



VOLUME : 5

NUMBER : 1

JAN. : 1976

THE JOURNAL OF TURKISH

PHYTOPATHOLOGY

Published by the Turkish Phytopathological Society

TURKISH PHYTOPATHOLOGICAL SOCIETY

President of Journal : Prof. Dr. İbrahim Karaca

Executive vice-president : Mustafa Copçu

Board of Editors : Doç. Dr. Tayyar Bora, Dr. Ülkü Yorgancı
Dr. Yıldız Sağmen, Dr. Coşkun Saydam, Yıldırım Arıncı

The Journal of Turkish Phytopathology is published once every four months. Three parts form a volume. The subscription price of a volume (which includes postage) is \$ 6.00

CONTENTS

Regional Disease Monitoring J. M. PRESCOTT.	1
Long Distance Transportation and Expansion of Wheat Rusts E. E. SAARI	7
Laboratory Studies of Three Species of Fungi Pathogenic to <i>Tetranychus urticae</i> Koch (Acarine: Tetranychidae) O. ECEVİT	13
Investigations on the Causal Agents and Their Pathogenicities and Chemical Control Methods of the Damping-Off of Carnation Seedlings Grown in Greenhouses of Izmir E. SEZGİN and İ. Karaca	21
Studies on Serology of Halo Blight (<i>Pseudomonas phaseolicola</i>) (Burkholder) (Dowson) of Beans Y. E. ÖKTEM	31
Elektronenmikroskopische Untersuchungen über die Stolburkrankheit an Tomaten Ü. YORGANCI	35

VOLUME : 5

NUMBER : 1

JAN. : 1976

Dizgi ve Baskı BİLGEHAN MATBAASI

Regional Disease Monitoring*

J. Michael PRESCOTT

CEMMYT/Pathologist

Cereals, especially wheat and barley, are widely grown in the region bounded by Bangladesh in the east and Morocco in the west. In fact they are the main crop of most of the countries within this region. Increased cereal production by way of growing varieties with higher yield potential, improved management practices, and better protection against pests and pathogens is the goal of a modern agriculture. Pests and pathogens annually account for the destruction of at least one-fifth of the world production. This is entirely too high a price to pay. Many of the production problems, especially the pathogens are similar throughout the region. These pathogens have been called "Shifty Enemies" because of their mobility and their ability to adapt to changes. Changes in the pathogen situation are usually evident at one or a few locations prior to appearing at another. With this changing situation in mind and since many airborne pathogens are quickly dispersed by weather sys-

tems, a disease monitoring system becomes a necessity. For this reason a regional disease surveillance program was initiated in 1971.

The actual beginnings were in the fall of 1970 when the First Turkish Trap Nursery was sent to many cooperators in Turkey and to seven locations within the region. This nursery was comprised of commercial varieties grown in Turkey and in the adjacent countries plus a number of resistant and susceptible checks. The response to this first monitoring or Trap Nursery was favorable and it was decided to expand this type of disease surveillance to more countries within the region. Thus with the first Turkish Trap Nursery as a nucleus the Regional Trap Nursery (RTN) came into existence in the fall of 1971.

The objectives for the RTN resulted from the experience gained in Wheat Improvement Programs in Turkey, India, Mexico, and the United States. They are:

* Paper presented at the First Turkish Plant Pathology Congress Izmir, Turkey 20-24 Oct. 1975

REGIONAL DISEASE MONITORING

1) to monitor the disease situation throughout the region using commercial and check varieties;

2) to determine the virulence potential of pathogens within the region;

3) to detect shifts or changes in virulence within the region and to provide advance warning of these shifts;

4) to assist in mapping the geographical distribution movement of the diseases within the region;

5) to determine the influence of the commercial varieties and their role in the selection and increase of virulent forms of the various diseases, particularly the rusts, within the region.

An effort was made to include in this nursery a representative number of the commercial wheat varieties, that is varieties grown on at least 100,000 ha, or more, along with appropriate checks. The number of entries selected for the nursery are not fixed and can be increased or decreased as the situation warrants each year. The RTN'71-'72 was comprised of 40 varieties, not replicated, and sent to 100 cooperators in 31 countries. The Turkish Wheat Improvement Program, CIMMYT, RF, ALAD/FF, and FAO were responsible for this regional effort. This program was continued in '72 - '74 program was continued in '72 - '73 and '3 - '74 in this format. In the third year of this program (1973-74), there

were 45 varieties included in the nursery and sent to 100 cooperators in 35 countries.

In 1974 the regional disease surveillance program was reorganized. the objectives remain the same but the manner of achieving these objectives were changed. During the first years of this program these were several specific disease monitoring nurseries being grown within the region. These nurseries included (1) The Yellow Rust Trial (Dr. R.W. Stubbs). (2) The European Leaf Rust Nursery (Dr. M. Boskovic), (3) The Egyptian Nursery (Dr. A.H. Kamel) and (4) The Regional Trap Nursery (Drs. J.M. Prescott and E.E. Saari). To avoid duplication of effort, to save time and needed nursery space, and to provide a stronger network for the regional disease surveillance program these four nurseries were combined into one nursery The Regional Disease Trap Nursery (RDTN). The identification of races and virulence of stripe, leaf, and stem rust will be performed by Drs. Stubbs, Boskovic, and Kamel respectively. The management, dispersal of seed, field note taking, and data summarization will be done by Drs. Saari and Prescott. This pooling of scientific effort will enhance the possibility of achieving the objectives of the regional disease surveillance program.

In 1974-75 the reorganized RDTN included 172 commercial and special rust identifying varieties or lines of

wheat and barley. The nursery was grown by 200 cooperators in 46 countries. This season, the RDTN is being grown by 145 cooperators in 41 coun-

tries (Table I). There are 115 varieties or lines in this nursery. These varieties have been grouped together as follows:

- I. Commercial Varieties
 - A. Durum Wheat
 - B. Bread Wheat
 - 1. Spring habit
 - 2. Winter habit
 - C. Barley
- II. Special Purpose Varieties/Lines
 - A. Stripe Rust (*Puccinia Striiformis*)
 - B. Leaf Rust (*Puccinia recondita*)
 - C. Stem Rust (*Puccinia graminis* f.sp. *tritici*)

TABLE I. COUNTRIES PARTICIPATING IN THE 1975-76 REGIONAL DISEASE SURVEILLANCE PROGRAM

1. Afganistan	11. Greece	21. Morocco	31. Switzerland
2. Algeria	12. India	22. Nepal	32. Sudan
3. Bangladesh	13. Iran	23. Netherlands	33. Syria
4. Bulgaria	14. Iraq	24. Nigeria	34. Tanzania
5. Cyprus	15. Italy	25. Oman	35. Tunisia
6. Czechoslovakia	16. Jordan	26. Pakistan	36. Turkey
7. Egypt	17. Kenya	27. Portugal	37. Yemen. North
8. England	18. Libya	28. Romania	38. Yemen, South
9. Ethiopia	19. Lebanon	29. Saudi Arabia	39. Yugoslavia
10. France	20. Madagascar	30. Spain	40. West Germany
	41. Zambia		

To illustrate the endemic virulence and resistance trends of the region, a comparison of the average coefficients of infection for the three rust among local, improved, and dwarf wheat varieties in four geographical areas for the three year pe-

riod 1972-74 is presented in Table II. A coefficient of infection greater than 10.0 suggests that the wheat varieties in this class have less adequate resistance and should be replaced.

REGIONAL DISEASE MONITORING

TABLE II. COMPARISON OF THE AVERAGE COEFFICIENT OF RUST INFECTION AMONG LOCAL, IMPROVED, AND DWARF WHEAT VARIETIES IN FOUR GEOGRAPHICAL REGIONS FOR THE THREE YEAR PERIOD 1972—74.

VARIETY* GROUP	INDIAN SUBCONTENT	MIDDLE EAST	NORTH AFRICA	SOUTH EUROPEA	AVE
A.) STEM RUST:					
LOCAL	22.35	20.16	15.94	18.67	20.41
IMPROVED	8.94	13.19	11.32	11.53	11.57
DWARF	1.13	4.30	2.95	2.95	3.12
B.) STRIPE RUST					
LOCAL	23.67	18.28	14.61	10.73	15.97
IMPROVED	8.07	9.46	5.24	8.23	8.39
DWARF	4.20	6.39	2.37	2.79	5.62
C.) LEAF RUST					
LOCAL	36.92	17.34	29.80	21.70	26.11
IMPROVED	27.92	14.74	18.40	12.03	18.84
DWARF	16.32	8.17	5.00	2.33	8.35

* DATA BASED ON:

1972 7 LOCAL, 12 IMPROVED, 12 DWARF WHEAT VARIETIES
 1973 7 LOCAL, 31 IMPROVED, 14 DWARF WHEAT VARIETIES
 1974 7 LOCAL, 21 IMPROVED, 15 DWARF WHEAT VARIETIES

A value greater than 5.0 but less than 10.0 suggests reasonable degrees of resistance however, new levels or sources of resistance may be needed shortly. A value less than 5.0 suggests the presence of adequate resistance.

From the data presented in Table II it is easily seen that the local varieties do not have adequate resistance to the three cereal rusts or in any of the four zones and could sustain heavy damage whenever the conditions for an epidemic are suitable. The improved varieties res-

ponse varies as to the species of rust involved. In this variety group stem rust resistance is barely adequate and in the possible damage area. Stripe rust resistance is adequate but new sources or degrees of resistance may be needed shortly. Leaf rust resistance is not sufficient and loss in production could occur in all zones within the region. In the case of the dwarf varieties, stem rust resistance is good to adequate in all zones., stripe rust resistance is reasonable but could require a change of resis-

tance genes, and leaf rust resistance reasonable in all zones of the region except the Indian Sub - continent where new sources of resistance are needed.

In general, the levels of resistance expressed by these commercial varieties follow a step-wise decrease with the dwarf commercial wheat varieties expressing the high-

est degree of resistance, the improved varieties in the middle, and the local commercially grown varieties expressing little or no resistance.

A large amount of data has now accumulated and is being assembled to that the goals of the Regional Disease Surveillance program will be attained and also be of value to the participating countries of the region.

Long Distance Transportation and Expansion of Wheat Rusts*

E.E. SAARI

Regional Plant Pathologist, CIMMYT, Beirut, Lebanon.

The nature of the wheat rusts dictate that a living host be present for the perpetuation of the uredospores cycle except for brief periods of time. Since the rust diseases in most areas survive via the uredospore cycle the cropping, accessory and adventive phenology is an important factor in the survival of the rust organism. In a number countries a continuing series of host plants are available throughout the year, and the uredospore cycle is restricted only by environmental factors. In such cases the situation is referred to as endemic.

There are, however, definite examples where the host plants are cyclic, the rust organism dies out in the area, and a manner of "geographical sterilization" occurs. In these situations the re-establishment of pathogen is dependent upon a new cycle of the host - usually represented by the sowing of wheat - the transport of viable initial (primary)

inoculum from a distant source, and favorable weather conditions for the deposition and infection process. This type of system is referred to as exodemic.

There are specific and well documented examples for both the endemic and exodemic models (2,3,4,5, 6,13,16,21). It should be remembered, however, that in most areas both systems probably function and tend to confuse our interpretation of results.

