Ni ğ de
A. caespitosum (Boiss.) Boiss.	C5 Kayseri
A. subulatum (Boiss. & Heldr.) Boiss.	C3 Antalya
A. glaucinum Greuter et al.	B5 Ni ğ de
A. membranaceum DC.	B9 Van, C3 Antalya
A. schistosum Boiss. & Kotschy	B4 Konya, C4 İçel
A. capitatum Boiss. & Balansa	B7 Kahramanmara ş
A. diastrophis Bunge	B8 Erzurum
A. coridifolium DC.	B6 Kahramanmara ş
A. munzurense Davis & Yıld.	B7 Tunceli
A. marashicum P.H.Davis	C6 Kahramanmara ş
A. huber-morathii P.H.Davis & Hedge	C5 Adana
A. turcicum H.Duman & Aytaç	B4 Ankara
A. dumanii Vural & Adıgüzel	B4 Ankara
A. alanyae H.Duman	C3 Antalya

Table 1. The information of specimens and localities

3. Results

The main palynological features of the *Aethionema* species examined are summarized in Table 2 and Figure 1.





3.1 Size, Symmetry and Shape

Pollen are isopolar, radialy symmetric. Pollen shape is prolate, prolatespheroidal, subprolate, spheroidal or suboblate; P: 9.4–29 μ m; E: 6.2–23.9 μ m; P/E: 0.82–1.67. Outline is elliptic or circular in equatorial view and usually semiangular, rarely more or less circular in polar view (Figures 2,3 and 4).

3.2 Apertures

The pollen grains are tricolpate, tetracolpate or syncolpate. The colpus is long, usually wide. Apertural membrane is generally psilate or rarely granular (Figures 2,3 and 4). Pollen grains of some taxa are surrounded by colpus (syncolpate pollen grains) (Figures 2,3 and 4).

Exine

Exine is semitectate with a reticulate sculpturing. Lumina are generally 0.1 μ m or less than 0.1 μ m wide (Table 2). Lumina shape is irregular, elliptic or polygonal (Figure 4). Muri is less than 0.01 μ m thick. Exine thickness is 0.8–2.3 μ m. Ectexine is slightly thicker than endexine. Intine is 0.8–2.3 μ m (Table 2). On the basis of reticulate exine sculpturing, 5 pollen types are recognised in the genus *Aethionema*:

Type I: The pollen grains of this type have a characteristic, vermiculate reticulation with relatively wide muri (mean 0.04 μ m) and irregularly shaped lumina. This type was observed in *A. lepidioides* and *A. marashicum* (Figures 2, 3 and 4). The species included in type I are characterised by tricolpate, spheroidal or prolate-spheroidal pollen grains (Table 2).

Type II: This type has characteristic pollen grains with wide muri (mean 0.06 μ m) and regularly shaped lumina. This type was characterized by tricolpate-syncolpate aperture types and observed in *A. trinervium*, *A. diastrophis* and *A. spicatum* (Figure 2, 3 and 4). All these species show heteromorphy.

Type III: Pollen grains have narrow muri (mean 0.025 μ m) and wide lumina (mean 0.05 μ m). This type was observed in *A. heterocarpum*, *A. lycium*, *A. caespitosum*, *A. subulatum*, *A. membranaceum*, *A. schistosum*, *A. dumanii* and *A. alanyae* (Figure 2, 3 and 4). Pollen grains are often tricolpate. **Type IV**: This type has characteristic pollen grains narrow muri (mean 0.025 μ m) and polygonal lumina. This type was observed in *A. virgatum, A. stylosum, A. glaucinum, A. capitatum, A. coridifolium, A. munzurense, A. huber-morathii and A. turcicum* (Figures 2, 3 and 4).

Various aperture types have been observed in this type: tricolpate (A. virgatum, A. stylosum, A. glaucinum, A. munzurense and A. turcicum), tricolpate-syncolpate (A. coridifolium), syncolpate (A. capitatum) and tetracolpate (A. huber-morathii) (Table 2). The pollen shape is often prolate-spheroidal or subprolate (Figure 3).

