



İSTANBUL
UNIVERSITY
PRESS

AQUATIC SCIENCES and ENGINEERING

VOLUME: 39 ISSUE: 1

2024

E-ISSN 2602-473X

Indexing and Abstracting

Web of Science - Emerging Sources Citation Index (ESCI)

SCOPUS

TÜBİTAK-ULAKBİM TR Dizin

Zoological Record

Biological Abstracts

BIOSIS Previews

CAB Abstract

SciLit

DOAJ

EBSCO Academic Source Ultimate

SOBIAD



Scopus®



Clarivate
Analytics

ZOOLOGICAL RECORD

Clarivate
Analytics

BIOLOGICAL ABSTRACTS

Clarivate
Analytics

BIOSIS PREVIEWS



Scilit

DOAJ
DIRECTORY OF
OPEN ACCESS
JOURNALS

EBSCO

SOBIAD

Owner

Prof. Dr. Melek İşinibilir Okyar
Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye

Responsible Manager

Prof. Dr. Devrim Memiş
Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye

Correspondence Address

Istanbul University Faculty of Aquatic Sciences,
Ordu Caddesi No: 8 34134 Laleli-Fatih / İstanbul, Türkiye
E-mail: ase@istanbul.edu.tr
<https://iupress.istanbul.edu.tr/tr/journal/ase/home>

Publisher

Istanbul University Press
İstanbul University Central Campus, 34452 Beyazıt,
Fatih / İstanbul, Türkiye
Phone: +90 (212) 440 00 00

Cover Photo

Cansu Saraçoğlu
E-mail: cansusaracoglu@istanbul.edu.tr

Authors bear responsibility for the content of their published articles.

The publication language of the journal is English.

This is a scholarly, international, peer-reviewed and open-access journal published quarterly in January, April, July, and October.

Publication Type: Periodical

EDITORIAL MANAGEMENT BOARD

Editor-in-Chief

Prof. Dr. Devrim Memiş

Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - mdevrim@istanbul.edu.tr

Co-Editor-in-Chief

Prof. Dr. Özkan Özden

Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - ozden@istanbul.edu.tr

Language Editors

Alan James Newson

Istanbul University, Department of Foreign Languages, İstanbul, Türkiye - alan.newson@istanbul.edu.tr

EDITORIAL BOARD

Prof. Dr. Genario Belmonte - University of Salento, Italy - genuario.belmonte@unisalento.it

Prof. Luis M. Botana - Santiago de Compostela University, Santiago-de-Compostela, Spain - luis.botana@usc.es

Prof. Dr. Carsten Harms- Applied University Bremerhaven, Germany - charms@hs-bremerhaven.de

Prof. Dr. Konstantinos Kormas - University of Thessaly, Greece - kkormas@uth.gr

Prof. Dr. Sergi Sabater - Institute of Aquatic Ecology, Spain - sergi.sabater@udg.edu

Prof. Dr. Maya Petrova Stoyneva-Gaertner - Sofia University "St Kliment Ohridski", Bulgaria - mstoyneva@abv.bg

Prof. Dr. Nuray Erkan - Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - nurerkan@istanbul.edu.tr

Prof. Dr. Reyhan Akçaalan - Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - akcaalan@istanbul.edu.tr

Prof. Dr. Firdes Saadet Karakulak - Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - karakul@istanbul.edu.tr

Assoc. Prof. Emine Gözde Özbayram - Istanbul University, Faculty of Aquatic Sciences, İstanbul, Türkiye - gozde.ozbayram@istanbul.edu.tr

Prof. Dr. Lukas Kalous Czech - University of Life Sciences, Czech - kalous@af.czu.cz

Dr. Pierantonio Addis - University of Cagliari, Italy - addisp@unica.it

Dr. Nico Salmaso - Research and Innovation Centre, Italy - nico.salmaso@fmach.it

Dr. Petra Visser - University of Amsterdam, Netherlands - P.M.Visser@uva.nl

Aims and Scope

Aquatic Sciences and Engineering is an international, scientific, open access periodical published in accordance with independent, unbiased, and double-blinded peer-review principles. The journal is the official publication of İstanbul University Faculty of Aquatic Sciences and it is published quarterly on January, April, July, and October. The publication language of the journal is English and continues publication since 1987.

Aquatic Sciences and Engineering aims to contribute to the literature by publishing manuscripts at the highest scientific level on all fields of aquatic sciences. The journal publishes original research and review articles that are prepared in accordance with the ethical guidelines.

The scope of the journal includes but not limited to; aquaculture science, aquaculture diseases, feeds, and genetics, ecological interactions, sustainable systems, fisheries development, fisheries science, fishery hydrography, aquatic ecosystem, fisheries management, fishery biology, wild fisheries, ocean fisheries, biology, taxonomy, stock identification, functional morphology freshwater, brackish and marine environment, marine biology, water conservation and sustainability, inland waters protection and management, seafood technology and safety.

The target audience of the journal includes specialists and professionals working and interested in all disciplines of aquatic sciences.

The editorial and publication processes of the journal are shaped in accordance with the guidelines of the International Committee of Medical Journal Editors (ICMJE), World Association of Medical Editors (WAME), Council of Science Editors (CSE), Committee on Publication Ethics (COPE), European Association of Science Editors (EASE), and National Information Standards Organization (NISO). The journal is in conformity with the Principles of Transparency and Best Practice in Scholarly Publishing (doaj.org/bestpractice).

Aquatic Sciences and Engineering is covered in Clarivate Analytics Web of Science Emerging Sources Citation Index

(ESCI), Clarivate Analytics Zoological Record, Biological Abstracts, BIOSIS Previews, Scopus, DOAJ, Scilit, TUBITAK ULAKBIM TR Index and CAB Abstracts.

Processing and publication are free of charge with the journal. No fees are requested from the authors at any point throughout the evaluation and publication process. All manuscripts must be submitted via the online submission system, which is available at <https://iupress.istanbul.edu.tr/en/journal/ase/home>. The journal guidelines, technical information, and the required forms are available on the journal's web page.

All expenses of the journal are covered by the İstanbul University Faculty of Aquatic Sciences. Potential advertisers should contact the Editorial Office. Advertisement images are published only upon the Editor-in-Chief's approval.

Statements or opinions expressed in the manuscripts published in the journal reflect the views of the author(s) and not the opinions of the İstanbul University Faculty of Aquatic Sciences, editors, editorial board, and/or publisher; the editors, editorial board, and publisher disclaim any responsibility or liability for such materials.

All published content is available online, free of charge at <https://iupress.istanbul.edu.tr/en/journal/ase/home>. Printed copies of the journal are distributed to the members of the İstanbul University Faculty of Aquatic Sciences, free of charge.

İstanbul University Faculty of Aquatic Sciences holds the international copyright of all the content published in the journal.

OPEN  ACCESS

Editor in Chief: Prof. Dr. Devrim MEMİŞ

Address: İstanbul Üniversitesi Su Bilimleri Fakültesi Yetiştiricilik Anabilim Dalı Kalenderhane Mah. 16 Mart Şehitleri caddesi No: 2 Vezneciler / İstanbul, Türkiye

Phone: +90 212 4555700/16448

Fax: +90 212 5140379

E-mail: mdevrim@istanbul.edu.tr

Contents/İçindekiler

Research Articles

Evaluation of Food Wastes in <i>Chlorella vulgaris</i> Cultivation for Remazol Brilliant Blue R Biosorption <i>Safiye Büşra Nazlı, Nazlıhan Tekin, Sevgi Ertuğrul Karatay, Gönül Sönmez</i>	1
Investigation of Growth Performance, Proximate and Fatty Acid Composition of Freshwater (<i>Euglena gracilis</i>, <i>Chlorella vulgaris</i>) and Marine (<i>Pavlova lutheri</i>, <i>Diacronema vlnanium</i>) Microalgae <i>Merve Sayar, Kamil Mert Eryalçın</i>	8
Growth Performance, Survival Rate, and Water Quality in an Aquaculture System Using Different Feeding Strategies for Juveniles of Nile Tilapia (<i>Oreochromis niloticus</i>) <i>Dagon Ribeiro, Emerson de Carvalho, Gustavo Fonseca</i>	17
The Relationship of Bristle Worm, <i>Protodorvillea kefersteini</i> (McIntosh, 1869) (Eunicida, Dorvilleidae) Abundance with Environmental Variables in Çardak Lagoon (Turkish Straits) Exposed to domestic Discharge <i>Ertan Dağlı, A. Suat Ateş, Seçil Acar, Yeşim Büyükkateş</i>	24
Validity of Visible Ectoparasite Intensity As a Non-invasive Biomarker for Fish welfare: Parasitic Copepod, <i>Lernantropus kroyeri</i> in Sea Bass As an Example <i>Hijran Yavuzcan</i>	30
Length – Weight Relationships, Meat Yield and Morphometric Indices of Five Commercial Bivalve Species Collected from the Çanakkale Strait (Türkiye) <i>Serhat Çolakoğlu, Fatma Çolakoğlu, İbrahim Ender Künili</i>	36
Records of Three Immature Gelatinous Specimens for the Turkish Mediterranean Coast with an Emphasis on Alternative Pathways <i>Erhan Mutlu, Doğukan Karaca</i>	43
Lakes of Turkey: Comprehensive Review of Lake Çıldır <i>Özgür Eren Zariç, Abuzer Çelekli, Sidar Yaygır</i>	54

Evaluation of Food Wastes in *Chlorella vulgaris* Cultivation for Remazol Brilliant Blue R Biosorption

Safiye Büşra Nazlı¹ , Nazlıhan Tekin¹ , Sevgi Ertuğrul Karatay¹ , Gönül Dönmez¹ 

Cite this article as: Nazlı, S.B., Tekin, N. Ertugrul Karatay, E., & Donmez, G. (2024). Evaluation of food wastes in *Chlorella vulgaris* cultivation for remazol brilliant blue R biosorption. *Aquatic Sciences and Engineering*, 39(1), 1-7. DOI: <https://doi.org/10.26650/ASE20241305101>

ABSTRACT

The current study demonstrates the biosorption efficiency of *Chlorella vulgaris* for the removal of Remazol Brilliant Blue R (RBBR), which is often used in the textile industry. For this, optimization of microalgal growth was investigated under photoautotrophic conditions including only BG-11 medium and photoheterotrophic conditions containing 0.5 g/L of pumpkin waste, apple pomace, or glucose. Some critical parameters for RBBR biosorption onto dry *C. vulgaris* biomass, such as pH (2-10), initial concentration of RBBR (100-800 mg/L), biosorbent concentration (1-3 g/L), and biosorption time (0-120 min) were optimized. As a result of the study, the best growth of microalgae was determined as 0.502 g/L under photoheterotrophic cultivation condition, including 0.5 g/L of pumpkin waste sugar. The highest dye removal was calculated as 99.49% in the presence of 3 g/L microalgal biosorbent and 103.38 mg/L RBBR concentration at pH 4. These results indicate that *C. vulgaris* has a promising biosorbent for waste management and dye removal.

Keywords: *Chlorella vulgaris*, agro-industrial waste, dye removal, biosorption, photoheterotrophic growth

INTRODUCTION

The growing textile industry and the use of synthetic dyes are the primary causes of environmental pollution. Moreover, water pollution caused by dyeing processes in the textile industry is approximately 17-20% (Premaratne et al., 2021). Synthetic dyes, defined as highly polluting, have toxic (cytotoxic, genotoxic, and mutagenic) effects (Verma, 2021). Up to 50% of dyes are used in different areas, such as skin and clothing, do not bind to the cloth's fibers, and are mixed into the aquatic environment as pollutants (Benkhaya, M'rabet & El Harfi, 2020). Therefore, the dyes used in the textile industry threaten the living ecosystem due to them mixing with natural water. Low amounts of dye in water can even significantly affect the photosynthetic activity of plants and toxicity for animals (Verma, 2021). Azo dyes constitute the biggest class (up to 60%) of textile industry

dyes. Of these dyes, 15-50% of them are mixed with effluent because they can not remain fixed into the product (Al-Tohamy et al., 2022). For example, Remazol Brilliant Blue R (RBBR), an anionic-azo dye utilized in polymeric material production and the textile industry, is quite dangerous for aquatic organisms due to its low biological disjunction and high toxicity (Araçagök, 2022).

For many years, chemical (electrochemical etc.), physical (coagulation etc.), and biological (enzyme or microorganisms) methods have been utilized to remove these dangerous dyes from natural life (Shabir et al., 2022). Among the methods used, biosorption from biological methods is a cheap, effective, and successful mechanism. Biosorption is a passive process, without the need for energy, in which xenobiotic chemicals from contaminated sources are removed by microbial biomass such as bacteria,

ORCID IDs of the author:
S.B.N. 0009-0000-5149-6067;
N.T. 0000-0003-1922-594X;
S.E.K. 0000-0001-9544-0276;
G.D. 000-0001-7972-5570

¹Ankara University, Science Faculty,
Biology Department, Beşevler,
Ankara, Türkiye

Submitted:
29.05.2023

Revision Requested:
11.08.2023

Last Revision Received:
23.08.2023

Accepted:
17.09.2023

Online Published:
00.00.0000

Correspondence:
Sevgi Ertuğrul Karatay
E-mail:
sertugrul@ankara.edu.tr



yeast, and algae (Gadd, 2009). Furthermore, the microbial biomass used in biosorption can be active (live) or inactive (dead) character. However, the use of dead microbial biomass is more effective than live biomass because of its features, such as reusability, high surface-to-volume ratio, storage convenience, metabolic independence, economic efficiency, and environmental friendliness (Goud et al., 2020). The biosorption mechanism occurs as a result of the interaction of azo dyes and different functional groups (such as carboxyl, hydroxyl, and amino) on the microbial cell surface. Among microorganisms used, microalgae can live adaptively in an environment contaminated with textile dyes since they are aquatic organisms. Phosphorus and nitrogen compounds in dyes can be used as nutrient sources for microalgae growth (Zohoorian et al., 2020). It is also indicated in the literature that it is a good biosorbent with its high binding affinity and high surface-to-volume ratio for dyes because of its functional groups (carboxyl, sulfhydryl, etc.) on the cell wall surface (Chu & Phang, 2019).

Furthermore, biosorption with microalgal biomass has a low carbon footprint profile due to its photosynthetic properties (Mustafa et al., 2021). Thus, it can be said that using microalgae in industrial dye removal is quite advantageous. *Chlorella vulgaris* is among the microorganisms frequently used in the literature because it has a high affinity for removing heavy metals (Joo, Lee & Choi, 2021) and dyes (Aksu & Tezer, 2005). Even though they typically grow photoautotrophically, microalgae can be cultivated in heterotrophic, mixotrophic, or photoheterotrophic conditions containing a carbon source (Saratale et al., 2022). In addition, in the literature, it is indicated that microalgae growth and yield significantly were supported in photoheterotrophic conditions using organic carbon and a light source. For microalgal growth, industrial and agricultural wastes with rich sugar content are often utilized in photoheterotrophic conditions as a carbon source (Isleten-Hosoglu et al., 2013). Apple pomace (AP) and pumpkin waste (PW) are important by-products of the food industry and are abundant in carbohydrates, protein, lipids, minerals, vitamins, and polyphenols. While pumpkin production is about 23 million tons worldwide, 93,144,358.17 tons of apples were generated in 2021 worldwide (FAOSTAT, 2021; Kido & Uwineza, 2022). Therefore, AP and PW are promising carbon sources for microalgae production.

Moreover, the contents and surface structures of microbial cells may be associated with the growth conditions (Joo et al., 2021). In addition, it is emphasized that *C. vulgaris* cells change the efficiency of their contents (macro (protein, carbohydrates, lipids etc.) and micro (phosphorus etc.) compounds) under different growth conditions (Joo et al., 2021). In this context, as a result of the growth of *C. vulgaris* under photoheterotrophic conditions, it can be expected to obtain high biomass and have a more effective cell surface for biosorption.

This work aimed to evaluate the biosorption of anionic RBBR dye onto dead *C. vulgaris* biomass produced in photoheterotrophic conditions in the presence of AP or PW. For this aim, carbon source and its concentrations were optimized for *C. vulgaris* growth. Using microalgal biosorbent cultivated at optimized photoheterotrophic conditions, important parameters such as

pH, initial dye concentration, biosorbent concentration, and time were optimized for RBBR biosorption. Thus, the present study aims to suggest an effective, eco-friendly, and cost-efficient biosorbent cultivated with AP or PW sugar in photoheterotrophic conditions for anionic dye biosorption to the literature for the first time.

MATERIAL AND METHODS

Microorganism and its cultivation conditions

C. vulgaris microalgae was supplied from the Ankara University culture collection. Microalgae stock culture was cultivated at 30°C with 2400 lx light in sterile BG-11 medium that contain of 1.5 g NaNO₃, 6 mg ferric ammonium citrate, 40 mg K₂HPO₄·3H₂O, 6 mg citric acid H₂O, 1 mg Na₂EDTA·2H₂O, 75 mg MgSO₄·7H₂O, 2.86 mg H₃BO₃, 20 mg Na₂CO₃, 1.81 mg MnCl₂·4H₂O, 0.22 mg ZnSO₄·7H₂O, 0.39 mg Na₂MoO₄·2H₂O, 0.049 mg Co(NO₃)₂·6H₂O, 0.08 mg CuSO₄·5H₂O, and 36 mg CaCl₂·2H₂O per liter (Rippka, 1988; Park et al., 2014). The experiments were carried out in photoautotrophic and photoheterotrophic conditions for 10 days. In photoautotrophic conditions, *C. vulgaris* was cultivated in the standard BG-11 medium under constant illumination. For photoheterotrophic condition, microalgae cultivation was carried out in BG-11 medium adding 0.5 g/L glucose, 0.5 g/L AP sugar, or 0.5 g/L PW sugar with constant illumination in the first part of the study. Furthermore, in further experiments, photoheterotrophic cultivation was carried out at different sugar concentrations of PW, the waste from which the highest microalgal biomass was obtained.

Pretreatment of agro-industrial wastes

PW and AP as carbon sources for photoheterotrophic conditions were provided by a local company in Sakarya and Aroma Bursa Fruit Juices and Food Industry, Turkey, respectively. Agro-industrial wastes were dried at 80°C in an oven, ground in a laboratory-type mill (Miprolab, Turkey), and stored at room temperature.

Before experiments, dried and ground wastes were pretreated with 1% H₂SO₄ by dilute acid pretreatment method. The solution was autoclaved for 15 min at 121°C to obtain monomeric sugars (Germec & Turhan, 2018). Hydrolyzates were filtered with Whatman No. 1 filter paper, and the solutions were stored at +4°C.

The effect of initial PW sugar concentration on microalgal growth

Different concentrations of PW sugar (0.25-1 g/L) were investigated for the growth of *C. vulgaris* under photoheterotrophic condition. For this, 10% (v/v) of microalgae culture was added into 250 mL flasks including 100 mL of BG-11 media containing desired PW sugar concentrations and cultivated at 2400 lx and 30°C for 10 days.

Preparation of microalgal biosorbent and stock dye solutions for biosorption

To obtain biosorbent, microalgae culture photoheterotrophically cultivated in the presence of PW sugar was harvested at 5000 rpm for 10 min by centrifugation and dried overnight at 70 °C using an oven. For stock biosorbent solution, 10-30 g of dried microalgal biomass was homogenized in 1L of distilled water with a homogenizer (IKA T18 digital Ultra Turrax) for 45 seconds at 13400 rpm.

To prepare the RBBR stock solution, 20g of dye was dissolved in 1L of distilled water and the solution was diluted to arrive desired RBBR (100-800 mg/L) concentration in experiments.

Biosorption studies

The experiments for biosorption were actualized in 150 mL flasks including 50 mL of RBBR dye solution. The flasks containing desired concentrations of dye and biosorbent were shaken at 100 rpm for 120 min. The samples taken at definite minutes were centrifuged for 4 min at 5000 rpm. After centrifugation, the supernatants were spectrophotometrically measured to determine the dye removal efficiency.

The effect of pH on RBBR biosorption

The pH effect was analyzed in the range of 2 to 10 in the dye solution containing about 100 mg/L of RBBR with 1 g/L microalgal biosorbent. The pH of dye solutions was brought to the determined value with 0.1 M NaOH or 0.1 M H₂SO₄ before mixing the microalgal biosorbent.

The effect of initial RBBR concentration on biosorption

To determine the effect of increasing dye concentrations, biosorption experiments were performed approximately from 100 to 800 mg/L (from 103.38 mg/L to 818.30 mg/L) RBBR at optimum pH value. The flasks containing dye solutions were shaken at 100 rpm for 120 min.

The effect of biosorbent concentration and time on RBBR biosorption

1, 2 and 3 g/L of biosorbent were tested for all dye concentrations at optimum pH value. In addition, during all the biosorption experiments, the samples were taken at 0, 5, 15, 30, and 120 minutes and spectrophotometrically analyzed.

Analytical methods

Reducing sugar concentration in agro-industrial wastes was analyzed according to Dinitrosalicylic acid Method (Miller, 1959) and spectrophotometrically measured at 540 nm using a Shimadzu UV-1201 model UV-VIS spectrophotometer.

Microalgal growth in different conditions was spectrophotometrically analyzed at 600 nm using a Shimadzu UV-1201 model UV-VIS spectrophotometer. Absorbance measurements for the concentration of RBBR in the dye solutions were spectrophotometrically carried out at 590 nm.

The biosorption percentage (%) and removal capacity (q_m) of RBBR were calculated using the equations below (Eq. 1 and Eq. 2) (Gül, 2022):

$$\text{Biosorption}(\%) = \frac{C_o - C_f}{C_o} \times 100 \quad (\text{Eq.1})$$

$$q_m(\text{mg/g}) = \frac{C_o - C_f}{X_m} \quad (\text{Eq.2})$$

Where C_o shows the initial RBBR concentration in solution (mg/L), C_f defines the final concentration of RBBR at any time (mg/L), and X_m shows the microalgal biosorbent concentration (g/L).

RESULT AND DISCUSSION

Microalgal growth in different cultivation conditions

Macro and microelements such as carbon, protein, mineral, and vitamins are important for microalgae production because they affect microbial metabolic activities. In this sense, agro-industrial wastes are unique sources containing substances required by microalgae. Therefore, the current study determined the effect of different cultivation conditions for the growth of microalgae with BG-11 media adding 0.5 g/L of AP, PW, or glucose. Figure 1 shows the growth of *C. vulgaris* in photoautotrophic (BG-11) and photoheterotrophic (agro-industrial wastes or glucose) condition. As shown in Figure 1, the highest microalgal biomass was 0.502 g/L in medium including 0.5 g/L sugar of PW after 10-days. According to this, the supplementation of PW sugar increased microalgal growth 2.3 times compared with photoautotrophic cultivation. The BG-11 medium containing PW sugar caused higher biomass production than the BG-11 medium containing synthetic glucose. This increase may be due to the fact that the pumpkin waste has compounds that support microbial growth, such as minerals, vitamins, protein and lipid, in addition to its sugar content (Valdez-Arjona & Ramírez-Mella, 2019). While proteins are important sources of nitrogen in cell growth (Wang et al., 2016), vitamins and minerals can participate as coenzymes or cofactors for enzymes involved in microalgal metabolism pathways such as carbon and lipid metabolism (Golub and Voyevoda, 2013). Thus, despite the same sugar concentration, microalgal growth was significantly increased thanks to the rich nutritional content of the pumpkin waste.