In any case, there are some specific examples of exodemic systems where the initial inoculum must be transported from some distant source. This aspect of long distance transport of wheat rusts has been the stimulus for international cooperation for many years. There is now additional interest in research efforts dealing with this subject, and for the purpose of this paper, I use long distance transport to mean distances in excess of 200 kilometers.

*) Paper presented at the First Turkish Plant Pathology Congress Izmir, Turkey 20-24 Oct 1975.

Probably the best documented long distance transport of rust spores is the example of *Puccinia graminis* f. sp. *tritici* in North America (13,20). Uredospore movement of 1000 km have been recorded on numerous occasions (20). Another well established system of long distance transport of stem rust uredospore occurs in Northwestern Europe. The disease cycle re-establishes itself each spring and it appears that the most likely source of inoculum is North Africa, or on occasion Southern Europe (19).

A specific exodemic situation which is being studied intensively is the Indian stem rust system. From extensive survey data re-establishment of stem rust into the plains of India occurs each year (6). The isochrones of rust appearance agree with crop phenology and there appears to be a close relationship with the isotherms of the monthly mean minimum temperature of 14°C. The environment and the absence of hosts for *P. graminis tritici* during the summer months eliminates the organism from the extensive plains area.

It has long been established that wheat and *P. graminis tritici* can be found at higher elevations in South India throughout the year (8). Stem rust also survives the summer period at the higher elevations in the northern hills of India, but the advent of cold weather shortly after sowing causes *P. graminis tritici* to "die out". The mean minimum for a three month period in the northern plains

of India is below 10°C. In most cases such a low temperature would eliminate the stem rust fungus, and survey data collected indicates that the fungus indeed does not survive this period in north India (6).

The only remaining source of inoculum would be the hills (Niligris) in southern India which extends to over 2,000 meters in elevation. Wheat grown in the Niligris during the summer period is at the elevations of 1500 meters or more. From this source area, stem rust uredospores must travel a considerable distance to the wheat growing areas of the plains (in excess of 300 km).

The long distance transport of spores requires that they reach heights of 1000 meters or more (5). Once uredospore have reached a height of several kilometers the principle deposition mechanisms are down-draughts or raindrops (5).

The association of the first appearance of stem rust with a previous rainfall period in India indicated that uredospore deposition probably occurred with the rain (9, 10). A study was undertaken to determine if this form of deposition was important (11). The sampling methods used to trap uredospores of stem rust were the same as those used in North America (17).

The technique involved the use of a millipore filter attached to the end of a rain funnel (15). After each rain the filter was removed by coo-

operators at the different sites throughout India, and mailed to the Cereal Disease Laboratory, Indian Agricultural Research Institute, New Delhi. There the filters were processed and counts were made for the presence and number of spores trapped.

A total of 135 filters were processed over a five year period. This represents 50 test site - years (see table 1). The number of testing locations were not constant or the same throughout the study, and the number of rain samples received from each station in any year varied. A total of 83 samples or 61.5 % of the samples were found to have stem rust uredospores. Spore counts varied from a few spores to more than 5000 per sample.

At 25 test sites the rain sampling was associated with spore trapping studies using the rod impaction method (14). At the remaining 25 test sites the National Disease Trap Nursery was grown and weekly inspections of the susceptible check variety were made for the first appearance of rust by cooperators (6).

Once stem rust spores were detected at a location subsequent rain samples with uredospores from that location were not counted in the data presented in Table 2. The percentage figures given are based upon the use of 50 test site-years. In 70 % of the cases a positive relationship exists. In 24 % of the cases spores were caught

but rust was not detected on the susceptible varieties in the nursery. The mere arrival of inoculum does not guarantee that the uredospores were viable or conditions were favorable for the infection process. In only 6 % of the cases did a negative relationship result. These negative examples could not be adequately explained but I feel confident that some unexplained experimental error is represented.

A comparison of the first date of the uredospores in rain samples and on the glass rod established an average difference of 22 days earlier detection for the rain sampling procedure. The date of rust appearance on the susceptible variety in the Trap Nursery averaged 36 days after the first rain sampling of uredospores.

The fact that uredospores are transported at high altitudes and become associated within cloud and rain systems suggests that Environmental Satellite Imagery or Satellite Television Cloud Photography may be a useful tool in studying long distance transport of uredospores (12). Cloud systems as viewed through such remote sensing programs provide a number of opportunities to map the movement of uredospores from a source to an area which is exodemic.

In this area of the world the most definitive exodemic wheat growing area is the Nile River Valley. Over summering of the host and the fungi (*P. graminis tritici*, *P. re-*

condita f. sp. *tritici* and *P. striiformis* f. sp. *tritici*) does not occur. All three rusts do, however, present a problem to wheat cultivation (1,5). Spore trapping data indicates that the rust arrives from some northerly sources (1). The rust isochrones also suggest a north to southward development in contrast to a general northward and eastward developmental pattern in the adjacent countries (5). The environmental factors on crop and pathogen development need to be studied, and the weather systems, especially, the common low pressure cell probably has a major influence on rust development in Egypt.

If one begins to analyse certain virulence patterns and compares varietal reactions the indications are that Western Asia - parts of Iran, Syria, possibly Iraq and portions of Afghanistan - may serve as the ino-

culum reservoirs for Egypt (18). This area includes parts of the ancestral home of wheat, and many related species are also present in this region. This provides for a bridging of inoculum through periods of reduced commercial acreage. In addition the cropping cycle at the higher elevations in this part of the world also provides a means by which the uredospores cycle can be perpetuate. The fact that many "islands of wheat" exists in this region, probably, provides for more than a single inoculum source. Unless all of the sources are identified and quantified, it will be difficult to use varietal response and race analysis data effectively.

Data and information is still too fragmented to draw any strong conclusions but if one reviews some of the weather systems common to this area (7) and considers both endemic and exodemic patterns a greater under-

Table 1. Summary for five years of rain sampling from Nagrajan et al. (11).

Years	Average No. Stations	Rain Samples	Samples with Uredospores
5	10	135	83

Table 2. Relationship of rain samples with spores (+) or no spores (-) and rust (+) or no rust appearance (-) from Nagarajan et al (11).

See text for details.

	Spores(+) Rust (+)	No spores(-) No rust (-)	Spores(+) No rust(-)	No spores(-) Rust (-)
Samples	24	11	12	3
%	48	22	24	6

standing of rust development seems possible. Turkey certainly must play a pivotal role in any wheat rust epidemiological patterns in the region because of all or the factors previously mentioned. A greater understanding of what is happening here in Turkey will provide opportunities for minimizing rust attacks and losses to production. This knowledge will also provide some of vital missing bits of information that will allow

other countries to develop the required background for controlling the losses caused by the wheat rusts.

I believe that First Turkish Phytopathological Society Congress represents progress toward this end. I will look forward to reading future proceedings, especially those articles contributing to our greater understanding of the wheat rust epidemiology.

LITERATURE CITED

- 1-ABDEL-HAK, T., A.H. KAMEL, S. KEDDIS and E. SHAFIK 1966. Epidemiology of wheat rusts in U.A.R. (Ergypt) Ministry of Agriculture, Plant Protection Department, Cereal Diseases Research Division Tech. Bull. No. 1, 45 p.
- 2-CHESTER, K.S. 1946. The Nature and Prevention of the Cereal Rusts as exemplified in the Leaf Rust of Wheat. *Chronica Botanica Co.*, Waltham, Mass, 269 p.
- 3-DINCOR, A. 1974. Role of wild and cultivated plants in the epidemiology of plant diseases in Israel. *Ann. Review Phytopath.* 12: 413-436.
- 4-HARDER, D.E., G.R. MATHENGE and L.K. MWAURA. 1972. Physiologic specialization and epidemiology of wheat stem rust in East Africa. *Phytopathology* 62:166-177.
- 5-HOGG, W.H., C.E. HOWNAM, A.K. MALIK and J.C. ZADOKS. 1969. Meteorological factors affecting the epidemiology of wheat rusts. *World Meteorological Organization Technical Note No.99 (WMO-No. 238 TP. 130)*. Geneva. 143 p.
- 6-JOSHI, L.M., E.E. SAARI, S.D. GARA and S. NAGARAJAN. 1974. Survey and epidemiology of wheat rusts in India p. 150-159. In *Current Trends in Plant Pathology*. (Ed) S.P. Raychaudhuri and J.P. Verma. Parnassus Publishers. New Delhi 341 p.
- 7-KENDREW W.G, 1961. The climates of the Continents, 5 th Ed., Oxford Press, London p. 348-377.
- 8-MEHTA,, K.C. 1952. Further studies on cereal rusts in India, Part III, Indian Council Agric. Res., *Scientific Monog.* 18, 368 p.
- 9-NAGARAJAN, S. and H. SINGH. 1975. The Indian stem rust rules - an epidemiological concept on the spread of wheat stem rust. *Plant Dis. Repr.* 59:133-136.
- 10-NAGARAJAN, G., H.SINGH and L.M. JOSHI 1975. Climatic factors in relationship to stem rust epidemiology. *Pl. Dis. Repr.* 59:670-672.

LONG DISTANCE TRANSPORTATION AND EXPANSION OF WHEAT RUSTS

- 11-NAGARAJAN, S., H.SINGH, L.M. JOSHI and E.E. SAARI. 1975. Long distance dissemination and deposition of uredospores of *Puccinia graminis* f. sp. *tritici* in India. (In press).
- 12-NAGARAJAN,S., H.SINGH, L.M.JOSHI and E.E. SAARI. 1975. The use of satellite television cloud photography in epidemiology investigation of *Puccinia graminis* f. sp. *tritici* in India (In press).
- 13-RAJARAM, S. and A. CAMPOS. 1974. Epidemiology of wheat rusts in the Western Hemisphere. CIMMYT Research Bull. No. 27 p.
- 14-ROELFS, A.P., V.A. DIRKS and R.W. ROMIG 1968. A comparison of rod and slide samples used in cereal rust epidemiology. Phytopathology 58:1150-1154.
- 15-ROELFS,A.P., J.B.ROWELL and R.W.ROMIG 1970. Sampler for monitoring cereal rust uredospores in rain. Phytopathology 0: 187-188.
- 16-ROELFS, A.P. 1974. Evidence for two populations of wheat stem and leaf rust in the USA. Plant Dis. Reprtr. 58:806-809.
- 17-ROWELL, J.B. and R.W. ROMIG. 1966. Detection of uredospores of wheat rust in spring rains. Phytopathology 56:807-811.
- 18-SAARI,E.E. G.KINGMA, and J.M.PRESCOTT 1972. Identifying sources of resistance through international testing. Proceed. European and Mediterranean Cereal Rusts Conf. Vol. II: 219-223.
- 19-SANTIAGO 1968. Differentiating biotypes within physiologic races and its importance on epidemiological studies. Proceed. Cereal Rust Conf., Portugal. p. 40.
- 20-STAKMAN E.C. and J.G.HARRAR, 1957. Principles of Plant Pathology. The Ronald Press Co., New York 581 p.
- 21-STUBBS ,R.W. 1972. The international survey of virulence of *Puccinia striiformis* virulence patterns in the Middle East and Africa and potential sources of resistance. Regional Wheat Workkshop, Beirut, Lebanon. 15 p.