Type V: Pollen grains have narrow muri and irregularly shaped lumina. This type was observed in *A. carneum* (Figure 2, 3 and 4). The pollen type is prolate-spheroidal (Figure 3) and tricolpate-syncolpate (Table 2).

4. Result & Discussion

The pollen of the Aethionema species examined in this study showed considerable variations with respect to features such as the number and type of apertures and thickness of muri and width of lumina. In Aethionema, 16 species were found with merely tricolpate (A. lepidioides, A. marashicum, A. heterocarpum, A. coespitosum, A. subulatum, A. membranaceum, A. schistosum, A. dumanii, A. virgatum, A. stylosum, A. glaucinum, A. munzurense, A. turcicum and A. syriacum), one with merely syncolpate (A. capitatum) and one with merely tetracolpate pollen (A. huber-morathii). The other species were tricolpate-syncolpate (A. trinervium, A. diastrophis, A. lycium, A. alanyae, A. caridifolium and A. carneum) (Table 2). Rollins and Banerjee have observed that the most common pollen type in the family Brassicaceae is tricolpate with grooves varying in length in different genera [20]. In addition, tetracolpate form in the pollen of Dithyrea californica and pentacolpate forms characteristic of many species of the genus Querella, Physaria and Dimorphocarpa of Brassicaceae have been seen. Doğan and İnceoğlu also found tricolpate, syncolpate, tetracolpate and pentacolpate grains in the genus Isatis L. of the family Brassicaceae [21]. Tricolpate pollen grains of the species A. stylosum, A. glaucinum, A. schistosum, A. turcica and A. dumanii show considerable variations in pollen size (Table 2 and Figure 1). In each specimen of these species both 50%larger and 50% smaller pollen grains have been observed. Variation in pollen size and aperture type was attributed to heteromorphy in pollen grains by some researchers [22,23,24,25]. Heteromorphy is caused by either anomalies in meiosis [22, 26], colchicine [27,28], or hybridization [29,30].

