



Comparison of Antioxidant Activities of Mono-, Di- and Tri-substituted Coumarins

Hülya Çelik Onar^{1*}  , Hasniye Yaşa¹   and Oktay Sin²  

¹ Istanbul University-Cerrahpaşa Engineering Faculty Chemistry Department Organic Chemistry Division, Avcılar/İstanbul/Turkey.

² Istanbul University-Cerrahpaşa Institute of Graduate Studies Avcılar/İstanbul/Turkey.

Abstract: In this study, numerous coumarin compounds were synthesized by Pechmann and Knoevenagel methods, and the substitution of the formyl group was provided by the Duff reaction. The FTIR spectra and melting points of the synthesized compounds were compared with the literature values. The structures were also confirmed by GC/MS analysis. Besides, the synthesized coumarin derivatives were compared in terms of antioxidant activity according to DPPH and CUPRAC methods. The main aim of the study is to determine the effects of substituents on antioxidant activity.

Keywords: Coumarin, Pechmann reaction, Knoevenagel reaction, Duff reaction, DDPH method, CUPRAC method.

Submitted: September 25, 2019. **Accepted:** October 30, 2019.

Cite this: Çelik Onar H, Yaşa H, Sin O. Comparison of antioxidant activities of mono-, di- and tri-substituted coumarins. JOTCSA. 2020; 7(1): 87–96.

DOI: <https://doi.org/10.18596/jotcsa.624265>.

Corresponding Author. E-mail: hcelik@istanbul.edu.tr.

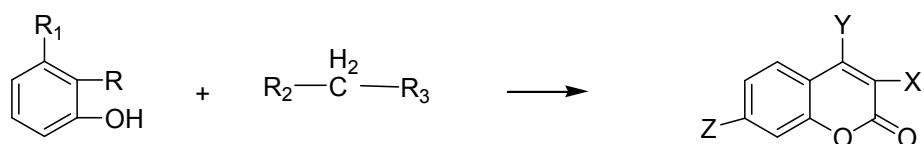
INTRODUCTION

Studies on heterocyclic compounds have intensified over the past decade because they have important medicinal properties, especially small-ring heterocyclic compounds. Coumarin derivatives are also one of these and some properties such as antibacterial, antifungal, and anticoagulant agents in pharmaceutical industry were studied (1). As well as being an optical whitening reagent, it is present in the structure of fluorescent and laser dyes (2), perfumes, soaps, cleansing products and food, because of their pleasant smell and sweetening effect (3).

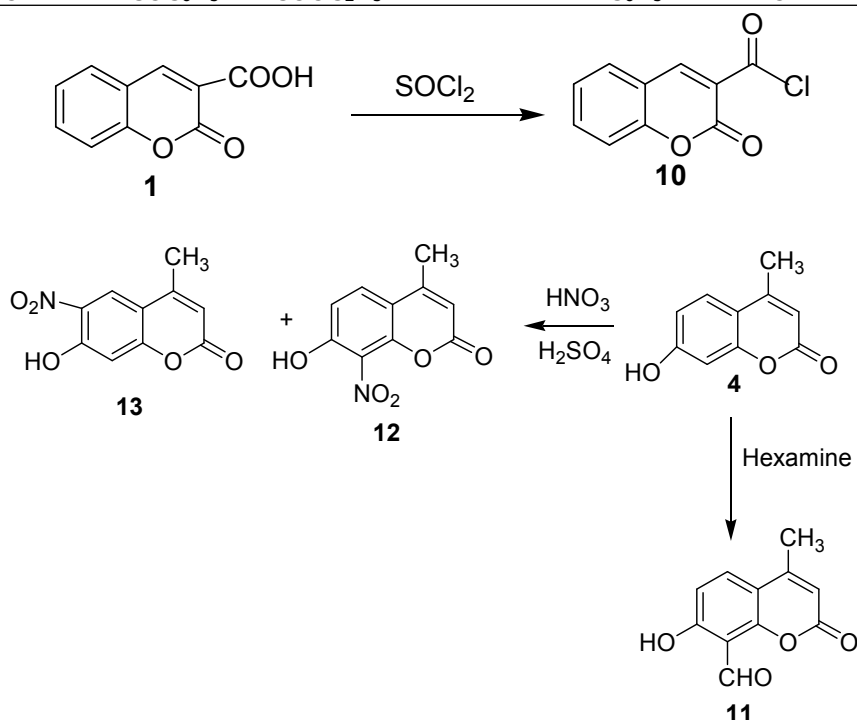
Coumarin is an oxygen-containing heterocyclic compound that is isolated from plants, especially

from green plants. Synthesis of coumarin is usually performed with one of the reactions known as Pechmann, Perkin, Knoevenagel, Reformatsky, and Wittig. Various catalysts such as H₂SO₄, H₃PO₄, CF₃COOH, p-toluenesulfonic acid, POCl₃, Bi(NO₃)₃·5H₂O, AlCl₃, TiCl₄, ZnI₂ and ionic liquids were used in these reactions.