Laboratory Studies of Three Species of Fungi Pathogenic to *Tetranychus urticae* Koch (Acarine: Tetranychidae)

Doç. Dr. Osman ECEVİT

SUMMARY

Three species of fungi identified as *Cladosporium* sp., *Aspergillus ochraceus* and *A. niger* are reported as causing mortality of two-spotted spider mites, *Tetranychus urticae* Koch. The fungi were isolated and the role of mite populations is discussed. *Cladosporium* species could be very important as a biological control agent for two-spotted spider mite. However, culture of the fungus is very difficult. *A. ochraceus* is readily disseminated in a variety of ways, easily penetrates the mite and is easy to culture. In spite, *A. niger* is easy culture, of secondary importance as a possible biological control agent for two-spotted spider mite.

INTRODUCTION

During the past 30 or more years many investigations have been reported concerning pathogenic fungi attacking plant - parasitic mites but relatively few experiments to observe the activity of the fungi have been performed. In the literature many species of Phycomycetes, Ascomycetes, Basidiomycetes, and Fungi Imperfecti are recorded as pathogenic to mites, some of which seem to be important in controlling them.

Petch (1940) reported that *Hyalotydeus destructor* became infected with *Entomophthora acaricida*. In 1944 the same author described another species, *E. acaridis*, from Acarina. Fisher (1950 a,b) observed that *Hirsutella thompsonii* caused injury to *Phyllocoptruta oleivora* and on non-sprayed citrus trees in June and July, these mites rapidly disappeared. In 1951 he reported that a species of *Entomophthora* caused

*) Atatürk University, Agricultural Faculty, Erzurum, TURKEY.

mortalities of adult citrus red mites *Panonychus citri*, ranging from 32 - 95 %. Muma (1955) observed two fungi pathogenic to citrus rust mites, *P. oleivora*. These were *Hirsutella thompsonii* and a species of *Entomophthora*. Leatherdale (1965) found that *Paecilomyces eriophytis* (= *Botrytis eriophytis*) originally described from an eriophyid mite, *Cecidophyopsis ribis*, was infective also to *Eriophyes padi*, *Aceria hippocastani*, and *Panonychus ulmi*.

A fungus attacking *Eutetranychus banksi* was first observed by Fisher (1954) and later described as *Entomophthora floridana* by Weiser and Muma (1966). This fungus produces single conidiophores with pyriform conidia. Selhime and Muma (1966) studied the biology of *E. floridana* and found that this pathogen had a 5-6 days life cycle. The death of the mites were caused by dense hyphal

growth inside the body. Also, Baker and Neunzing (1968) reported that *H. thompsonii* attacked the blueberry bud mite, and Muma (1969) found that *E. floridana* caused the death of *Eutetranychus banksi* during the summer and fall, mite populations being reduced by 36.8-86.3 per cent.

Carner and Canerday (1968) observed an entomogenous fungus infecting *Tetranychus telarius* and *T. urticae*. They identified it as *Entomophthora fresenii*, however this identification has been questioned by Wilding and Weiser (Carner, pres. comm. 1969, cf., Lipa, 1971). Recently Sannasi and Oliver (1971) studied the integument of the velvet mite, *Dinathrombium giganteum* and the histological changes caused by the fungus, *Aspergillus flavus*. Also, Kenneth, et al. (1972) found that *Triplosporium floridanum* caused mortality of spider mites in Israel.

MATERIAL and METHODS

In our laboratory, cultures of phytoseiid mites were being isolated in petri plates and two-spotted spider mites used as prey. Fungi were observed attacking the spider mites in these cultures and also it was noted that the stock culture of the spider mites was at a very low density and declining. Examination of infested leaf samples under the stereomicroscope showed that the spider mites were infected with fungi. Slide

mounts of the fungi were made using Hoyer's medium as the mountant. Examination of the slides indicated that at least two different species were present.

Isolation of each fungus was made on potato-dextrose medium in the Biological Control of Insects Research Laboratory at Columbia, Mo., and identification of the made by a specialist at the Peoria, Illinois USDA laboratory.

Uninfected spider mite cultures were established in our laboratory and infected with each of the fungus isolated taken from the P-D media. Also, fungi isolated from the infected

stock culture were found capable of infecting live mites. Unfortunately, the original source of the infected stock culture of two-spotted spider mites is unknown.

RESULTS and DISCUSSION

Three species of fungi were found to be infecting the two-spotted spider mites. They have been identified as *Cladosporium* sp.; *Aspergillus ochraceus*; and *Aspergillus niger*. Frequently it was found that more than one species of fungi were grow-

ing on the same mite specimen. For this reason, isolation and establishing pure cultures had been difficult.

Cladosporium sp.- In our opinion this pathogen must be the most important of the three. Examination of numerous slides reveals that spores

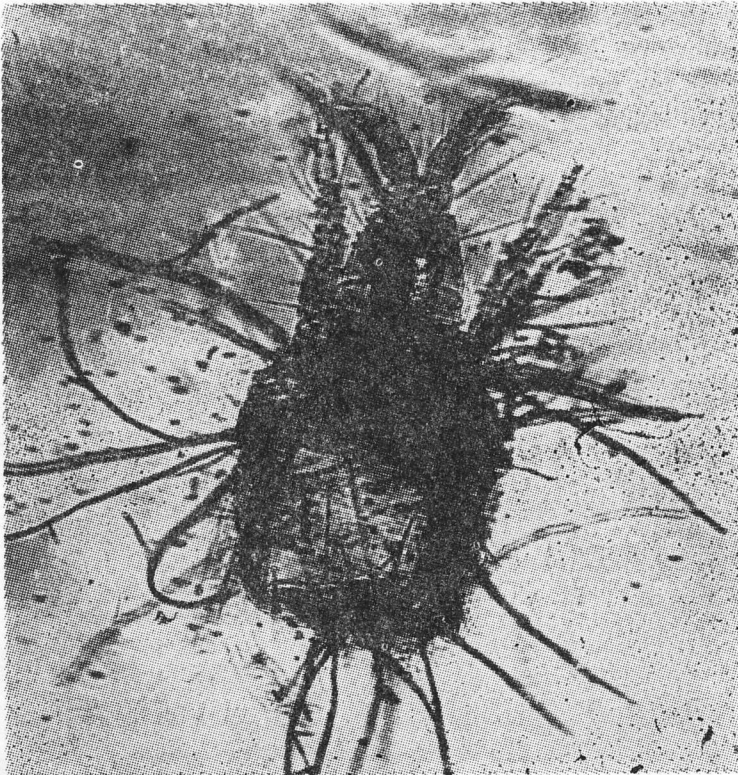


Fig.1. Two-Spotted Spider Mite killed by *Cladosporium* sp.

of *Cladosporium* grow on the surface of the mite and the hyphae enter through the anus or mouthparts. It seems probable also this fungus is able to dissolve the body wall of the mite. Death of the mite follows penetration, by whatever method, and the mite, inside and out, is completely covered (Fig. 1). The fungus is brown in color.

This fungus seems also to cause mortality of mite eggs. The fungus first forms a covering over the egg, then penetrates inside (Fig. 2). There are two ways in which the fungus might penetrate the egg, first, it may dissolve the shell and penetrate in that manner or, second, it may penetrate via the micropylar area. The probability seems to favor the latter

because the fungus is almost always found at only one point inside the egg.

This *Cladosporium* species could be very important as a biological control agent for the two-spotted spider mite. However, culture of the fungus is very difficult. In our experience only a pair of culture plates were achieved and none of them grew very well. Another drawback is that dissemination of the fungus is very slow and depends on mite movement primarily and very little by spreading over the leaf surface.

Aspergillus ochraceus.- One of the most important genera of fungi attacking insects and mites in *Aspergillus*. Hundreds of spores of *A. ochraceus* have been observed on the

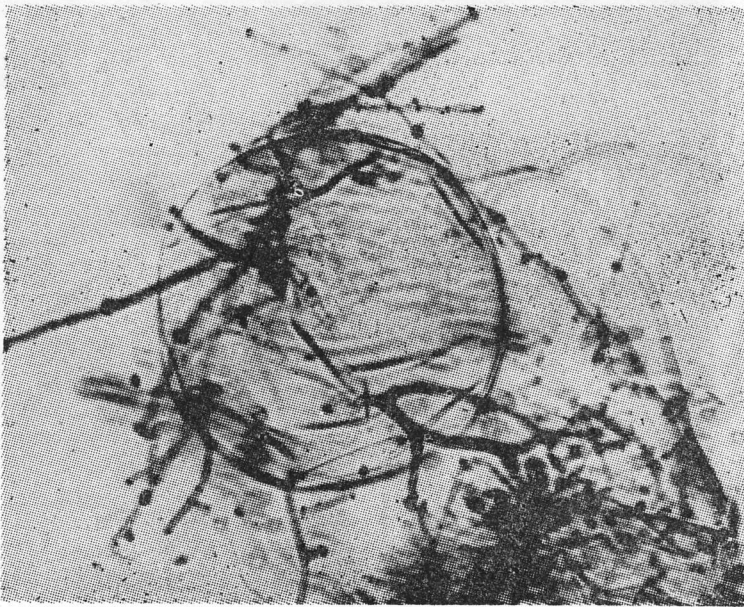


Fig. 2. The egg of Tw-Spotted Spider Mite killed by *Cladosporium* sp.

body surfaces of the mites. This fungus seems able to penetrate the body wall of the spider mite and invade the body. From the spore a structure grows for a short distance and penetrates the cuticle. From this a stalk (Fig. 3) emerges on the end of which a spore-bearing body forms. A single dead mite may have from 1 - 5 such structures.

This fungus is readily disseminated in a variety of ways, easily penetrates the mite, and is easy to culture.

Aspergillus niger.- This species is very similar in penetrating ability, dissemination characteristics, and easy of culture to **A. ochraceus** (Fig. 4). There is also very little difference in conidiophore structure between the two.

This fungus usually appears in cultures later than the two preceding and is therefore believed to be a more secondary cause of mite mortality. However, it has the same ability to penetrate into mites and to spread rapidly once it becomes established. In spite of its easy culture it is, in our opinion, of secondary importance as a possible biological control agent for spider mites.

In summary, many attempts have been made to use fungi for spider mite control and unquestionably fungi play a very important role in regulating mite populations. However, fungi are dependent to a high degree on local weather conditions and on the microenvironment in the immediate vicinity of the target spe-

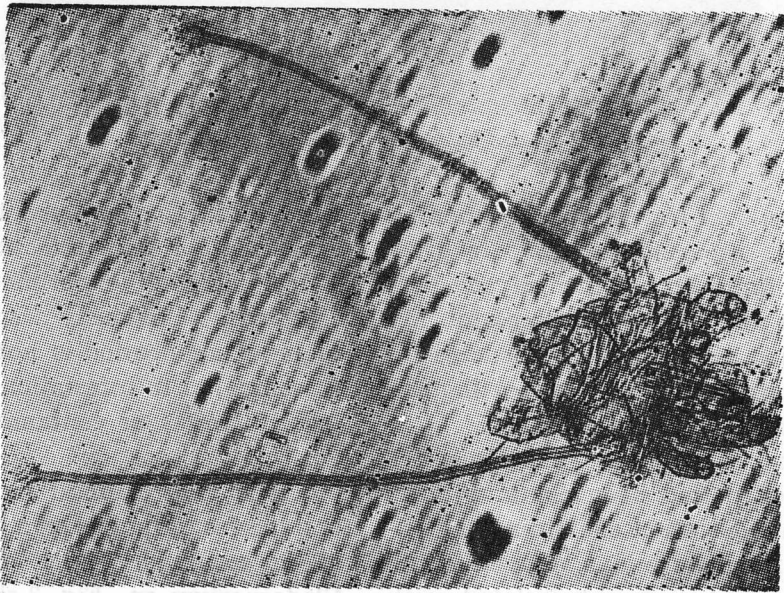


Fig.3. Two-Spotted Spider Mite killed by *Aspergillus ochraceus*.

