Research article Eurasian J Bio Chem Sci, 2(2): 52-55, 2019



Eurasian Journal of Biological and Chemical Sciences

EJBCS Tornan Brunna Egner (Brustan Brunna Egner (Brustan I Bie Charr, 54)

Journal homepage: www.dergipark.org.tr/ejbcs

Glucose effect on biofilm formations of S. aureus strains

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Abstract: Biofilm formations of 25 *S.aureus* strains were investigated on microtitration plates using TSB and TSB media supplemented with glucose (0.25%, 1.0%, or 2.0%, w/v) for 48 hours. The biofilm formations of *S.aureus* isolates incubated in TSB were generally found to be moderate (84%). Majority of strong biofilm formations (84-96%) were determined by *S.aureus* strains incubated in the presence of glucose. It was shown that the presence of glucose had positive effect on *S.aureus* biofilm formations.

Keywords: S.aureus, biofilm formation, D-glucose, polystyrene, microtitration plates.

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1. Introduction

Staphylococci are the most important pathogens causing major problems in food industry (Tauxe, 2002). The predominant *Staphylococcus* species are *S. epidermidis, S. haemolyticus* and *S. aureus* (Otto, 2013; Becker et al., 2014; Méric et al., 2015). Especially *S.aureus* is known as the third common cause of foodborne illness worldwide (Aydin et al., 2011). *S.aureus* can be easily isolated from surfaces in the food and dairy industries (Ateba et al., 2010; Santos et al., 2014). The products can also be contaminated during food processing by *S.aureus* enterotoxins that cause foodborne illnesses (Cha et al., 2006; Sudagidan and Aydin, 2009). Moreover, *S.aureus* strains create biofilms on different types of surfaces such as stainless steel, plastic, glass, and rubber (Akbas and Kokumer, 2015; Lee et al., 2014, 2016, 2017; Unlu et al., 2018).

In this study, the effects of glucose on biofilm formations by *S. aureus* strains isolated from raw milk samples were investigated.

2. Materials and Method

2.1. Bacterial strains

In this work, 25 *S.aureus* isolates (S1-S25) were evaluated for their biofilm formation abilities in TSB (Tryptic soy broth) or TSB_{Glc} (TSB enriched with D-glucose) media. *S. aureus* strains ATCC 25923 and ATCC 29213 were used as biofilm positive and biofilm negative strains, respectively.

2.2. Biofilm Formation

For determining the biofilm formations, TSB and TSB_{Glc} (TSB enriched with D-glucose at different concentrations, 0.25% TSB_{Glc0.25}; 1.0% TSB_{Glc1} and 2.0% TSB_{Glc2}; w/v) media were used. The isolates and reference strains were incubated overnight at 37°C in TSB. Following the incubation, the cultures were diluted at a rate of 1:100 in fresh TSB or TSB_{Glc} media. The cultures were transferred into flat-bottomed 96-well polystyrene microtitration plates (150µL; about 10⁵cfu/mL, final concentration) and incubated at 37°C for 48 h. At the end of the incubation, bacterial cells were removed and then the wells were stained with crystal violet solution (0.5% w/v, Sigma, St. Louis, MO, USA) for 20 min. Then, the wells were decanted and treated with glacial acetic acid (33% v/v, Merck) for 10 min. All samples were transferred into a new microtitration plate wells and measured at 570 nm using an ELISA plate reader (Christensen et al., 1985). The absorbance values of TSB or TSB supplemented with D-glucose (OD_c) were used as a blank. Each experiment was performed in triplicate for all isolates.

Biofilm formations were determined according to Stepanović et al. (2000).

- Nonbiofilm producer (-, $OD \le OD_c$),
- Weak biofilm producer (+, $OD_c < OD \le 2xOD_c$),
- Moderate biofilm producer (++, $2xOD_c < OD \le 4xODc$), and
- Strong biofilm producer (+++, $4xOD_c < OD$)

3. Results

In this study, 25 *S. aureus* isolates (S1-S25) and 2 reference strains (ATCC 25923 and ATCC 29213) were used for determination of biofilm formations. The negative reference *S. aureus* ATCC 29213 and the positive reference strain *S. aureus* ATCC 25923 were shown as non and strong biofilm producers in the all media, respectively (Table 1). In addition, S3 and S4 were considered as strong biofilm producers (Table 1).