Species	Polar axis (P)		(P)	Equatorial axis		P/E	Colpus length		Colpus width			Shape	SS	SS	Aperture type					
					(E) Ratio (Clg)						(Clgt)			Exine Thickne	Intine Thicknes		Muri width	Lumen Width	Pollen Type	
	min	mean	max	min	mean	max	mean	min	mean	max	min	mean	max					mean	mean	
A. lepidioides(A1)	16,2	16.6	16.8	16.3	16.6	16.7	1	9.2	11.4	12.4	3.7	4.2	4.8	Spheroidal	1	0.4	tricolpate	0.04	0.07	I
A. marashicum(A2)	15.6	17.3	18.7	12.5	14.6	15.6	1.18	12.3	15.6	16.9	2	2.6	3.1	Prolate- Spheroidal	1	0.5	Tricolpate, 6 % tetracolpate	0.04	0.04	I
A. trinervium(A3)	17.7	18.2	19.8	17.7	18.2	19.8	1	14.4	16.1	17.6	3.7	4.2	4.9	Spheroidal	1.1	0.3	Tricolpate, 40% syncolpate	0.06	0.03	п
A. spicatum(A4)	16.6	17.9	18.7	17.7	17.9	18.7	1	10.4	12.2	14.1	3.9	4.9	5.2	Spheroidal	0.8	0.3	Tricolpate, 3% tetracolpate	0.06	0.05	II
A. diastrophis(A5)	13.5	15.2	17.7	8.3	9.6	11.4	1.58	9.4	11.2	12.1	0.9	1.1	1.7	Subprolate	1	0.3	Tricolpate, 10% syncolpate	0.06	0.05	П
A. heterocarpum (A6)	22.9	27.3	29	17.9	17.8	18.7	1.53	15.7	22.4	23.4	1.9	2.8	3.0	Subprolate	1	0.3	tricolpate	0.02	0.05	Ш
A. lycium(A7)	13.5	15.6	16.6	9.4	11.4	12.5	1.37	9.7	12.5	13.7	1.9	2.1	2.8	Subprolate	1.6	0.3	Tricolpate, 2% syncolpate	0.03	0.04	ш
A. coespitosum(A8)	13.5	16.4	17.7	14.6	15.1	16.6	1.09	9.1	12	13.4	4.8	5.2	6.1	Prolate- Spheroidal	1.4	0.3	tricolpate	0.02	0.06	Ш
A. subulatum(A9)	14.6	15	15.6	13.5	14.6	15.6	1.03	9.2	10.4	11.1	3.1	3.5	3.9	Prolate- Spheroidal	0.9	0.3	tricolpate	0.01	0.06	Ш
A. membranaceum (A10)	14.6	17.1	19.8	8.3	10.4	13.5	1.64	9.7	13.3	14.7	1.9	2.1	3.2	Subprolate	0.9	0.3	tricolpate	0.02	0.06	Ш
A. schistosum(A11)	11.4	15.2	26	6.2	11.1	18.7	1.38	8.1	12.5	21.4	4.2	5.3	6.1	Subprolate	0.85-	0.3	tricolpate	0.007-0.01	0.05- 0.07	ш
A. dumanii(A12)	17.7	20	25	12.5	14.6	15.6	1.45	12.4	16.6	20.2	1.8	2.6	3.0	Subprolate	1	0.3	tricolpate	0.04	0.07	III
A. alanyae(A13)	13.5	16.1	16.6	12.5	15.4	15.6	1.05	8.6	15.1	16.2	2.0	2.3	2.7	Prolate- Spheroidal	0.9	0.3	Tricolpate,	13.5	16.1	IV
A. virgatum(A14)	16.6	17	17.7	12.5	15.9	17.7	1.07	10.9	12	13.1	4.8	5.2	5.7	Prolate- Spheroidal	1	0.6	tricolpate	0.03	0.2	IV
A. stylosum(A15)	15.6	21.1	23.9	17.7	20.6	23.9	1.02	10.2	18	20.4	5.1	9	9.8	Prolate- Spheroidal	1.3	0.3	tricolpate	0.02	0.06	IV
A. glaucinum(A16)	14.6	20.7	23.9	15.6	20.2	20.9	1.02	11.1	17.3	20.2	3.9	4.6	5.7	Prolate- Spheroidal	1	0.5	tricolpate	0.03	0.2	IV
A. capitatum(A17)	14.6	15.9	17.7	8.3	11	12.5	1.45	10.3	14.1	15.2	1.2	2.1	2.8	Subprolate	1.6	0.3	syncolpate	0.02	0.07	IV
A. caridifolium (A18)	16.6	17.6	18.7	14.6	15.3	15.6	1.15	13.1	15.6	16.2	2.4	3.1	3.8	Prolate- Spheroidal	2.1	0.3	Tricolpate, 10% syncolpate	0.02	0.09	IV
A. munzurense(A19)	19.8	20.5	21.8	17.7	18.5	19.8	1.11	14.1	14.8	16.1	3.4	4.2	5.1	Prolate- Spheroidal	2	0.3	tricolpate	0.02	0.1	IV
A. huber-morathii (A20)	14.6	15	15.6	15.6	15.8	16.6	0.9	8.3	10.4	11.2	4.3	5.2	5.9	Oblate- Spheroidal	0.8	0.3	tetracolpate	0.02	0.05	IV
A. turcicum(A21)	17.7	21.7	27.1	15.6	20	22.9	1.09	14.7	18.7	24.2	2.0	2.4	3.2	Prolate- Spheroidal	1.8	0.5	tricolpate	0.03	0.1	IV
A. carneum(A22)	17.7	18.7	20.8	15.6	18.4	19.8	1.02	14.2	15.6	16.7	5.8	6.2	7.1	Prolate-	1	0.3	Tricolpate, 2% Syncolpate	0.015	0.03	v
A. syriacum (A23)	25	26	27	14	15.6	17	1.67	19.7	20.8	21.2	1.1	1.6	2.0	Subprolate	1	0.3	tricolpate	0.01	0.03	П