Coumarin is a compound suitable for substitution at six points in the structure. The attachment of different substituents at different points adds different properties to the coumarins. Therefore, various mono-, di-, and tri-substituted coumarins have been synthesized (Scheme 1) and compared with standards for antioxidant activity according to DPPH and CUPRAC methods.



R	R ₁	R ₂	R ₃	X	Y	Z	Compound
CHO	H	COOH	COOH	COOH	H	H	1
CHO	H	COCH ₃	COOC ₂ H ₅	COCH ₃	H	H	2
CHO	H	COC ₆ H ₅	COOC ₂ H ₅	COC ₆ H ₅	H	H	3
H	OH	COCH ₃	COOC ₂ H ₅	H	CH ₃	OH	4
H	NH ₂	COCH ₃	COOC ₂ H ₅	H	CH ₃	NH ₂	5
H	OCH ₃	COCH ₃	COOC ₂ H ₅	H	CH ₃	OCH ₃	6
H	NH ₂	COC ₆ H ₅	COOC ₂ H ₅	H	C ₆ H ₅	NH ₂	7
H	OH	COC ₃ H ₇	COOC ₂ H ₅	H	C ₃ H ₇	OH	8
H	OH	COC ₆ H ₅	COOC ₂ H ₅	H	C ₆ H ₅	OH	9



Scheme 1. Synthesized compounds.

EXPERIMENTAL SECTION

Instruments

Recovery of the solvents during the recovery of the products and during the crystallization processes was carried out in a BÜCHI Rotavapor R-200 rotary evaporator. In thin layer chromatography (TLC), a Silicagel 60F₂₅₄ layer (Merck) was used. The plates were illuminated with a UV lamp to make the stains on the film sheets visible. Melting points of the coumarin compounds were examined in a Buchi Melting Point B-540 melting point apparatus. Fourier Transform Infrared (FT-IR) spectra of the synthesized coumarin compounds were taken on a Mattson 1000 Series FT-IR spectrometer. Mass spectra were checked with a Shimadzu GC-MS

2010 device.

Coumarin-3-carboxylic acid synthesis (1)

The synthesis was carried out by Knoevenagel reaction. 1.05 mL (10 mmol) of salicylaldehyde and 1.56 g (15 mmol) of malonic acid were refluxed with 1 mL of KSF catalyst with 3.3 mL of H₂O at 110-120 °C for 24 hours. The reaction was checked by TLC and when a new product was formed, the reaction was terminated with TLC control about the termination of the starting material. It was cooled down to room temperature. The remaining solid was triturated with 60 mL of methanol or ethyl acetate (4).

Yield(%): 95 m.p. 192 °C. Analysis of the GC / MS revealed MS peaks at m/z 190, 173, 146,

118, 89, 77, 63. $M=(190)$, $M-OH = M^{+1}(173)$, $M^{+1}-CO = M^{+2}(145)$, $M^{+2}-CO = M^{+3}(117)$. FTIR spectrum shows a 3174 cm^{-1} (-OH) stretching, 3057 cm^{-1} (Ar, C-H) aryl stretching, 1737 cm^{-1} (C=O) lactone carbonyl stretching, 1672 cm^{-1} (C=O) acid carbonyl stretching, 1607 cm^{-1} , 1566 cm^{-1} (C=C) double band stretching, and 1226 cm^{-1} (C-O) stretching (4,5).

Synthesis of 3-acetylcoumarin (2)

It was synthesized by Knoevenagel reaction with triethylamine as catalyst. 2.65 mL (0.025 mol) of salicylaldehyde and 3.175 mL (0.025 mol) of ethyl acetoacetate were refluxed in 15 mL of ethanol and 5 mL of triethylamine as catalyst at $80\text{ }^{\circ}\text{C}$ for 12 hours. The reaction was continued until TLC showed that a new product was formed. When the starting material completely finished, the reaction was terminated and cooled to room temperature, then filtered through a Büchner funnel, then the remaining solid was crystallized from ethanol (6).