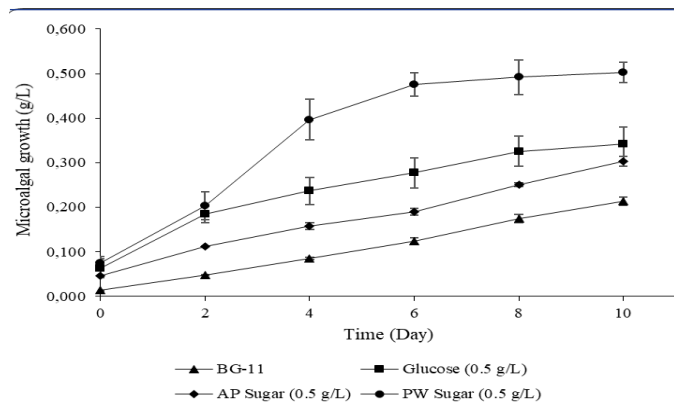


Figure 1. The effects of photoautotrophic and photoheterotrophic conditions on *C. vulgaris* growth (BG-11 media; 2400 lx; 30 °C; pH 7; 10 days).

Moreover, AP sugar supported microalgae production but was less effective than glucose. This situation may be due to the sugar and composition of apple pomace. Furthermore, 0.25 - 1 g/L of PW sugar was investigated in BG-11 Medium to determine the optimum PW sugar concentration and the results were demonstrated in Figure 2. According to the obtained results, when sugar concentration increased from 0.25 to 0.5 g/L, biomass production significantly enhanced and reached from 0.357 g/L to 0.502 g/L. Moreover, 1 g/L PW sugar caused a slight decrease in mi-

coalgal growth. This decline can be explained by the fact that high organic matter content causes a decrease in biomass productivity (Manzoor et al., 2020).

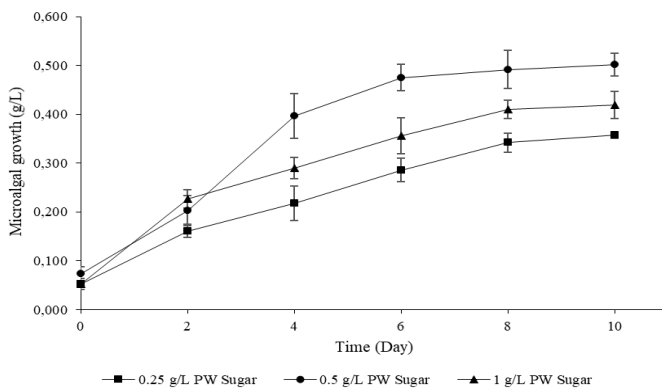


Figure 2. The effects of different PW sugar concentrations on *C. vulgaris* growth (BG-11 medium; 2400 lx; 30 °C; pH 7; 10 days).

In previous studies, the addition of various carbon sources has been shown to increase microalgal biomass production. For instance, Kassim et al. (2022) determined that the highest biomass of *Tetraselmis suecica* and *Halochlorella rubescens* was 0.669 ± 0.01 g/L and 0.653 ± 0.009 g/L, respectively, when molasses were used in photoheterotrophic conditions. Mohammad Mirzaie et al. (2016) observed that under the mixotrophic conditions using cane molasses and corn steep liquor for carbon and nitrogen sources, the dry weight of *C. vulgaris* was 4 and 2.5 times higher than the heterotrophic and autotrophic conditions, respectively. Moreover, adding a carbon source improves microalgae growth and its valuable content, such as carbohydrates, lipids, and protein, for fields such as biofuel production and food supplementation. For example, supplementation of carbon sources in growth of different *Chlorella* species has been stated to increase lipid accumulation for biodiesel production compared to the photoautotrophic condition (Sharma et al., 2016).

Thus, according to the results obtained, for use as a biosorbent in biosorption experiments, *C. vulgaris* biomass was produced under the photoheterotrophic cultivation conditions containing 0.5 g/L PW sugar.

The effect of pH on RBBR biosorption

The pH of the solution is a critical parameter in the biosorption activity of the biosorbent, depending on whether the dyes are anionic or cationic. Therefore, in the current study, the effect of pH on RBBR biosorption was tested in the range of 2-10 in the presence of 1 g/L microalgal biosorbent and about 100 mg/L RBBR. Figure 3 shows that the pH change significantly affected the biosorption of RBBR. For this dye, which has an anionic character, the increase at pH value caused a decrease on biosorption onto *C. vulgaris* biomass, and the highest biosorption percentage was obtained at pH 4 as 93.06% at the end of 120 min (Figure 3). In addition, there was not a significant difference between pH 2 and pH 4. Higher biosorption activity at low pH values for

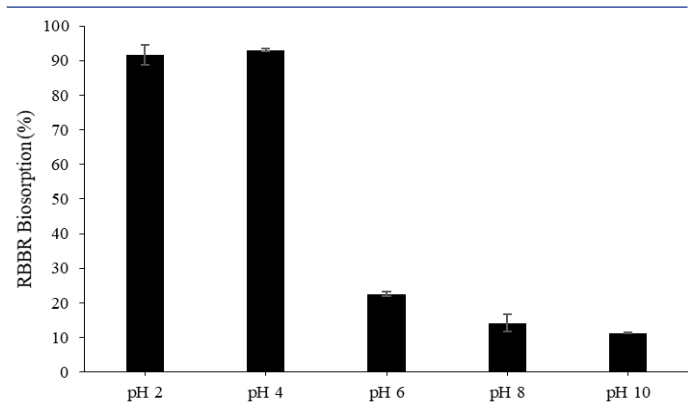


Figure 3. The effect of pH on RBBR biosorption (1 g/L biosorbent, 100 mg/L RBBR, 30 °C; 100 rpm, 120 min).

an anionic dye is due to the ionized dye molecules producing electrostatic charges. When the pH of the solution is low, the solution positively charges and the biosorbent surface also becomes protonated, and the adsorption of negative-charged dye increases (Salleh et al., 2011). In addition, the decrease in biosorption at high pH can be explained by decreasing attraction force between negative-charged cell surfaces and anionic dye molecules with the increase of negative charges in the environment (Yu et al., 2018).

Furthermore, in the studies where *C. vulgaris* was used as a biosorbent, Aksu & Tezer (2005) determined the highest biosorption capacities for Remazol Red RR, Remazol Black B and Remazol Golden Yellow at pH 2 while Kumar, Ahluwalia & Charaya (2019) obtained the maximum biosorption value at pH 5 in dye solution containing 5 ppm Orange G (an anionic dye) and 50 mg biosorbent. Similar effects of pH were shown in other studies using microalgal biosorbent. For example, Khataee, Vafaei & Jannatkah (2013) used *Spirogyra* sp. biomass as a biosorbent for the removal of acid orange 7. In the study, the highest percentage of dye removal at pH values of 2, 4, 6, 8 and 10 was determined as 42% at pH 4, and a significant decrease in dye removal was observed after pH 4. Gunasundari et al. (2020) investigated the adsorption of Naphthol green-B using *Spirulina platensis* biomass. pH 3 was determined as the best solution pH for biosorption. The dye removal was noticeably decreased when the pH increased from 3 to 7. In another study, the removal of anionic Methyl orange dye using *Chlorella* biomass decreased from 96.3% to 18.7% when pH increased from 2.5 to 11, and the decrease was evident after pH 4 (El Amri, Elkacmi & Boudouch, 2023).

Moreover, in the current study, the biosorption percentage for RBBR sharply decreased when the pH value was increased from 4 to 10. This trend of RBBR biosorption was demonstrated in the study of Ergene et al. (2009) performed with *Scenedesmus quadricauda* biomass, and a significant decrease was determined after pH 4.

The effect of initial RBBR concentration on biosorption

In the present study, the effect of initial dye concentration was investigated approximately from 100 to 800 mg/L (103.38 mg/L -

818.30 mg/L RBBR concentrations at pH 4 in the presence of 1 g/L biosorbent, and the results were demonstrated in Figure 4. It is clearly seen in Figure 4 that increasing RBBR concentration from 100 to 800 mg/L decreased the biosorption from 93.06% to 16.11% at the end of 120 min. RBBR biosorption efficiency was better for low initial dye concentrations due to the availability of active binding sites on the adsorbent (Aracagök, 2022). Similarly, Hernández-Zamora et al. (2015) observed that biosorption of the anionic azo dye Congo red onto inactive *C. vulgaris* biomass decreased when the initial concentration of Congo red increased from 5 mg/L to 25 mg/L. The authors stated that this is due to the fact that low concentrations of dye molecules in solution interact more easily with the binding sites of biosorbent.

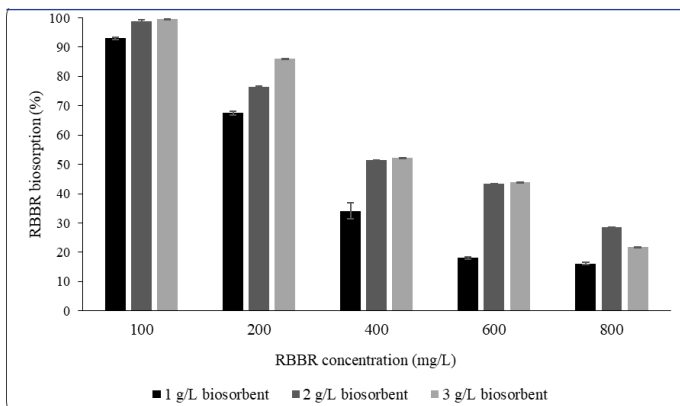


Figure 4. The highest biosorption values obtained with different biosorbent concentrations in increasing initial dye concentrations (pH 4; 30 °C; 100 rpm; biosorption time: the 30th min for 100 and 200 mg/L RBBR, the 120th min for 400, 600 and 800 mg/L RBBR in the presence of 3 g/L biosorbent; the 15th min for 100 mg/L RBBR, 30th for 400 and 800 mg/L RBBR, the 120th min for 200 and 600 mg/L RBBR in the presence of 2 g/L biosorbent; the 0th min for 400 mg/L RBBR, the 30th min 200 mg/L RBBR, the 120th min for 100, 600 and 800 mg/L RBBR in the presence of 1 g/L biosorbent).

In the current study, 600 and 800 mg/L RBBR removals were concluded with similar biosorption values at the end of the process. This situation can be explained by the fact that the amount of fixed dye molecules decreases the attractiveness of biosorbent surface's functional groups (Lai, 2021).

According to results obtained from the current study and literature, it has been seen that effective results can be obtained using *C. vulgaris* as a biosorbent for the biosorption of azo dyes.

The effect of initial biosorbent concentration and time on biosorption

Time and biosorbent concentration parameters for the removal of RBBR were found to be correlated with each other in the current study. The effect of biosorbent concentration (1, 2, and 3 g/L) on the RBBR biosorption is also shown in Figure 4. RBBR re-

moval accelerated when biosorbent concentration in dye solution was risen from 1 to 3 g/L. According to this, the highest RBBR biosorption was 98.91% at 15th min in 100 mg/L dye solution in the presence of 2 g/L biosorbent. Furthermore, for 3 g/L biosorbent concentration, the highest dye removal was calculated as 99.49% at the 30th min while it was similarly examined as 99.18% at the 15th min, and no significant change was observed between these values. Correspondingly with the current study, Radwan et al. (2020) showed that the biosorption rate of the 10 mg/L reactive yellow 145 dye was accelerated when the concentration of *C. vulgaris* biosorbent modified with citric acid increased from 0.1 to 0.5 g/L. For example, while equilibrium was reached at 60 min with 0.5 g/L modified- *C. vulgaris*, the dye uptake process continued for 0.1 g/L modified- *C. vulgaris*. The authors also stated that this increase is due to the increase in active binding sites on surface with increasing biosorbent concentration.

In addition, for 200 mg/L initial RBBR concentration, an increase from 1 to 3 g/L in biosorbent concentration reduced biosorption time up to the 15th minute and the biosorption percentage ranged from 67.45% to 85.88%. However, at dye concentrations after 200 mg/L, removal percentages were slightly decreased at the end of 30 min when biosorbent concentration increased from 2 to 3 g/L. This decline can be clarified by the crowding of the biosorbent and by releasing some dye molecules from the biosorbent surface (Mohd Khori et al., 2018).

Behl et al. (2019) examined the range of 0.25-1.5 g/L of *C. pyrenoidosa* biomass in Direct Red 31 removal. As a result of the study, no increase in dye removal efficiency was observed at concentrations after 1 g/L biosorbent. Furthermore, Revathi et al. (2017) demonstrated that moderate *C. vulgaris* concentration is more effective than higher cell concentrations.

According to the results of the present study, a significant increase in RBBR biosorption was observed at biosorbent concentrations above 1 g/L. At low RBBR concentrations (about 100 and 200 mg/L), the most effective biosorbent concentration was detected as 3 g/L *C. vulgaris* biomass. In addition, although there is no significant difference between 2 and 3 g/L biosorbent concentrations for approximately 400 and 600 mg/L RBBR concentrations, the best biosorption of 800 mg/L RBBR was observed in the presence of 2 g/L biosorbent. The decrease for 800 mg/L RBBR biosorption percentage in the solution containing 3 g/L biosorbent concentration may be due to the crowding of the biosorbent particles, which declined the number of active binding sites on the surface for adsorption, and the adsorption sites overlapped. This causes a decrease in the dye biosorption percentage (Mohd Khori et al., 2018).

Biosorption capacity of microalgal biosorbent on biosorption

Table 1 shows q_m values at the 15th min on RBBR biosorption in the presence of increasing biosorbent concentrations and increasing RBBR dye concentrations. As seen in Table 1, maximum q_m values at the 15th min were obtained in 200 mg/L RBBR for 1 g/L microalgal biosorbent and in 600 mg/L RBBR for 2 and 3 g/L microalgal biosorbent. When the q_m values for the RBBR concentrations of about 100, 200, and 400 mg/L were evaluated, a de-

Table 1. q_m values at 15th min on RBBR biosorption in the presence of increasing biosorbent concentrations and increasing RBBR dye concentrations (30 °C; 100 rpm; pH 4)

100	Initial RBBR dye concentration (mg/L)					
		100	200	400	600	800
Biosorbent concentration (g/L)	1	90.14±2.72	135.92±0.11	132.92±0.01	81.34±4.37	79.21±5.67
	2	52.42±0.27	68.23±0.01	100.28±0.43	112.47±0.08	102.77±0.01
	3	34.18±0.04	55.97±0.04	65.31±0.04	78.72±1.61	56.31±0.15

crease in the q_m was observed as the concentration of biosorbent increased. The highest q_m values for 100, 200 and 400 mg/L dye concentrations were observed at 1 g/L biosorbent concentration. Furthermore, at 600 and 800 mg/L initial RBBR dye concentrations, the highest q_m value was observed at 2 g/L biosorbent concentration. The lowest q_m value in all dye concentrations was at 3 g/L biosorbent concentration.

Similarly, studies in the literature show that the biosorption capacities of microalgal biosorbents for the removal of dyes decreased by increasing biosorbent concentration. da Rosa et al. (2018) investigated the *C. pyrenoidosa* biosorbent amount effect on the biosorption of rhodamine B at pH 4.5 in the presence of 100 mg/L dye. The biosorption capacities decreased from about 20 mg/g to 3.6 mg/g due to increasing biosorbent concentration from 0.1 to 2.0 g. Seth et al. (2022) determined that the increasing biosorbent concentrations (1-5 g/L) caused the decrease on sorption (mg/g) of 100 mg/L anionic dye Indigo Carmine. The authors also indicated that a decrease in sorption with the increase in biomass concentration is associated with a decrease in the availability of dye molecules per unit of biomass. In addition, the aggregation of biomass particles as another factor can decrease sorption (mg/g) due to the slowing of the intraparticle diffusion of dye molecules. Another study also showed that biosorption capacity decreased from 34.89 mg/g to 9.61 mg/g with increasing concentrations (1-5 g/L) of wet-torrefied *Chlorella* biochar on Congo Red biosorption (Yu et al. 2021). Thus, the results from the current study are consistent with the mentioned studies.

CONCLUSION

In this study, the maximum *C. vulgaris* biomass for RBBR biosorption was obtained as 0.502 g/L under photoheterotrophic cultivation conditions containing 0.5 g/L PW sugar. After optimization of microalgal growth, the highest RBBR biosorption percentage was determined as 99.49% at pH 4 in the presence of 3 g/L biosorbent and 103.38 mg/L initial RBBR concentration. Thus, the current study shows that PW for microalgae production and *C. vulgaris* for RBBR removal are potent, eco-friendly, and cost-effectively materials.

Acknowledgment: Nazlıhan TEKİN was awarded a PhD Scholarship by Council of Higher Education (YOK) and the Scientific and Technological Research Council of Turkey (TUBITAK).

Conflict of interests: The authors do not declare any conflict of interest.

Ethics Committee approval: No ethical approval is required for this study.

Financial Disclosure: There are no relevant financial or non-financial interests that the authors can disclose.

REFERENCES

- Aksu, Z., & Tezer, S. (2005). Biosorption of reactive dyes on the green alga *Chlorella vulgaris*. *Process Biochemistry*, 40(3-4), 1347-1361.
- Al-Tohamy, R., Ali, S. S., Li, F., Okasha, K. M., Mahmoud, Y. A. G., Elsamahy, T., ... & Sun, J. (2022). A critical review on the treatment of dye-containing wastewater: Ecotoxicological and health concerns of textile dyes and possible remediation approaches for environmental safety. *Ecotoxicology and Environmental Safety*, 231, 113160.
- Aracagök, Y. D. (2022). Biosorption of remazol Brilliant Blue R dye onto chemically modified and unmodified *Yarrowia lipolytica* biomass. *Archives of Microbiology*, 204(2), 128.
- Behl, K., Sinha, S., Sharma, M., Singh, R., Joshi, M., Bhatnagar, A., & Nigam, S. (2019). One-time cultivation of *Chlorella pyrenoidosa* in aqueous dye solution supplemented with biochar for microalgal growth, dye decolorization and lipid production. *Chemical Engineering Journal*, 364, 552-561.
- Benkhaya, S., M'rabet, S., & El Harfi, A. (2020). A review on classifications, recent synthesis and applications of textile dyes. *Inorganic Chemistry Communications*, 115, 107891.
- Chu, W. L., & Phang, S. M. (2019). Biosorption of heavy metals and dyes from industrial effluents by microalgae. *Microalgae biotechnology for development of biofuel and wastewater treatment*, 599-634.
- da Rosa, A. L. D., Carissimi, E., Dotto, G. L., Sander, H., & Feris, L. A. (2018). Biosorption of rhodamine B dye from dyeing stones effluents using the green microalgae *Chlorella pyrenoidosa*. *Journal of Cleaner Production*, 198, 1302-1310.
- El Amri, R., Elkacmi, R., & Boudouch, O. (2023). Removal of Methyl Orange from Water Using Microalgae: Effect of Operating Parameters, Equilibrium, Kinetic and Thermodynamic Studies. *Chemistry Africa*, 1-12.
- Ergene, A., Ada, K., Tan, S., & Katircioğlu, H. (2009). Removal of Remazol Brilliant Blue R dye from aqueous solutions by adsorption onto immobilized *Scenedesmus quadricauda*: Equilibrium and kinetic modeling studies. *Desalination*, 249(3), 1308-1314.
- FAOSTAT (2021) Crops and livestock products. Retrieved from <https://www.fao.org/faostat/en/#data/QCL> (accessed 14.05.23)
- Gadd, G. M. (2009). Biosorption: critical review of scientific rationale, environmental importance and significance for pollution treatment. *Journal of Chemical Technology & Biotechnology: International Research in Process, Environmental & Clean Technology*, 84(1), 13-28.
- Germec, M., & Turhan, I. (2018). Ethanol production from acid-pretreated and detoxified tea processing waste and its modeling. *Fuel*, 231, 101-109.
- Golub, N. B., & Voyevoda, D. V. (2013). Effect of sulphur compounds on cultivation process of microalgae *Chlorella vulgaris*. *Chem. Technol. Appl. Subst*, 761, 151-158.
- Goud, B. S., Cha, H. L., Koyyada, G., & Kim, J. H. (2020). Augmented biodegradation of textile azo dye effluents by plant endophytes: a sustainable, eco-friendly alternative. *Current Microbiology*, 77, 3240-3255.

- Gunasundari, E., Kumar, P. S., Rajamohan, N., & Vellaichamy, P. (2020). Feasibility of naphthol green-b dye adsorption using microalgae: Thermodynamic and kinetic analysis. *Desalination and Water Treatment*, 192, 358-370.
- Gül, U. D. (2022). Utilization of Surfactants to Augment Decolorization Process by Biosorbent. *NanoWorld J*, 8(4), 107-112.
- Hernández-Zamora, M., Cristiani-Urbina, E., Martínez-Jerónimo, F., Perales-Vela, H. V., Ponce-Noyola, T., Montes-Horcasitas, M. D. C., & Cañizares-Villanueva, R. O. (2015). Bioremoval of the azo dye Congo Red by the microalga *Chlorella vulgaris*. *Environmental Science and Pollution Research*, 22, 10811-10823.
- Isleten-Hosoglu, M., Ayyıldız-Tamis, D., Zengin, G., & Elibol, M. (2013). Enhanced growth and lipid accumulation by a new *Ettlia texensis* isolate under optimized photoheterotrophic condition. *Bioresource technology*, 131, 258-265.
- Joo, G., Lee, W., & Choi, Y. (2021). Heavy metal adsorption capacity of powdered *Chlorella vulgaris* biosorbent: effect of chemical modification and growth media. *Environmental Science and Pollution Research*, 28, 25390-25399.
- Kassim, M. A., Ramli, S. H., & Meng, T. K. (2022). Analysis of microalgal growth kinetic model and carbohydrate biosynthesis cultivated using agro-industrial waste residuals as carbon source. *Preparative biochemistry & biotechnology*, 52(5), 514-524.
- Khataee, A. R., Vafaei, F., & Jannatkah, M. (2013). Biosorption of three textile dyes from contaminated water by filamentous green algal *Spirogyra* sp.: Kinetic, isotherm and thermodynamic studies. *International Biodeterioration & Biodegradation*, 83, 33-40.
- Kidoń, M., & Uwineza, P. A. (2022). New Smoothie Products Based on Pumpkin, Banana, and Purple Carrot as a Source of Bioactive Compounds. *Molecules*, 27(10), 3049.
- Kumar, S., Ahluwalia, A. S., & Charaya, M. U. (2019). Adsorption of Orange-G dye by the dried powdered biomass of *Chlorella vulgaris* Beijerinck. *Current Science*, 116(4), 604-611.
- Lai, H. J. (2021). Adsorption of remazol brilliant violet 5R (RBV-5R) and remazol brilliant blue R (RBBR) from aqueous solution by using agriculture waste. *Tropical Aquatic and Soil Pollution*, 1(1), 11-23.
- Manzoor, M., Ahmad, Q. U. A., Aslam, A., Jabeen, F., Rasul, A., Schenk, P. M., & Qazi, J. I. (2020). Mixotrophic cultivation of *Scenedesmus dimorphus* in sugarcane bagasse hydrolysate. *Environmental Progress & Sustainable Energy*, 39(2), e13334.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical chemistry*, 31(3), 426-428.
- Mohammad Mirzaie, M. A., Kalbasi, M., Mousavi, S. M., & Ghobadian, B. (2016). Investigation of mixotrophic, heterotrophic, and autotrophic growth of *Chlorella vulgaris* under agricultural waste medium. *Preparative Biochemistry & Biotechnology*, 46(2), 150-156.
- Mohd Khor, N. K. E., Hadibarata, T., Elshikh, M. S., Al-Ghamdi, A. A., & Yusop, Z. (2018). Triclosan removal by adsorption using activated carbon derived from waste biomass: Isotherms and kinetic studies. *Journal of the Chinese Chemical Society*, 65(8), 951-959.
- Mustafa, S., Bhatti, H. N., Maqbool, M., & Iqbal, M. (2021). Microalgae biosorption, bioaccumulation and biodegradation efficiency for the remediation of wastewater and carbon dioxide mitigation: Prospects, challenges and opportunities. *Journal of Water Process Engineering*, 41, 102009.
- Park, W. K., Moon, M., Kwak, M. S., Jeon, S., Choi, G. G., Yang, J. W., & Lee, B. (2014). Use of orange peel extract for mixotrophic cultivation of *Chlorella vulgaris*: increased production of biomass and FAMES. *Bioresource technology*, 171, 343-349.
- Premaratne, M., Nishshanka, G. K. S. H., Liyanaarachchi, V. C., Nimarshana, P. H. V., & Ariyadasa, T. U. (2021). Bioremediation of textile dye wastewater using microalgae: current trends and future perspectives. *Journal of Chemical Technology & Biotechnology*, 96(12), 3249-3258.
- Radwan, E. K., Abdel-Aty, A. M., El-Wakeel, S. T., & Abdel Ghafar, H. H. (2020). Bioremediation of potentially toxic metal and reactive dye-contaminated water by pristine and modified *Chlorella vulgaris*. *Environmental Science and Pollution Research*, 27, 21777-21789.
- Revathi, S., Kumar, S. M., Santhanam, P., Kumar, S. D., Son, N., & Kim, M. K. (2017). Bioremoval of the indigo blue dye by immobilized microalga *Chlorella vulgaris* (PSBDU06). *Journal of Scientific & Industrial Research*, 76:50-56.
- Rippka, R. (1988). [1] Isolation and purification of cyanobacteria. *Methods in enzymology*, 167, 3-27.
- Salleh, M. A. M., Mahmoud, D. K., Karim, W. A. W. A., & Idris, A. (2011). Cationic and anionic dye adsorption by agricultural solid wastes: a comprehensive review. *Desalination*, 280(1-3), 1-13.
- Saratale, R. G., Ponnusamy, V., Sirohi, R., Piechota, G., Shobana, S., Dharmaraja, J., ... & Veermuthu, A. (2022). Microalgae cultivation strategies using cost-effective nutrient sources: recent updates and progress towards biofuel production. *Bioresource Technology*, 127691.
- Seth, B. M., Uniyal, V., Kumar, D., & Singh, A. (2021). Sorption of cationic and anionic dyes by dead biomass of filamentous green alga *Cladophora* sp. (Chlorophyceae). *International Journal of Environmental Science and Technology*, 19:12079-12090.
- Shabir, M., Yasin, M., Hussain, M., Shafiq, I., Akhter, P., Nizami, A. S., ... & Park, Y. K. (2022). A review on recent advances in the treatment of dye-polluted wastewater. *Journal of Industrial and Engineering Chemistry*, 112:1-19.
- Sharma, A. K., Sahoo, P. K., Singhal, S., & Patel, A. (2016). Impact of various media and organic carbon sources on biofuel production potential from *Chlorella* spp. *3 Biotech*, 6, 1-12.
- Valdez-Arjona, L. P., & Ramirez-Mella, M. (2019). Pumpkin waste as livestock feed: Impact on nutrition and animal health and on quality of meat, milk, and egg. *Animals*, 9 (10), 769.
- Verma, R. K., Sankhla, M. S., Rathod, N. V., Sonone, S. S., Parihar, K., & Singh, G. K. (2021). Eradication of fatal textile industrial dyes by wastewater treatment. *Biointerface Res. Appl. Chem*, 12, 567-587.
- Wang, Y., Guo, W., Cheng, C. L., Ho, S. H., Chang, J. S., & Ren, N. (2016). Enhancing bio-butanol production from biomass of *Chlorella vulgaris* JSC-6 with sequential alkali pretreatment and acid hydrolysis. *Bioresource technology*, 200, 557-564.
- Yu, J., Zhang, X., Wang, D., & Li, P. (2018). Adsorption of methyl orange dye onto biochar adsorbent prepared from chicken manure. *Water Science and Technology*, 77(5), 1303-1312.
- Yu, K. L., Lee, X. J., Ong, H. C., Chen, W. H., Chang, J. S., Lin, C. S., ... & Ling, T. C. (2021). Adsorptive removal of cationic methylene blue and anionic Congo red dyes using wet-torrefied microalgal biochar: Equilibrium, kinetic and mechanism modeling. *Environmental pollution*, 272, 115986.
- Zohoorian, H., Ahmadzadeh, H., Molazadeh, M., Shourian, M., & Lyon, S. (2020). Microalgal bioremediation of heavy metals and dyes. *In Handbook of algal science, technology and medicine* (pp. 659-674). Academic Press.