Table 2. Pollen morphological parameters of Aethionema (values in $\mu \rm{m})$



FIGURE 2. LM pollen microphotograph of Aethionema taxa. 1–3: A. carneum, 1; Meridional, optical section, outline, 2; Aperture and ornamentation, 3; Polar view. 4–6: A. heterocarpum. 4: Meridional, optical section, outline. 5: Aperture and ornamentation. 6: Polar view. 7–9: A. stylosum. 7: Meridional, optical section, outline. 8: Ornamentation, 9: Polar view. 10–12: A. trinervium. 10: Meridional, optical section, outline. 11: Aperture and ornamentation. 12: Polar view. 13-15: A. caespitosum, 13; Meridional, optical section, outline, 14; Ornamentation. 15; Polar view. 16-18: A. membraneceum, 16; Meridional, optical section, outline, 17; Ornamentation, 18; Polar view. 19-24: A. schistosum. 19; Meridional, optical section, outline, 20; Aperture. 21; Polar view, 22; Meridional, optical section, outline. 23; Ornamentation. 24; Polar view (Scale bar: 10 μm).



FIGURE 3. LM pollen microphotograph of Aethionema taxa. 1-3: A. capitatum, 1; Meridional, optical section, outline. 2; Aperture and ornamentation, 3; Polar view. 4-6: A. diastrophis, 4; Meridional, optical section, outline. 5; Aperture and ornamentation, 6; Polar view. 7-9: A. coridifolium, 7; Meridional, optical section, outline. 9; Syncolpate aperture, 9; Polar view. 10-11: A. munzurense, 10; Meridional, optical section, outline. 11; Polar view. 12-13: A. marashicum, 12; Meridional, optical section, outline. 13; Ornamentation. 14-15: A. huber-morathii, 14; Meridional, optical section, outline. 15; Polar view. 16-17: A turcica, 16; Meridional, optical section, outline. 17; Polar view. 18-20: A. dumanii, 18: Meridional, optical section, outline. 19; Ornamentation, 20; Polar view. 21-22: A. alanyae, 21; Meridional, optical section, outline. 22; Aperture and ornamentation (Scale bar: 10 μm)



FIGURE 4. SEM pollen microphotograph of *Aethionema* taxa. A-C: *A. marashicum* (type I), A; Polar view, B; equatorial view, C; ornamentation. D-F: *A. trinervium* (type II), D; Syncolpate and tricolpate pollen in polar view, E; equatorial view, F; ornamentation. G-I: *A. schistosum* (type III), G; Syncolpate pollen in polar view, H; equatorial view, I; ornamentation. J-L: *A. capitatum* (type IV), J; Syncolpate pollen in polar view, K; equatorial view, L; ornamentation. M-O: *A. carneum* (type V), M-N; equatorial view, O; ornamentation.

There has been no chromosomal study of the *Aethionema* species so far. Thus, future cytological studies in the genus may provide further information about heteromorphy in pollen grains. Inceoğlu and Karamustafa investigated 32 Brassicaceae species pollen grains and these were divided into 3 reticulate types, according to the diameters of the lumina (faintly reticulate, reticulate and coarsely reticulate), using LM [13].