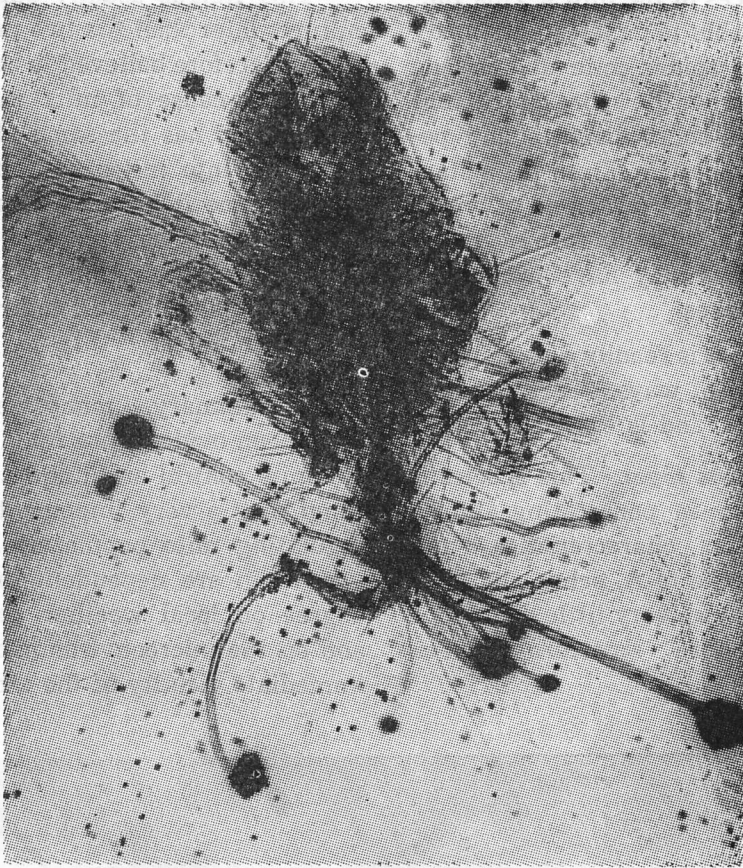


Fig. 4. Two-Spotted Spider Mite killed by *Aspergillus niger*.

cimens. As Carner and Canerday (1968) reported, attempts to grow these fungi on common mycological media are usually unsuccessful. My isolation attempts were unsuccessful more often than not, supporting the conclusion of these authors.

In a greenhouse experiment, I infested potted bean plants with *T. urticae*. Solution containing the several fungi were then sprayed on the plants, and each pot isolated with polyethylene plastic covers. Every

week leaf samples were taken and examined for mite infection by the fungi but the experiment failed completely. Obviously a spray technique of application is not effective as compared to infestation by infected live mites or contaminated leaves.

It is very difficult to find only one species of fungus on a given mite as, for example, Fig. 1 shows only *Cladosporium* sp., Fig. 3 shows only *A. ochraceus*, and Fig. 5 shows only

A. niger. Usually **A. ochraceus**, **A. niger**, and **Cladosporium** sp. are found in combination. The associations in

all probability have considerable, but at present unknown, significance.

ACKNOWLEDGEMENTS

This research has been done in Department of Entomology Missouri State University, Mo., USA. The writer gratefully acknowledge Dr. Wilbur R. Enns and the assistance of Don L. Hostetter of the USDA Biological Control of Insects Research Laboratory, Columbia, Mo., for isolating

the fungus cultures and helping with illustrations; Dorothy I Fennell, microbiologist, of the USDA Northern Regional Research Laboratories, Peoria, Ill. for identifying the fungi; and Tom A. Donahoo, University of Missouri - Columbia photographer for the photomicrographs.

Ö Z E T

Tetranychus urticae (ACARINA: TETRANCYHIDAE)'DA PATHOGENİK ÜÇ FUNGUS TÜRÜ ÜZERİNDE LABORATUVAR ÇALIŞMALARI

Tetranychus urticae (Koch)'da ölüm meydana getiren üç tür patojenik fungus bulunarak **Cladosporium** sp., **Aspergillus ochraceus** ve **Aspergillus niger** olarak tanılanmıştır. Funguslar izole edilerek akar popülasyonları üzerindeki tesirleri tesbit edilmiştir. **Cladosporium** sp., **T. urticae** ile yapılacak mücadelede çok önemli olacağı benzetmektedir. Bu

nunla beraber bu fungusun kültüre alınmasının çok zor olduğu görülmüştür. **A. ochraceus** muhtelif yollardan kolaylıkla yayılabilen, kolay kültüre alınabilen ve akar'ın vücuduna nüfuz edebilen bir fungustur. Buna karşılık, **A. niger** kolay kültüre alınabilmekle beraber, kanaatıma göre **T. urticae**'nin kontrolunda ikinci derecede rol oynayacaktır.

LITERATURE CITED

- BAKER, J.R., and H.H. NEUZING. 1968. *Hirsutella thompsonii* as a fungus parasite of the blueberry bud mite. J. Econ. Entomol. 61 (4):1117 - 18.
- CARNER, G.R., and T.D. CANERDAY. 1968. Field and Laboratory investigations with *Entomophthora fresenii*, a pathogen of *Tetranychus* spp. J. Econ. Entomol. 61 (4) : 956 - 59.
- FISHER, F.E. 1950 a. Entomogenous fungi attacking scale insects and rust mites on citrus in Florida. J. Econ. Entomol. 43 (3) : 305 - 09.
- . 1950 b. Two new species of *Hirsutella* Patonillard. Mycologia 42:290-93.
- . 1951. An *Entomophthora* attacking citrus red mites. Florida Entomol. 34(3): 83 - 88.

TETRANYCHUS URTICAE KOCH (ACARINE : TETRANYCHIDAE)

- _____. 1954. Diseases of citrus insects. Florida Agr. Exp. Sta. Ann. Rpt. for 1953, 191.
- KENNETH R., G. WALLIS, U. GERSON, and N. H. PLANT. 1972. Observations and experiments on *Triplosporium* (Entomophthorales) attacking spider mites in Israel. J. Invert. Pathol. 19 : 366 - 369.
- LEATHERDALE, D. 1965 Fungi infecting rust and gall mites (Acarina: Eriophyidae). J. Invert. Pathol. 7 : 325 - 328.
- LIPA, J.J. 1971. Microbial control of mites and ticks. Academic Press. London and New-York. 357 - 373.
- MUMA, M.H. 1955. Factors contributing to the natural control of citrus insects and mites in Florida. J. Econ. Entomol. 48(4):437-438.
- _____. 1969. Coincidence and incidence of *Entomophthora floridana* with and in *Eutetranychus banksi* in Florida citrus groves. Florida Entomol. 57(2): 107-112.
- PETCH, T. 1940. An *Empusa* on a mite. Proc. Linnaean Soc. N.S. Wales 65:259-260.
- _____. 1944. Notes on entomogenous fungi Trans. British Mycol. Soc. 27:81-93.
- SANNASI, A., and J.H. OLIVER. 1971. Integument of the velvet mite, *Dinathrombium giganteum*, and histopathological changes caused by the fungus *Aspergillus flavus*. J. Invert. Pathol. 17 : 354-365.
- SELHIME, A.G., and M.H. MUMA. 1966. Biology of *Entomophthora floridana* attacking *Eutetranychus banksi*. Florida Entomol. 49(3): 161-168.
- WEISER, J., and M.H. MUMA. 1966. *Entomophthora floridana* n. sp. (Phycomycetes: Entomophthoraceae), a parasite of the Texas citrus mite, *Eutetranychus banksi*. Florida Entomol. 49(3) : 155 - 159.

LITERATURE CITED

BAKER, I.B. and M.H. MUMA. 1966. *Entomophthora floridana* n. sp. (Phycomycetes: Entomophthoraceae), a parasite of the Texas citrus mite, *Eutetranychus banksi*. Florida Entomol. 49(3) : 155 - 159.

CAMMERMEYER, D.R. and T.D. CARRETERO. 1968. Field and laboratory investigations with *Entomophthora floridana*, a pathogen of *Tetranychus* spp. J. Econ. Entomol. 61(4) : 522 - 529.

_____. 1969. Coincidence and incidence of *Entomophthora floridana* with and in *Eutetranychus banksi* in Florida citrus groves. Florida Entomol. 57(2) : 107 - 112.

_____. 1972. Observations and experiments on *Triplosporium* (Entomophthorales) attacking spider mites in Israel. J. Invert. Pathol. 19 : 366 - 369.

LEATHERDALE, D. 1965. Fungi infecting rust and gall mites (Acarina: Eriophyidae). J. Invert. Pathol. 7 : 325 - 328.

LIPA, J.J. 1971. Microbial control of mites and ticks. Academic Press. London and New York. 357 - 373.

MUMA, M.H. 1955. Factors contributing to the natural control of citrus insects and mites in Florida. J. Econ. Entomol. 48(4) : 437 - 438.

_____. 1954. Diseases of citrus insects. Florida Agr. Exp. Sta. Ann. Rpt. for 1953, 191.

PETCH, T. 1940. An *Empusa* on a mite. Proc. Linnaean Soc. N.S. Wales 65 : 259 - 260.

_____. 1944. Notes on entomogenous fungi. Trans. British Mycol. Soc. 27 : 81 - 93.

SANNASI, A., and J.H. OLIVER. 1971. Integument of the velvet mite, *Dinathrombium giganteum*, and histopathological changes caused by the fungus *Aspergillus flavus*. J. Invert. Pathol. 17 : 354 - 365.

SELHIME, A.G., and M.H. MUMA. 1966. Biology of *Entomophthora floridana* attacking *Eutetranychus banksi*. Florida Entomol. 49(3) : 161 - 168.

WEISER, J., and M.H. MUMA. 1966. *Entomophthora floridana* n. sp. (Phycomycetes: Entomophthoraceae), a parasite of the Texas citrus mite, *Eutetranychus banksi*. Florida Entomol. 49(3) : 155 - 159.

Investigations on the Causal Agents and Their Pathogenicities and Chemical Control Methods of the Damping-Off of Carnation Seedlings Grown in Greenhouses of Izmir

Emel SEZGİN* and İbrahim KARACA**

ABSTRACT

This study was conducted on the causal agents and their pathogenicities and chemical control methods of the damping-off of carnation seedlings grown in greenhouses of İzmir in 1972-1974. In this study 41 genera of fungi were isolated from diseased samples collected at two different times.