Investigation of Growth Performance, Proximate and Fatty Acid Composition of Freshwater (*Euglena gracilis*, *Chlorella vulgaris*) and Marine (*Pavlova lutheri*, *Diacronema vlnanium*) Microalgae

Merve Sayar¹ , Kamil Mert Eryalçın¹ 

Cite this article as: Sayar, M., & Eryalçin, K.M. (2024). Investigation of growth performance, proximate and fatty acid composition of freshwater (*Euglena gracilis*, *Chlorella vulgaris*) and marine (*Pavlova lutheri*, *Diacronema vlnanium*) microalgae. *Aquatic Sciences and Engineering*, 39(1), 8-16. DOI: <https://doi.org/10.26650/ASE20241303511>

ABSTRACT

This work is focused on investigating the nutrient compositions, growth, and fatty acid composition of *Chlorella vulgaris*, *Euglena gracilis*, *Pavlova lutheri*, and *Diacronema vlnanium*, which are natural diets of bivalve, crustaceans, live prey such as rotifer, copepods, daphnia and feed ingredients in aquaculture nutrition. Microalgae culture was performed in a live feed laboratory under controlled physical and chemical conditions. The initial concentration of microalgae species was adjusted as 2×10^6 cells/mL and growth performance was calculated by Neubauer Hemocytometer daily. The maximum growth performance was detected in *Diacronema vlnanium* culture with 1.78×10^7 cells/mL. In the case of proximate composition, the highest dry matter content was found in *Pavlova lutheri* (6.21%). Freshwater microalgae species *Chlorella vulgaris* (50.5%) and *Euglena gracilis* (42.5%) had high crude protein compared to *Pavlova lutheri* and *Diacronema vlnanium*. Fatty acid compositions of microalgae were also determined. The highest EPA (C20:5n-3) content was found in *Pavlova lutheri* (6.85%) whereas arachidonic acid (C20:4n-6) and docosahexaenoic acid (C22:6n-3) contents were only found with a level of (3.32%) and (1.79%) in *Euglena gracilis*, respectively. Microalgal culture should have high biomass in a short time of culture and in this study, *E.gracilis* and *P.lutheri* showed high growth and essential nutrients gain in laboratory scale production and this result could be applied in larger volume photobioreactor.

Keywords: Microalgae, growth, fatty acids, proximate, biomass

ORCID IDs of the author:
M.S. 0000-0002-2628-0608;
K.M.E. 0000-0002-8336-957X

¹Faculty of Aquatic Sciences, Department of Aquaculture and Fish Diseases, Phytoplankton-Zooplankton Culture Laboratory, Istanbul University, Istanbul, Turkiye

Submitted:
27.05.2023

Revision Requested:
11.08.2023

Last Revision Received:
16.08.2023

Accepted:
19.09.2023

Online Published:
13.11.2023

Correspondence:
Kamil Mert Eryalçın
E-mail:
eryalcin@istanbul.edu.tr

INTRODUCTION

Microalgae contribute greatly to both the marine and freshwater food-web and they are able to synthesize inorganic matter into organic compounds such as lipids, polysaccharides and pigments (Chiu et al., 2011). They are used for live prey enrichment and feeding (Eryalçın, 2018; Eryalçın, 2019; Turcihan et al., 2021; Turcihan et al., 2022), wastewater treatment (Wollmann et al., 2019), biodiesel production (Goh et al., 2019), fish diet ingredients (Eryalçın et al., 2013; Eryalçın and Yıldız, 2015; Eryalçın et al., 2015; Camacho-Rodríguez et al., 2018; Soto-Sánchez et al.,

2023) and bivalve culture (Shah et al., 2018). Microalgae must be nutritionally riched in essential biochemical compounds such as polyunsaturated fatty acids (PUFAs), highly unsaturated fatty acids (HUFAs), essential amino acids (EAA), and pigments (Raja et al., 2004; Patil et al., 2005; Patil et al., 2007; Hemaiswarya et al., 2011; Singh et al., 2015; Peltomaa et al., 2017). Moreover, they have antagonistic effects on bacterial communities in culture tanks (Spolaore et al., 2006; Neori, 2011). The first priority of microalgae culture is to get fast high biomass gain in a short time. The fast growth performance of microalgae is based on several parameters. The rapid proliferation of



microalgae contributes to the high biomass in wet weight and this leads to the possible production of nutrients such as lipid, protein, carbohydrate, and pigment. For example, dinoflagellate *Crypthecodinium cohnii* can contain DHA up to 40% in dry weight that is necessary for both growth and stress resistance at fish larval cultivation (Eryalçın et al., 2013). Nutrient contents of microalgae such as protein, lipids, and pigments can be species-specific which means each alga can contain specific nutrients (Das et al., 2012; Eryalçın, 2019; Gharajeh et al., 2020). For instance, *Nannochloropsis oculata* contains a high amount of EPA whereas dinoflagellate *Crypthecodinium cohnii* is famous for DHA. In comparison, freshwater microalgae are rich in essential 18C chain fatty acids such as linoleic (C18:2n-6), and α -linolenic acid (C18:3n-3) which are also important for freshwater fish.

Moreover, microalgae are the main energy source and substantial for enhancing the survival and growth of bivalve larvae (Parrish et al., 1998; Pazos et al., 1997; Budge et al., 2001). The nutritional value of microalgae is changed during their culture time. There are two main phases of the culture period called the exponential and stationary phases where algae should be harvested (da Silva Ferreira and Anna, 2017). Not only the culture phase but also the culture medium affects the nutritional value of microalgae biomass. These nutrient profiles consist of macronutrients (nitrogen, phosphorus, and sulphur) and micronutrients (iron, manganese, sodium molybdenum oxide, zinc, copper, and selenium) (Aslam et al., 2021; Shaaban et al., 2010). The nitrogen source of microalgae increases the growth performance and nutritional content by synthesizing large nutrient molecules like minerals and proteins (Procházková et al., 2014; Kumaran et al., 2023).

The growth performance and nutritional composition of microalgae also depend on physical and chemical parameters such as light, temperature, salinity, pH, and cultivation methods such as heterotrophic, autotrophic, and mixotrophic culture (Bashir et al., 2019; Zhao et al., 2011). Salinity and light conditions are very important in the cultivation of microalgae. For instance; *Nannochloropsis* sp. shows high growth performance at high salinity levels but it shows slow growth performance in heterotrophic culture (Bashir et al., 2019). In particular, autotrophic microalgae species directly affect the synthesis of biochemical substances and growth performance due to the intensity of the light (Sandnes et al., 2005). The reason is because these microalgae species use light as an energy source. As a result, biomass, proximate and fatty acid composition change depending on the light intensity. Another agent affecting microalgal growth performance, and fatty acid composition, microalgae cell metabolism, and the initial enzyme used for photosynthesis is culture temperature (Chaisutyakorn et al., 2018; Chiu et al., 2011). The increasing temperature in the culture adversely affects the fatty acid composition of microalgae. Most importantly, higher biomass gain and growth performance as well as protein and lipid contents in most microalgae are linearly related to light intensity and photoperiod such as *Chlorella vulgaris*, *Ankistrodesmus falcatus*, *Monoraphidium* sp., *Botryococcus braunii* (He et al., 2015; Metsoviti et al., 2019).

The other reason that affects biomass gain of microalgae depends on the growth potential of the species. As the growth performance increases, the biomass recovery rate also increases

(Lau et al., 2022). The size of microalgae cells also affects their growth performance. Small-sized microalgae show higher growth performance than larger cell microalgae due to doubling time. For example, Arkronrat et al. (2016) have stated that *Nannochloropsis* sp. are smaller microalgae, and its growth performance is faster than *Tetraselmis* sp. due to its size. The unicellular freshwater eukaryote *Euglena gracilis* obtains flagellates and instead of a cell wall it has a pellicle based protein layer by a substructure of microtubules (Zhang et al., 2023). *Euglena gracilis* is rich in Paramylon which is a linear β -1,3-glucan polysaccharide polymer, antioxidants such as β -carotene, α -tocopherol and L-ascorbic acid, and PUFAs (Kottuparambil et al., 2019). The other freshwater microalgae *Chlorella vulgaris* belongs to Chlorophyceae with the thick cell walls and contains a high level of protein, minerals, vitamins, and pigments (Spinola et al., 2023). *Pavlova lutheri* is unicellular motile marine prymnesiophyte algae containing flagellate and it is known for high sterols, EPA and, DHA contents (Ahmed et al., 2015). *Diacronema vlkianum* is another marine green microalgae that belongs to the Haptophyceae family and is also rich in high levels of EPA and DHA (Fradique et al., 2013).

In this study, growth performance, proximate, and fatty acid compositions two unicellular both freshwater (*Euglena gracilis* and *Chlorella vulgaris*) and two marine microalgae species (*Pavlova lutheri*, *Diacronema vlkianum*) were investigated under laboratory conditions for biomass utilization.

MATERIAL AND METHODS

Microalgae strains and stock culture

Culture mediums f/2 and 3N-BBM-V were sterilized at 121 °C for 15 min before they were used (Guillard, 1975). Stock culture of microalgae was cultured in 50 mL test tubes to 250 mL, followed by 1-L, and 5-L erlenmeyers. Microalgae were counted in each experimental flask during the experiment. The microalgae growth trial was conducted in the Phytoplankton and Zooplankton Laboratory of the Faculty of Aquatic Sciences of Istanbul University, for 32 days. Four microalgae species were obtained from CCAP (Culture collection of algae and Protozoa, Scotland) which are *Pavlova lutheri* (Strain number: CCAP940/2), *Diacronema vlkianum* (Strain number: CCAP914/1), *Euglena gracilis* (Strain number: CCAP1224/38), *Chlorella vulgaris* (Strain number: CCAP211/110).

Experimental design and growth performance

In this study, the microalgae species were cultured from the initial to 15 days under 250 mL volume. Each experimental group was studied in three replicates. The initial cell density of the microalgae species was adjusted at 2×10^6 cells/mL for the second part of the culture experiment after the 15th day culture, all volumes were inoculated into 1000 mL erlenmeyers with gentle aeration till the 32nd day. This method was chosen by the same culture procedure at commercial hatcheries where first culture occurred in small-scale flasks and then continuously up-scaled in larger volumes. At the end of the culture, all biomass was harvested and stored at -80 °C in the refrigerator. During culture, the growth performance of microalgae species was calculated daily with a Neubauer Hemocytometer.

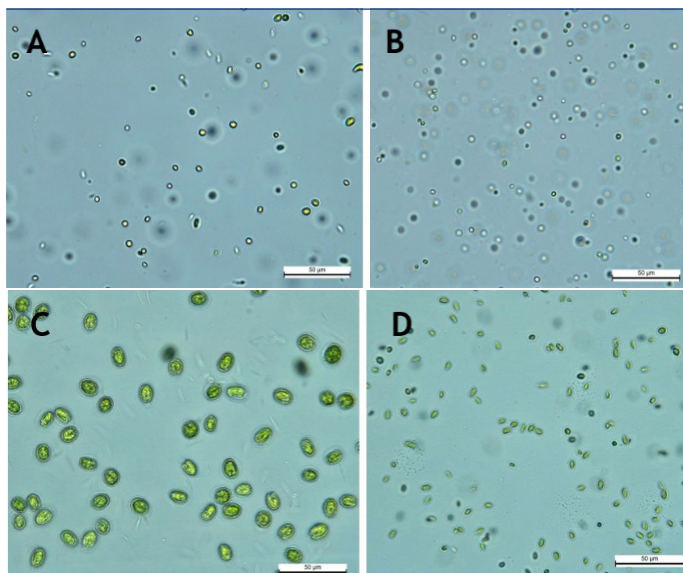


Figure 1. *Chlorella vulgaris* (A), *Diacronema vlkanium* (B), *Pavlova lutheri* (C) and *Euglena gracilis* (D).

Proximate analysis

Dry matter analysis of microalgae, the samples were first filtered using a vacuum filtration system. Vacuum filter papers (Schleicher&Schuell GF-52, 47 mm, nominal pore opening $0.7 \mu\text{m}$.) were dried in an oven at 105°C for 3 hours. When the papers were cooled at room temperature, their empty weights were weighed. Microalgae (100 mL) were filtered by a vacuum filtration system. The filter papers obtained after the filtration process were taken back to the oven at 105°C and the drying process was carried out. After drying methods, the papers were taken into a desiccator. The dry matter (%) was calculated by measuring the papers that were cooled in the desiccator (AOAC, 1995). Kjeldahl method was preferred for crude protein analysis of microalgae. Microalgae samples were weighed 0.5 g–0.8 g and placed in Kjeldahl tubes (AOAC, 1995). Two pieces of Kjeldahl tablets and 20 mL sulphuric acid (H_2SO_4) were added to the microalgae samples placed in the tubes. The samples burned at 450°C for 120 minutes. The tubes were placed in the Kjeldahl device (Gerhardt VAPODEST®), and distillation and titration were performed. The amount of crude protein in the samples was calculated by determining the amount of 0.1N HCl consumed in the titration. Microalgae samples to be analyzed for crude lipid were weighed around 1 g and placed in the lipid extraction device. The glass VELPs (VELP® Scientifica), were previously dried in an oven at 105°C and kept in the desiccator. The samples were completed in the Soxhlet device for 60 minutes. After extraction, the glass VELPs were placed in an oven at 105°C . The weights of the glass VELPs were weighed, and the percentage of crude lipid was calculated (Folch et al., 1957). Microalgae samples were placed in ceramic and burned in a muffle furnace at 550°C for 5-6 hours. The samples were taken into a desiccator to come to room temperature. The samples at room temperature were weighed. After weighing, the amount of ash was calculated (AOAC, 1995).

Fatty acid analysis

Fatty acid methyl esters were analyzed by GC (GC-2030; Shimadzu, Tokyo, Japan) in a Supercolvax-10 fused silica capillary column

(constant pressure with 100KPa, length: 100 m; internal diameter: 0.25 mm; 0.20 i.d (Ref.: 24080-U) Supelco, Bellefonte, PA, USA) using H_2 as a carrier gas. Fatty acid methyl esters in algae biomass were gained by the transmethylation method with 1% sulfuric acid in methanol (Christie, 1982). The column temperature was 180°C for the first 10 min, increasing to 260°C at a rate of 2°C min^{-1} and then held at 260°C for 15 min. Then they were quantified by FID following the conditions described by Izquierdo et al. (1990) and identified by comparison with external standards well-characterized fish oils (EPA 28, Nippai, Ltd Tokyo, Japan).

Statistical analysis

Each sampling was conducted in triplicate and all data were treated with one-way analysis of variance (ANOVA) and the averages in the study were compared with the Duncan test ($p < 0.05$) method in the SPSS program (SPSS for Windows 11.5; SPSS Inc., Chicago, IL, USA) and significance was adjusted at $p < 0.05$.

RESULTS AND DISCUSSION

Growth performance of microalgae

In this study, the growth performance of *Chlorella vulgaris*, *Pavlova lutheri*, *Euglena gracilis* and *Diacronema vlkanium* microalgae species were investigated. Growth performance was measured in two different volumes and time-lapse. The first measurement was between the initial and 15th days of culture and had a 250 mL culture volume while the second culture process was upscaled to 1000 mL culture volume with stable aeration between the 15th and 32nd days. The highest growth was determined in DV (*Diacronema vlkanium*) culture with 1.68×10^7 and 1.59×10^7 cells/mL density at 13th days and 30th days, respectively. Freshwater microalgae *Chlorella vulgaris* (CV) showed the highest growth rate on the 15th day of culture in 250 mL and cell density continuously increased until the end of the experiment with gentle aeration in 1000 mL erlenmeyer. In terms of *Euglena gracilis* (EG) rapid growth was obtained between the 18th and 28th days in the presence of 1000 mL volume and regular aeration. The cell density is higher from the 20th and 28th days (3.50×10^6) compared to the culture of between the 2nd and 17th days (9.50×10^6). *Pavlova lutheri* (PL) maximum cell density was recorded at (7.77×10^6) at 15 days in and (8.50×10^6) at 32th days of culture, respectively. Microalgae growth performances are shown the Figure 2.

Proximate composition of microalgal biomass

In this study, the nutritional compositions were examined and it was reported that the highest crude protein content was found in *Chlorella vulgaris* ($50.05 \pm 0.01\%$) and *Euglena gracilis* ($42.15 \pm 0.52\%$) had the second highest level ($p < 0.05$). However, the lowest crude protein ($38.4 \pm 0.55\%$) was detected in marine microalgae *Diacronema vlkanium* ($p < 0.05$). In terms of crude lipid content, the highest crude lipid ($19.96 \pm 0.97\%$) was found in marine microalgae *Pavlova lutheri* species whereas the lowest value was found in freshwater microalgae *Chlorella vulgaris* ($11.2 \pm 0.02\%$) ($p < 0.05$). *Pavlova lutheri* had the highest dry matter ($6.21 \pm 0.33\%$) content among groups ($p < 0.05$). The table below shows the nutritional content of microalgae (Table 1).

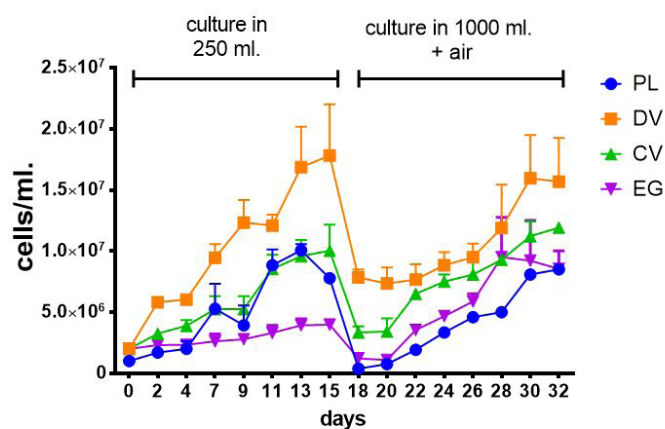


Figure 2. Growth performance of microalgae species; PL (*Pavlova lutheri*), DV (*Diacronema vlkanium*), CV (*Chlorella vulgaris*), and EG (*Euglena gracilis*).

feeding and formulated diets in marine fish. The nutritional value and growth performance of microalgae are also essential for biomass production. In microalgae culture, growth (doubling time), fatty acid content, and nutritional values are directly affected by the cultivation method. Moreover, the ingredients of the culture medium and stress conditions also affect the growth and proximate composition of the microalgae. For instance, *Scenedesmus* sp. can accumulate high levels of lipids under stress conditions (Khatoun et al., 2019). The other halophilic microalgae *Dunaliella salina* can contain a high amount of pigments under high salinity conditions (de Souza Celente et al., 2022), and freshwater microalgae *Chlorella vulgaris* may have high protein under low salinity conditions (Liu et al., 2008). In this study, the growth performance, proximate, and fatty acid composition of four microalgae cultured under constant laboratory conditions were investigated for potential aquaculture purposes such as live prey feeding or microalgae biomass.

Jeong et al. (2016) reported that the highest growth performance in *E. gracilis* was obtained by mixotrophic cultivation compared

Table 1. Proximate composition of microalgal biomass.

Proximate Analysis (%)	<i>Chlorella vulgaris</i>	<i>Euglena gracilis</i>	<i>Pavlova lutheri</i>	<i>Diacronema vlkanium</i>
Crude Protein	50.05±0.01 ^a	42.15±0.52 ^b	39.02±0.32 ^c	38.4±0.55 ^c
Crude Lipid	11.2±0.02 ^c	15.35±0.31 ^b	19.96±0.97 ^a	18.01±0.99 ^a
Crude Ash	7.2±0.00 ^c	5.01±0.22 ^d	10.01±0.88 ^b	18.45±0.83 ^a
Dry Matter	3.00±0.01 ^c	2.44±0.00 ^d	6.21±0.33 ^a	5.89±0.96 ^b

Dissimilar lettering show significant differences among groups (* $p < 0.05$; Duncan's multiple range test).