In general, 21 of 25 *S. aureus* isolates (84%) were considered as moderate biofilm producers when they were

incubated in TSB medium (Table 2). The biofilm formation capacities of strains gradually increased by increasing concentrations of glucose in TSB media supplemented with glucose. 24 of 25 *S. aureus* isolates (96%) cultured in TSB_{Glc2} media were determined as strong biofilm producers (Table 2).

According to obtained results, it was shown that the content of media was important parameter for biofilm productions. For this reason, the effects of media components should be investigated in more detail for biofilm formation studies.

Table 1. Biofilm formations (OD_{570nm}) of *S. aureus* strains on TSB and TSB_{Glc} (TSB_{Glc0.25}, TSB_{Glc1}, and TSB_{Glc2}) media on microtitration plates after 48 hours. OD, optical density; standard deviations are in parentheses.

Strain	Media				
	TSB	TSBGlc0.25	TSB _{Glc1}	TSBGlc2	
S1	0.42 (0.00)	0.88 (0.01)	1.25 (0.02)	1.09 (0.04)	
S2	0.46 (0.02)	0.52 (0.15)	0.77 (0.21)	0.69 (0.00)	
S 3	0.99 (0.20)	1.51 (0.04)	1.49 (0.05)	1.52 (0.04)	
S4	0.57 (0.07)	0.76 (0.09)	1.32 (0.01)	1.16 (0.17)	
S 5	0.43 (0.06)	0.98 (0.07)	1.36 (0.00)	1.31 (0.02)	
S6	0.39 (0.11)	0.87 (0.12)	0.87 (0.12)	0.97 (0.01)	
S7	0.64 (0.17)	0.70 (0.00)	0.77 (0.23)	0.91 (0.10)	
S8	0.31 (0.04)	0.80 (0.04)	0.75 (0.03)	0.82 (0.23)	
S9	0.57 (0.03)	0.94 (0.02)	1.26 (0.01)	1.23 (0.15)	
S10	0.56 (0.11)	1.03 (0.09)	1.08 (0.04)	1.31 (0.01)	
S11	0.55 (0.04)	1.03 (0.10)	0.99 (0.07)	1.16 (0.10)	
S12	0.69 (0.02)	0.94 (0.10)	1.60 (0.15)	1.72 (0.23)	
S13	0.63 (0.09)	0.89 (0.08)	0.84 (0.00)	0.93 (0.04)	
S14	1.13 (0.11)	1.54 (0.05)	1.16 (0.06)	1.64 (0.18)	
S15	0.63 (0.01)	0.89 (0.13)	0.94 (0.12)	0.93 (0.05)	
S16	0.78 (0.13)	0.97 (0.10)	1.15 (0.39)	1.08 (0.00)	
S17	0.50 (0.08)	1.00 (0.00)	1.13 (0.18)	0.97 (0.10)	
S18	0.49 (0.05)	0.86 (0.08)	1.06 (0.05)	1.13 (0.08)	
S19	0.50 (0.05)	0.99 (0.05)	0.92 (0.12)	1.16 (0.11)	
S20	0.62 (0.09)	1.03 (0.07)	0.96 (0.10)	0.90 (0.36)	
S21	0.79 (0.10)	1.02 (0.00)	0.94 (0.12)	1.11 (0.01)	
S22	0.60 (0.10)	1.05 (0.01)	1.22 (0.04)	1.09 (0.02)	
S23	0.51 (0.06)	1.05 (0.06)	1.01 (0.19)	0.98 (0.18)	
S24	0.69 (0.19)	0.91 (0.05)	1.10 (0.02)	0.95 (0.00)	
S25	0.47 (0.00)	0.98 (0.01)	1.14 (0.14)	1.02 (0.12)	
ATCC 29213	0.22 (0.04)	0.25 (0.03)	0.26 (0.04)	0.28 (0.07)	
ATCC 25923	1.18 (0.19)	1.04 (0.06)	1.46 (0.05)	1.45 (0.00)	

	S.aureus (n)				
	TSB	TSB _{Glc0.25}	TSB _{Glc1}	TSB _{Glc2}	
Nonbiofilm (-)	-	-	-	-	
Weak (+)	2	-	-	-	
Moderate (++)	21	4	3	1	
Strong (+++)	2	21	22	24	

Table 2. The biofilm formations of *S.aureus* isolates (n) with different types of TSB media (TSB, $TSB_{Glc0.25}$, TSB_{Glc1} and TSB_{Glc2}) on microtitration plates after 48 hours.