Yield(%): 60 m.p. $120\text{ }^{\circ}\text{C}$. Analysis of the GC / MS revealed MS peaks at 188, 173, 145, 118, 101, 89, 63. $M=(188)$, $M-CH_3 = M^{+1}(173)$, $M^{+1}-CO = M^{+2}(145)$, $M^{+2}-CO = M^{+3}(117)$. FTIR spectrum shows a 3031 cm^{-1} (Ar, C-H) aryl stretching, 2983 cm^{-1} (C-H) aliphatic carbon - hydrogen stretching, 1740 cm^{-1} (C=O) lactone carbonyl stretching, 1675 cm^{-1} (C=O) ketone carbonyl stretching, 1612 cm^{-1} (C=C) double band stretching, 1265 cm^{-1} (C-O) stretching (6,7).

Synthesis of 3-benzoylcoumarin (3)

It was synthesized by Knoevenagel reaction with pyridine as catalyst. 1.05 mL (0.01 mol) of salicylaldehyde and 3.47 mL (0.020 mol) of ethylbenzoylacetate were refluxed with 10 mL of ethanol with 0.08 mL of pyridine as catalyst at $80\text{ }^{\circ}\text{C}$ for 12 hours. The reaction was checked by TLC and when a new product was formed and the starting material completely finished, the reaction was terminated. It was cooled to room temperature and filtered through a Büchner funnel. The remaining solid was crystallized from ethanol (8).

Yield(%): 60 m.p. $138\text{ }^{\circ}\text{C}$. Analysis of the GC / MS revealed MS peaks at 250, 222, 194, 173, 105, 77, 51. $M=(250)$, $M-CO = M^{+1}(222)$, $M^{+1}-CO = M^{+2}(194)$. IR spectrum shows a 3046 cm^{-1} (Ar, C-H) aryl stretching, 1716 cm^{-1} (C=O) lactone carbonyl stretching, 1658 cm^{-1} (C=O) ketone carbonyl stretching, 1608 cm^{-1} (C=C) double band stretching, 1264 cm^{-1} (C-O) stretching (8).

Synthesis of 4-methyl-7-hydroxycoumarin (4)

It was synthesized by Pechmann reaction in the presence of a catalytic amount of oxalic acid. 1.1 g (10 mmol) of resorcinol (1,3-

dihydroxybenzene) and 2.54 mL (20 mmol) ethylacetoacetate were reacted at 0.09 g (1 mmol) of oxalic acid catalyst and at $80\text{ }^{\circ}\text{C}$ for 12 hours with some molecular sieve. The reaction was checked by TLC and when a new product was formed and the starting materials completely finished, the reaction was stopped and allowed to cool. As the reaction cooled, ice was added and then filtered on a Büchner funnel. The remaining solid was crystallized from ethanol (9).

Yield(%): 60. m.p. $118\text{ }^{\circ}\text{C}$. Analysis of the GC / MS revealed MS peaks at 176, 148, 133, 116, 105, 91, 74, 65. $M=(176)$, $M-CO = M^{+1}(148)$, $M^{+1}-CH_3 = M^{+2}(133)$, $M^{+2}-OH = M^{+3}(116)$. FTIR spectrum shows a 3225 cm^{-1} (-OH) stretching, 3077 cm^{-1} (Ar, C-H) aryl stretching, 1673 cm^{-1} (C=O) lactone carbonyl stretching, 1586 cm^{-1} (C=C) double band stretching, 1272 cm^{-1} (C-O) stretching (9,10).

Synthesis of 4-methyl-7-amino-, 4-methyl-7-methoxy-, 4-phenyl-7-aminocoumarin

They were synthesized by Pechmann reaction with oxalic acid as catalyst. (10 mmol) of 3-substituted phenol and (12 mmol) of ethyl acetoacetate or ethylbenzoyl acetate were reacted at 0.09 g (1 mmol) of oxalic acid catalyst and a quantity of molecular sieve at $80\text{ }^{\circ}\text{C}$ for 12 hours. The reaction was checked by TLC and when a new product was formed and the starting materials completely finished, the reaction was turned off and allowed to cool. As the reaction cooled, ice was added and then filtered through a spinner funnel. The remaining solid was crystallized from a mixture of ethanol-water (9:1) (11-13).