Fatty acid composition of microalgae biomass

Microalgae is an important source of essential fatty acids such as EPA (C20:5n-3), DHA (C22:6n-3), and ARA (C20:4n-6) for aquaculture. *Euglena gracilis* had the highest EPA level (0.32±0.01%) among groups ($p < 0.05$). The highest DHA level (1.79±0.02%) was found in *Euglena gracilis* biomass ($p < 0.05$). *Euglena gracilis* and *Pavlova lutheri* had the highest ARA levels (3.32±0.05% and 3.16±0.00%) ($p < 0.05$). Oleic acid (C18:1n-9) content highest values had *Chlorella vulgaris* and *Diacronema vlkanium* and the lowest value was *Euglena gracilis*. Freshwater microalgae had the highest (9.24±0.14% and 8.27±0.06%) linoleic acid (C18:2n-6) content ($p < 0.05$). ALA (α -linolenic acid) (C18:3n-3) was found in *Euglena gracilis*, *Pavlova lutheri* and *Diacronema vlkanium* microalgae species with a level of 14.98±0.10%, 6.62±0.14% and 0.21±0.00%, respectively. γ -linolenic acid (C18:3n-6) was only found in *Pavlova lutheri*. The highest Σ n-3 (24.87±0.03), Σ n-6 (17.28±0.56) fatty acid contents were found in *Euglena gracilis*. Additionally, the highest Σ n-3 HUFA (8.47±0.15%) and Σ PUFA contents (39.58±1.33%) were found in *Euglena gracilis*. The lowest value (0.30±0.02%) was found in *Chlorella vulgaris* ($p < 0.05$).

Microalgae are rich in lipids (Fields et al., 2014), carbohydrates (Chen et al., 2013), proteins (Becker, 2007), pigments (Begum et al., 2016), and fatty acids such as; PUFA, EPA and ARA (Eryalçın et al., 2013; Eryalçın et al., 2015) and therefore, they are very important future food supply not only for aquaculture purpose but also direct utilization of their biomass for human and animal diets. From this respect, recent studies are focused on the utilization of microalgae in both live prey

to phototrophic and heterotrophic cultivation methods with values of 2.48×10^6 , 0.61×10^6 and 0.49×10^6 (cells/mL), respectively. A similar study was conducted by Gu et al. (2022) that calculated the growth performance of *E. gracilis* at autotrophic and mixotrophic culture methods and they reported autotrophic culture had a higher growth result (0.6×10^6 cell/mL) at 12 days of culture. In our study, *E. gracilis* was cultivated in a phototrophic way and found higher growth than other studies was obtained at a larger volume (1000 mL) with 9.2×10^6 (cells/mL), and lower growth was detected 3.97×10^6 (cells/mL) cell density in smaller culture (250 mL). In terms of fatty acid results, the highest levels of ALA, and EPA (14.92±0.10% and 4.79±0.08%) were obtained in phototrophic cultivation whereas the highest ARA, and DHA levels (3.39±0.03%, 1.74±0.02%) were found in mixotrophic cultivation method (Jeong et al., 2016). In our study, the highest levels of ALA, ARA, and DHA (14.98±0.10%, 3.32±0.05% and 1.79±0.02%) were found in *E. gracilis* biomass among microalgae whereas the EPA level (4.56±0.03%) was found the lowest in other microalgae species. This result could be related to different cultivation methods of the *Euglena gracilis*. Similar to Jeong et al. (2016) results, our study also showed that the phototrophic cultivation method enhanced fatty acid contents of *E. gracilis*. *Chlorella vulgaris* has a high potential for biomass production in both indoor and outdoor culture systems. It has been evaluated as feed ingredients in aquaculture (Ahmad et al., 2020). At laboratory scale production, we obtained the highest cell density at 1.05×10^7 (cells/mL)

Table 2. Fatty acid composition of microalgae species.

Fatty Acid Compositions (% total fatty acid)	<i>Euglena gracilis</i>	<i>Pavlova lutheri</i>	<i>Diacronema vlkanium</i>	<i>Chlorella vulgaris</i>
C8:0	-	-	0.19±0.01	-
C10:0	-	0.06±0.00 ^b	0.11±0.01 ^a	0.03±0.00 ^c
C11:0	-	0.62±0.02	-	-
C12:0	-	0.86±0.00 ^a	0.44±0.03 ^b	0.16±0.00 ^c
C14:0	12.30±0.15 ^b	1.01±0.01 ^c	3.69±0.07 ^a	0.60±0.02 ^c
C14:1	-	-	-	0.11±0.00
C15:0	2.24±0.04 ^a	0.14±0.01 ^c	0.39±0.03 ^b	0.26±0.01 ^b
Iso16:0	5.81±0.03	-	-	-
C16:0	27.25±0.23 ^b	24.50±0.09 ^b	33.59±0.27 ^a	18.80±0.21 ^c
C16:1n-5	3.08±0.01	-	-	-
C16:2n-4	0.46±0.02	-	-	-
C16:1	-	0.57±0.03 ^b	26.33±0.08 ^a	28.76±0.18 ^a
C17:0	0.90±0.01 ^a	n.d.	0.35±0.03 ^b	0.23±0.02 ^b
C16:3n-4	1.26±0.02	-	-	-
C18:0	2.42±0.01 ^b	1.14±0.01 ^b	1.86±0.02 ^b	14.38±0.05 ^a
C18:1n-9	1.32±0.01 ^d	15.94±0.02 ^c	22.24±0.15 ^b	28.21±0.13 ^a
C18:2n-6	8.27±0.06 ^b	9.24±0.14 ^a	3.18±0.03 ^d	7.46±0.01 ^b
C18:2n-4	0.28±0.00	-	-	-
C18:3n-3	14.98±0.10 ^a	6.62±0.14 ^b	0.21±0.00 ^c	n.d.
C18:3n-6	-	1.05±0.01	-	-
C20:0	0.16±0.02 ^a	-	-	0.14±0.01 ^a
C20:1	-	0.70±0.01 ^a	-	0.09±0.01 ^b
C20:2	-	1.12±0.01	-	-
C20:2n-6	2.57±0.13	-	-	-
C20:3n-3	1.42±0.05 ^a	-	-	0.24±0.00 ^b
C20:3n-6	0.65±0.02	-	-	-
C20:4n-6	3.32±0.05 ^a	3.16±0.00 ^a	0.08±0.00 ^b	-
C20:4n-3	1.72±0.02	-	-	-
C20:5n-3	4.56±0.03 ^b	6.84±0.03 ^a	2.20±0.03 ^c	0.06±0.01 ^d
C22:0	-	-	-	0.06±0.01
C22:4n-6	0.28±0.05	-	-	-
C22:5n-6	2.19±0.01	-	--	-
C22:4n-3	0.08±0.01	-	-	-
C22:5n-3	0.32±0.01	-	-	-
C22:6n-3	1.79±0.02 ^a	-	-	0.09±0.02 ^b
C24:0	-	0.08±0.00 ^b	-	0.16±0.01 ^a
Σ Monounsaturated	4.40±0.05 ^d	17.21±0.04 ^c	22.24±0.15 ^b	29.0±0.16 ^a
Σ Saturated	51.08±0.23 ^a	28.61±0.13 ^d	40.60±0.25 ^b	34.8±0.18 ^c
Σ n-3	24.87±0.03 ^a	6.84±0.03 ^b	2.40±0.03 ^c	0.3±0.02 ^d
Σ n-6	17.28±0.56 ^a	13.45±0.13 ^b	0.08±0.00 ^d	7.5±0.01 ^c
Σ n-9	1.32±0.22 ^d	15.94±0.02 ^c	22.24±0.15 ^b	28.2±0.13 ^a
Σ n-3 HUFA	8.47±0.15 ^a	6.84±0.03 ^b	2.40±0.03 ^c	0.30±0.02 ^d
EPA/ARA	1.37±0.02	-	-	-
DHA/EPA	0.39±0.01	-	-	-
DHA/ARA	0.54±0.04	-	-	-
n-3/n-6	1.44±0.02 ^b	0.51±0.01 ^b	30.00±0.38 ^a	0.04±0.02 ^c
Σ PUFA	39.58±1.33 ^a	26.90±0.03 ^b	27.90±0.21 ^b	7.61±0.04 ^c

Dissimilar lettering shows significant differences among groups (*p<0.05; Duncan's multiple range test).

in 1000 mL volume at phototrophic culture. Taş and Dalkıran (2022) reported that the *C. vulgaris* initial cell density was 1.1×10^6 (cells/mL) and the highest cell density obtained was 2.4×10^7 (cells/mL) on the 3rd day of mixotrophic culture. This higher algal productivity might be related to the nutrient composition of cultures medium by affecting the metabolism of microalgae cells (Fields et al., 2014). Light, temperature, and cultivation methods are important factors in microalgal growth and proximate composition. In our study, all parameters were constant therefore we assumed that growth and nutrient compositions were positively affected by culture mediums even when we used phototrophic culture methods.

Total lipid and protein accumulation should be higher in microalgae cells in order to be evaluated as feed ingredients. *E. gracilis* has distinctive features in the phototrophic cultivation method such as high protein content and high digestibility (Nwoye et al., 2017). In our study, the crude protein content of *Euglena gracilis* was higher than the marine microalgae species. On the other hand, the highest crude lipids were found in marine microalgae both *Diacronema vlkanium* and *Pavlova lutheri*. Yeh et al. (2010) found the crude protein and lipid contents of *C. vulgaris* as 25-30% and 30-40%, respectively. In our study, crude protein (50.05%) was found to be higher compared to Yeh et al. (2010), moreover, the crude lipid (11.2%) value was lower. This result could be related to cultured microalgae in photobioreactor culture. Moreover, salinity highly affects of fatty acid contents of microalgae. Teh et al. (2021) investigated the fatty acid content of *C. vulgaris* at different salinity levels and oleic acid (C18:1n-9), linoleic acid (C18:2n-6), and α -linolenic acid (C18:3n-3) levels were found as 24.6%, 15% and 4.7%, respectively. In our study, we had higher oleic acid (C18:1n-9) (28.21%) and lower linoleic acid (C18:2n-6) (7.46%) levels compared to Teh et al. (2021). Marine haptophyte species *Pavlova lutheri* is known rich in protein and lipid content due to their large cell and ability to accumulate nutrients from culture water. *Pavlova lutheri* is known as rich in protein content among microalgae species (Shah et al., 2014). In our study, the crude protein content ($39.02 \pm 0.32\%$) was detected highest value among four microalgae species. The other nutrients also showed good levels of crude lipid, crude ash, and dry matter contents at a level of $19.96 \pm 0.9\%$, $10.01 \pm 0.88\%$, and $6.21 \pm 0.33\%$, respectively.

Fradique et al. (2013) reported crude protein ($38.4 \pm 0.2\%$), crude lipid ($17.9 \pm 0.5\%$), crude ash ($18.04 \pm 0.8\%$), and dry matter content ($91.03 \pm 0.01\%$) determined in ‰25 salinity culture conditions of *Diacronema vlkanium*. In our study, salinity was adjusted at ‰30 – ‰32 salinity, the contents of crude protein, lipid, ash, and dry matter were found as $38.4 \pm 0.55\%$, $18.01 \pm 0.99\%$, $18.45 \pm 0.83\%$, $5.89 \pm 0.96\%$, respectively. In another study, Cañavate and Fernández-Díaz (2022) showed lipid and fatty acid composition of *D. vlkanium* at different salinity levels. According to this study, EPA and DHA levels were found 7.6% and 6.6% of total fatty acids between ‰20 - ‰35 different salinity ranges in *D. vlkanium* production. In our study, essential fatty acids showed moderate levels of EPA and DHA ($0.06 \pm 0.01\%$ and $0.09 \pm 0.02\%$) at similar salinity levels. This result could be related to the fatty acid elongation of microalgae as long as salinity increases (Cañavate and Fernández-Díaz, 2022). We assume that EPA and DHA levels were lin-

early correlated with a high salinity in *D. vlkanium* species. In terms of marine haptophyte *Pavlova lutheri* has essential fatty acids such as EPA and ARA of total fatty acids. We obtained high accumulation of EPA (6.84%) and ARA (3.16%) levels in this haptophyte algae.

As a result, *D. vlkanium* can be cultured with the highest growth performance under phototrophic cultivation when compared to other microalgae species that were examined. However, all microalgae enhanced cell density after the 18th day of the experiment due to gentle aeration and flow current of culture water. The aeration positively effects the microalgal cell density and growth performance due to increases in the amount of CO₂ and nutrient content in the phototrophic culture conditions (Mohsenpour and Willoughby, 2016). *Euglena gracilis* and *Chlorella vulgaris* are productive species together with high protein and biomass contents. Dry matter is important when powder product is concerned with microalgae. The highest dry matter content was found in *Pavlova lutheri*. From this point, *P. lutheri* is a suitable species for the production of biomass and turn into dry material which features of highest dry matter content. *Euglena gracilis* have high content of ALA, ARA, EPA, and DHA which are important for fish feed raw materials (Wang et al., 2018). Microalgae fatty acid contents depend on the aeration, amount of CO₂, light intensity, temperature, and culture medium (Schwarzans et al., 2015, Guedes et al., 2010, Go et al., 2012). In our study, the highest ALA, ARA, and DHA fatty acids contents were found in *E. gracilis*. This result could be related to, the cultivation of microalgae by phototrophic methods. However, the highest EPA content was found in *Pavlova lutheri*. EPA is highly important for fish feeding and larval development. EPA and DHA fatty acids are very difficult to synthesize from fish (Guedes et al., 2010). That's the reason why the aquaculture industry has to use rich EPA and DHA contents from microalgae species.

CONCLUSION

In conclusion, within this study, we applied the same microalgae culture procedure at commercial hatcheries in our laboratory where microalgae culture start with small vessels and then continuously inoculated a large volume with gentle aeration. The purpose of this work was to investigate both the growth of algae during 32 days of culture (250 mL and 1000 mL glass flasks) and nutritional value and fatty acid composition at the end of the 32nd day of culture just before they were inoculated in 30 L plastic bags. The data obtained from our laboratory is valuable for both commercial hatcheries where those microalgae are utilized. To sum up, the success of microalgal up-scale culture in both freshwater and marine microalgae species are strongly related to inoculation time and volume. As a result, all microalgae have a high potential for biomass gain in a very short time with good enough nutrients. Moreover, *Pavlova lutheri* and *Euglena gracilis* can supply promising levels of highly unsaturated essential fatty acids such as ARA, EPA, and DHA. Most importantly, we suggest based on obtained data these two microalgae have high potential for dry biomass production due to their high dry matter content. Therefore, those biomass have high potential to use feed ingredients in aquaculture and this can lead to a positive effect on sustainable production. Additionally, we conclude that laboratory-scale production of

those four microalgae should be inoculated from a 250 mL culture flask to 1000 mL flasks at two weeks. Microalgae biomass production and its nutrient compositions are affected by culture systems like photobioreactors, volumes, and culture types such as phototrophic, myxotrophic, and heterotrophic culture. Further studies are needed for larger photobioreactor production and biomass investigations.

Acknowledgement: -

Conflict of Interest: The authors declared that they have no conflict of interest.

Ethics Committee approval: Ethics committee approval is not required.

Financial Disclosure: -

REFERENCES

- AOAC (1995). Official methods of analysis of the association of analytical chemistry (15th ed.). Arlington, VA: AOAC.
- Ahmed, F., Zhou, W., & Schenk, P. M. (2015). Pavlova lutheri is a high-level producer of phytosterols. *Algal Research*, 10, 210-217. <https://doi.org/10.1016/j.algal.2015.05.013>
- Ahmad, M. T., Shariff, M., Md. Yusoff, F., Goh, Y. M., & Banerjee, S. (2020). Applications of microalga *Chlorella vulgaris* in aquaculture. *Reviews in Aquaculture*, 12(1), 328-346. <https://doi.org/10.1111/raq.12320>
- Arkonrat, W., Deemark, P., & Oniam, V. (2016). Growth performance and proximate composition of mixed cultures of marine microalgae (*Nannochloropsis* sp. & *Tetraselmis* sp.) with monocultures. *Songklanakarin Journal of Science and Technology*, 38(1), 1-5.
- Aslam, A., Rasul, S., Bahadar, A., Hossain, N., Saleem, M., Hussain, S., Rasool, L. & Manzoor, H. (2021). Effect of micronutrient and hormone on microalgae growth assessment for biofuel feedstock. *Sustainability*, 13(9), 5035. <https://doi.org/10.3390/su13095035>
- Bashir, K. M. I., Mansoor, S., Kim, N. R., Grohmann, F. R., Shah, A. A., & Cho, M. G. (2019). Effect of organic carbon sources and environmental factors on cell growth and lipid content of *Pavlova lutheri*. *Annals of Microbiology*, 69(4), 353-368. <https://doi.org/10.1007/s13213-018-1423-2>
- Begum, H., Yusoff, F. M., Banerjee, S., Khatoon, H., & Shariff, M. (2016). Availability and utilization of pigments from microalgae. *Critical Reviews in Food Science and Nutrition*, 56(13), 2209-2222. <https://doi.org/10.1080/10408398.2013.764841>
- Becker, E. W. (2007). Micro-algae as a source of protein. *Biotechnology Advances*, 25(2), 207-210. <https://doi.org/10.1016/j.biotechadv.2006.11.002>
- Budge, S. M., Parrish, C. C., & McKenzie, C. H. (2001). Fatty acid composition of phytoplankton, settling particulate matter and sediments at a sheltered bivalve aquaculture site. *Marine Chemistry*, 76(4), 285-303. [https://doi.org/10.1016/S0304-4203\(01\)00068-8](https://doi.org/10.1016/S0304-4203(01)00068-8)
- Cañavate, J. P., & Fernández-Díaz, C. (2022). Salinity induces unique changes in lipid classes and fatty acids of the estuarine haptophyte *Diacronema vlkianum*. *European Journal of Phycology*, 57(3), 297-317. <https://doi.org/10.1080/09670262.2021.1970234>
- Camacho-Rodríguez, J., Macías-Sánchez, M. D., Cerón-García, M. C., Alarcón, F. J., & Molina-Grima, E. (2018). Microalgae as a potential ingredient for partial fish meal replacement in aquafeeds: nutrient stability under different storage conditions. *Journal of Applied Phycology*, 30, 1049-1059. <https://doi.org/10.1007/s10811-017-1281-5>
- Chaisutyakorn, P., Praiboon, J., & Kaewsuralikhit, C. (2018). The effect of temperature on growth and lipid and fatty acid composition on marine microalgae used for biodiesel production. *Journal of Applied Phycology*, 30, 37-45. <https://doi.org/10.1007/s10811-017-1186-3>
- Chen, C. Y., Zhao, X. Q., Yen, H. W., Ho, S. H., Cheng, C. L., Lee, D., Bai, F. & Chang, J. S. (2013). Microalgae-based carbohydrates for biofuel production. *Biochemical Engineering Journal*, 78, 1-10. <https://doi.org/10.1016/j.bej.2013.03.006>
- Chiu, S. Y., Kao, C. Y., Huang, T. T., Lin, C. J., Ong, S. C., Chen, C. D., Chang, J. & Lin, C. S. (2011). Microalgal biomass production and on-site bioremediation of carbon dioxide, nitrogen oxide and sulfur dioxide from flue gas using *Chlorella* sp. cultures. *Bioresource Technology*, 102(19), 9135-9142. <https://doi.org/10.1016/j.biortech.2011.06.091>
- Christie, W. W. (1982). Lipid analysis (2nd revised ed., p. 201). Oxford: Pergamon Press.
- Das, P., Mandal, S. C., Bhagabati, S. K., Akhtar, M. S., & Singh, S. K. (2012). Important live food organisms and their role in aquaculture. *Frontiers in Aquaculture*, 5(4), 69-86.
- da Silva Ferreira, V., & Sant'Anna, C. (2017). Impact of culture conditions on the chlorophyll content of microalgae for biotechnological applications. *World Journal of Microbiology and Biotechnology*, 33(1), 20. <https://doi.org/10.1007/s11274-016-2181-6>
- de Souza Celente, G., Rizzetti, T. M., Sui, Y., & de Souza Schneider, R. D. C. (2022). Potential use of microalga *Dunaliella salina* for bioproducts with industrial relevance. *Biomass and Bioenergy*, 167, 106647. <https://doi.org/10.1016/j.biombioe.2022.106647>
- Eryalçın, K. M., Roo, J., Saleh, R., Atalah, E., Benítez, T., Betancor, M., Hernandez-Cruz, M. & Izquierdo, M. (2013). Fish oil replacement by different microalgal products in microdiets for early weaning of gilthead sea bream (*Sparus aurata*, L.). *Aquaculture Research*, 44(5), 819-828. <https://doi.org/10.1111/j.1365-2109.2012.03237.x>
- Eryalçın, K. M., & Yıldız, M. (2015). Effects of long-term feeding with dried microalgae added microdiets on growth and fatty acid composition of gilthead sea bream (*Sparus aurata* L., 1758). *Turkish Journal of Fisheries and Aquatic Sciences*, 15(4), 905-915. https://doi.org/10.4194/1303-2712-v15_4_14
- Eryalçın, K. M., Ganuza, E., Atalah, E., & Hernández Cruz, M. C. (2015). *Nannochloropsis gaditana* and *Cryptocodinium cohnii*, two microalgae as alternative sources of essential fatty acids in early weaning for gilthead seabream. *Hidrobiológica*, 25(2), 193-202.
- Eryalçın, K. (2018). Effects of different commercial feeds and enrichments on biochemical composition and fatty acid profile of rotifer (*Brachionus plicatilis*, Muller 1786) and *Artemia franciscana*. *Turkish Journal of Fisheries and Aquatic Sciences*, 18. https://doi.org/10.4194/1303-2712-v18_1_09
- Eryalçın, K. M. (2019). Nutritional value and production performance of the rotifer *Brachionus plicatilis* Müller, 1786 cultured with different feeds at commercial scale. *Aquaculture International*, 27(3), 875-890. <https://doi.org/10.1007/s10499-019-00375-5>
- Fields, M. W., Hise, A., Lohman, E. J., Bell, T., Gardner, R. D., Corredor, L., Characklis, G. & Gerlach, R. (2014). Sources and resources: importance of nutrients, resource allocation, and ecology in microalgal cultivation for lipid accumulation. *Applied Microbiology and Biotechnology*, 98, 4805-4816. <https://doi.org/10.1007/s00253-014-5694-7>
- Folch, J., Lees, M., & Sloane Stanley, G. H. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226(1), 497-509.
- Fradique, M., Batista, A. P., Nunes, M. C., Gouveia, L., Bandarra, N. M., & Raymundo, A. (2013). Isochrysis galbana and *Diacronema vlkianum* biomass incorporation in pasta products as PUFA's source. *LWT-Food Science and Technology*, 50(1), 312-319. <https://doi.org/10.1016/j.lwt.2012.05.006>
- Guillard, R. R. (1975). Culture of phytoplankton for feeding marine