Brochmann divided the genus Draba L. of the family Brassicaceae into 5 pollen types with respect to exine sculpturing resembling Aethionema in pollen morphology. In this study 5 pollen types are recognised according to pollen exine sculpturing (types I, II, III, IV and V) These pollen type described as verniculate reticulation with relatively wide muri and irregularly shaped lumina (Type I), reticulate pollen grain with wide muri and regularly shaped lumina (Type II), pollen grain have narrow muri and wide lumina (Type III), pollen grain of this type have narrow muri and polygonal lumina (Type IV), pollen grains have narrow muri and irregularly shaped lumina (Type V). Heteromorphic pollen grains are mostly observed in the species belonging to type IV [8]. Inceoğlu and Karamustafa described the general pollen morphological features of A. arabicum and A. armenum using LM. They described the sculpturing of these 2 species as faintly reticulate [13]. Karaismailoğlu, determined reticulate, microreticulate and coarsely reticulate exine sculpturing in eleven taxa of Aethionema [15]. Similarly Atceken et al. determined reticulate and microreticulate ornamentation in four taxa of Aethionema [14]. In addition, in a study which is about pollen morphology of 39 species belonging to 23 genera of the tribes Arabideae, Euclidieae, Hesperideae, Lunarieae, Matthioleae and Sisymbrieae from Brassicaceae from Egypt were studied by using scanning electron microscope (SEM), the pollen grains were tricolpate and their shape varies from prolate spheroidal, subprolate to prolate. Three pollen types can be distinguished based on the size of the lumina. The ornamentation varies between genera within the tribes and between species within the same genus [31].

This study results show that pollen shape, size, surface ornamentation, muri and lumina shape and size are important and useful characters contributing to the differentiation of the taxa.

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References

- I.C Hedge and J.M. Lamond . Aethionema R.Br. In: C.C. Townsend and E. Guest (ed.), Flora of Iraq 4 (1980) 915-922.
- [2] Ş. Yıldırımlı, Ö. Kılıç, New infrageneric taxa and species of Aethionema W.T.Aiton (Brassicaceae) and their current key from Turkey. Ot Sistematik Botanik Dergisi 23(1-2) (1935-2016) 1-66.
- [3] I.C. Hedge. *Aethionema* R.Br. In: Davis PH (ed.). Flora of Turkey and The East Aegean Islands. Edinburgh: Edinburgh University Press. (1965).
- [4] P.H. Davis, R.R Mill and K. Tan, Flora of Turkey and the East Aegean Islands 10. Edinburgh : Edinburgh University Press. (1988).
- [5] A. Güner, N. Özhatay, T. Ekim and K.H.C. Başer. Flora of Turkey and The East Aegean Islands vol 11. Edinburgh: Edinburgh University Press. (2000).
- [6] A. Güner, S. Aslan, T. Ekim, M. Vural and M.T. Babac Türkiye bitkileri listesi (damarlı bitkiler). A Checklist of the Flora of Turkey (Vascular Plants) Nezahat Gokyigit Botanik Bahcesive Flora Arastırmaları Dernegi Yayını, Istanbul. (2012).
- [7] K. Sorkun. Türkiye'nin Nektarlı Bitkileri, Polenleri ve Balları. Ankara. Palme Yayınları (2008) 462.
- [8] C. Brochmann, Pollen and seed morphology of Nordic Draba (Brasicaceae) phylogenetic and ecological implications, Nordic Journal of Botany 12/6 (1992) 657-673.
- [9] N.M. Pinar, A. Duran, T. Çeter and G.N. Tuğ, Pollen and Seed Morphology of the Genus Hesperis L. (Brassicaceae) in Turkey. Turkish Journal Botany (2009) 33(2) / 83-96.
- [10] T. Çeter, N.M. Pınar, Y. Bağcı ve L. Tutar, Türkiyede yayılış gösteren Rorippa Scopp. (Brassicaceae) türlerinin polen ve tohum morfolojisi. 20. Ulusal Elektron Mikroskopi Kongresi. Kemer/Antalya. Ekim 2011 25-28.
- [11] T. Çeter, N.M. Pınar, Y. Bağcı and A. Savran, -Türkiyede Yayılış Gösteren Barbarea (Brassicaceae) Türlerinin Polen Morfolojisi 21. Ulusal Biyoloji Kongresi, Ege Üniversitesi, İzmir. (3-7 Eylül 2012).
- [12] B. Mutlu and S. Erik, Pollen morphology and its taxonomic significance of the genus Arabis (Brassicaceae) in Turkey. Plant Systematics and Evolution. 298 (2012) 1931–1946.
- [13] Ö. İnceoğlu and F. Karamustafa. The pollen morphology of plants in Ankara region II. Cruciferae. Communacation. Faculty of Science University of Ankara Series. İstanbul University Press C2, 21 (1977) 111–118.