The genera of *Sclerotium*, *Macrophomina*, *Fusarium*, *Acremoniella*, *Cladorrhinum*, *Stilbella*, *Rhizoctonia*, *Nigrospora*, *Melanospora*, *Verticillium*, *Alternaria*, *Sordaria*, *Pythium* and *Actinomucor* were selected for pathogenicity tests among all of the isolated fungi according to the results of the preliminary laboratory tests. In the pathogenicity tests, these fungi were used alone and combined with saprophytic fungi such as *Aspergillus*, *Myrothecium*, *Penicillium*, *Trichoderma*, *Chaetomium* and *Gliocladium*. When all the tested pathogens were combined with saprophytic fungi, it was generally determined that the incidence of the disease increased.

In the chemical control tests the genera of *Sclerotium*, *Rhizoctonia*, *Fusarium*, *Macrophomina*, *Cladorrhinum*, *Melanospora*, *Pythium*, *Acremoniella*, *Stilbella* and *Actinomucor* and Methyl bromide, Orthocide soil treater Dexamal and Benlate chemicals were used. There have not been obtained any satisfactory effectivenesses from the chemicals against the causal agents of the damping-off carnation seedlings.

*) Regional Plant Protection Research Institute, Bornova, İzmir, TURKEY.

**) Department of Phytopathology and Agricultural Botany, University of Ege, İzmir, TURKEY.

INTRODUCTION

Commercial flower growing has become a great importance in Turkish agriculture. Flower growing were done in three sections such as cuttings flower growing, greenhouse flower growing and flower bulbs and corms growings. The value of these sections were 12.000.000, 35.000.000 TL. respectively in 1968-1972 and 1974 (OSMANLIOĞLU, 1974).

In Ege Region green house flower growing began in 1936 and it was done in an area of 2000 m² and soon become enlarged According to 1971-1972 statistics, total flower growing area is 220.766m² in Balçova and Narlıdere. Generally flowers are grown

in greenhouses which covered with polyethylene as a family enterprise. The most growing flower is carnation (*Dianthus, caryophyllus* L.). There are many diseases and pests problems of carnation in our region. One of them is damping-off disease which causes damage in early stage of carnations. In some years this damage is very high because of growers don't use any chemicals against soil born fungi.

This study was done to determine root mycoflora, their pathogenicities and chemical control methods of the damping-off disease of carnation in 1972-1974.

MATERIAL AND METHODS

Survey were done at two times in order to collect diseased cuttings and seedlings. During the surveys 2925 samples were collected from 117 greenhouses. The roots of diseased samples were washed with tap water and cut into small pieces with a sterilized scalpel. Their surface sterilized with 0.5 % sodium hypochloride for one minute and washed in sterilized water and then dried between sterile blotting papers. The root pieces placed in petri dishes. The isolation mediums were %2 PDA, czapeck-dox agar and 8 % water agar. The petri dishes were incubated at 20°C

Pathogenicity tests were performed in pots. Test plants were grown from cuttings taken from healthy plants and rooted in sterilized soil. Genera of *Fusarium*, *Rhizoctonia*, *Pythium*, *Sclerotium*, *Macrophomina*, *Alternaria*, *Cladorrhinum*, *Nigrospora*, *Acremonia*, *Stilbella*, *Actinomucor*, *Sordaria*, *Verticillium* and *Melanospora* were selected for pathogenicity tests among all of the isolated fungi according to the preliminary laboratory tests. These genera were both applied singly and in mixture with saprophytic fungi such as *Gliocladium*, *Aspergillus*, *Chaetomium*, *Penicillium*,

Myrothecium and Trichoderma.

Inoculations were done according to the WILES (1952) method. Tested fungi were cultured on PDA for 10 days then 5 petri dishes of each fungi were blended in 250 cc sterilized water for 5 min. at high speed in waring's blender. Roots of the experimental seedlings were dipped in these inoculum for 5 min. and the seedlings were transplanted to pots. Roots of check plants were dipped only in sterilized water. Results of pathogenicity tests were obtained 50 days after inoculation.

In the chemical control test, the genera of *Sclerotium*, *Rhizoctonia*, *Melanospora*, *Fusarium*, *Macrophoma*, *Cladorrhinum*, *Pythium*, *Acremonia*, *Stilbella* and *Actinomucor* which they shown above 30 % patho-

genicity in the pathogenicity tests were used. These and 6 saprophytic fungi (*Aspergillus*, *Myrothecium*, *Penicillium*, *Trichoderma* and *Gliocladium*) were combined and then added to pots. After 7 days when the fungi colonized in the pot soil methyl bromide (100 cc / 1m³), Benlate (methyl 1- (butyl-carbomyl)- 2-benzimidazole carbomate) 0,5 /lt. Orthocide soil treater (10 % N-trichloromethyl - mercapto - 4- cyclohexene-1,2 dicarboximide (captan) and 60 % PCNB) 10 Kg/hect. and Dexonal (P-Dimethyl ominolbenzol - diazonat - rium sulfanat 2,5 % + 10 % PCNB) 200 mg /kg soil. Were applied. 5 seedlings per pot were sown. After two months from applications effects of the chemicals were determined by counting the healthy and diseased seedlings.

RESULTS AND DISCUSSION

A — Isolated genera and their rates of occurrence :

The genera of fungi isolated from diseased samples were collected at two different times and their percentage, occurrence in the total isolates are shown in table.

41 genera of fungi were isolated from diseased samples. Among these genera the most commonly fungus was *Fusarium* which were isolated from all of the greenhouses which were surveyed. The genera of *Fusarium* is common for soils and rhizos-

phere in Turkey and other countries (MENON, WILLIAMS, 1957; PARKINSON et al., 1963; KUBIKOVA, 1968; BAGGA, 1970; BORA, 1972; TURHAN, 1973; SARIBAY, 1974).

The other genera of fungi followed *Fusarium* have different rates of occurrences at both surveys. This can be attributed to the ecological conditions especially to soil temperature. For example *Sclerotium* species give damage to seedlings especially during the summer months (ANDREUCCI 1955) and *Aspergillus* spe-

Table. Isolated genera and their rates of occurrence in total number of isolates and sampled in the greenhouses.

Genera of Fungi	Survey I (1-15.5.1972)			Survey II (15-7-15.8.1972)		
	No of recovered isolates	Relative intensity (%)	Rates of presence in the nurseries (%)	No of recovered isolates	Relative intensity (%)	Rates of presence in the nurseries (%)
Fusarium	177	32.89	100	849	38.22	100
Rhizoctonia	47	8.73	60.4	350	15.75	91.8
Aspergillus	6	0.92	2.3	234	10.53	33.7
Pythium	45	8.36	48.8	125	5.62	48.7
Gliocladium	66	12.26	74.4	102	4.59	64.8
Melanospora	1	0.18	2.3	101	4.54	35.1
Macrophomina	17	3.15	23.2	97	4.36	63.5
Sclerotium	2	0.37	4.6	65	2.92	29.7
Alternaria	64	11.89	88.3	42	1.89	44.5
Mucor	7	1.30	11.6	30	1.35	24.3
White steril mycel	3	0.55	6.9	28	1.26	27.02
Chaetomium	—	—	—	27	1.21	16.2
Cladorrhinum	16	2.97	20.9	23	1.03	17.5
Trichoderma	3	0.55	4.6	19	0.85	25.6
Papulospora	2	0.37	2.3	18	0.81	16.2
Rhizopus	5	0.74	6.9	17	0.76	13.5
Actinomucor	32	5.94	23.2	16	0.72	12.1
Myrothecium	—	—	—	14	0.63	12.1
Penicillium	6	1.11	4.6	14	0.63	13.5
Stilbella	7	1.30	11.6	12	0.54	9.4
Helminthosporium	4	0.74	9.3	9	0.40	12.1
Sordaria	2	0.37	2.3	8	0.36	8.1
Cladosporium	3	0.55	6.9	4	0.18	9.4
Dark steril mycel	2	0.37	2.3	4	0.18	2.7
Doratomyces	1	0.18	2.3	2	0.09	5.4
Stemphyllum	4	0.74	6.9	2	0.09	2.7
Stachybotris	—	—	—	1	0.04	1.3
Beauveria	—	—	—	1	0.04	1.3
Acremoniella	—	—	—	1	0.04	1.3
Volutina	—	—	—	1	0.04	1.3
Basidiomycet	—	—	—	1	0.04	1.3
Epicoccum	—	—	—	1	0.04	1.3
Monocrosporium	—	—	—	1	0.04	1.3
Chlamydomyces	—	—	—	1	0.04	1.3
Ulocladium	3	0.55	6.9	—	—	—
Trichothecium	2	0.37	2.3	—	—	—
Nigrospora	1	0.18	2.3	—	—	—
Phoma	1	0.18	2.3	—	—	—
Verticillium	1	0.18	2.3	—	—	—
Heterosporium	1	0.18	2.3	—	—	—
Cephalosporium	1	0.18	2.3	—	—	—

cies were more common in hot soils than cold soils. These two fungus were obtained commonly in the second survey which the soil temperature was higher (31,2 °C) than during the first survey.

Among these fungi *Stilbella*, *Monocrosporium* and *Volutina* are new genera and *Acremoniella atra* Sacc. and *Beaveria alba* are new species for Turkish mycoflora (Fig. 1,2,3).

B — Pathogenicity tests :

In pathogenicity tests, genera of *Fusarium*, *Rhizoctonia*, *Pythium*, *Sclerotium*, *Alternaria*, *Verticillium*, *Macrophomina*, *Cladorrhinum*, *Acremoniella*, *Nigrospora*, *Stilbella*, *Melanospora*, *Sordaria* and *Actinomyces* caused various degree of discoloration and decay of the roots and drying and death on above ground soil parts of the plants.

According to the results of the pathogenicity tests the genera of fungi showed wide ange of differences in their pathogenicities. The isolates of *Sclerotium*, *Fusarium*, *Macrophomina*, *Stilbella*, *Cladorrhinum*, *Rhizoctonia*, *Nigrospora*, *Melanospora*, *Verticillium*, *Alternaria*, *Sordaria*, *Pythium*, and *Actinomyces* were shown % 52, 48, 48, 44, 40, 40, 36, 36, 32, 20, 16, 16, 12 and 12 pathogenicity respectively. The isolates of *Pythium*, *Alternaria*, and *Verticillium* has shown slight pathogenicity. According to HENDRIX and CAMPBELL (1973) not all of the isolates

of *Pythium* species are pathogenic on plants. *Verticillium* and *Alternaria* species are more important on latter stage than seedling stage on carnations (GUBA, 1945; SCHMIDT, 1952; HELMERS, 1960; ROBINSON, 1961). There were no report about the pathogenicity of *Nigrospora*, *Melanospora*, *Cladorrhinum*, *Acremoniella* and *Stilbella* on carnation seedlings in literature. But they were found pathogenic in our tests. However *Melanosporia* and *Nigrospora* may caused lesions and decline on some plant roots (VIENNOT - BOURGIN, 1949; DICKSON, 1956; TARR, 1963). Also *Cladorrhinum foecodissimum* caused necrosis on pea roots in vitro conditions (DOMSCH und GAMS, 1970).