- invertebrates. In *Culture of marine invertebrate animals: proceedings—1st conference on culture of marine invertebrate animals greenport* (pp. 29-60). Boston, MA: Springer US.
- Go, S., Lee, S. J., Jeong, G. T., & Kim, S. K. (2012). Factors affecting the growth and the oil accumulation of marine microalgae, *Tetraselmis suecica*. *Bioprocess and Biosystems Engineering*, 35, 145-150. <https://doi.org/10.1007/s00449-011-0635-7>
- Goh, B. H. H., Ong, H. C., Cheah, M. Y., Chen, W. H., Yu, K. L., & Mahlia, T. M. I. (2019). Sustainability of direct biodiesel synthesis from microalgae biomass: A critical review. *Renewable and Sustainable Energy Reviews*, 107, 59-74. <https://doi.org/10.1016/j.rser.2019.02.012>
- Gharajeh, N. H., Valizadeh, M., Dorani, E., & Hejazi, M. A. (2020). Biochemical profiling of three indigenous *Dunaliella* isolates with main focus on fatty acid composition towards potential biotechnological application. *Biotechnology Reports*, 26, e00479. <https://doi.org/10.1016/j.btre.2020.e00479>
- Gu, G., Ou, D., Chen, Z., Gao, S., Sun, S., Zhao, Y., Hu, C., & Liang, X. (2022). Metabolomics revealed the photosynthetic performance and metabolomic characteristics of *Euglena gracilis* under autotrophic and mixotrophic conditions. *World Journal of Microbiology and Biotechnology*, 38(9), 160. <https://doi.org/10.1007/s11274-022-03346-w>
- Guedes, A. C., Meireles, L. A., Amaro, H. M., & Malcata, F. X. (2010). Changes in lipid class and fatty acid composition of cultures of *Pavlova lutheri*, in response to light intensity. *Journal of the American Oil Chemists' Society*, 87(7), 791-801. <https://doi.org/10.1016/j.btre.2020.e00479>
- He, Q., Yang, H., Wu, L., & Hu, C. (2015). Effect of light intensity on physiological changes, carbon allocation and neutral lipid accumulation in oleaginous microalgae. *Bioresource Technology*, 191, 219-228. <https://doi.org/10.1016/j.biortech.2015.05.021>
- Hemaiswarya, S., Raja, R., Ravi Kumar, R., Ganesan, V., & Anbazhagan, C. (2011). Microalgae: a sustainable feed source for aquaculture. *World Journal of Microbiology and Biotechnology*, 27, 1737-1746. <https://doi.org/10.1007/s11274-010-0632-z>
- Izquierdo, M. S., T. Watanabe, T. Takeuchi, T. Arakawa & C. Kitajima. (1990). Optimal EFA levels in *Artemia* to meet the EFA requirements of red seabream (*Pagrus major*). In: Takeda, M. & T. Watanabe. (Eds.). *The Current Status of Fish Nutrition in Aquaculture*. Tokyo University Fisheries, Tokyo, pp. 221-232.
- Jeong, U., Choi, J. K., Kang, C. M., Choi, B. D., & Kang, S. J. (2016). Effects of culture methods on the growth rates and fatty acid profiles of *Euglena gracilis*. *Korean Journal of Fisheries and Aquatic Sciences*, 49(1), 38-44.
- Khatoun, H., Rahman, N. A., Suleiman, S. S., Banerjee, S., & Abol-Munafi, A. B. (2019). Growth and proximate composition of *Scenedesmus obliquus* and *Selenastrum bibrainum* cultured in different media and condition. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 89, 251-257. <https://doi.org/10.1007/s40011-017-0938-9>
- Kottuparambil, S., Thankamony, R. L., & Agusti, S. (2019). *Euglena* as a potential natural source of value-added metabolites. A review. *Algal Research*, 37, 154-159. <https://doi.org/10.1016/j.algal.2018.11.024>
- Kumaran, M., Palanisamy, K. M., Bhuyar, P., Maniam, G. P., Rahim, M. H. A., & Govindan, N. (2023). Agriculture of microalgae *Chlorella vulgaris* for polyunsaturated fatty acids (PUFAs) production employing palm oil mill effluents (POME) for future food, wastewater, and energy nexus. *Energy Nexus*, 9, 100169. <https://doi.org/10.1016/j.nexus.2022.100169>
- Lau, Z. L., Low, S. S., Ezeigwe, E. R., Chew, K. W., Chai, W. S., Bhatnagar, A., Yap, Y. & Show, P. L. (2022). A review on the diverse interactions between microalgae and nanomaterials: growth variation, photosynthesis performance and toxicity. *Bioresource Technology*, 127048. <https://doi.org/10.1016/j.biortech.2022.127048>
- Liu, Z. Y., Wang, G. C., & Zhou, B. C. (2008). Effect of iron on growth and lipid accumulation in *Chlorella vulgaris*. *Bioresource Technology*, 99(11), 4717-4722. <https://doi.org/10.1016/j.biortech.2007.09.073>
- Metsoviti, M. N., Papapolymerou, G., Karapanagiotidis, I. T., & Katsoulas, N. (2019). Effect of light intensity and quality on growth rate and composition of *Chlorella vulgaris*. *Plants*, 9(1), 31. <https://doi.org/10.3390/plants9010031>
- Mohsenpour, S. F., & Willoughby, N. (2016). Effect of CO₂ aeration on cultivation of microalgae in luminescent photobioreactors. *Biomass and Bioenergy*, 85, 168-177. <https://doi.org/10.1016/j.biombioe.2015.12.002>
- Neori, A. (2011). "Green water" microalgae: the leading sector in world aquaculture. *Journal of Applied Phycology*, 23, 143-149. <https://doi.org/10.1007/s10811-010-9531-9>
- Nwoye, E. C., Chukwuma, O. J., Obisike, N. O., Shedrack, O. I., & Nwuche, C. O. (2017). Evaluation of some biological activities of *Euglena gracilis* biomass produced by a fed-batch culture with some crop fertilizers. *African Journal of Biotechnology*, 16(8), 337-345. <https://doi.org/10.5897/AJB2016.15651>
- Parrish, C. C., Wells, J. S., Yang, Z., & Dabinett, P. (1998). Growth and lipid composition of scallop juveniles *Placopecten magellanicus* fed the flagellate *Isochrysis galbana* with varying lipid composition and the diatom *Chaetoceros muelleri*. *Marine Biology*, 133, 461-471. <https://doi.org/10.1007/s002270050486>
- Patil, V., Reitan, K. I., Knutsen, G., Mortensen, L. M., Källqvist, T., Olsen, E., Vogt, G. & Gislerød, H. R. (2005). Microalgae as source of polyunsaturated fatty acids for aquaculture. *Plant Biology*, 6(6), 57-65.
- Patil, V., Källqvist, T., Olsen, E., Vogt, G., & Gislerød, H. R. (2007). Fatty acid composition of 12 microalgae for possible use in aquaculture feed. *Aquaculture International*, 15, 1-9. <https://doi.org/10.1007/s10499-006-9060-3>
- Pazos, A. J., Román, G., Acosta, C. P., Sánchez, J. L., & Abad, M. (1997). Lipid classes and fatty acid composition in the female gonad of *Pecten maximus* in relation to reproductive cycle and environmental variables. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 117(3), 393-402. [https://doi.org/10.1016/S0305-0491\(97\)00135-1](https://doi.org/10.1016/S0305-0491(97)00135-1)
- Peltomaa, E., Johnson, M. D., & Taipale, S. J. (2017). Marine cryptophytes are great sources of EPA and DHA. *Marine Drugs*, 16(1), 3. <https://doi.org/10.3390/md16010003>
- Procházková, G., Brányiková, I., Zachleder, V., & Brányik, T. (2014). Effect of nutrient supply status on biomass composition of eukaryotic green microalgae. *Journal of Applied Phycology*, 26, 1359-1377. <https://doi.org/10.1007/s10811-013-0154-9>
- Raja, R., Anbazhagan, C., Lakshmi, D., & Rengasamy, R. (2004). Nutritional studies on *Dunaliella salina* (Volvocales, Chlorophyta) under laboratory conditions. *Seaweed Resources Utilization*, 26(1&2), 127-146.
- Sandnes, J. M., Källqvist, T., Wenner, D., & Gislerød, H. R. (2005). Combined influence of light and temperature on growth rates of *Nannochloropsis oceanica*: linking cellular responses to large-scale biomass production. *Journal of Applied Phycology*, 17, 515-525. <https://doi.org/10.1007/s10811-005-9002-x>
- Schwarzthans, J. P., Cholewa, D., Grimm, P., Beshay, U., Risse, J. M., Friehs, K., & Flaschel, E. (2015). Dependency of the fatty acid composition of *Euglena gracilis* on growth phase and culture conditions. *Journal of Applied Phycology*, 27, 1389-1399. <https://doi.org/10.1007/s10811-014-0458-4>
- Shaaban, M. M., El-Saady, A. M., & El-Sayed, A. B. (2010). Green microalgae water extract and micronutrients foliar application as promoters to nutrient balance and growth of wheat plants. *Journal of American Science*, 6(9), 631-636.
- Shah, S. M. U., Che Radziah, C., Ibrahim, S., Latiff, F., Othman, M. F., &

- Abdullah, M. A. (2014). Effects of photoperiod, salinity and pH on cell growth and lipid content of *Pavlova lutheri*. *Annals of Microbiology*, 64(1), 157-164. <https://doi.org/10.1007/s13213-013-0645-6>
- Shah, M. R., Lutz, G. A., Alam, A., Sarker, P., Kabir Chowdhury, M. A., Parsaeimehr, A., Liang, Y. & Daroch, M. (2018). Microalgae in aquafeeds for a sustainable aquaculture industry. *Journal of Applied Phycology*, 30, 197-213. <https://doi.org/10.1007/s10811-017-1234-z>
- Singh, J., & Saxena, R. C. (2015). An introduction to microalgae: diversity and significance. In *Handbook of marine Microalgae* (pp. 11-24). Academic Press. <https://doi.org/10.1016/B978-0-12-800776-1.00002-9>
- Soto-Sánchez, O., Hidalgo, P., González, A., Oliveira, P. E., Hernández Arias, A. J., & Dantagnan, P. (2023). Microalgae as raw materials for aquafeeds: Growth kinetics and improvement strategies of polyunsaturated fatty acids production. *Aquaculture Nutrition*, 2023. <https://doi.org/10.1155/2023/5110281>
- Spínola, M. P., Costa, M. M., & Prates, J. A. (2023). Enhancing Digestibility of *Chlorella vulgaris* Biomass in Monogastric Diets: Strategies and Insights. *Animals*, 13(6), 1017. <https://doi.org/10.3390/ani13061017>
- Spolaore, P., Joannis-Cassan, C., Duran, E., & Isambert, A. (2006). Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, 101(2), 87-96. <https://doi.org/10.1263/jbb.101.87>
- Taş, B., & Dalkıran, T. G. (2022). Investigation of the Effect of Zero-Valent Iron Nanoparticle on *Chlorella* sp. Growth in Autotrophic, Mixotrophic and Heterotrophic Cultures. *Review of Hydrobiology*, 15, 1-20.
- Teh, K. Y., Loh, S. H., Aziz, A., Takahashi, K., Effendy, A. W. M., & Cha, T. S. (2021). Lipid accumulation patterns and role of different fatty acid types towards mitigating salinity fluctuations in *Chlorella vulgaris*. *Scientific Reports*, 11(1), 1-12. <https://doi.org/10.1038/s41598-020-79950-3>
- Turcihan, G., Turgay, E., Yardımcı, R. E., & Eryalçın, K. M. (2021). The effect of feeding with different microalgae on survival, growth, and fatty acid composition of *Artemia franciscana* metanauplii and on predominant bacterial species of the rearing water. *Aquaculture International*, 29(5), 2223-2241. <https://doi.org/10.1007/s10499-021-00745-y>
- Turcihan, G., Isinibilir, M., Zeybek, Y. G., & Eryalçın, K. M. (2022). Effect of different feeds on reproduction performance, nutritional components and fatty acid composition of cladocer water flea (*Daphnia magna*). *Aquaculture Research*, 53(6), 2420-2430. <https://doi.org/10.1111/are.15759>
- Wang, Y., Seppänen-Laakso, T., Rischer, H., & Wiebe, M. G. (2018). *Euglena gracilis* growth and cell composition under different temperature, light and trophic conditions. *PLoS One*, 13(4), e0195329. <https://doi.org/10.1371/journal.pone.0195329>
- Wollmann, F., Dietze, S., Ackermann, J. U., Bley, T., Walther, T., Steingroewer, J., & Krujatz, F. (2019). Microalgae wastewater treatment: Biological and technological approaches. *Engineering in Life Sciences*, 19(12), 860-871. <https://doi.org/10.1002/elsc.201900071>
- Yeh, K. L., Chang, J. S., & Chen, W. M. (2010). Effect of light supply and carbon source on cell growth and cellular composition of a newly isolated microalga *Chlorella vulgaris* ESP-31. *Engineering in Life Sciences*, 10(3), 201-208. <https://doi.org/10.1002/elsc.200900116>
- Zhang, K., Wan, M., Bai, W., He, M., Wang, W., Fan, F., Guo, J., Yu, T. & Li, Y. (2023). A novel method for extraction of paramylon from *Euglena gracilis* for industrial production. *Algal Research*, 71, 103058. <https://doi.org/10.1016/j.algal.2023.103058>
- Zhao, B., Zhang, Y., Xiong, K., Zhang, Z., Hao, X., & Liu, T. (2011). Effect of cultivation mode on microalgal growth and CO₂ fixation. *Chemical Engineering Research and Design*, 89(9), 1758-1762. <https://doi.org/10.1016/j.cherd.2011.02.018>

Growth Performance, Survival Rate, and Water Quality in an Aquaculture System Using Different Feeding Strategies for Juveniles of Nile Tilapia (*Oreochromis niloticus*)

Dagon Ribeiro¹ , Emerson de Carvalho² , Gustavo Fonseca³ 

Cite this article as: Ribeiro, D., Carvalho, E., & Fonseca, G. (2024). Growth performance, survival rate, and water quality in an aquaculture system using different feeding strategies for juveniles of Nile tilapia (*Oreochromis niloticus*). *Aquatic Sciences and Engineering*, 39(1), 17-23. DOI: <https://doi.org/10.26650/ASE20241338060>

ABSTRACT

Aquaculture is a rapidly growing industry worldwide, with Nile tilapia (*Oreochromis niloticus*) being one of the most intensively farmed fish species. This study aimed to evaluate the growth performance and water quality parameters in different culture systems for Nile tilapia. Six treatments were tested, including variations in feed type (commercial or microalgae), aeration, and their combinations. The results showed that the presence of commercial feed and aeration (T2) resulted in the highest weight gain and specific growth rates, while treatments without commercial feed showed lower growth performance. The addition of microalgae supplementation did not significantly improve growth compared to commercial feed alone. Water quality parameters, particularly nitrite levels and dissolved oxygen, played crucial roles in the production of tilapia. It was observed that high nitrite levels were associated with decreased growth and survival rates. Proper monitoring and management of water quality, including nitrite levels and dissolved oxygen, are essential to ensure the survival and growth of tilapia in aquaculture systems. These findings highlight the importance of implementing sustainable practices and appropriate feeding strategies to optimize the growth and well-being of farmed tilapia while minimizing environmental impacts.

Keywords: Fish nutrition, fish production, fish farming, water quality, microalgae, *Oreochromis niloticus*

ORCID IDs of the author:

D.R. 0000-0001-6179-2855;
E.D.C. 0000-0002-4865-6784;
G.F. 0000-0002-8784-661X

¹Federal University of Grande Dourados, Brazil

²Federal University of Southern Bahia, Brazil

³University of Akureyri, Faculty of Natural Resource Sciences, Borgir v. Nordurslod, Akureyri, Iceland

Submitted:
04.08.2023

Revision Requested:
07.09.2023

Last Revision Received:
21.09.2023

Accepted:
27.09.2023

Online Published:
13.11.2023

Correspondence:
Gustavo Graciano Fonseca
E-mail:
gustavo@unak.is

INTRODUCTION

Aquaculture is a thriving industry worldwide, characterized by diversity and expansion (Sarker et al., 2016; Verdegem et al., 2023). In developing countries, this sector has experienced significant growth due to ongoing efforts in regulation, professionalization, and modernization. Farmers have become increasingly knowledgeable about management practices and essential inputs (Scorvo-Filho et al., 2010; Moreira et al., 2012).

Fish farming is expanding at a faster rate compared to other livestock commodities (FAO, 2022). However, despite notable progress, the

adoption of technology and technical capabilities still restricts overall achievements and expectations. Addressing the reduction of production costs and environmental impact is crucial. These are critical obstacles that need to be overcome for a more optimistic and sustainable outlook (Leonardo et al., 2009).

In Brazil, Nile tilapia (*Oreochromis niloticus*) is the most intensively farmed fish species. This is primarily due to its rapid growth, efficient feed conversion, desirable meat quality, and market acceptability both locally and internationally (Schwarz et al., 2010). In fact, tilapia is the second most cultivated fish worldwide, following carp (Fonseca et al., 2013).



Successful fish farming in ponds relies on effective management of nutrition and water quality. In most cases, the use of complete diets is necessary to meet the nutritional requirements of fish, ensuring growth, high productivity, and profitability (Furuya et al., 2001; Carvalho et al., 2012). This becomes particularly crucial when fish are bred or fattened under confined conditions, such as cages, ponds, and raceways, where high biomass per unit area is present.

In an attempt to increase income, farmers often increase stocking rates (density), but this may negatively impact growth and result in poor water quality for the fish (Lima et al., 2016). Such conditions affect survival rates since suitable water quality is essential for the physiological functions of fish, including breathing, reproduction, feeding, and defecation. Water quality in fishponds may also depend on the availability and nutritional quality of food (Bhatnagar and Devi, 2013).

At the farm level, intensification typically leads to deteriorating water quality in fish ponds, posing hazards and increased risks to surrounding ecosystems. Effluents from aquaculture activities can contain substantial amounts of feces, leftover feed, and bacterial biomass (Gorlach-Lira et al., 2013). Therefore, ensuring constant monitoring is crucial to maintain the sustainability of fish farming, preventing downstream contamination and environmental degradation. In ponds with limited water renewal, the effects of intensification become even more problematic. Aeration techniques offer alternatives to increase the carrying capacity of a pond for fish (Boyd et al., 2018). Nonetheless, water renewal remains important to dilute the concentration of toxic metabolites.

The concentration of microalgae in fishponds also influences fish performance and water quality. Microalgae are responsible for recycling nutrients excreted by fish (Gorlach-Lira et al., 2013). They also contribute to the dissolved oxygen content in the water through photosynthesis and can adsorb significant amounts of heavy metals from contaminated water sources (Coelho et al., 2014).

Moreover, certain microalgae serve as direct or indirect sources of food for fish and their ecosystem (Sarker et al., 2016). This is particularly interesting considering that production costs, especially the high cost of feed, pose major constraints in the global fish farming scenario (Taelman et al., 2013). Consequently, research efforts have focused on identifying alternative feeding strategies (Leonardo et al., 2009; Carvalho et al., 2012). In general, these alternatives aim to promote

growth, minimize stress, and enhance the efficiency of the fish immune system (Ungsethaphand et al., 2010; Sarker et al., 2016; Zeinab et al., 2015).

The aim of this study was to evaluate the growth performance and the survival rates of Nile tilapia (*Oreochromis niloticus*) and the water quality in an aquaculture system using different feeding strategies. For that, we examined the ability of fish to thrive in ponds without water renewal under various experimental conditions and monitored water quality to assess variations based on treatments involving feed, aeration, and microalgae supplementation.

MATERIALS AND METHODS

An aquaculture system consisting of circular open ponds without water renewal was utilized for raising Nile tilapia (*Oreochromis niloticus*). The ponds were made of polyethylene plastic, with a capacity of 1000 L, filled with 500 L of water. Juvenile Nile tilapia (53.2 ± 5.4 g; 8.0 ± 1.0 cm) were bred at an initial stocking density of 30 fish per pond ($n=30$), resulting in an average density of $3,192 \text{ g m}^{-3}$. The system was built on a fish farm located in Dourados, MS, Brazil.

Six treatments were designed to evaluate the impact of different factors on the growth and development of juvenile Nile tilapia. These treatments included: commercial feed (T1), commercial feed with aeration (T2), microalgae supplementation (T3), microalgae supplementation with aeration (T4), commercial feed with microalgae supplementation (T5), and commercial feed with microalgae supplementation and aeration (T6) (Table 1).

The commercial feed contained 40% crude protein. The microalgae used in the study was *Chlorella sorokiniana* CTT 7727 (3.9×10^6 cells mL^{-1} ; 0.39 g L^{-1}), obtained from the André Tosselo Foundation (FAT). The strain was cultivated in a separate open pond using the Bold Basal medium (Bischoff and Bold, 1963). The temperature of the ponds was maintained at $28 \pm 2^\circ\text{C}$ using thermostats, while the photoperiod followed a 12 h light/12 h dark cycle.

The experiment was conducted for two weeks. In the first week, the amount of commercial feed provided was fixed at 4% of the initial live weight (g) for treatments T1 and T2. For treatments involving multiple food sources (T5 and T6), the amount of feed was reduced to 50% of the other treatments as per dilution criteria (Table 1). In the second week, the feed amount was halved for all treatments. Feed was administered daily during four feeding periods throughout the entire experiment.

Table 1. Treatments offered to juvenile tilapia bred in open ponds with no water renewal during the early stages of development.

Treatment	Day 0 – 7	Day 8 – 14
T1 Commercial feed	64 g	32 g
T2 Commercial feed and aeration	64 g	32 g
T3 Microalgae	2,000 mL	1,000 mL
T4 Microalgae and aeration	2,000 mL	1,000 mL
T5 Commercial feed and microalgae	32 g + 1,000 mL	16 g + 500 mL
T6 Commercial feed, microalgae, and aeration	32 g + 1,000 mL	16 g + 500 mL

Water parameters such as pH, hardness, dissolved oxygen, toxic ammonia, and nitrite were measured daily using a commercial colorimetric disk kit. At the end of the experiment (day 14), water samples were collected for microbiological analysis, including total thermo-tolerant coliforms, *Escherichia coli*, and mesophilic bacteria, using the classical multiple-tube technique (APHA, 1998).

To evaluate the growth performance, fish were length measured and weighed before being immediately released into the ponds without inflicting any harm. The other parameters monitored were the specific growth rate (SGR) (Eq. 1), the daily weight gain (DWG) (Eq. 2) (Haque et al., 2023), and the condition factor (K) (Eq. 3) (Pauly, 1983):

$$\text{SGR (\% day}^{-1}\text{)} = [(W_2 - W_1) / W_1] \times (100 / T) \quad 1$$

$$\text{DWG (\% day}^{-1}\text{)} = [(W_2 - W_1) / W_1] \times 100 \quad 2$$

$$K (\text{g cm}^{-3}) = 100 W / L^3 \quad 3$$

Where: W = fish weight (g); W_1 = initial fish weight (g); W_2 = final fish weight (g); L = fish length (cm); T = the experimental period in days.

The survival rate (SR) was evaluated at the end of the experiment and expressed as follows (Eq. 4) (Haque et al., 2023):

$$\text{SR (\%)} = (\text{FH} / \text{FS}) \times 100 \quad 4$$

Where: FH = number of fish harvested; FS = number of fish stocked.

Variance tests (one-way ANOVA) and comparison of means (Tukey test, 5%) were conducted separately according to data of weight (g) and total length (cm) during the experimental trial. All analyzes were performed in triplicate.

RESULTS AND DISCUSSION

Development of Nile tilapia

The weight of the fish varied among treatments on both the 7th and 14th days of the experiment (Fig. 1). During the first week (day 7), treatments T2 and T5 exhibited better results. Subsequently, in the second week, T2 and T6 showed superior outcomes. Conversely, the absence of commercial feed (T3, T4) was associated with lower weight gain in both experimental periods (days 7 and 14) (Fig. 1). Similarly, fish body length was highest in T2 and T6, while it was lowest in T3 and T4 during the final evaluation trial (Fig. 2).

These results showed that the growth performance of Nile tilapia was influenced by various factors, primarily associated with the type of food provided (commercial or natural). However, it was expected that due to the efficient capacity of Nile tilapia to harvest microalgae from the water through their gill rakers (Turker et al., 2003), T6 and T5 would present, respectively, the best results for weight by supplying an extra source of nutrients to the fish diet (T5) in addition to an increment in the oxygen availability (T6), which is critical for fish to achieve their full growth and activity potential (Obirikorang et al., 2020).