4-Methyl-7-aminocoumarin (5): Yield(%): 80 m.p. $223,8\text{ }^{\circ}\text{C}$. Analysis of the GC/MS revealed MS peaks at 175, 147, 132, 116, 91, 73, 65, 44. $M=(175)$, $M-CO = M^{+1}(147)$, $M^{+1}-CH_3 = M^{+2}(132)$, $M^{+2}-NH_2 = M^{+3}(116)$. FTIR spectrum shows a 3249 cm^{-1} (-NH₂) stretching, 1661 cm^{-1} (C=O) lactone carbonyl stretching, 1603 cm^{-1} (C=C) double band stretching, and 1294 cm^{-1} (C-O) stretching (9,14).

4-Methyl-7-methoxycoumarin (6): Yield(%): 80 m.p. $159,2\text{ }^{\circ}\text{C}$. Analysis of the GC / MS revealed MS peaks at 190, 162, 147, 116, 91, 77, 65. $M=(190)$, $M-CO = M^{+1}(162)$, $M^{+1}-CH_3 = M^{+2}(147)$, $M^{+2}-OCH_3 = M^{+3}(116)$. FTIR spectrum shows a 3028 cm^{-1} (Ar, C-H) aryl stretching, 1711 cm^{-1} (C=O) lactone carbonyl stretching, 1605 cm^{-1} (C=C) double band stretching, 1264 cm^{-1} (C-O) stretching (12,15).

4-Phenyl-7-aminocoumarin (7): Yield(%): 75 m.p. $222,6\text{ }^{\circ}\text{C}$. Analysis of the GC / MS revealed MS peaks at 237, 209, 193, 116. $M=(237)$, $M-CO = M^{+1}(209)$, $M^{+1}-NH_2 = M^{+2}(193)$, $M^{+2}-C_6H_5 =$

M⁺³ (116). FTIR spectrum shows a 3245 cm⁻¹(N-H) stretching, 1661 cm⁻¹ (C=O) lactone carbonyl stretching, 1598 cm⁻¹ (C=C) double band stretching, 1293 cm⁻¹(C-O) stretching (13).

Synthesis of 4-propyl-7-hydroxycoumarin (8)

It was synthesized by Pechmann reaction in the presence of oxalic acid as catalyst. 1.1 g (10 mmol) of resorcinol (1,3-dihydroxybenzene) and 1.6 mL (10 mmol) of ethylbutyryl acetoacetate were added to the reaction mixture at 0.08 g (1 mmol) of oxalic acid and a quantity of molecular sieve were introduced. The reaction was checked by TLC and when a new product was formed and the starting materials completely finished, the reaction was turned off and allowed to cool. As the reaction cooled, ice was added and then filtered on a Büchner funnel. The remaining solid was crystallized in ethanol. Column chromatography was performed for purifying using chloroform: methanol (5: 1) as the eluent phase (16).

Yield(%): 65 m.p.133 °C. Analysis of the GC/MS revealed MS peaks at 204, 189, 176, 161, 148, 133, 105, 91, 77, 44. M=(204), M-CH₃ = M⁺¹ (189) M-CO = M⁺¹(176), M⁺¹- C₃H₇ = M⁺² (133). FTIR spectrum shows a 3253 cm⁻¹ (O-H) stretching, 1691 cm⁻¹ (C=O) lactone carbonyl stretching, 1614 cm⁻¹ (C=C) double band stretching, and 1310 cm⁻¹(C-O) stretching (16,17).

Synthesis of 4-phenyl-7-hydroxycoumarin (9)

It was synthesized by Pechmann reaction with oxalic acid as catalyst. 2.0 g (12 mmol) of ethylbenzoyl acetate were reacted with 1.1 g (10 mmol) of resorcinol (1,3-dihydroxybenzene) and 0.314 g (1 mmol) of amberlyte 15 catalyst and a quantity of molecular sieve at 80 °C for 12 hours. The reaction was checked by TLC and in case a new product was formed, the starting materials were waited to finish completely, the reaction was turned off and allowed to cool. As the reaction cooled, ice was added and then filtered on a Büchner funnel. The remaining solid was crystallized from ethanol (18).