4. Discussion

In this study it was shown that *S. aureus* strains could produce moderate biofilm formations in the TSB, or strong biofilms in TSB supplemented with glucose, The obtained results were consistent with previous studies (Stepanovic et al., 2000; Rode et al., 2007; Manandhar et al., 2018). The environmental factors including the presence of glucose, or salt, temperature, and pH could be effective on the biofilm formations (Michu et al., 2011; Khangholi and Jamalli, 2016; Kyoui et al., 2016).

Biofilm formations on a solid surface also depends on the type of the surface materials (Jo et al., 2010). Generally, glass and stainless steel are described as hydrophilic materials while rubber and plastic are assumed as hydrophobic materials (Sinde and Carballo, 2000; Donlan and Costerton, 2002). The biofilm formations were found to be higher on hydrophobic surfaces than on hydrophilic surfaces (Simões et al., 2008). In this study, *S.aureus* isolates produced biofilms on hydrophobic polystyrene microtitration plates. Therefore, the biofilm formation of *S.aureus* isolates should also be assessed on various surfaces such as stainless steel, glass, or polypropylene in the presence of different media supplements.

5. Conclusions

In this work, the biofilm formations of 25 *S.aureus* isolates and 2 biofilm reference *S.aureus* strains were investigated by using TSB and TSB_{Glc} media on microtitraiton plates. When *S.aures* isolates were cultured in the TSB media, 21 of 25 isolates (84%) produced moderate biofilm formations. On the other hand, nearly all of isolates (96%) cultivated in TSB supplemented with glucose produced strong biofilm formations. According to these results, it can be concluded that presence of glucose is an important factor for *S.aureus* biofilm formations.

Acknowledgements

This work was supported by the Gebze Technical University (2012-A05), Turkey.

Authors' contributions:

Taner Sar: Performed the experiment, analyzed data, and wrote the manuscript.

Meltem Yesilcimen Akbas: Analyzed data, supervised the experiment's progress, and wrote the manuscript.

Conflict of interest disclosure:

There are no ethical issues after the publication of this manuscript.

References

- Akbas MY, Kokumer T 2015. The prevention and removal of biofilm formation of *Staphylococcus aureus* strains isolated from raw milk samples by citric acid treatments. J Food Sci Technol, 50(7): 1666-1672.
- Ateba CN, Mbewe M, Moneoang MS, Bezuidenhout CC 2010. Antibiotic-resistant *Staphylococcus aureus* isolated from milk in the Mafikeng Area, North West province, South Africa. S Afr J Sci, 106(11-12): 1-6.
- Aydin A, Sudagidan M, Muratoglu K 2011. Prevalence of staphylococcal enterotoxins, toxin genes and geneticrelatedness of foodborne *Staphylococcus aureus* strains isolated in the Marmara Region of Turkey. Int J Food Microbiol, 148(2): 99-106.
- Becker K, Heilmann C, Peters G 2014. Coagulase-negative staphylococci. Clin Microbiol Rev, 27(4): 870-926.
- Bonsaglia ECR, Silva NCC, Júnior AF, Júnior JA, Tsunemi MH, Rall VLM 2014. Production of biofilm by *Listeria monocytogenes* in different materials and temperatures. Food Control, 35(1): 386-391.
- Cha JO, Lee JK, Jung YH, Yoo JI, Park YK, Kim BS, Lee YS 2006. Molecular analysis of *Staphylococcus aureus* isolates associated with staphylococcal food poisoning in South Korea. J Appl Microbiol, 101(4): 864-871.
- Christensen GD, Simpson WA, Younger JJ, Baddour LM, Barrett FF, Melton DM, Beachey EH 1985. Adherence of coagulasenegative staphylococci to plastic tissue culture plates: a quantitative model for the adherence of staphylococci to medical devices. J Clin Microbiol, 22(6): 996-1006.
- Donlan RM, Costerton JW 2002. Biofilms: survival mechanisms of clinically relevant microorganisms. Clin Microbiol Rev, 15(2): 167-193.
- Flemming HC, Wingender J 2010. The biofilm matrix. Nat Rev Microbiol, 8(9): 623.
- Jo SH, Baek SB, Ha JH, Ha SD 2010. Maturation and survival of *Cronobacter* biofilms on silicone, polycarbonate, and stainless steel after UV light and ethanol immersion treatments. J Food Prot, 73(5): 952-956.