The best treatment in terms of total weight gain was obtained by combining feed and aeration (T2), which resulted in an increase

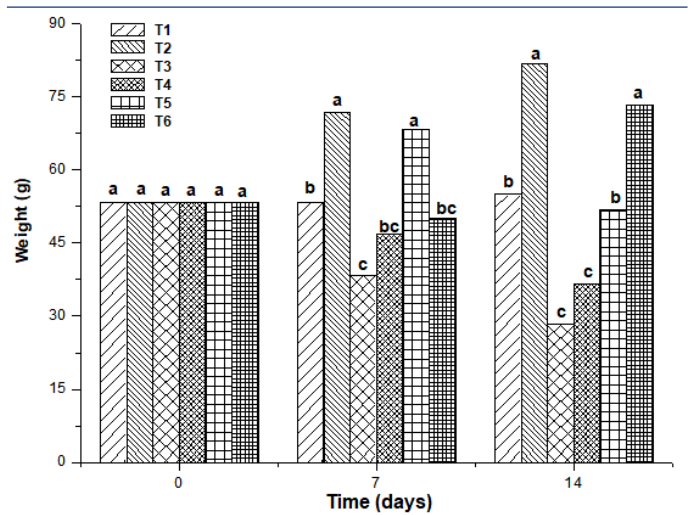


Figure 1. Analysis of variance of the increased weight (g) of Nile tilapia in relation to six treatments and time (Table 1). The results presented are the average of the weighted juveniles (N=5). *Means followed by the same letter do not differ by the Tukey test at the 5% probability level.

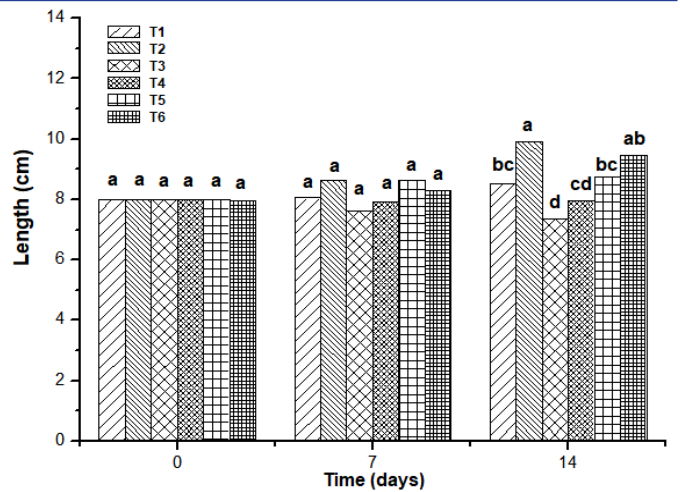


Figure 2. Analysis of variance of the increased total body length (cm) of juvenile Nile tilapia in relation to six treatments and time (Table 1). The results presented are the average of the measured juveniles (N=5). *Means followed by the same letter do not differ by the Tukey test at the 5% probability level.

of 53.33%. The treatment involving feed, microalgae, and aeration (T6) demonstrated the second-highest weight gain rate of 37.77%. However, this treatment also had the lowest survival rate of 53.3%. Solely relying on commercial feed (T1) was associated with poor performance in daily weight gain. In fact, treatments T3, T4, and T5 exhibited negative daily weight gain throughout the experiment. Thus, weight loss was not correlated with lower survival rates ($S_{\%}$) (Table 2). Weight loss can occur when fish are transitioning to a different feed or adjusting to new dietary con-

ditions. Moreover, environmental stressors, such as water quality issues, handling stress, or competition, can indeed lead to weight loss in fish without negatively impacting survival rates. These stressors can affect fish health and behavior, potentially causing reduced feeding, increased energy expenditure, or physiological changes that result in weight loss. The impact on survival rates will depend on the severity and duration of these stressors (Martins et al., 2012; Toni et al., 2017; Yavuzcan et al., 2017; Hvas et al., 2020; Canosa and Bertucci, 2023).

On the other hand, the specific growth rates of tilapia cultivated in T2 (3.80% day⁻¹) and T6 (2.69% day⁻¹) (Table 2) were higher or comparable to those reported in the literature for juveniles (Table 3).

Finally, the condition factor seems to better explain the growth behavior by combining information from weight and length (Table 2). The results underline that T3 and T4 were the worst treatments for the Nile tilapia development, *i.e.*, microalga is not a good enough source of nutrients to sustain growth, at least at the utilized concentration. However, the other treatments (T1, T2, T5, and T6) did not show statistical differences ($P>0.05$) between them for this parameter.

Various studies have reported success in the partial replacement of commercial fish feed by microalgae biomass, but the inclusion of *Chlorella* sp. into feed in most cases evolves biomass processing, pre-treatment, and extrusion into pellets (Teuling et al., 2017; Tibbetts et al., 2017; Batista et al., 2018; de Cruz et al., 2018). However, a study conducted to evaluate the use of *Chlorella vulgaris* and *Scenedesmus obliquus* as an autotrophic bio floc technology

during Nile tilapia cultivation with commercial feed showed a positive effect on water purification and to prevent fish mortality. Nevertheless, their system utilized water exchange every 10 days, which included the microalgae supply of 0.01 g L⁻¹ (equivalent to 2 g per pond per day). Here it was utilized the maximum concentration of 0.78 g L⁻¹ (equivalent to 0.78 g per pond per day) (2000 mL of microalgae solution per pond per day; Table 1).

Evaluation of water quality

Table 4 presents the profile of the physical and chemical parameters of the water during two stages of the experimental trial. In the initial stage, the water in all ponds met the appropriate standards outlined in Resolution 357 (Brazil, 2005). This condition persisted until the end of the experiment, with only one exception.

By the end of the experiment, water in T1 was associated with low levels of dissolved oxygen, falling below the recommended thresholds. This is likely due to T1 ponds receiving twice the amount of commercial feed compared to treatments involving commercial feed (T2, T5, and T6). The concentration of dissolved oxygen (DO₂ (mg L⁻¹)) may gradually decrease with increased feed input. In terms of nitrite (NO₂-), treatments exhibited critical levels in the final stage of the experiment, except for T1.

Microbiological indicators of water quality revealed the presence of thermotolerant coliforms in all treatments at the end of the experiment. Regarding fecal coliforms (*E. coli*) and mesophilic bacteria, treatments involving commercial feed (T1, T2, T5, and T6) exhibited the highest values, ranging from 10⁴ to 10⁵ CFU 100 mL⁻¹ (Table 5).

Table 2. Specific growth rate, total weight gain, survival rate, and condition factor of Nile tilapia juveniles during 14 days of cultivation.

Treatment	Specific growth rate (% day ⁻¹)	Weight gain (% day ⁻¹)	Survival rate (%)	Condition factor (g cm ⁻³)
T1	0.34 (± 0.29) ^b	4.80 (± 3.1) ^b	86.7 ^a	8.96 ^a
T2	3.80 (± 0.37) ^a	53.33 (± 5.25) ^a	96.7 ^a	8.42 ^{ab}
T3	-3.26 (± 0.60) ^c	-45.70 (± 8.41) ^c	93.3 ^a	7.11 ^c
T4	-2.18 (± 0.52) ^c	-30.55 (± 7.40) ^c	93.3 ^a	7.58 ^c
T5	-0.19 (± 0.60) ^b	-2.72 (± 8.48) ^b	83.3 ^{ab}	7.67 ^{bc}
T6	2.69 (± 0.58) ^a	37.77 (± 8.14) ^a	53.3 ^b	8.64 ^{ab}

Different letters in the same column differ (Tukey test, 5% significance level). Treatments are according to Table 1.

Table 3. Specific growth rates (% day⁻¹) of juvenile Nile tilapia depending on treatment (feed), time period (days), and culture system reported elsewhere.

Feed	Time (days)	System	Specific growth rates (% day ⁻¹)		Reference
			Min.	Max.	
Microalgae supplementation	28	pond/constant aeration	0.21	0.35	Costa et al. (2011)
Organic wheat	15	pond/constant aeration	0.3	0.3	Lui et al. (2012)
Alternative protein	180	pond/constant aeration	1.0	1.1	Assano et al. (2011)
Commercial fed (CF)	29	pond/constant aeration	1.34	1.82	Coêlho et al. (2014)
Agroindustrial byproducts	60	cages/water recirculation	2.03	2.27	Carvalho et al. (2012)
CF and enzyme complex	62	pond/water recirculation	2.28	2.37	Signor et al. (2010)
Agroindustrial byproducts	70	pond/constant aeration	3.43	3.96	Workagegn et al. (2014)

Table 4. Physical and chemical parameters of water depending on feed and the presence of microalgae and aeration in Nile tilapia culture systems.

Treatment	pH		DO ₂ (mg L ⁻¹)		NH ₃ (mg L ⁻¹)		NO ₂ ⁻ (mg L ⁻¹)	
	Day 1	Day 14	Day 1	Day 14	Day 1	Day 14	Day 1	Day 14
T1	7.2	6.6	7.9	8.3	< LD	0.02	< LD	0.2
T2	7.2	6.2	7.9	7.9	< LD	0.001	< LD	2.8
T3	7.2	6.2	7.9	8.0	< LD	0.02	< LD	2.8
T4	7.2	6.2	7.9	7.9	< LD	0.02	< LD	2.8
T5	7.2	6.6	7.9	7.9	< LD	0.003	< LD	2.8
T6	7.2	6.4	7.9	7.9	< LD	0.02	< LD	2.8
Rv	6.0-9.0		≥ 5 mg L ⁻¹		≤ 2.0 mg L ⁻¹		≤ 1.0 mg L ⁻¹	

< LD = Values below the limit of detection. RV = reference values (guidelines of CONAMA 357/05); pH = hydrogen potential; DO₂ = dissolved oxygen; NH₃ = toxic ammonia; NO₂⁻ = nitrite. Treatments are according to Table 1.

Table 5. Counts of mesophilic total bacteria present in the water from ponds used to raise tilapia fish under different experimental conditions.

Treatment	TTC		<i>E. coli</i>	Mesophilic bacteria
	24 h	48 h	(CFU 100 mL ⁻¹)	(CFU 100 mL ⁻¹)
T1	+	+	6.06 x 10 ⁴ a	8.67 x 10 ⁴ c
T2	-	+	7.00 x 10 ⁴ a	4.70 x 10 ⁵ a
T3	+	+	1.45 x 10 ⁴ b	5.20 x 10 ⁴ d
T4	-	+	1.04 x 10 ⁴ b	6.50 x 10 ⁴ d
T5	+	+	1.80 x 10 ⁴ b	2.28 x 10 ⁵ b
T6	+	+	1.30 x 10 ⁴ b	2.23 x 10 ⁵ b

TTC: total thermo-tolerant coliforms after 24 and 48 h (+ presence or - absence); CFU: Colony-Forming Unit. *Means followed by the same letter do not differ by the Tukey test at the 5% probability level. Treatments are according to Table 1.

The level of dissolved oxygen (DO₂) in ponds with low water renewal is likely the major constraint in tilapia production. Thus, monitoring the feed ratio and regularly testing for dissolved oxygen levels is advisable. Tilapia can survive at low levels of dissolved oxygen as long as temperature, pH, and stocking density remain favorable (Caldini et al., 2013; Abdel-Tawwab et al., 2015).

High levels of nitrite can challenge the immune system of fish, leading to diseases and sudden death (Kroupova et al., 2005). Surprisingly, the nitrite levels in this study were at least twice as high as the reference values (RV) (Table 2). However, the reported levels of nitrite in the literature vary significantly and are frequently higher than the recommended values. Pereira and Lapolli (2009), Gorlach-Lira et al. (2013), Coêlho et al. (2014), and Lima et al. (2016) found lower values ranging from 0.1-2.0, 0.004-0.005, 0.0-0.27, and 0.26-0.46 mg L⁻¹, respectively. Interestingly, Santos et al. (2013) reported higher values ranging from 1.03-3.61 mg L⁻¹ in cultivation systems using water recirculation.

Overall, similar to the results observed in this study, the data reported in the literature show a discreet association between high

nitrite values and mortality or weight trends in tilapia (Pereira and Lapolli, 2009; Gorlach-Lira et al., 2013; Santos et al., 2013; Coêlho et al., 2014; Lima et al., 2016). However, this association may not be straightforward, limiting our understanding and comparisons between the cultivation strategies.

Recirculation systems are often more effective in controlling nitrite levels in fishponds. However, these systems require larger amounts of water, which can increase environmental footprints. Exploring alternatives to remove nitrite from the water may enhance production while also limiting the adoption of technology for family farmers. On the other hand, biotransformation techniques such as denitrification may reduce the investment required to mitigate the toxicity of effluents derived from tilapia production.

As per the reference values established in Resolution 357 (Brazil, 2005), confined animals should not be exposed to thermo-tolerant coliforms beyond 1000 per 100 milliliters (1.0 x 10³ CFU 100 mL⁻¹); *Escherichia coli* may be used as a substitute for thermo-tolerant coliforms based on the limits set by the competent environmental agency. The contamination of aquaculture systems and fish by human pathogens such as *E. coli* is usually attributed to infected handlers or storage (Rocha et al., 2014; Dewi et al., 2022).

The relatively high bacterial contamination observed in the experiment reflects the nature of fish farming conducted in ponds, which involves a considerable volume of dietary inputs in a relatively small area (Gorlach-Lira et al., 2013). Bacterial contamination was higher than the values recorded by Ahmed and Naim (2003) in tilapia culture ponds and by Gorlach-Lira et al. (2013) in tilapia culture floating net cages, ranging from 10³-10⁴ and 10⁴-10⁵ CFU 100 mL⁻¹, respectively. The results were similar to the values reported by Ntengwe and Edema (2008) in tilapia culture ponds (10⁶ CFU 100 mL⁻¹).

However, bacterial contamination is comparable to the critical values observed for dissolved oxygen and nitrite. The low levels of dissolved oxygen may be attributed to the decomposition of leftover feed (excess of commercial feed). The levels of total mesophilic aerobic bacteria support the evaluation of water in terms

of decomposition activity (Gorlach-Lira et al., 2013). Microbiological decomposition requires dissolved oxygen from the water, which can lead to critical levels of DO₂ that threaten the survival of fish. High nitrite values may result from this type of decomposition. Lima et al. (2016) observed that higher loads of organic matter are associated with greater amounts of total ammonia nitrogen, which serves as a substrate for *Nitrossomonas* to release nitrite into the water.

CONCLUSION

In conclusion, this study examined the growth performance and water quality parameters of Nile tilapia (*Oreochromis niloticus*) in different culture systems. The results demonstrated the significant impact of various factors, particularly the type of food provided, on the growth and development of the fish. The treatments combining commercial feed with aeration (T2) and commercial feed with microalgae supplementation and aeration (T6) showed the highest weight gain rates, while treatments without commercial feed exhibited lower weight gains. The specific growth rates of tilapia in T2 and T6 were comparable to those reported in the literature for juveniles. Proper monitoring and management of water quality, including nitrite levels and dissolved oxygen, played a crucial role in the survival and growth of Nile tilapia.

Acknowledgments: The authors gratefully acknowledge the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Fundação de Apoio ao Desenvolvimento do Ensino, Ciência e Tecnologia do Estado de Mato Grosso do Sul (FUNDECT) and Coordenação de Aperfeiçoamento Pessoal de Nível Superior (CAPES) for their financial support.

Ethics Committee Approval: All procedures used in experiments with animals followed the ethical standards of the Ethical Committee from the Federal University of Grande Dourados, Brazil.

Financial Disclosure: The study was financed by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq).

Conflict of interest: The authors declare that they have no conflicts of interest.

REFERENCES

- Abdel-Tawwab, M., Hagra, A. E., Elbaghdady, H. A. M. & Monier, M. N. (2015). Effects of dissolved oxygen and fish size on Nile tilapia, *Oreochromis niloticus* (L.): growth performance, whole-body composition, and innate immunity. *Aquacul. Int.* 23(5), 1261-1274. doi:10.1007/s10499-015-9882-y.
- Ahmed, H. A. & Naim, U. (2003). Quantitative and qualitative studies on bacterial flora of hybrid tilapia (*Oreochromis niloticus* x *O. arcus*) cultured in earthen ponds in Saudi Arabia. *Aquacul. Res.* 34(1), 43-48. doi:10.1046/j.1365-2109.2003.00791.x.
- APHA (1998). *Standard Methods for the Examination of Water and Wastewater*, 20th edn. Washington, DC.
- Assano, M., Ramirez, A. P. M., Stech, M. R., Honorato, C. A., Malheiros, E. B. & Carneiro, D. J. (2011). Desempenho de tilápia-do-Nilo cultivadas em viveiros alimentadas com diferentes fontes e níveis proteicos. *Ens. Ciê.* 15(5), 81-90.
- Batista, S., Pintado, M., Marques, A., Abreu, H., Silva, J. L., Jessen, F., Tulli, F. & Valente, L. M. (2020). Use of technological processing of seaweed and microalgae as strategy to improve their apparent digestibility coefficients in European seabass (*Dicentrarchus labrax*) juveniles. *J. Appl. Phycol.* 32, 3429-3446. doi:10.1007/s10811-020-02185-2.
- Bischoff, H. W. & Bold, H. C. (1963). Phycological studies. IV. Some soil algae from enchanted rock and related algal species. *Univ. Texas Pub.* 6318, 1-95.
- Bhatnagar, A. & P. Devi. 2013. Water quality guidelines for the management of pond fish culture. *Int. J. Environ. Sci.*, 3(6):1980-1997. doi:10.6088/ijes.2013030600019.
- Boyd, C. E., Torrans, E. L. & Tucker, C. S. (2018). Dissolved oxygen and aeration in ictalurid catfish aquaculture. *World Aquacul. Soc.* 49(1):7-70. doi:10.1111/jwas.12469.
- Brazil (2005). National Council for the Environment (CONAMA) Resolution n. 357. Available at: <<http://www.mma.gov.br/port/conama/res/res05/res>>.
- Caldini, N. N., Pereira, N. V., Rebouças, V. T. & Sá M. V. C. (2013). Can a small change in the tilapia's on-going feeding strategy impair its growth? *Acta Scient. Anim. Sci.* 35(3), 227-234. doi:10.4025/actascianimsci.v35i3.18290.
- Canosa, L. F. & Bertucci, J. I. (2023). The effect of environmental stressors on growth in fish and its endocrine control. *Front. Endocrinol.* 14, 1109461. doi: 10.3389/fendo.2023.1109461.
- Carvalho, J. S. O., Azevedo, R. V., Ramos, A. P. S. & Braga, L. G. T. (2012). Agroindustrial by-products in diets for Nile tilapia juveniles. *Rev. Bras. Zootec.* 41(3), 479-484. doi:10.1590/S1516-35982012000300002.
- Coelho, A. A. C., Bezerra, J. H. C., Silva, J. W. A., Moreira, R. T., Albuquerque, L. F. G. & Farias, W. R. L. (2014). Growth performance of Nile tilapia cultured in a recirculating water system with microalgae *Spirulina platensis*. *Rev. Bras. Saúde Prod. Anim.* 15(1), 149-159. doi:10.1590/S1519-99402014000100024.
- Costa, F. T. M., Reis, F. R. C., Santos, J. M. S., Maciel, S. M., Biserra, T. S., Moreira, R. L. & Farias, W. R. L. (2011). *Chlorella* sp. as a food supplement during the Nile tilapia larviculture. *Rev. Bras. Saúde Prod. Anim.* 12(4), 1103-1115.
- de Cruz, C. R., Lubrano, A. & Gatlin, D. M. (2018). Evaluation of microalgae concentrates as partial fishmeal replacements for hybrid striped bass *Morone* sp. *Aquaculture* 493, 130-136. doi:10.1016/j.aquaculture.2018.04.060.
- Dewi, R. R., Hassan, L., Daud, H. M., Matori, M. F., Nordin, F., Ahmad, N. I. & Zakaria, Z. (2022). Prevalence and antimicrobial resistance of *Escherichia coli*, *Salmonella* and *Vibrio* derived from farm-raised red hybrid tilapia (*Oreochromis* spp.) and Asian sea bass (*Lates calcarifer*, Bloch 1970) on the West Coast of Peninsular Malaysia. *Antibiotics* 11(2):136. doi:10.3390/antibiotics11020136.
- FAO (2022). *The State of World Fisheries and Aquaculture 2022. Towards Blue Transformation*. Rome, FAO. doi:10.4060/cc0461en.
- Fonseca, G. G., Cavenaghi-Altério, A. D., Silva, M. F., Arcanjo, V. & Sanjinez-Argandoña, E. J. (2013). Influence of treatments in the quality of Nile tilapia (*Oreochromis niloticus*) filets. *Food Sci. Nutr.* 1(3):246-253. doi:10.1002/fsn3.33.
- Furuya, W. M., Pezzato, L. E. & Miranda, E. C. (2001). Digestibility coefficients and digestible amino acids values of some ingredients for Nile tilapia (*Oreochromis niloticus*). *Rev. Bras. Zootec.* 30(4), 1186-1192. doi:10.1590/S1516-35982001000500002.
- Gorlach-Lira, K., Pacheco, C., Carvalho, L. C. T., Melo-Júnior, H. N. & Crispim, M. C. (2013). The influence of fish in floating net cages on microbial indicators of water quality. *Braz. J. Biol.* 73(3), 457-463. doi:10.1590/S1519-69842013000300001.
- Haque, M.R., Das, D.R., Sarkar, M.R., Begum, N., Pandit, D. & Jaman, A. (2023). Effect of stocking densities on growth and production performance of Bheda (*Nandus nandus*) in pond aquaculture. *Aquat. Sci. Eng.*, 38(2), 97-105. doi:10.26650/ASE20231247849.