Although we used some antagonistic fungi according to the various report (WILSON, 1955; CATANI and PETERSON, 1963) generally the incidence of the disease increased when the tested pathogens were combined with saprophytic fungi (Fig.4). These results were shown that the saprophytic fungi have not antagonistic effect on pathogenic genera in this study. It depends on the various causes. However, antagonism were developed easily on artificial medium between two organisms (GAUMANN, 1950) it can only take place under some favourable conditions in soil. Such as pH, soil temperature humidity, organic matter content, inoculum potential of the antagonistic organisms and persistence of their toxic substances (WAKSMAN 1952, DEVAY

1956). Also some saprophytic fungi may actually be pathogenic under certain conditions or combinations. On the other hand saprophytic fungi may help penetration of pathogen by increasing the sensitivity of the host plant through certain biochemical activities

C — Chemical control tests :

The results of the chemical tests were not satisfactory. The effective-

ness of Methyl bromide, Orthocide soil treater, Banlate and Dexonal were 57,7 %, 43,3 %, 25,1 % and 21,2 % respectively.

Different effectiveness were recorded from the same chemicals used in our studies according to the genus of the fungi and the inoculum density of the same species (DROSIHN et al., 1968; TORGESON, 1969).

These findings were also confirmed by our results.

Ö Z E T

İZMİR İLİ ÇİÇEK SERALARINDA KARANFİL FİDELERİNDE ÇÖKERTEN HASTALIĞINA SEBEB OLAN FUNGAL ETMENLERİN SAPTANMASI, PATOJENİSİTELERİ VE İLAÇLI SAVAŞ YOLLARI ÜZERİNDE ARAŞTIRMALAR

Araştırma, 1972 ve 1974 yılları arasında İzmir ili çiçek seralarında karanfil fidelerinde çökerten hastalığına sebep olan fungal etmenleri, patojenisitelerini ve kimyasal savaş yollarını araştırmak amacıyla yapılmıştır. Çalışma sırasında 117 sera gezilmiş ve bunlardan toplam 2925 hastalıklı bitki örneği alınmıştır.

İzolasyon çalışmaları sonunda 41 fungus genusu izole edilmiştir. Bu genuslardan *Stilbella*, *Volutina*, *Monocrosporium* yurdumuz için yeni genuslar, *Acremoniella atra* ile *Beauveria alba* ise yeni türlerdir.

Patojenisite testlerine alınan *Sclerotium*, *Macrophomina*, *Fusarium* *Acremoniella*, *Cladorrhinum*, *Stilbella*, *Rhizoctonia*, *Nigrospora*, *Me-*

lanospora, *Verticillium*, *Alternaria*, *Pythium* ve *Actinomucor* genusları survey sırasında izole edilen tüm genuslar arasından bir ön test sonucu seçilmişlerdir. Bu genuslar tek başlarına ve ayrı bir karakter olarakda *Aspergillus*, *Myrothecium*, *Penicillium*, *Trichoderma* ve *Gliocladium* gibi survey sırasında en çok izole edilen saprofit genuslarla birlikte patojenisite testlerine alınmışlardır. Patojenisite testlerine alınan tüm genuslar saprofit genuslarla birlikte verildiklerinde karanfil fidelerinde oluşan kuruma oranı artmıştır.

İlaçlı savaş denemelerine patojenisite testlerinde % 30 un üzerinde patojenisite gösteren *Sclerotium*, *Rhizoctonia*, *Fusarium*, *Macrophomi-*

na, *Cladorrhinum*, *Melanospora*, *Pythium Acremoniella*, *Stilbella* ve *Actinomucor* genusları ile metil bromit, orthocide soil treater dexonal ve benlate preparatları alınmıştır.

Deneme sonuçlarına göre ilaçlar karanfii fidelerinde çökerten hastalığına karşı yüksek bir etki göstermişlerdir.

LITERATURE CITED

- ANDREUCCI, E., 1965. (Basal rot of carnation caused by *Sclerotium rolfsii* Sacc.) Riv. Ortoflorofrutic. ital., 39,7-8, pp:344-348. (R.A.M. 35 : 679).
- BAGGA, D.K., 1970. Soil fungi of the Delta of Mississippi. Soil Science 109 : 247-249.
- BORA, T., 1972. İzmir, Manisa, Muğla illerinde tütün fideliklerinde çökerten hastalığının zarar derecesi, fungal etmenlerin genuserlarının tanımı, dağılışı ve patojenisiteleri üzerinde arařtırmalar. Ege Üniversitesi Ziraat Fakültesi Yayınları No. 183. IV+82.
- CATANI S.C., J.L. PETERSON, 1963. Antagonistic effects of some soil fungi on *Verticillium* sp. isolated from maple trees. Phytopathology 53 : 872 (Abot).
- DEVAY, Y.E., 1956. Mutual relationship in fungi. Ann. Rew of microbiology, 10 :115-146.
- DICKSON, Y.G., 1956. Diseases of field crops. Second edition Mc Graw-Hill Book Comp Inc. New-York, Toronto, London 517.
- DOMSCH, K.L., W. GAMS. 1970. Pilze aus Agrarböden. Gustav Fischer verlag. Stuttgart XI+222.
- DROSIHN, U.G., B.R. STEPHAN and G.M. HOFFMANN, 1968. (Observations on soil disinfection with methyl bomidre) Z. Pflkrankh. Pfl. Path. Pfl. Sifhutz, 75(5):272-287. (R.A.M. 48: 21).
- GAUMANN, E., 1950. Principles of Plant Infections. Hafner Publishing Co. New York XVI+543.
- GUBA, E.F., 1945. Carnation wilt diseases and their control. Bull. Mass. Agric. Exp. Sta. 427. p 64, (R.A.M. 25 : 344)
- HELMERS, E., 1958. Four wilt diseases of perpetual flowering carnations in Denmark Denks. bot. Ark. 18,2 pp 0 1-1200. (R.A.M. 37 : 481).
- HENDRIX, F. F., W. A. CAMPBELL, 1973. *Pythium* as Plant pathogens. Annual Rew. of Phytopath. 11. 77-98.
- KUBIKOVA, Z., 1968. *Fusarium oxysporum* (Schlecht) et Hans. a dominant fungus species on the root-surface of woody plant seedlings. Plant and soil, 28 : 306-312.
- MENON, K.S., L.E. WILLIAMS., 1957. Effect of crop, crop residues, temperature and moisture on soil fungi Phytopathology, 47 : 559-564.
- OSMANLIOĞLU, E., 1974. Türkiye'de çiçekçilik endüstrisinin durumu ile Yalova civarındaki çiçekçilik işletmelerin ekonomik yapısı ve gelişmeleri için gerekli kredi ihtiyaçları. Yalova Bahçe Kùltürleri Arařtırma Enst. Matbaası.

INVESTIGATIONS ON THE DAMPING-OFF OF CARNATION

- PARKINSON, D. G. S., TAYLOR and R. PEARSON, 1963, Studies on fungi in the root region I. The development of fungi on young root. *Plant and soil*, 19 : 332-349.
- ROBINSON, J. A., 1961. Wilt and dieback of the carnation in New Zealand. *N. Z. J. Agric. Res.*, 4,5,6, pp. 660-666. (R.A.M. 42 : 20).
- SARIBAY, A., 1975. Investigations on the determination and pathogenicity of the fungal flora of Fruit Nursery Soils in İzmir. *The Journal of Turkish Phytopathology*, 4 : 9-12.
- SCHMIDT, T., 1952. (Carnations disease and their causes.) *Pflanzenarzt*, 5, 11 pp:5-6; 12, p. 6 (R.A.M., 32 : 190).
- TARR, S.A.J., 1962. Disease of Sorghum, Sudan grass and broom corn. *C.M.I., Kew, Surrey*, X+380.
- TORGESON, D.C., 1969. Fungicides. An advanced treatise. Vol. II. Chemistry and Physiology. Academic Press, New York and London, 742.
- TURHAN, G., 1973. Bazı sebze fidelerinin köklerinden izole edilen fungusların taksonomileri üzerinde araştırmalar. Doktora tezi (Yayınlanmamıştır.)
- VIENNOT - BOURGIN, G., 1949. Les champignons Parasites des Plantes Cultives. Masson C. Editeurs Libraries de L'Académie de Medicine, XVI+1950.
- WAKSMAN, S.A., 1952. Soil microbiology. John Wiles and Sons. Inc., New York, 356.
- WILES, A.B., 1952. A seedling inoculation technique for testing cotton varieties for resistance to *Verticillium* wilt. *Phytopathology*, 42 : 288 (Abst.).
- WILSON, K.S., 1955. The fate of *Verticillium albo atrum* R. and B., in much soils as affected by various species of fungi. *Diss. Abst.*, 14 : 1516 (R.A.M., 34 : 320).

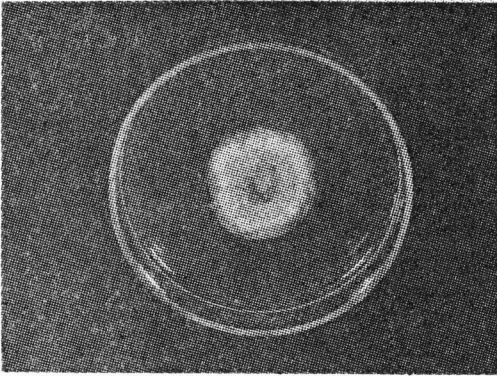


Fig. 1. The colony of *Stilbella* sp. On PDA

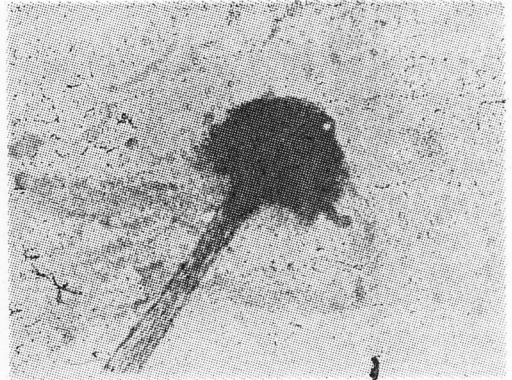


Fig. 2. The synnema of *Stilbella* sp. (appr. X 115)

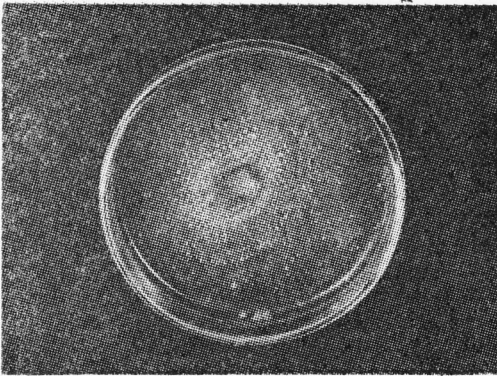


Fig. 3. The colony of *Volutina* sp. On PDA

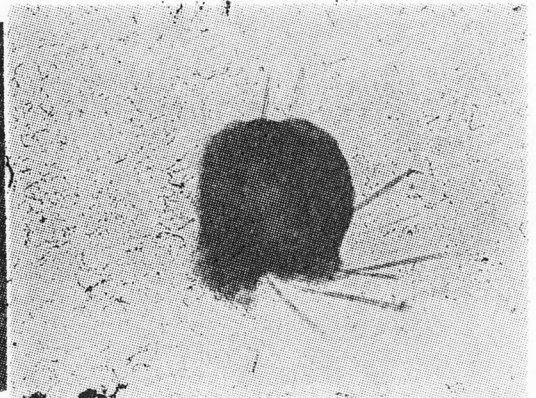


Fig. 4. The sporadachia of *Volutina* sp. (appr. X 115)