Yield(%): 65 m.p. 249 °C. Analysis of the GC / MS revealed MS peaks at 238, 210, 181, 152, 139, 127, 76, 63, 51. M=(238), M-CO = M⁺¹(210), C₃H₇⁺ = (76). FTIR spectrum shows a 3115 cm⁻¹(O-H) stretching, 1684 cm⁻¹ (C=O) lactone carbonyl stretching, 1592 cm⁻¹ (C=C) double band stretching, 1268 cm⁻¹(C-O) stretching (18).

Synthesis of coumarin-3-carboxylic acid chloride (10)

To 1.9 g (0.1 mol) of coumarin-3-carboxylic acid,

10 mL of thionyl chloride (SOCl₂) was added to a round bottom flask, a few drops of pyridine was introduced, and the reboiler was fitted with a 5% NaOH solution in a hose over the cooler. It was refluxed at 80°C for three hours and cooled to room temperature. The white precipitate was washed several times with dry n-hexane (19).

Yield(%): 95 m.p. 145 °C. Analysis of the GC / MS revealed MS peaks at 208, 173, 145, 101, 89, 63. M=(208), M-Cl = M⁺¹(173), M⁺¹-CO = M⁺² (145). FTIR spectrum shows a 3038 cm⁻¹(Ar, C-H) aryl stretching, 1747cm⁻¹ (C=O) lactone carbonyl stretching, 1693 cm⁻¹ (C=O) acyl carbonyl stretching, 1605 cm⁻¹ (C=C) double band stretching, 1179 cm⁻¹(C-O) stretching and 759 cm⁻¹ (C-Cl) stretching (19,20).

Synthesis of 4-methyl-7-hydroxy-8-formylcoumarin (11)

This compound was synthesized by Duff reaction. 6.5 g of 4-methyl-7-hydroxycoumarin, 12 g of hexamethylene tetramine (hexamine), 61.5 mL of glacial acetic acid were added and the mixture was refluxed at 85-90 °C for 12 hours. 92.18 mL of 20% HCl solution (49.83 mL of HCl, 42.33 mL of distilled water) was added and refluxed for a further 4 hours. The reaction was checked by TLC and when a new product was formed, it was cooled to room temperature and extracted with diethyl ether. The resulting extract was dried in sodium sulfate (Na₂SO₄) for 15 min. The solvent was removed on the evaporator to give a pale yellow solid. The residue was purified by crystallization from a solvent mixture of ethanol-1,4-dioxane (9:1) (21).

Yield(%): 65 m.p. 176 °C. Analysis of the GC/MS revealed MS peaks at 204, 176, 148, 119, 91, 77, 65, 44 M=(204), M-CO = M⁺¹(176), M⁺¹ -CO = M⁺²(148). FTIR spectrum shows a 3125 cm⁻¹(O-H) stretching, 1742 cm⁻¹ (C=O) lactone carbonyl stretching, 1644 cm⁻¹ (C=O) aldehyde carbonyl stretching, 1518 cm⁻¹ (C=C) double band stretching, 1268 cm⁻¹(C-O) stretching (21-23).

Syntheses of 4-methyl-7-hydroxy-8-nitrocoumarin and 4-methyl-7-hydroxy-6-nitrocoumarin

1.2 g of 4-methyl-7-hydroxycoumarin was dissolved in 10 mL of H₂SO₄ at 0-5 °C. 1.5 mL of H₂SO₄ and 0.5 mL of HNO₃ were added dropwise to the reaction flask. It was stirred at 0-5 °C for four hours and then filtered on a Büchner funnel. When crystallized from ethanol, the solid residue on the filter paper was separated as 4-methyl-7-hydroxy-6-nitrocoumarin. The 4-methyl-7-hydroxy-8-nitrocoumarin which remains in the filtrate is cooled in the refrigerator and crystallized, then filtered through Büchner funnel (24).

4-Methyl-7-hydroxy-8-nitrocoumarin (12):

Yield(%): 40 m.p. 254,6 °C. Analysis of the GC/MS revealed MS peaks at 221, 193, 176, 147, 135, 119, 91, 77, 65, 51, 44. M=(221), M-CO = M⁺¹(193), M⁺¹- OH = M⁺²(176). FTIR spectrum shows a 3265 cm⁻¹(O-H) stretching, 1703 cm⁻¹ (C=O) lactone carbonyl stretching, 1569 cm⁻¹ (C=C) double band stretching, and 1314 cm⁻¹(C-O) stretching (24,25).