- Hvas, M., Stien, L. H. & Oppedal, F. (2020). The metabolic rate response to feed withdrawal in Atlantic salmon post-smolts. *Aquaculture* 529, 735690. doi: 10.1016/j.aquaculture.2020.735690.
- Jung, J.-Y., Damusaru, J. H., Park, Y., Kim, K., Seong, M., Je, H.-W., Kim, S. & Bai, S. C. (2017). Autotrophic biofloc technology system (ABFT) using *Chlorella vulgaris* and *Scenedesmus obliquus* positively affects performance of Nile tilapia (*Oreochromis niloticus*). *Algal Res.*, 27, 259-264. doi:10.1016/j.algal.2017.09.021.
- Kroupova, H., Machova, J. & Svobodova, Z. (2005). Nitrite influence on fish: a review. *Vet. Med.* 50(11), 461-471. doi:10.17221/5650-VETMED.
- Leonardo, A. F. G., Tachibana, L., Corrêa, C. F., Gonçalves, T. G. & Baccarin, A. E. (2009). Water quality of rearing ponds and productive performance of Nile tilapia juveniles using three feeding systems. *Rev. Acad.: Ciên. Agrár. Amb.* 7(4), 383-393.
- Lima, F. R. S., Cavalcante, D. H., Rebouças, V. T. & Sá, M. V. C. (2016). Interaction between afternoon aeration and tilapia stocking density. *Acta Scient. Anim. Sci.* 38(1), 23-30. doi:10.4025/actascianimsci.v38i1.27093.
- Lui, T. A., Neu, D. H., Boscolo, W. R., Bittencourt, F., Freitas, J. M. A., Feiden, A. (2012). Use of organic wheat in the diet of Nile tilapia juveniles. *Pesq. Agrop. Trop.* 42(4), 383-389. doi:10.1590/S1983-40632012000400015.
- Martins, C. I., Galhardo, L., Noble, C., Damsgård, B., Spedicato, M. T., Zupa, W., Beauchaud, M., Kulczykowska, E., Massabuau, J. C., Carter, T., Planellas, S. R. & Kristiansen, T. (2012). Behavioural indicators of welfare in farmed fish. *Fish Physiol Biochem.* 38, 17-41. doi: 10.1007/s10695-011-9518-8.
- Moreira, r. l., silveira, l. p., teixeira, e. g., moreira, a. g. l., moura, p. s. & farias, w. r. l. (2012). growth and gastrointestinal indices in Nile tilapia fed with different diets. *Acta Scient. Anim. Sci.* 34(3), 223-229. doi:10.4025/actascianimsci.v34i3.13327.
- Ntengwe, F. W. & Edema, M. O. (2008). Physico-chemical and microbiological characteristics of water for fish production using small ponds. *Phys. Chem. Earth, Parts A, B and C* 33(1), 701-707. doi:10.1016/j.pce.2008.06.032.
- Obirikoranga, K. A., Acheamponga, J. N., Duodua, C. P., Skov, P. V. (2020). Growth, metabolism and respiration in Nile tilapia (*Oreochromis niloticus*) exposed to chronic or periodic hypoxia. *Comp. Biochem. Physiol. Part A* 248, 110768. doi:10.1016/j.cbpa.2020.110768.
- Pauly D. (1983). Some simple methods for the assessment of tropical fish stocks. *FAO Fisheries Tech. Pap.*, FAO Rome, 234, 52.
- Pereira, C. M. & Lapolli F. R. (2009). Nile tilapia culture on domestic effluent treated in stabilization ponds. *Biotemas* 22(1), 93-102.
- Rocha R. S., Leite, L. O., de Sousa, O. V. & Vieira, R. H. (2014). Antimicrobial susceptibility of *Escherichia coli* isolated from fresh-marketed Nile tilapia (*Oreochromis niloticus*). *J. Pathog.* 2014, 756539. doi:10.1155/2014/756539.
- Santos, V. B., Mareco, E. A. & Silva, M. D. P. (2013). Growth curves of Nile tilapia (*Oreochromis niloticus*) strains cultivated at different temperatures. *Acta Scient. Anim. Sci.* 35(3), 235-242. doi:10.4025/actascianimsci.v35i3.19443.
- Sarker, P. K., Gamble, M. M., Kelson, S. & Kapuscinski, A. R. (2016). Nile tilapia (*Oreochromis niloticus*) show high digestibility of lipid and fatty acids from marine *Schizochytrium* sp. and of protein and essential amino acids from freshwater *Spirulina* sp. feed ingredients. *Aquacul. Nutr.* 22(1), 109-119. doi:10.1111/anu.12230.
- Schwarz, K. K., Furuya, W. M., Natali, M. R. M., Michelato, M. & Gualdezi, M. C. (2010). Mannanligosaccharides in diets for Nile tilapia, juveniles. *Acta Scient. Anim. Sci.* 32(2), 197-203. doi:10.4025/actascianimsci.v32i2.7724.
- Scorvo-Filho, J. D. S., Frasca-Scorvo, C. M. D., Alves, J. M. C. & Souza, F. R. A. (2010). Tilapia culture and its inputs, economic relations. *Rev. Bras. Zootec.* 39(sp.), 112-118. doi:10.1590/S1516-35982010001300013.
- Signor, A. A., Boscolo, W. R., Bittencourt, F., Feiden, A., Gonçalves, G. S. & Freitas, J. M. A. (2010). Performance of juvenile Nile tilapia fed diets with enzymatic complex. *Rev. Bras. Zootec.* 39(5), 977-983. doi:10.1590/S1516-35982010000500006.
- Taelman, S. E., Meester, S. D. e., Roef, L., Michiels, M. & Dewulf, J. (2013). The environmental sustainability of microalgae as feed for aquaculture: A life cycle perspective. *Biores. Technol.* 150(1), 513-522. doi:10.1016/j.biortech.2013.08.044.
- Teuling, E., Schrama, J. W., Gruppen, H. & Wierenga, P. A. (2017). Effect of cell wall characteristics on algae nutrient digestibility in Nile tilapia (*Oreochromis niloticus*) and African catfish (*Clarus gariepinus*). *Aquaculture* 479, 490-500. doi:10.1016/j.aquaculture.2017.06.025.
- Tibbetts, S. M., Mann, J. & Dumas, A. (2017). Apparent digestibility of nutrients, energy, essential amino acids and fatty acids of juvenile Atlantic salmon (*Salmo salar* L.) diets containing whole-cell or cell-ruptured *Chlorella vulgaris* meals at five dietary inclusion levels. *Aquaculture*, 481, 25-39. doi:10.1016/j.aquaculture.2017.08.018.
- Toni, M., Angiulli, E., Malavasi, S., Alleva, E. & Cioni, C. (2017). Variation in environmental parameters in research and aquaculture: Effects on behaviour, physiology and cell biology of teleost fish. *J Aquac. Mar. Biol.* 5(6), 00137. doi:10.15406/jamb.2017.05.00137.
- Turker, H., Eversole, A. G. & Brune, D. E. (2003). Filtration of green algae and cyanobacteria by Nile tilapia, *Oreochromis niloticus*, in the Partitioned Aquaculture System. *Aquaculture* 215, 93-101. doi:10.1016/S0044-8486(02)00133-3
- Ungsethaphand, T., Peerapornpisal, Y., Whangchai, N. & Sardud, U. (2010). Effect of feeding *Spirulina platensis* on growth and carcass composition of hybrid red tilapia (*Oreochromis mossambicus* × *O. niloticus*). *Maejo Int. J. Sci. Technol.* 4(2), 331-336.
- Verdegem, M., Buschmann, A. H., Latt, U. W., Dalsgaard, A. J. T. & Lovatelli, A. (2023). The contribution of aquaculture systems to global aquaculture production. *J. World Aquacult. Soc.* 54(2), 206-250. doi:10.1111/jwas.12963.
- Workagegn, K. B., Ababboa, E. D., Yimer, G. T. & Amare, T. A. (2014). Growth performance of the Nile tilapia (*Oreochromis niloticus* L.) fed different types of diets formulated from varieties of feed ingredients. *J. Aquacul. Res. Develop.* 5(3), 1000235(1-4). doi:10.4172/2155-9546.1000235.
- Yavuzcan, Y. H., Robaina, L., Pirhonen, J., Mente, E., Domínguez, D. & Parisi, G. (2017). Fish welfare in aquaponic systems: Its relation to water quality with an emphasis on feed and faeces - A review. *Water* 9(1), 13. doi:10.3390/w9010013.
- Zeinab, A. K., Aly, M. S., Faiza, A. K. & Fatma, E. M. (2015). Effect of *Spirulina platensis* and *Lactobacillus rhamnosus* on growth and biochemical performance of Nile tilapia (*Oreochromis niloticus*) fingerlings. *Int. J. Curr. Microbiol. Appl. Sci.* 4(4), 747-763.

The Relationship of Bristle Worm, *Protodorvillea kefersteini* (McIntosh, 1869) (Eunicida, Dorvilleidae) Abundance with Environmental Variables in Çardak Lagoon (Turkish Straits) Exposed to domestic Discharge

Ertan Dağlı¹ , A. Suat Ateş² , Seçil Acar² , Yeşim Büyükkateş² 

Cite this article as: Dağlı, E., Ates, A.S., Acar, S., & Buyukates, Y. (2024). The relationship of bristle worm, *Protodorvillea kefersteini* (McIntosh, 1869) (Eunicida, Dorvilleidae) abundance with environmental variables in Çardak Lagoon (Turkish Straits) exposed to domestic discharge. *Aquatic Sciences and Engineering*, 39(1), 24-29. DOI: <https://doi.org/10.26650/ASE20241322577>

ABSTRACT

This study presents the correlations between opportunistic polychaeta, *Protodorvillea kefersteini* (McIntosh, 1869) abundance, and environmental variables in Çardak Lagoon. Samplings were carried out on the bottoms using a 30x30 cm quadrat seasonally between 1 and 1.8 m depths of seven sampling points in October 2018, February, April, and June 2019. A total of 1094 specimens belonging to *P. kefersteini* were collected. Environmental variables such as gravel content in sediment, pH, and salinity levels in the water had the highest correlations with the abundance through the sampling periods. Considering the sampling points, the highest correlation value was between water salinity and the abundance. Sediment gravel content, pH, salinity, temperature, anionic surfactant levels, and NO₂+NO₃ were major environmental variables affecting *P. kefersteini* abundance in the study area spatially and temporally.

Keywords: *Protodorvillea kefersteini*, abundance, environmental variables, Çardak Lagoon, Turkish Straits

ORCID IDs of the author:
E.D. 0000-0001-6579-505X;
A.S.A. 0000-0002-4682-1926;
S.A. 0000-0002-6426-8095;
Y.B. 0000-0002-4402-4587

¹Ege University, Fisheries Faculty,
İzmir, Türkiye

²Çanakkale Onsekiz Mart University
Faculty of Marine Sciences and
Technology, Çanakkale, Türkiye

Submitted:
04.07.2023

Revision Requested:
18.09.2023

Last Revision Received:
18.09.2023

Accepted:
18.09.2023

Online Published:
17.11.2023

Correspondence:
A. Suat Ateş
E-mail:
asuataates@yahoo.com

INTRODUCTION

Anthropogenic environmental variables have negative effects on marine biodiversity, especially on some benthic communities dominant in polluted environments that are sensitive to disturbances (Santos *et al.* 2021). Sudden daily and seasonal fluctuations in the environmental variables in marine systems, mainly in lagoon areas, may change the abundance of benthic organisms (Koutsoubas *et al.* 2000). Water movement, dissolved oxygen concentration, salinity, sediment grain size, and organic matter content effectively affect the polychaeta abundance and distribution in marine environments (Guerra-Garcia & Garcia-Gomez 2004). Among these, organic matter accumulation in sediment with anthropogenic origin controls the macrofaunal communities (Magni *et al.* 2004).

Sewage discharges are one of the most common anthropogenic disturbances on marine benthos. There is important information in the relevant literature on the temporal and spatial status of benthic assemblages, especially on conspicuous disturbances caused by organic enrichment in sediments (Del-Pilar-Ruso *et al.* 2009). Opportunistic macrozoobenthic species adapt easily to a new marine environment and can form dense populations in a short time (Carlton 1985).

Polychaetas with high sensitivity to changes on soft bottoms (Del-Pilar-Ruso *et al.* 2009) may be preferred for monitoring studies (Muxika *et al.* 2005). Some polychaetas are also highly tolerant to pollution and low oxygen stress caused by excessive accumulation of organic matter. Besides, they are important in detecting the ef-



fect of pollutants between water and sediment (Elias *et al.* 2003). Additionally, they can tolerate high organic matter content in marine soft bottoms (Fernández-Romero *et al.* 2019) and play an important role in the bioturbation of marine sediments (Hutchings 1998).

Effects of pollutants on polychaetes were previously studied by many researchers (Karakassis *et al.* 2000; Warwick 2001; Ergen *et al.* 2004; Como *et al.* 2007; Afli *et al.* 2008; Dauvin 2008; Zaaiba *et al.* 2009; Terlizzi *et al.* 2010; Zaâbi *et al.* 2010; Martins *et al.* 2013; Zaâbi-Sendi 2013; Cabral-Oliveira *et al.* 2014; Hamdy *et al.* 2023). Those dorvilleid polychaetes are known as opportunistic communities in marine sediments enriched with organic matter (Alalykina & Polyakova 2022). Dorvilleid polychaeta, *Protodorvillea kefersteini* which is an important biotope descriptive and pollution indicator (MES 2010), is one of the 14 known species of the genus, *Protodorvillea* (Worms 2023). *P. kefersteini* is a species that is 1-3 cm long, lives on mud, gravel, or sandy bottoms at depths of 10-30 m, and mostly uses empty tubes of serpulids under stones. *P. kefersteini* is distributed in the North Atlantic to the North Sea and English Channel, Mediterranean, and Black Sea (Tillin 2016). *P. kefersteini* is known from the Mediterranean (Núñez *et al.* 2013; Çınar *et al.* 2015; Mikac 2015) and the Black Sea (Kurt Şahin *et al.* 2017; Kopyı 2018). This study presents the relationships between *P. kefersteini* abundance and environmental variables measured in Çardak Lagoon affected by domestic discharges.

MATERIALS AND METHODS

The sampling area included the depths of 1 to 1.8 m of 7 different sampling points (GPS Coordinates: 40° 23'14" N, 26° 43'30" D) chosen in Çardak Lagoon located in the northeast of Çanakkale Strait (Figure 1).

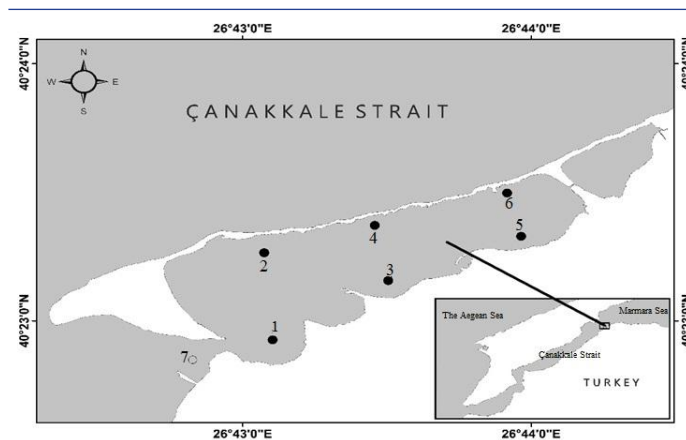


Figure 1. Map of the study area showing sampling points.

Benthos material including *Protodorvillea kefersteini* specimens was collected by a SCUBA diver with a 30x30 cm metal framed quadrat system in October 2018, February, April, and June 2019. Samples were fixed in 4% neutralized formaldehyde in 5 L plastic bottles. In the laboratory, the bottom samples were washed with the help of pressurized water and passed

through a triple sieve system with apertures of 0.5, 1, and 2 mm. Faunal species remaining on the sieves were extracted at macro and micro levels and fixed in 70% ethanol in 50 cc glass tubes on a group basis. *P. kefersteini* specimens were identified on the trinocular stereomicroscope according to definitions based on previous studies, and all were counted. The definitions of Fauchald (1977) and Fauchald and Rouse (1997) were used for diagnosis.

The correlations between values of environmental variables and *P. kefersteini* abundance recorded for 7 different sampling stations in 4 seasons were calculated according to Pearson coefficient correlation (r) in the PAST program.

Water quality variables of lagoon water were measured *in situ* using a YSI 650 MDS. The number of nutrients (NO_2 , NO_3 , NH_4 , $\text{PO}_4\text{-P}$, SiO_2) and total suspended solids (TSS) in the lagoon water were measured based on the analysis method proposed by Strickland and Parsons (1972) at different wavelengths in the spectrophotometer. Analyses were made using the Jasco Brand UV spectrophotometer in the Laboratory of the Faculty of Marine Sciences and Technology. Analyses of % organic matter and % particle content in the sediment were performed in Çanakkale Onsekiz Mart University Central Laboratory. Particle size analyses in the sediment were done according to Allen (1997).

RESULTS

A total of 1094 specimens belonging to *P. kefersteini* were sampled in the study area (Figure 2).



Figure 2. General view of *Protodorvillea kefersteini* (Photographed by E. Dağlı).

Although it is known that *P. kefersteini* is tolerant to pollution, reference station (Stn. 7) with low sediment organic matter content and water chl.-a amount (OM%= 1.73% and chl.-a= 1.49 $\mu\text{g L}^{-1}$, respectively) was the most abundant in terms of specimen number (1001 out of a total of 1094 specimens) in the study area. Except for October 2018, *P. kefersteini* showed a regular distribution in terms of specimen number in other sampling periods (the lowest 34 individuals, di%= 3.10) (Table 1).

Considering the relationships between environmental variables and *P. kefersteini* abundance, the highest positive correlation ($r=0.93$; $p < 0.05$) was found between *P. kefersteini* abundance and

Table 1. Mean water quality, nutrient, chl-a, total suspended solids (TSS), chemical oxygen demand (COD), anionic surfactant (AS), and sediment variables values measured at sampling periods and points.

Sampling period	A	OM (%)	WOM (mg L ⁻¹)	T (°C)	S (‰)	O ₂ (mg L ⁻¹)	ORP	pH	Chl.-a (µg L ⁻¹)	SiO ₂
Autum 2018	34	6.42	11.02	14.76	20.77	8.07	80.41	8.14	3.79	0.34
Winter 2019	338	5.98	12.2	7.99	23.46	7.45	-83.37	8.15	1.19	0.39
Spring 2019	382	6.61	10.11	15.5	22.53	6.48	-90.98	8.31	1.83	0.17
Summer 2019	340	6.76	11.48	25.66	21.04	7.83	-100.52	8.29	5.31	0.83
Mean		6.44±0.15	11.2±0.38	15.98±3.15	21.95±0.55	7.46±0.30	-48.62±37.3	8.22±0.04	3.03±0.81	0.43±0.12
Sampling point										
Station 1	58	10.65	11.1	12.53	22.45	8.19	-27.63	7.89	3.63	0.36
Station 2	7	3.03	11.65	13.12	22.35	7.95	-31.6	8.32	2.64	0.56
Station 3	7	15.52	11.62	12.4	22.13	6.9	-31.23	8.14	3.38	0.29
Station 4	20	3.49	10.85	12.68	22.35	6.93	-30.16	8.27	2.96	0.57
Station 5	1	7.91	11.2	12.41	22.36	6.74	-35.23	8.22	2.59	0.55
Station 6	-	2.75	11.17	12.93	21.87	6.93	-35.1	8.27	4.47	0.18
Station 7 (ref.)	1001	1.73	10.85	13.1	22.66	7.7	-31.16	8.29	1.49	0.48
Mean		6.44±1.79	11.2±0.11	12.73±0.10	22.31±0.08	7.33±0.20	-31.73±0.94	8.2±0.05	3.02±0.32	0.42±0.05
Total number	1094									

Table 1. Continue.

Sampling period	TP	NO ₂ +NO ₃ (mg L ⁻¹)	TN	TSS (mg L ⁻¹)	COD (mg L ⁻¹)	AD (mg L ⁻¹)	PO ₄ (mg L ⁻¹)	Sand (%)	Gravel (%)	Mud (%)
Autum 2018	0.02	0.13	0.61	9.66	197.83	0.027	0.01	75.36	16.56	8.11
Winter 2019	0.02	0.09	0.23	35.28	178.71	0.029	0.01	69.29	19.9	10.74
Spring 2019	0.04	0.06	0.41	13.57	80.75	0.051	0.01	68.87	23.14	7.96
Summer 2019	0.09	0.03	0.21	6.64	74	0.032	0.02	72.91	21.1	5.95
Mean	0.04±0.01	0.08±0.02	0.37±0.08	16.29±5.62	132.82±27.95	0.03±0.0	0.01±0.0	71.61±1.34	20.18±1.19	8.19±0.85
Sampling point										
Station 1	0.048	0.083	0.48	11	127.87	0.045	0.03	61.8	24.1	14.16
Station 2	0.029	0.071	0.23	8.07	152.12	0.027	0.02	78.6	18.1	3.17
Station 3	0.026	0.071	0.42	22.15	126.66	0.032	0.02	55.2	31.6	13.24
Station 4	0.035	0.036	0.16	9.35	103.33	0.04	0.02	86.2	8.49	5.27
Station 5	0.051	0.089	0.49	19.07	201	0.034	0.01	57.6	25.6	16.71
Station 6	0.065	0.089	0.49	28.41	148.25	0.035	0.03	92.8	3.98	3.29
Station 7 (ref.)	0.053	0.094	0.26	15.98	163	0.032	0.01	69.1	29.4	1.49
Mean	0.04±0.004	0.07±0.006	0.36±0.04	16.29±2.60	146.03±10.95	0.035±0.002	0.02±0.002	71.61±5.10	20.18±3.68	8.19±2.19
Total number										

average % gravel content in sediment for the sampling periods. For sampling points, the highest correlation ($r=0.64$; $p < 0.05$) was between abundance and salinity. There were positive correlations between *P. kefersteini* abundance and seawater pH, salinity, and anionic detergent amount ($r = 0.67, 0.62, 0.58$; $p < 0.05$, re-

spectively) in the sampling points. The weakest positive relationships were between the % of mud content in sediment and the amount of organic matter in water and sediment for the sampling points ($r = 0.01, 0.02, 0.09$; $p < 0.05$, respectively) (Table 2).

Table 2. Correlations between total abundance and mean environmental variables for all sampling periods and sampling points (Pearson coefficient correlation, $p < 0.05$).

EV	Sampling points	
	A (ind. 0.09 m ⁻²)	A (ind. 0.09 m ⁻²)
OM (%)	0.09	-0.39
WOM (mg L ⁻¹)	0.02	-0.50
Temperature (°C)	0.10	0.50
Salinity (‰)	0.62	0.64
O ₂ (mg L ⁻¹)	-0.68	0.31
ORP	-0.99	0.14
pH	0.67	0.22
Chl.-α (µg L ⁻¹)	-0.31	-0.72
SiO ₂	0.13	0.15
TP	0.43	0.28
NO ₂ +NO ₃ (mg L ⁻¹)	-0.82	0.39
TN	-0.81	-0.31
TSS (mg L ⁻¹)	0.30	-0.05
COD (mg L ⁻¹)	-0.71	0.21
AD (mg L ⁻¹)	0.58	-0.18
PO ₄ (mg L ⁻¹)	0.28	-0.52
Sand (%)	-0.84	-0.09
Gravel (%)	0.93	0.39
Mud (%)	0.01	-0.46

A: Abundance, EV: Environmental variable, OM: Organic matter in sediment, WOM: organic matter in water, ORP: Oxygen reduction potential, TP: total phosphate, TN: Total nitrogen, TSS: Total suspended solids, COD: Chemical oxygen demand, AD: Anionic detergent.

DISCUSSION

Effects of environmental variables such as temperature and salinity were previously studied at the population level in opportunistic polychaetas such as *Capitella* sp., *Ophryotrocha diadema*, and *Streblospio benedicti* in polluted areas (Simonini & Prevedelli, 2003). Based on the previous studies regarding *P. kefersteini* which is another opportunistic polychaeta, *P. kefersteini* is a recognizable species in areas affected by hypoxia (Leonhard 2006). Similarly, Warwick *et al.* (1986) stated that *P. kefersteini* is abundant in organic matter-rich habitats with sewage discharge. Hiscock *et al.* (2004) also indicated that *P. kefersteini* is an increasingly abundant species on sea bottoms where slight organic enrichment is observed. *P. kefersteini* abundance on the organic matter-rich bottoms of the Mediterranean and its relationship with other environmental variables were previously studied (Karakassis *et al.* 2000; Ergen *et al.* 2004; Como *et al.* 2007; Afli *et al.* 2008; Zaaiba *et al.* 2009; Terlizzi *et al.* 2010; Zaâbi *et al.* 2010; Martins *et al.* 2013; Zaâbi-Sendi 2013; Hamdy *et al.* 2023). Among these studies, Karakassis *et al.* (2000) found that *Protodorvillea kefersteini* and *Cirrophorus lyra* were dominant (more than 20% of the total abundance) on the bottoms of fish farms established at the Sounion coast (eastern Greece) at a depth of 13-20 m, with 80% silt and high carbon content.

Similarly, Ergen *et al.* (2004) found dense populations of *P. kefersteini* (3060 ind. m⁻²) on the polluted or semi-polluted bottoms of cage farms in the eastern Aegean Sea. In addition, regarding organic matter content in sediment, Como *et al.* (2007) found that *P. kefersteini* dominated the inlet area of Oristano Bay (Sardinia, Italy) with high organic matter content (mean value of 25% of the total sediment dry weight). Terlizzi *et al.* (2010) also observed *P. kefersteini* in abundance on mud-character bottoms rich with organic matter where fish cages are located, approximately 100 to 500 m offshore of Corsica Island (France, Mediterranean). On the contrary, Hamdy *et al.* (2023) recorded *P. kefersteini* only on the bottoms with low organic matter content (max. 1.25%) off the coast of Alexandria (Egypt, Mediterranean). Considering the sediment type depending on the sediment grain size, Zaaiba *et al.* (2009) found *P. kefersteini* to be the most dominant (1900 ind. m⁻²) at Cap Bon Peninsula coasts (northern Tunisia) in the bottoms where the sediment-gravel ratio varies between 0.2 and 60.6%. *P. kefersteini* is known as of the few polychaeta species that preferred the bottoms with a coarser particle in the Bizerte Lagoon (southern Tunisia) (Afli *et al.* 2008). The Bizerte Lagoon example presented for *P. kefersteini* specimens was observed in this study. We found the highest number of individuals on the bottoms where the average sand+gravel content was the highest and mud content was the lowest. Similarly, Zaâbi-Sendi (2013) stated that *P. kefersteini* was well represented in coarse sand bottoms where the coarse fraction between 500 and 630 µm dominates the Tunisia coasts. Further, *P. kefersteini*, characteristic of the fine and coarse sandy bottoms of Cap Bon Peninsula coasts, was abundant in all seasons with an average abundance of 66 ± 32.73 (ind. m⁻²) (Zaâbi *et al.* 2010).