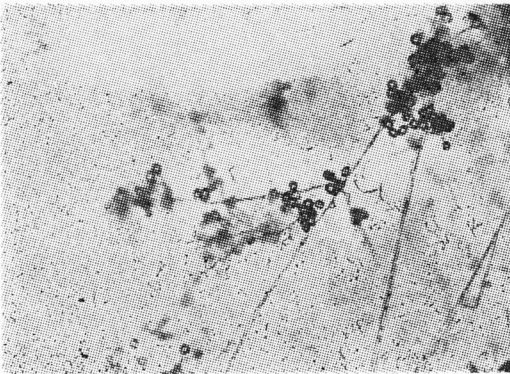


Fig. 5. The conidia of *Acremoniella atra* (appr. X 465)

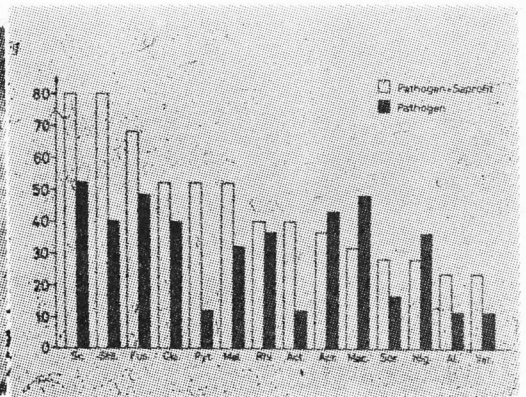


Fig. 6. The severity of disease of the pathogen and their combinations with the saprophytes.

Studies on Serology of Halo Blight (*Pseudomonas phaseolicola*) (Burkholder) (Dowson) of Beans*

Yavuz Emin ÖKTEM

Regional Plant Protection Research Institute, Ankara, TURKEY

INTRODUCTION

Bean production in Turkey shows a considerable increasing trend. In 1974 bean production was 230.000 tons pink calavance production was 60.000 tons and dried beans was 145.000 tons the yield being 145 kgs. per decar. (ANONYMUS 1974).

KARAHAN (1971) reports that *Pseudomonas phaseolicola* (Burkholder) (Dowson) exists in Turkey and the primary infections of the disease caused by infested seeds.

GUTRIE et al. (1965) declare that the causal organism of Halo Blight can easily be spread to long distances with seed, dust, rain, animals and equipment. They also add that one infested plant in 16.000 is

good enough to result in a complete crop loss under epiphytotic conditions. They also indicate that while the pathogen bacteria isolated from leaves, pods and seeds of the bean can be detected by classical bacteriological methods shorten this period. According to the same authors, rabbit or sheep can be used in producing antiserum and it can be obtained by injecting rabbit twice a week, totally 8 times, and four days after the last injection antiserum is obtained. Thus obtained antiserum can be used to detect the *P. phaseolicola* in infested seeds.

This study has been realized to enable the identification of Halo Blight by serological techniques.

* This paper was summarized from first phytopathological congress report which organized by the Turkish Phytopathological Society on 20-24 October, 1975 in İzmir.

MATERIAL AND METHODS

One year old female new Zealand rabbits are used in the course of the study. *P. phaseolicola* culture with number 52 which has been obtained from NCPPB is used as antigen. Preparation of the antigen and injections were made according to the method of GUTRIE et al. (1965) *P. phaseolicola* culture was grown on NA (Nutrient Agar) for 36 hours.

Then colonies of bacteria were taken, suspended in sterile saline and added formalin with concentration of 0.5 %. The concentration of antigen which is preparat was adjusted to the Mc. Farland tube no.3. Injections were interveinously made. In Table I the amount of the antigen and the type of the injections made are given.

Table I. Numbers of Injection and the Amount of Antigen Inject

Numbers of Injection	method	Amount
1	Interveinous	0.25 ml.
2	»	0.25 ml.
2	»	0.50 ml.
3	»	1.0 ml.
4	»	1.0 ml.
5	»	1.0 ml.
6	»	1.0 ml.
7	»	1.0 ml.
8	»	1.0 ml.

Four days after the last injection rabbit was bled and the antibody

concentration was checked with tube agglutination method.

RESULTS

Until the seventh injection, the method was strictly followed both for the amount of the antigen injected and for the recommended days. However, during the seventh injection the rabbit was subject to a shock, therefore injection were postponed

for some time. Later when rabbit recovered, injection started again. Four days after the eighth injection, antibody concentration of the blood taken from the ear of the rabbit, was determined (Table 2).

Table 2. Control of antibody concentration with tube agglutination method

Series	1/50	1/100	1/400	1/800	1/1600	1/3200	1/6400	Kontrol
Reaction	+	+	+	+	+	+	-	-

In the control of the antibody concentration by this method, results was found to be 1/3200. A very vague agglutination was found in 1/6400 which is evaluated as negative. There was no change in control tubes. Finding the result obtained satisfactory the rabbit was sacrificed. Blood taken from arteries were separated. The

antiserum thus prepared was controlled with slide agglutination method against different groups of bacteria such as *Agrobacterium tumefaciens*, *A. radiobacter*, *E. coli* and *Pseudomonas* sp. All of the above mentioned cultures gave negative reaction with antiserum. Stock antiserum is kept in deep-freeze.

DISKUSSION

The antiserum gave a strong reaction in the slide agglutination against *P. phaseolicola* with ratio 1/10 and 1/20. In the control process with different groups of bacteria the negative result achieved from the cultures apart from *P. phaseolicola* proves that the antiserum is specific thus the identification of the pathogen

can be made in a short period. However, GUTRIE et al. (1965) recommend to use the gel diffusion test and other tests for the most satisfactory result with large number of seed samples. The antiserum has been prepared for this purpose and we do believe that it will give satisfactory result when it is started to be used.

Ö Z E T

FASULYA YAPRAK YANIKLIĞI ETMENİ (*Pseudomonas phaseolicola*) (Burkholder) (Dowson)'NİN SEROLOJİSİ ÜZERİNDE ÇALIŞMALAR

Yurdumuzda taze ve kuru fasulye üretimi sürekli olarak artmaktadır. İklim koşullarının, bakteriyel hastalıkların gelişmesine uygun olduğu yıllarda üretim kayıpları meydana gelmektedir. Bu çalışma fasulye

yenin önemli hastalıklarından olan Fasulye Yaprak Yanıklığı etmeni (*Pseudomonas phaseolicola*)'nin serolojik yöntemlerle saptanması amacıyla yapılmıştır.

Çalışmada bir yaşlı Yeni Zelanda

tavşanı, antijen olarakta NCPPB'dan temin edilen *Ps. phaseolicola* kültürü kullanılmıştır. Enjeksiyonlar damardan (Interveinous) haftada iki defa uygulanmıştır.

Birinci enjeksiyonda 0.25 ml., ikincide 0.5 ml. ve diğer altı enjeksiyonda 1'er ml. antijen enjekte edilmiştir. Sekizinci enjeksiyondan dört gün sonra kulaktan kan alınıp antibady konsantrasyonu tüp aglütinasyon metoduyla 1/3200 olarak saptanmıştır. Konsantrasyon yeterli görülmüş ve tavşan kesilmiştir. Arterlerden alınan gerekli seperasyonu yapılmıştır. Elde edilen antiserum deep-freeze'de saklanmıştır.

Antiserumun bir kısmı 1/10 ve 1/20 oranında serum fizyolojikle seyreltildikten sonra lam aglütinasyon metoduyla *P. phaseolicola* kültürü kontrol edilmiş ve kuvvetli bir reaksiyon vermiştir. Buna karşılık *A. tumefaciens*, *A. radiobacter*, *E. coli* ve *pseudomonas* sp. gibi değişik gruplara ait bakterilerde aynı metodla uygulama yapılmış ve kültürlerin hiç birinde aglütinasyon görülmemiştir.

Alınan sonuçlar hazırlanmış olan antiserumun *P. phaseolicola* için spesifik olduğunu göstermektedir. Patojenin teşhisi kolaylıkla kısa süre içerisinde yapılabilmektedir.

LITERATURE CITED

- ANONYMUS, 1974. Tarım İstatistikleri Özeti. Devlet İstatistikleri Enstitüsü Matbaası-Ankara.
- GUTRIE, J.W., D.M. HUBER, and H.S. FENWICK, 1965. Serological detection of halo blight. *Plant Disease Repr.* 49, 297-299.
- KARAHAN, O., 1971. Sebze hastalıkları ve mücadele usulleri. Tarım Bakanlığı Ziraat Mücadele ve Karantina Genel Müdürlüğü. Mesleki Kitaplar Serisi. Ayyıldız Matbaası Ankara.

Elektronenmikroskopische Untersuchungen über die Stolburkrankheit an Tomaten

Von Ülkü YORGANCI*

ZUSAMMENFASSUNG

In dieser Arbeit wurden die stolburkranken Tomatenpflanzen elektronenmikroskopisch untersucht. Diese Krankheit wurde in der Türkei vorher symptomatologisch beobachtet und berichtet.

Die Blütenknospen von Tomaten mit typischen Stolbursymptomen wurden mit Glutaraldehyd und OSO_4 fixiert, in Epon eingebettet. Die hergestellten ultradünnen Schnitte wurden mit Uranylacetat und Bleicitrat nachkontrastiert, mit dem Elektronenmikroskop aufgenommen. Im Phloem besonders in den Siebröhren von kranken Tomatenpflanzen wurden pleomorphe Gebilde beobachtet. Die Dimensionen dieser Gebilde variierten zwischen etwa 70-630 nm. Die kleineren Partikel enthalten dichtes, granuliertes Plasma. Dagegen haben die Grösseren ribosomenähnliche Strukturen im Plasma. Verschiedene Formen kommen nebeneinander in einer Siebröhre vor.

Das Auftreten von Ackerwinde (*Convolvulus arvensis*) in grossen Mengen im Plasma. Verschiedene Formen kommen nebeneinander in einer fallend.

Die Mycoplasmen sind als Erreger von manchen Pflanzenkrankheiten erst im Jahre 1967 entdeckt worden, und diese Entdeckung ist wohl eines der wichtigsten Ereignisse der letzten Jahre. Als Krankheitserreger

bei Tieren und Menschen sind die Mycoplasmen schon sehr lange bekannt. Obwohl sie aber überall zu finden sind, (d.h. dass sie ubiquiter sind) wurden die Veröffentlichungen von Doi et al. (1967), die Mycoplas-

*) Institut für Phytopathologie und landwirtschaftliche Botanik, Ege Universität, Bornova/İZMİR

Als Vortrag gehalten im I. Phytopathologischen Kongress, 20-24 Oktober, 1975 — İZMİR.

men können auch Krankheiten an Pflanzen verursachen, mit grosser Überraschung aufgenommen. Diese Autoren stellten die Mycoplasmen im Phloem von kranken Pflanzen elektronenmikroskopisch fest. Später folgten weitere Arbeiten über andere Pflanzenkrankheiten, bei denen die Mycoplasmen als Erreger eine Rolle spielen.