4-Methyl-7-hydroxy-6-nitrocoumarin (13):

Yield(%): 40 m.p. 261 °C. Analysis of the GC/MS revealed MS peaks at 221, 193, 147, 135, 119, 91, 77, 65, 51, 44. M=(221), M-CO = M⁺¹(193), M⁺¹ - NO₂ = M⁺²(147). FTIR spectrum shows a 3244 cm⁻¹(O-H) stretching, 1727 cm⁻¹ (C=O) lactone carbonyl stretching, 1615 cm⁻¹ (C=C) double band stretching, 1290 cm⁻¹(C-O) stretching (24,25).

DPPH assay

Concentrations of standards and coumarin compounds were studied at 250, 500, and 1000 µg/mL. In the experiment, 1 mL of sample solution was mixed with 1 mL of DPPH solution in 0.002% concentration prepared in methanol. After standing for 30 minutes in the dark, absorbance was measured at 517 nm. The spectrophotometer was made zero with methanolic solution. The mixture prepared from 1 mL of methanol with 1 mL of DPPH solution was measured as control sample (26).

The DPPH solution was prepared daily, fresh and kept in the dark and at 4 °C, and covered with aluminum foil around the flask during use. DPPH inhibition is calculated by the following formula:

$$IC(\%) = \left[\frac{(A_0 - A_t)}{A_0} \right] \times 100$$

A₀: Absorbance of the control sample

A_t: The absorbance value of the sample being tested

CUPRAC assay

1 mL of a solution of 0.01.10⁻² M copper(II) chloride, 1 mL of a solution of 7.5.10⁻³ M neocuproine, 1 mL of a 0.1 M ammonium acetate buffer (pH= 7.0) and 1 mL of a sample solution (250, 500, 1000 µg/mL) were placed in the tube and mixed. The mixture was allowed to stand at room temperature for 30 minutes, in the dark and absorbance was read at 450 nm. The reading was performed by zeroing against the blank, which was prepared by placing 1 mL of methanol instead of 1 mL of sample (27).

RESULTS AND DISCUSSION

The physical and spectroscopic values of the synthesized coumarin derivatives are in accordance with the literature values. The DPPH activity results of the coumarin derivatives are shown in Table 1, and the CUPRAC activity results are shown in Table 2.

In DPPH antioxidant capacity determination, as shown in Table 1, DPPH activity was observed to increase more as the concentration of only coumarin-3-carboxylic acid was decreased. DPPH activities decreased as concentration decreased in other substances. Especially some of the coumarin compounds which are coumarin-3-carboxylic acid chloride, 4-methyl-7-aminocoumarin and 4-phenyl-7-aminocoumarin, showed values close to DPPH activity at concentrations of 1000 µg /mL of α- tocopherol, NDGA, BHT, BHA and ascorbic acid as standard antioxidants.

In the determination of CUPRAC antioxidant capacity, as shown in Table 2, as the concentration decreased, CUPRAC reduction activities decreased as well. The compounds 4-Methyl-7-hydroxy-8-formylcoumarin and 4-methyl-7-hydroxy-8-nitrocoumarin showed values close to the CUPRAC reduction activity exhibited by BHA and ascorbic acid. CUPRAC reduction activity of only four of the synthesized compounds was observed. In other compounds, values are too small to be considered, so they are not added to the table.

In particular, 3-carboxylic acid chloride, 4-methyl-7-amino coumarin, 4-phenyl-7-aminocoumarin, 4-methyl-7-hydroxy-8-formyl, 4-methyl-7-hydroxy-8-nitro coumarin compounds have been found to be more effective in terms of antioxidant activity. These coumarin compounds can be evaluated with antioxidant activity properties.

ACKNOWLEDGMENTS

Part of this work is from the master's thesis of Oktay Sin. It was supported by the Research Fund of the İstanbul University-Cerrahpaşa. Project number: FYL-2017-23395.

REFERENCES

1. Khan KM, Saify ZS, Hayat S, Khan MZ, Noor F, Makhmoor T, Choudhary MI, Ziaullah MI, Perveen S. Synthesis, Antioxidant and Insecticidal Activities of Some Coumarin Derivatives. J. Chem. Soc. Pak. 2002; 24 (3): 226-30.