- [14] M.M. Atçeken, H. Dural, and B. Yilmaz Citak, The morphological, anatomical and palynological investigations on some taxa of genus *Aethionema* A.T. Waiton (Brassicaceae). Biology Diversity Conservation 9 (2016) 55–68.
- [15] M.C Karaismailoğlu, Palynological features of eleven Aethionema taxa from Turkey and their systematic implications. Bangladesh Journal of Plant Taxon 24/2 (2017) 197-204.
- [16] R.P. Wodehouse, *Pollen grains*. Newyork , Mc Graw-Hill Press.
- [17] K. Faegri and J. Iversen. Textbook of Pollen Analysis. Munksgaard, Copenhagen, 3rd ed. (1975) 295.
- [18] A. Van der Pluym and M. Hideux, Application d'une mèthodologie quantitative á la palynologie d'*Eryngium maritimum* (Umbelliferae). Plant Systematics and Evolution (1977) 127 / 55-85.
- [19] P.H. Davis, Flora of Turkey and the East Aegean Islands, Vol. 1Edinburgh, Edinburgh Univ. Press. (1965).
- [20] R.C. Rollins and U.C Banerjee. Pollen of the Cruciferae. Publ.Bussey Inst. Harvard Univ. (1979) 33–64.
- [21] C. Doğan and O. İnceoğlu, Pollen morphology of some *Isatis* L. taxa in Turkey. Turkish Journal of Botany 14 (1990) 12-31.
- [22] S. Mukherji, Pollen analysis in *Mangifera* in relation to fruit set and taxonomy. Journal of Indian Botanical Society 30 (1951) 49 -56.
- [23] P.K Nair and K.N Kaul, Pollen grain in a gigantic of *Rauwolfia serpentina*. Current Sciences. 34 (1965) 256-257.
- [24] B.D. Sharma, Pollen abnormalities in *Helicteres isora* L. Botanical Survey of India Calcutta (1967) 53-61.
- [25] Ö. İnceoğlu. Asyneuma canescens (W.K.) Griseb. & Schenk' in pollen morfolojisi ve heteromorf polenler. Türk Biyoloji Dergisi 23 (1973) 89-94.
- [26] E. Wagner, Cytological researches on meiosis and pollen development of female grape varieties. Chromosoma 4 (1951) 439-455.
- [27] E.J Olden, Giant pollen grains in fruit trees from colchicine treatment in vacuum. Hereditas 40 (1954) 526-529.
- [28] H. Laws, Pollen grains morphology of polyploid Oenotheras. Journal of Heredity 56/1 (1965) 18-21.
- [29] H. Matsuda, On the origin of big pollen grains with abnormal numbers of choromosomes. Cellules 38 (1928) 213-243.
- [30] B. Aytug, S. Aykut, N. Merev, G. Edis, Pollen Atlas of Istanbul and Its Surrounding Plants, Istanbul University Faculty of Forestry Publication Number: 1650 / 174 (1971) Kutulmuş Press, Istanbul, [Turkish].
- [31] K. Abdel Khalik, R. G. Van Den Berg, L J. G. Van Der Maesen, & M. N. El Hadidi, Pollen morphology of some tribes of Brassicaceae from Egypt and its

systematic implications. Feddes Repertorium: Zeitschrift für botanische Taxonomie und Geobotanik, 113/3-4 (2002) 211-223.

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