An Tomaten gibt es eine Krankheit die als Stolbur oder big bud bekannt ist und im späteren Stadium mit grossen, unfruchtbaren Blütenknospen ins Auge springt. Als Ursache dieser Krankheit wurde bis vor einigen Jahren ein Stamm der Falschblütigkeit von *Vaccinium* (*Vaccinium false blossom virus*) angenommen.

Aber in den letzten Jahren berichteten verschiedene Autoren darüber, die Erreger der Stolburkrankheit seien mycoplasmaähnliche Gebilde, (Gianotti et al., 1968; Bor-

ges V. de Lourdes, 1969; Bowyer et al., 1969; Marchoux et al., 1969; Panjan et al., 1969; Granett et Provvidenti, 1974), was wurde das von diesen Autoren elektronenmikroskopisch festgestellt .

In der Türkei wurde über das Vorhandensein dieser Krankheit an Tomaten von Tanrikut (1953) und Tekinel (1973) und an Kartoffeln von Sahtiyancı (1966) symptomatologisch berichtet.

Für die Feststellung der Stolburkrankheit wurden die Tomatenpflanzen im August kontrolliert. Die Symptome stimmten mit denen, die von Klinkowski et Uschdraweit (1968) gegeben wurden, völlig überein. Die Blätter von infizierten Pflanzen waren löffelförmig verbeult. Ihre Nerven und der Rand der Blattunterseiten waren rötlich. Die Blütenknospen standen aufrecht. Die Kelchsegmente



Abb 1. Die Symptome an den Blütenknospen der mit Stolbur infizierten Tomatenpflanze

waren ganz oder teilweise verwachsen der Kelch war blasenförmig erweitert und viel grösser als normal. Dagegen waren Petalen und die Stamina sehr klein und teilweise grün. Das Gynaceum war häufig vergrössert, vor allem war das Gynophor ausserordentlich verlängert.

In dieser Arbeit wurden im August kleine Stücke von den Basalregionen der Blütenknospen und Kelchblätter ausgeschnitten. Die Fixierung erfolgte in Glutaraldehyd und die Nachfixierung in 1 % igem OSO_4 . Die Gewebestücke wurden nach Luft

in Epon eingebettet (Köktürk, 1967) Die ultradünnen Schnitte wurden mit dem Reichertultramikrotom OmU, hergestellt, mit Uranylacetat und Bleicitrat (Reynolds, 1963) nachkontrastiert und mit dem Elektronenmikroskop der Fa. Zeiss 9a aufgenommen.

Besonders in Schnitten von den Basalregionen der vergrösserten Blüten enthielten einige Zellen im Phloem vorwiegend in den Siebröhren, pleomorphe Gebilde. Die Dimensionen dieser Gebilde variierten zwischen etwa 70-630 nm. Die kleineren

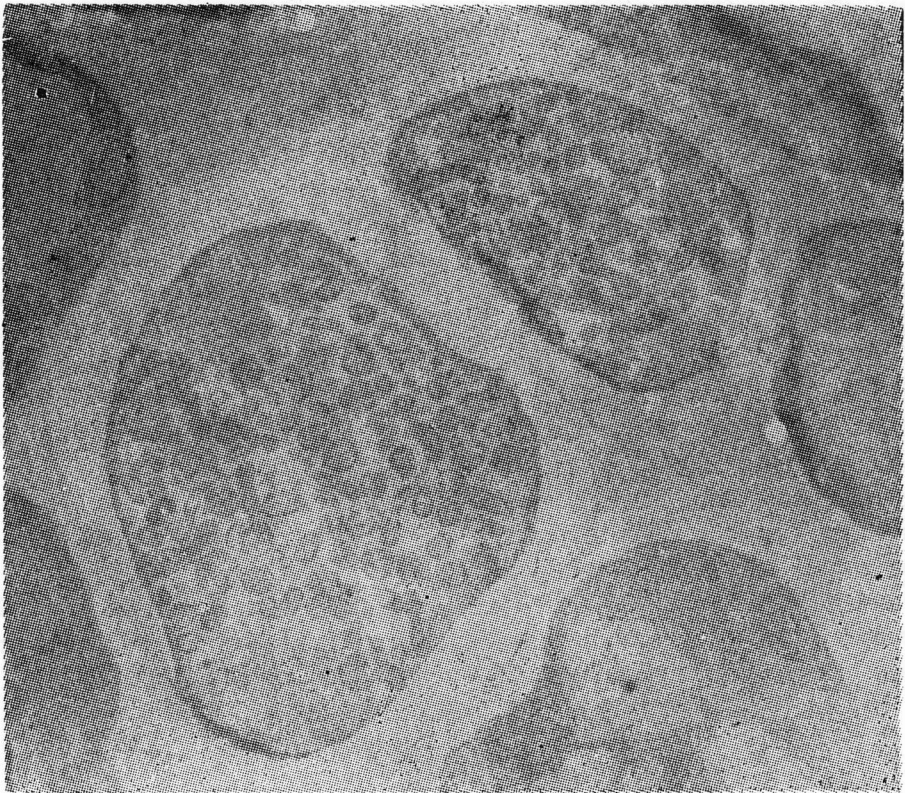


Abb 2. Pleomorph mycoplasmaähnliche Gebilde in den Siebröhren von Tomatepflanzen nach Infektion mit Stolburkrankheit

Partikel enthalten dichtes, granuliertes Plasma. Dagegen haben die grösseren ribosomenähnliche Strukturen im Plasma. Verschiedene Formen kommen nebeneinander in einer Siebröhre vor (Abb. 2).

Sinha et Paliwal beschrieben die kleineren, dichten Gebilde als Elementarkörperchen, die Grösseren, mit fädigen Inneren als reife Zellen.

Das Auftreten von Ackerwinde (*Convolvulus arvensis*) in grossen Massen in Tomatenfeldern mit typischen Stolbursymptomen ist sehr auffallend. Ackerwinde ist in der Li-

teratur als Wirtzpflanze und Krankheitsreservoir von Stolbur angegeben. Ausserdem war *Cuscuta campestris*, die auch als Vektor bekannt ist, auf einem Tomatenfeld in Muradiye sehr verbreitet.

Diese Arbeit wurde technisch und finanziell vom Zentrum für Elektronenmikroskopie und vom Institut für Morphologie der Medizinischen Fakultät der Ege Universität unterstützt. Herrn Prof. Dr. E. Cireli und Fräulein Ş. Kürşat sei an dieser Stelle herzlich gedankt.

Ö Z E T

DOMATESLERDE STOLBUR HASTALIĞI ÜZERİNDE ELEKTRONMİKROSKOBİK İNCELEMELER

Bu çalışmada daha önce Türkiye'de simptomatolojik olarak gözlenmiş olan Stolbur hastalığı elektronmikroskopik olarak incelenmiştir.

Tipik Stolbur belirtisi gösteren domates çiçek tomurcukları glutaldehyit ve Osmium tetroksit'le fikse edildikten sonra Epon'a gömülmüştür. Hazırlanan ince kesitler uranylasetat ve kurşun sitratla boyandıktan sonra elektronmikroskopunda incelenmişlerdir. Stolburla infekteli domates bitkilerinin kalbur borularında çok sayıda, büyüklükleri 70 -

630 nm arasında değişen mikroplazma benzeri pleomorf cisimcikler gözlenmiştir. Küçük partiküller koyu, granüle plazma içermektedir. Daha büyük olanlarda, plazmada ribozom benzeri yapılar bulunur. Çeşitli şekillere bir kalbur borusunda yanyana rastlanabilir.

Stolburla bulaşık tarlalarda çok miktarda literatürde konukçu ve hastalık kaynağı olarak bildirilen *Convolvulus arvensis* (tarla sarmaşığı) bulunması ilginçtir.

LİTERATÜR

- BORGES, V. de LORES, Maria, 1969. La Mal Azul' de la tomate au Portugal et ses relations avec *Mycoplasma* sp. Ann. de Phytopathol. 1, Deuxième Congrès de l'union Phytopathologique Méditerranéenne, 443
- BOWYER, J.W.; J.G. ATHERTON; D.S. TEAKLE; and Gabrielle. A. AHERN. 1969. Mycoplasma - Like bodies in plants affected by legume little leaf, tomato big bud, and lucerne witches broom diseases. Austr. J. biol. Sci. 22. 271-274.
- DOI, Y.; M. TERANAKA; K. YORA; and H. ASUYAMA, 1967. Mycoplasma - or PLT group-like microorganisms found in the phloem elements of plants infected with mulberry dwarf, potato witches broom, aster yellows, or Paulownia witches broom. Ann. Phytopath. Soc. Japan, 33, 259-266.
- GIANOTTI, J.; G. MARCHOU; C. VAGO et J.L. DUTHOIT. 1968. Micro - organismes de type mycoplasme dans les cellules libériennes de *Solanum lycopersicum* L. atteinte de Stolbur. Comp. rend. Acad. Sci. (Paris) 267, (Sér. D), 454-456.
- GRANOTT, A.L. and R. PROVVIDENTI. 1971. mato big bud in NewYork State. Plant Dis. Roptr. 58, 211-214..
- KÖKTÜRK, İ. 1967. Elektron Mikroskop ve Genel Araştırma Metodları. Ege Üniversitesi Tıp Fakültesi Yayınları, No: 66. Ege Üniversitesi Matbaası ,Bornova, pp 184
- MARCHOUX, G.; J. GIANOTTI; H. LATERROT 1969. Le Stolbur P, une nouvelle maladie de type jaunisse chez la tomate. Symptomes et examen cytologique. Ann. Phytopath. 1, 633-640.
- PANJAN, M.; A. SARIC' and M. WRISCHER. 1970. Mycoplasmaähnliche Gebilde in Tomatenpflanzen nach Infektion mit Kartoffelgelbsucht. Phytopath. Z., 69, 31-35.
- REYNOLDS, E.S., 1963. The use of lead citrate at high pH as an electron-opaque stain in electron microscopy, J. cell. Biol. 17, 208-212.
- SAHTİYANCI, Ş., 1966. Patates Stolbur-Virozu ve Türkiye'de ilk Müşahadesi. Bitki Koruma Bülteni, 6, 24-30.
- SINHA, R.C. and Y.C. PALIWAL, 1969. Association, development, and growth of Mycoplasma-like organism in plants affected with clover phyllody. Virology, 39, 759-767.
- TANRIKUT, S., 1953. Domates Yetiştiriciliği için Tehlikeli bir Hastalık. Bitki Koruma Bülteni, 5, 22-28.
- TEKİNEL, N., 1973. Adana, Antalya, Hatay ve İçel İllerinde Domates Virus Hastalıklarının Yayılış Alanlarının ve Oranlarının Tesbiti Üzerinde Araştırmalar. Bitki Koruma Bülteni 13, 107-141.

All Correspondance Should Be Made To

FİTOPATOLOJİ DERNEĞİ

Ege Üniversitesi Ziraat Fakültesi

Fitopatoloji ve Ziraat Botanik Kürsüsü

Bornova - İzmir,

TURKEY