2. Yao C, Zeng H. Synthesis and Spectroscopic Study of Coumarin Derivatives, UV-Vis Absorption Spectroscopy and Fluorescence Spectroscopy, 2011; 27 (4): 599-03.
3. O'Kennedy R, Thornes RD, Coumarins: Biology, Applications and Mode of Action, John Wiley and Sons, Chichester, ISBN: 978-0-471-96997-6, p. 348-60 (1997).
4. Bigi F, Chesini L, Maggi R, Sartori G, Montmorillonite KSF as an Inorganic Water Stable, and Reusable Catalyst for the Knoevenagel Synthesis of Coumarin-3-carboxylic Acids, *J. Org. Chem.* 1999; 64 (3): 1033-5.
5. Moussaoui Y, Ben Salem R, Catalyzed Knoevenagel reactions on inorganic solid supports: Application to the synthesis of coumarin compounds. *C.R. Chim.* 2007; 10 (12): 1162-9.
6. Bolakatti GS, Maddi VS, Mamledesai SN, Ronad PM, Palkar MB, Swamy S. Synthesis and evaluation of antiinflammatory and analgesic activities of a novel series of coumarin Mannich bases. *Arzneim. Forsch.* 2008; 58 (10): 515-20.
7. Tanaka T, Yamashita K, Hayashi M. Titanium Tetraisopropoxide Promoted Reactions for the Synthesis of Substituted Coumarins. *Heterocycles.* 2010; 80 (1): 631-6.
8. Yuan HY, Wang M, Liu YJ, Liu Q. Copper(II)-Catalyzed C-C Bond-Forming Reactions of α -Electron-Withdrawing Group-Substituted Ketene S,S-Acetals with Carbonyl Compounds and a Facile Synthesis of Coumarins. *Adv. Synth. Catal.* 2009; 351 (1-2): 112-6.
9. Kokare ND, Sangshetti JN, Shinde DB. Oxalic acid catalyzed solvent-free one pot synthesis of coumarins. *Chinese Chem. Lett.* 2007; 18 (11): 1309-12.
10. Montazeri N, Khaksar S, Nazari A, Alavi SS, Vahdat SM, Tajbakhsh M. Pentafluorophenylammonium triflate (PFPAT): An efficient, metal-free and Reusable catalyst for the von Pechmann reaction. *J. Fluor. Chem.* 2011; 132 (7): 450-2.
11. Pozdnev VF. Improved method for synthesis of 7-amino-4-methylcoumarin, *Chem. Heterocycl. Compd.* 1990; 26 (3): 264-5.
12. Murty KS, Rao PS, Seshadri TR. Geometrical Inversion in the Acids derived from The Coumarins Part 6 The Behaviour of the Acids derive from 4-Methyl-Coumarin, *Proceedings-Indian Academy of Sciences.* 1937; 6A: 316-27.
13. Reszka P, Schulz R, Methling K, Lalk M, Bednarski PJ. Synthesis, Enzymatic Evaluation, and Docking Studies of Fluorogenic Caspase and Tetrapeptide Substrates. *Chem. Med. Chem.* 2010; 5 (1): 103-17.
14. Anand B, Roy N, Sai SSS, Philip R. Spectral dispersion of ultrafast optical limiting in Coumarin-120 by white-light continuum Z-scan. *Appl. Phys. Lett.* 2013; 102 (20): 23302.
15. Jung ME, Allen DA. Use of 4-Cyanocoumarins as Dienophiles in a Facile Synthesis of Highly Substituted Dibenzopyranones. *Org. Lett.* 2009; 11 (3): 757-60.
- Mentzer C, Gley P, Molho D, Billet D. Estrogenic substances of the coumarin series II. *Bull. Soc. Chim. Fr.* 1946; 271-6.
17. Timonen J, Aulaskari P, Hirva P, Vainiotalo P. Coumarin M. Spectrometry. *Eur. J. Mass Spectrom.* 2009; 15 (5): 595-603.
18. Timonen JM, Nieminen RM, Sareila O, Goulas A, Moilanen LJ, Haukka M. Synthesis and anti-inflammatory effects of a series of novel 7-hydroxycoumarin Derivatives. *Eur. J. Med. Chem.* 2011; 46 (9): 3845-50.
19. Adreani LL, Lapi E. One some new esters of coumarin-3-carboxylic acid with balsamic and bronchodilator action. *Boll. Chim. Farm.* 1960; 99: 583-6.
20. Polyansky DE, Necker DC. Photodecomposition of Organic Peroxides Containing Coumarin Chromophore: Spectroscopic Studies. *J. Phys. Chem.* 2005; 109/12: 2793-800.
21. Dong Y, Mao X, Jiang X, Hau J, Cheng Y, Zhu C. Electronic Supplementary Information. *Roy. Soc. Chem. China.* 2011; 1-10.
22. Kulkarni A, Patil SA, Badami PS. DNA cleavage and in vitro antimicrobial studies of Co(II), Ni(II), and Cu(II) complexes with ONNO donor Schiff bases: Synthesis, spectral characterization and electrochemical studies. *J. Enzym. Inhib. Med. Chem.* 2010; 25 (1): 87-96.
23. Huang X, Dong Y, Huang Q, Cheng Y. Hydrogen bond induced fluorescence recovery of coumarin-based sensor system. *Tetrahedron Lett.* 2013; 54 (29): 3822-5.