- Khangholi M, Jamalli A 2016. The effects of sugars on the biofilm formation of *Escherichia coli* 185p on stainless steel and polyethylene terephthalate surfaces in a laboratory model. Jundishapur J Microbiol, 9(9): e40137.
- Koutina G, Skibsted LH 2015. Calcium and phosphorus equilibria during acidification of skim milk at elevated temperature. Int Dairy J, 45: 1-7.
- Kyoui D, Hirokawa E, Takahashi H, Kuda T, Kimura B 2016. Effect of glucose on *Listeria monocytogenes* biofilm formation, and assessment of the biofilm's sanitation tolerance. Biofouling, 32(7): 815-826.
- Lee SHI, Barancelli GV, Camargo TM, Corassin CH, Rosim RE, Cruz AG, Cappato LP and Oliveira CA 2017. Biofilmproducing ability of *Listeria monocytogenes* isolates from Brazilian cheese processing plants. Food Res Int, 91: 88-91.
- Lee SHI, Cappato LP, Corassin CH, Cruz AG, Oliveira CAF 2016. Effect of peracetic acid on biofilms formed by *Staphylococcus aureus* and *Listeria monocytogenes* isolated form dairy plants. J Dairy Sci, 99: 2384-2390.
- Lee SH, Mangolin BL, Gonc
 ßalves JL, Neeff DV, Silva MP, Cruz AG, Oliveira CA 2014. Biofilm-producing ability of *Staphylococcus aureus* isolates from Brazilian dairy farms. J Dairy Sci, 97:1812-1816.
- Manandhar S, Singh A, Varma A, Pandey S, Shrivastava N 2018. Evaluation of methods to detect in vitro biofilm formation by Staphylococcal clinical isolates. BMC Res Notes, 11(1): 714.
- Méric G, Miragaia M, de Been M, Yahara K, Pascoe B, Mageiros L, Mikhail J, Harris LG, Wilkinson TS, Rolo J, Lamble S, Bray JE, Jolley KA, Hanage WP, Bowden R, Maiden MCJ, Mack D, de Lencastre H, Feil EJ, Corander J, Sheppard SK 2015. Ecological overlap and horizontal gene transfer in *Staphylococcus aureus* and *Staphylococcus epidermidis*. Genome Biol Evol, 7(5): 1313-1328.
- Michu E, Cervinkova D, Babak V, Kyrova K, Jaglic Z 2011. Biofilm formation on stainless steel by *Staphylococcus epidermidis* in milk and influence of glucose and sodium

chloride on the development of ica-mediated biofilms. Int Dairy J, 21(3): 179-184.

- Otto M 2013. Coagulase-negative staphylococci as reservoirs of genes facilitating MRSA infection. Bioessays, 35(1): 4-11.
- Rode TM, Langsrud S, Holck A, Møretrø T 2007. Different patterns of biofilm formation in *Staphylococcus aureus* under food-related stress conditions. Int J Food Microbiol, 116(3): 372-383.
- Santos VM, Martins HB, Rezende IS, Barbosa MS, Andrade EF, Souza SG, Campos GB, Oliveira PS, Sousa DS, da Silva DCC, Amorim AT, Timenetsky J, Cruz MP, Yatsuda R, Marques LM 2014. Virulence factor profile of *Staphylococcus aureus* isolated from bovine milk from Brazil. Food Nutr Sci, 5(15): 1496-1505.
- Simões M, Simões LC, Cleto S, Pereira MO, Vieira MJ 2008. The effects of a biocide and a surfactant on the detachment of *Pseudomonas fluorescens* from glass surfaces. Int J Food Microbiol, 121(3): 335-341.
- Sinde E, Carballo J 2000. Attachment of *Salmonella* spp. and *Listeria monocytogenes* to stainless steel, rubber and polytetrafluorethylene: the influence of free energy and the effect of commercial sanitizers. Food Microbiol, 17(4): 439-447.
- Stepanović S, Vukovic D, Dakic I, Savic B. Svabic-Vlahovic M 2000. A modified microtiter-plate test for quantification of Staphylococcal biofilm formation. J Microbiol Methods, 40: 175-179.
- Sudagidan M, Aydin A 2009. Screening virulence properties of staphylococci isolated from meat and meat products. Wien Tierärztl Monat, 96: 128-134.
- Tauxe RV 2002. Emerging foodborne pathogens. Int J Food Microbiol, 78(1-2): 31-41.
- Unlu A, Sar T, Seker G, Erman AG, Kalpar E, Akbas MY 2018. Biofilm formation by *Staphylococcus aureus* strains and their control by selected phytochemicals. Int J Dairy Tech, 71(3): 637-646.