24. Hussien FAH, Keshe M, Alzobar K, Merza J, Karam A. Synthesis and Nitration of 7-Hydroxy-4-Methyl Coumarin via Pechmann Condensation Using Eco-Friendly Medias. *Int. Lett. Chem. Phys. Astron.* 2016; 69: 66-73.

25. Kanodia S, Thapliyal PC. Regioselective mononitration of coumarins using claycop reagent. *J.Indian Chem. Soc.* 2011; 88 (2): 241-4.

26. Al-Majedy YK, Al-Duhaidahawi DL, Al-Azawi KF, Kadhum AAH, Mohamad AB. Coumarins as

Potential Antioxidant Agents Complemented with suggested Mechanisms and Approved by Molecular Modeling Studies. *Molecules.* 2016; 21 (2): 135-9.

27. Apak R, Güçlü K, Özyürek M, Karademir SE. A novel total antioxidant capacity index for dietary polyphenols, vitamin C and E, using their cupric ion reducing capability in the presence of neocuproine, CUPRAC method. *J. Agric. Food Chem.* 2004; 52: 7970-81.

Table 1. The DPPH activity results of the coumarin derivatives

Entry	Sample	%DPPH (1000 µg/mL)	%DPPH (500 µg/mL)	%DPPH (250 µg/mL)
1	3-Benzoylcoumarin	5.6	0.4	0
2	Coumarin-3-carboxylic acid	3.2	4.4	5.2
3	3-Acetylcoumarin	6	2.8	0
4	Coumarin-3-carboxylic acid chloride	84	64.4	42.8
5	4-Methyl-7-hydroxycoumarin	18.4	11.2	10.4
6	4-Propyl-7-hydroxycoumarin	13.6	6.4	5.2
7	4-Phenyl-7-hydroxycoumarin	22	11.6	6.4
8	4-Methyl-7-methoxycoumarin	4.4	2	1.6
9	4-Methyl-7-hydroxy-8-formylcoumarin	14.4	11.6	7.6
10	4-Methyl-7-hydroxy-8-nitrocoumarin	28	13.6	3.6
11	4-Methyl-7-aminocoumarin	82	78	70.8
12	4-Phenyl-7-aminocoumarin	82.8	75.2	69.6
13	α-Tocopherol	96.4	93.6	93.2
14	NDGA*	95.6	93.2	92
15	BHT*	95.6	95.2	94.8
16	BHA*	96.8	94	93.6
17	Ascorbic acid	97.2	97.2	96.8

*BHA (Butylated Hydroxy Anisole) BHT (Butylated Hydroxy Toluene) NDGA (Nordihydroguaiaretic Acid)

Table 2. The CUPRAC activity results of the coumarin derivatives

Entry	Sample	%Absorbance (1000 µg/mL)	%Absorbance (500 µg/mL)	%Absorbance (250 µg/mL)
9	4-Methyl-7-hydroxy-8-formylcoumarin	0.664 ± 0.0202	0.199 ± 0.0263	0.022 ± 0.0021
10	4-Methyl-7-hydroxy-8-nitrocoumarin	0.457 ± 0.0025	0.226 ± 0.0036	0.116 ± 0.001
11	4-Methyl-7-aminocoumarin	0.209 ± 0.0035	0.136 ± 0.0062	0.095 ± 0.0043
12	4-Phenyl-7-aminocoumarin	0.196 ± 0.0020	0.142 ± 0.0037	0.098 ± 0.0049
16	BHA*	0.734 ± 0.0117	0.489 ± 0.001	0.456 ± 0.0035
17	Ascorbic acid	0.476 ± 0.0041	0.443 ± 0.001	0.432 ± 0.0007

*BHA (Butylated Hydroxy Anisole) Values are means ± SD.