Considering the northeastern Atlantic specimens of *P. kefersteini*, the species was defined as a characteristic species for Portugal coast bottoms with a mean depth of 50 m, 68.8% sand, 27.2% gravel, and 1% mean organic matter content (Martins *et al.* 2013). Lourido *et al.* (2008) also found *P. kefersteini* in abundance on the bottoms of the Ria de Aldan coasts (northwest Spain) with an average content of 1.46% organic matter, 19.57% gravel, and 78.06% sand. On the other hand, although *P. kefersteini* prefers shallow waters as its habitat, it was also recorded at depths between 984 and 1113 m in Capbreton Canyon (Biscay Bay, NE Atlantic) (Aguirrezabalaga & Ceberio 2003). Furthermore, *P. kefersteini* was dominant in surface waters with an average redox potential (ORP) value of $+61 \pm 177$ mV on the western Scottish coast (Pearson & Stanley 1979). In this study, no significant relationship was found between ORP and *P. kefersteini* abundance.

Considering only *P. kefersteini* abundance, Kopy (2018) observed *P. kefersteini* on Sevastopol coasts (Crimea, Black Sea) throughout the year. Besides, there are many studies regarding polychaeta assemblages including *P. kefersteini* in Turkish Seas. Among these, Çınar *et al.* (2011) found *P. kefersteini* was most abundant (2900 ind. m⁻²) at 17 m depth of Erdek coasts (the southern Marmara Sea). Then, Kurt Şahin *et al.* (2017) recorded *P. kefersteini* (8675 ind. m⁻²) during spring in fine particle sand bottoms at 3 m depths of the Sinop Peninsula (southern Black Sea) being the most abundant species.

Apart from all these studies, a study examining polychaeta communities and their relationship with environmental vari-

ables was conducted by Can Yılmaz (2009) in Homa Lagoon (Turkish Aegean Sea). Can Yılmaz (2009) has found negative correlations between polychaeta abundance and sediment surface temperature in winter, and pH and abundance in spring ($p = -0.730$; $p = -0.782$; $p < 0.05$, respectively) in Homa Lagoon. Can Yılmaz (2009) has also stated that the environmental variable that shows a positive correlation with polychaeta abundance ($p = 0.697$; $p < 0.05$) in Homa Lagoon was % sand content in the sediment for the summer period. In this study, on the contrary, while % sand content of bottoms in the study area was negatively correlated with abundance temporally and spatially, seawater salinity moderately affected *P. kefersteini* abundance both temporally and spatially. In addition, very weak positive and negative correlations were recorded between *P. kefersteini* abundance and % sediment organic matter content in this study. This may be because the abundance of *P. kefersteini* was greatest at the reference point outside the lagoon area with very low organic matter bottoms.

Our study did not agree with the results regarding the correlation between *P. kefersteini* abundance and % sediment organic matter content presented by Warwick *et al.* (1986), Ergen *et al.* (2004), and Como *et al.* (2007). On the contrary, findings by Hiscock *et al.* (2004), Lourido *et al.* (2008), Martins *et al.* (2013), and Hamdy *et al.* (2023) were supported in the present study. Moreover, based on our results, we may point out that excessive accumulation of organic matter in the sediment may not be a major environmental variable that positively affects *P. kefersteini* abundance since the abundance was the greatest in the sediment with the lowest % organic matter content in the study area.

CONCLUSION

Sediment gravel content is one of the most important environmental variables that positively correlated with *P. kefersteini* abundance temporally. According to the results of this study and other studies performed on the Mediterranean and eastern Atlantic coasts, we may state that salinity, temperature, sediment particle size, and % sediment organic matter content had the most effect on *P. kefersteini* abundance. Although *P. kefersteini* is known as a pollution indicator species, we think that it cannot be concluded that the sediment of an area is polluted based only on *P. kefersteini* abundance.

Conflict of Interests: The authors declare that they have no financial interests or personal relationships that could affect this work, hence no conflict of interest.

Financial disclosure: This study is within the scope of the COST Action Project supported by TUBITAK coded 117Y510.

REFERENCES

- Afli, A., Ayari, R., Zaabi, S. 2008. Ecological quality of some Tunisian coast and lagoon locations. by using benthic community parameters and biotic indices. *Estuarine, Coastal and Shelf Sciences*, 80, 269–280.
- Aguirrezabalaga, F. & Ceberio, A. (2003). Dorvilleidae (Polychaeta) from the Capbreton Canyon (Bay of Biscay, NE Atlantic) with the description of *Pettiboneia sanmartini* sp. nov. *Cahiers de Biologie Marine*, 44 (1), 41-48.
- Alalykina, I.L. & Polyakova, N.E. (2022). New species of Ophryotrocha (Annelida: Dorvilleidae) associated with deep-sea reducing habitats in the Bering Sea, Northwest Pacific. *Deep-Sea Research Part II*, 206, 105217. <https://doi.org/10.1016/j.dsr2.2022.105217>.
- Allen, J.R.L. (1997). Subfossil mammalian tracks (Flandrian) in the Severn Estuary. S.W. Britain: *mechanics of formation, preservation and distribution*, *Philosophical Transactions of the Royal Society, London B*. 352, 481–518.
- Cabral-Olivera, J., Mendes, S., Maranhão, P. & Pardal, M.A. (2014). Effects of sewage pollution on the structure of rocky shore macroinvertebrate assemblages. *Hydrobiologia*, 726, 271-283. <http://doi.org/10.1007/s10750-013-1773-5>.
- Can Yılmaz, E. (2009). The polychaeta of the Homa lagoon (Izmir Bay). Dokuz Eylül University, Institute of Natural and Applied Sciences, PhD. Thesis, 60 pp.
- Carlton, J.T. (1985). Transoceanic and interoceanic dispersal of coastal marine organisms: the biology of ballast water. *Oceanography and Marine Biology Annual Review* 23: 313-371.
- Como, S., Magni, P., Casu, D., Floris, A., Giordani, G., Natale, S., Fenzi, G.A., Signa, G., & De Falco, G. (2007). Sediment characteristics and macrofauna distribution along a human-modified inlet in the Gulf of Oristano (Sardinia, Italy). *Marine Pollution Bulletin*, 54, 733–744.
- Çınar, M.E., Dağlı, E. & Açıık, Ş. (2011). Annelids (Polychaeta and Oligochaeta) from the Sea of Marmara, with descriptions of five new species. *Journal of Natural History*, 45 (33-34), 2105-2143. <http://doi.org/10.1080/00222933.2011.582966>.
- Çınar, M.E., Dağlı, E., Çağlar, S. & Albayrak, S. (2015). Polychaetes from the northern part of the Sea of Marmara with the description of a new species of *Polydora* (Annelida: Polychaeta: Spionidae). *Mediterranean Marine Science*, 16, 524–532. <http://doi.org/10.12681/mms.1226>.
- Dauvin, J.C. (2008). Effects of heavy metal contamination on the macrobenthic fauna in estuaries: the case of the Seine estuary. *Marine Pollution Bulletin*, 57 (1–5), 22-40. <http://doi.org/10.1016/j.marpolbul.2007.10.016>.
- Del-Pilar-Ruso, Y., De-La-Ossa-Carretero, J.A., Loya-Fernandez, A., Ferrero-Vicente, L. M., Giménez-Casaldueiro, F. & Sánchez-Lizaso, J.L. (2009). Assessment of soft-bottom Polychaeta assemblage affected by a spatial confluence of impacts: Sewage and brine discharges. *Marine Pollution Bulletin*, 58 (5), 776-782. <http://doi.org/10.1016/j.marpolbul.2009.03.002>.
- Elias, R., Rivero, M.S. & Vallarino, E.A. (2003). Sewage impact on the composition and distribution of Polychaeta associated to intertidal mussel beds of the Mar del Plata rocky shore, Argentina. *Iheringia, Sér. Zool.*, Porto Alegre, 93 (3), 309-318.
- Ergen, Z., Çınar M.E. & Dağlı E. (2004). Effects of fish farming in the distribution of polychaetes in the Aegean Sea, *Rapport de la Commission International Exploration de la Mer Méditerranée*, 37, 350.
- Fauchald, K. & Rouse, G. (1997). Polychaeta systematics: Past and present. *Zoologica Scripta*, 26 (2), 71-138.
- Fauchald, K. (1977). The Polychaete Worms. Definitions and Keys to the Orders. Families and Genera. Natural History Museum of Los Angeles County Allan Hancock Found. Science Series, Los Angeles, 28, 1-188.
- Fernández-Romero, A., Moreira, J. & Guerra-García, J. M. (2019). Marinas: An overlooked habitat for exploring the relation among polychaete assemblages and environmental factors. *Marine Pollution Bulletin*, 138, 584-597. <https://doi.org/10.1016/j.marpolbul.2018.11.064>.
- Guerra-García, J.M. & García-Gomez, J.C. (2004). Polychaete assemblages and sediment pollution in a harbour with two opposing entrances. *Helgoland Marine Research*, 58, 183–191.
- Hamdy, R., Elebiary, N., Abdel Naby, F., Borghese, J., Dorgham, M., Hamdan, A. & Musco, L. (2023). Hard-bottom polychaetes exposed to multiple human pressure along the Mediterranean coast of Egypt. *Water*, 15, 997. <https://doi.org/10.3390/w15050997>.

- Hiscock, K., Southward, A., Tittley, I. & Hawkins, S. (2004). Effects of changing temperature on benthic marine life in Britain and Ireland. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 14 (4), 333-362.
- Hutchings, P. (1998). Biodiversity and functioning of polychaetes in benthic sediments. *Biodiversity and Conservation*, 7, 1133-1145.
- Karakassis, I., Tsapakis, M., Hatziyanni, E., Papadopoulou, K.-N., & Plaiti, W. (2000). Impact of cage farming of fish on the seabed in three Mediterranean coastal areas. *ICES Journal of Marine Science*, 57 (5), 1462–1471. <https://doi.org/10.1006/jmsc.2000.0925>.
- Kopiç, V. (2018). Some aspects of the biology and the present state of the population of *Protodorvillea kefersteini* (Polychaeta: Dorvilleidae) in the coastal zone of the Crimea (The Black Sea). Diversity in Coastal Marine Sciences, Edited by C. Finkl and C. Makowski, Coastal Research Library book series, Springer Publ., 23, 405–412.
- Koutsoubas D., Dounas, C., Arvanitidis C., Kornilios S., Petihakis G., Triantafyllou G. & Eleftheriou, A. (2000). Macrobenthic community structure and disturbance assessment in Gialova Lagoon, Ionian Sea. *ICES Journal of Marine Science*, 57, 1472–1480.
- Kurt Şahin, G., Dağlı, E. & Sezgin, M. (2017). Spatial and temporal variations of soft bottom polychaetes of Sinop Peninsula (southern Black Sea) with new records. *Turkish Journal of Zoology*, 41, 89-101, <http://doi:10.3906/zoo-1510-15>.
- Leonhard, S.B. (2006). Horns Rev offshore wind farm. Environmental impact assessment of sea bottom and marine biology. Technical Report, 54 Environmental Sciences; 17 Wind Energy; Seabed; Environmental Impact Statements; Wind Turbines; Offshore Sites, 43 pp.
- Lourido, A. Cacabelos, E. & Troncoso, J.S. (2008). Patterns of distribution of the polychaete fauna in subtidal soft sediments of the Ría de Aldán (north-western Spain). *Journal of the Marine Biological Association of the United Kingdom*, 88 (2), 263–275. <http://doi.org/10.1017/S0025315408000696>.
- Magni, P., Micheletti, S., Casu, D., Floris, A., de Falco, G. & Castelli, A. (2004). Macrofaunal community structure and distribution in a muddy coastal lagoon. *Chemistry and Ecology*, 20 (1), 397–S409.
- Martins, R., Sampaio, L., Rodrigues, A.M. & Quintino, V. (2013). Soft-bottom Portuguese continental shelf polychaetes: Diversity and distribution. *Journal of Marine Systems*, 123–124, 41-54. <https://doi.org/10.1016/j.jmarsys.2013.04.008>.
- MES (2010). Marine Macrofauna Genus Trait Handbook. Marine Ecological Surveys Limited. <http://www.genustraithandbook.org.uk/>
- Mikac, B. (2015). A sea of worms: Polychaete checklist of the Adriatic Sea. *Zootaxa* 3943, 001–172. <http://doi.org/10.11646/zootaxa.3943.1.1>.
- Muxika, I., Borja, Á. & Bonne, W. (2005). The suitability of the marine biotic index (AMBI) to new impact sources along European coasts. *Ecological Indicators*, 5 (1), 19-31.
- Núñez, J., Maggio, Y. & Riera, R. (2013). Dorvilleidos (Polychaeta, Dorvilleidae) recolectados durante el proyecto "Faunalbérica" y catálogo de las especies ibero-baleares. *Graellsia*, 69, 275–282. <http://doi.org/10.3989/graellsia.2013.v69.i2>.
- Pearson, T.H., & Stanley, S.O. (1979). Comparative measurement of the redox potential of marine sediments as a rapid means of assessing the effect of organic pollution. *Marine Biology*, 53, 371-379.
- Santos, M.E.M., Silva, C.C. & de Azevedo-Cutrim, A.C.G. (2021). Spatial-temporal distribution of polychaeta in urbanized sandy beaches of northeastern Brazil: Tools for environmental assessment. *Oecologia Australis*, 25 (4), 834–845. <https://doi.org/10.4257/oeco.2021.2504.04>.
- Simonini, R. & Prevedelli, D. (2003). Effects of temperature on two Mediterranean population of *Dinophilus gyrotiliatus* (Polychaeta: Dinophilidae) II. Effects on demographic parameters. *Journal of Experimental Marine Biology and Ecology*, 291, 95–110.
- Strickland, J.D.H. & Parsons, T.R. (1972). A Practical Handbook of Seawater Analysis. Fisheries Research Board of Canada. Ottawa, 167, 310 p.
- Terlizzi, A., De Falco, G., Fellingine, S., Fiorentino, D., Gambi, M.C. & Cancemi, G. (2010). Effects of marine cage aquaculture on macrofauna assemblages associated with *Posidonia oceanica* meadows. *Italian Journal of Zoology*, 77, 362-371.
- Tillin, H.M. (2016). *Protodorvillea kefersteini* and other polychaetes in impoverished circalittoral mixed gravelly sand. In Tyler-Walters H. and Hiscock K. (eds) Marine Life Information Network: Biology and Sensitivity Key Information Reviews, [on-line]. Plymouth: Marine Biological Association of the United Kingdom. <https://dx.doi.org/10.17031/marlinhab.1115.1>.
- Warwick, R.M., Collins, N.R., Gee, J.M. & Geaorge, C.L. (1986). Species size distributions of benthic and pelagic Metazoa: evidence for interaction? *Marine Ecology Progress Series*, 34, 63-68.
- Warwick, R.M. (2001). Evidence for the effects of metal contamination on the intertidal macrobenthic assemblages of the Fal Estuary. *Marine Pollution Bulletin*, 42 (2), 145-148.
- WORMS, 2023. World Register of Marine Species. Accessed by <http://https://www.marinespecies.org>.
- Zaâbi, S., Gillet, P., Afli, A. & Boumaiza, M. (2009). Biodiversity of polychaetous annelids from the peninsula of Cap Bon, northeast coast of Tunisia. *Zoosymposia*, 2, 587–600.
- Zaâbi, S., Gillet, P., Afli, A., & Boumaiza, M. (2010). Structure and diversity of polychaetous annelids population along the eastern coast of the Cap Bon Peninsula (north-east coast of Tunisia, western Mediterranean). *Marine Biodiversity Records*, page 1 of 11, 3, e100. <http://doi.org/10.1017/S1755267210000850>.
- Zaabi-Sendi, S. (2013). Etude faunistique et ecologique des annelides polychètes de la cote nord-est de la Tunisie (Peninsule du Cap Bon, Mediterranee Ouest). These de Doctorat, Université de Carthage, Faculté des Sciences de Bizerte, 494 pp.

Validity of Visible Ectoparasite Intensity As a Non-invasive Biomarker for Fish welfare: Parasitic Copepod, *Lernantropus kroyeri* in Sea Bass As an Example

Hijran Yavuzcan¹ 

Cite this article as: Yavuzcan, H. (2024). Validity of visible ectoparasite intensity as a non-invasive biomarker for fish welfare: parasitic copepod, *Lernantropus kroyeri* in sea bass as an example. *Aquatic Sciences and Engineering*, 39(1), 30-35. DOI: <https://doi.org/10.26650/ASE20241334425>

ABSTRACT

Ensuring fish welfare is essential from the ethical, legal, environmental, economic, and social perspectives. It plays a vital role in maintaining the health and sustainability of aquaculture practices while respecting the intrinsic value and welfare of the fish themselves. The presence of reliable welfare assessment schemes is of utmost importance to appraise the well-being of animals in aquaculture and uphold stringent welfare standards. In determining fish welfare, conducting welfare assessments with non-invasive biomarkers is crucial thus the primary objective of this study is to explore the potential usability of visible parasites as welfare biomarkers in fish without causing any harm to the fish. In this research, certain secondary stress indicators (hematocrit, plasma glucose and lactate) were employed as biomarkers for assessing the well-being of European sea bass (*Dicentrarchus labrax*). The study aimed to investigate whether there is a possible correlation between the presence of visible ectoparasites on the gills (specifically, the Copepod parasite, *Lernantropus kroyeri*) and the aforementioned stress parameters. Thus, in this study, the examination was conducted to establish the validity of ectoparasites as non-invasive biomarkers for evaluating the welfare of fish. The results showed that there was a statistically significant relationship between the intensity of ectoparasites and the stress parameters used as indicators of welfare. The observable presence and intensity of ectoparasites on the gills of the fish can be proposed as a non-invasive biomarker for evaluating fish welfare in aquaculture.

Keywords: Fish welfare, non-invasive biomarker, ectoparasite intensity, validity

ORCID IDs of the author:
H.Y. 0000-0001-6567-7467

¹Ankara University, Faculty of
Agriculture, Department of Fisheries and
Aquaculture, Ankara, Turkiye

Submitted:
29.07.2023

Revision Requested:
17.09.2023

Last Revision Received:
20.09.2023

Accepted:
03.10.2023

Online Published:
17.11.2023

Correspondence:
Hijran Yavuzcan
E-mail:
yavuzcan@ankara.edu.tr

INTRODUCTION

The importance of animal welfare in the aquaculture industry, which is responsible for producing 52% of the fish consumed by humans (FAO, 2022) has started to be better understood. Poor welfare conditions in aquaculture can lead to negative outcomes for the environment, animal well-being, and global food security. By promoting fish well-being, the aquaculture industry can contribute to improving sustainability and fish food safety, which is beneficial for the environment and human health.

Therefore, it is crucial to ensure sustainable growth in aquaculture while prioritizing fish welfare.

Non-invasive biomarkers for assessing fish welfare have gained significant attention in recent years. These biomarkers allow for the evaluation of fish well-being without causing unnecessary stress or harm to the animals. The need for non-invasive fish welfare biomarkers arises from the increasing concern and recognition of the ethical and practical importance of fish welfare in various industries, including aquaculture,



fisheries, and research. Fish are sentient beings capable of experiencing pain, stress, and suffering, and as such, there is a growing responsibility to ensure their well-being and minimize any potential harm they may experience (Garrath & McCulloch, 2022). Assessing fish welfare has been challenging due to their environment and the fact that they are not as expressive as some land animals. Many conventional welfare assessment methods involve capturing, handling, or even sacrificing fish for analysis, which can cause additional stress and harm to the animals (Browning, 2023). Non-invasive biomarkers offer a more humane and sustainable approach to monitoring and assessing fish welfare. These biomarkers can be indicators of physiological, biochemical, or molecular changes in fish, providing valuable insights into their well-being without causing harm or stress. For example, cortisol concentration in fish skin mucus has been proposed as a non-invasive technique to assess fish welfare and stress (Guardiala et al., 2016; Carbajal et al., 2019). Similarly, the cutaneous stress response system (CSRS) in fish skin has been reported as a new source of information on the welfare status of farmed fish (Kulcykowska, 2019). Although not strictly biomarkers, observing fish behavior can provide valuable insights into their welfare. Non-invasive methods such as video recording or automated image analysis can be used to monitor behaviors such as feeding patterns, swimming activity, aggression, or abnormal behaviors, which can indicate stress or welfare issues (Martins et al., 2012). The establishment of standardized indicators and assessment of welfare needs in aquaculture has been an ongoing discussion (Segner et al., 2019; Magalhães et al., 2020; Barreto et al., 2021; Yavuzcan Yildiz et al., 2021). The use of welfare indicators provides crucial insights into fish welfare, reflecting the extent to which their needs are met. These indicators should possess characteristics such as usability, reliability, scalability, recognizability, minimal damage, and feasibility within a reasonable timeframe. An ideal assessment framework should encompass both operational and laboratory-based welfare markers (Segner et al., 2019; Browning, 2023).

Fish health assessments are one component of on-farm welfare appraisals, particularly in intensive aquaculture systems. Segner et al. (2012) explained that good welfare is reflected in the ability of the fish to cope with various stressors, thereby maintaining homeostasis associated with good health, while stressful aquaculture conditions will cause the loss of coping capacity and impaired health. The welfare status of fish, as measured by the indices of stress can be unveiled. The biochemical and physiological changes result from the effects of the factors released during the primary stress response (Ellis et al., 2012; Schreck & Tort, 2016). The secondary stress response refers to the physiological and behavioral adaptations that occur in response to stressful conditions. These adaptations involve the activation of various metabolic pathways, leading to significant changes in blood chemistry, hematology, respiration, acid-base balance, and ion losses in the gills (Iwama, 2007). Hematocrit, blood lactate, and glucose are considered secondary stress biomarkers in fish. These biomarkers are used to assess the physiological responses of fish to stress and provide valuable information about their overall well-being and health (Seibel et al., 2021).

It is generally recognized that parasitic diseases in fish are one of the most important indicators in relation to fish health (Segner et al., 2012; Stien et al., 2013; Bui et al., 2019). The visible parasites are considered in operational indicators and assessment of fish welfare (Bui et al., 2019), and visible ectoparasites can provide insights into fish well-being without necessarily causing the death of the fish. Thus, the study by Overli (2014) has contributed to our understanding of the relationship between parasites, stress, and welfare. This study revealed that infestation by sea lice led to increased brain stem levels of the 5-Hydroxytryptamine(5-HT) catabolite 5-Hydroxyindolacetic acid (5-HIAA) in Atlantic salmon, indicating a general stress response in infested fish. It was also found that infected fish showed depressed feeding and reduced locomotion, suggesting a negative impact on animal welfare.

The European sea bass (*D. labrax*), one of the most commonly farmed fish in the Mediterranean, faces challenges from the ectoparasite *Lernanthropus* (Yavuzcan Yildiz & Korkmaz, 2021). *Lernanthropus kroyeri*, belonging to the genus of parasitic copepods, is visible in the gills of sea bass and causes several pathologies such as erosion and necrosis on gill filaments (Henry et al., 2009). The parasite *L. kroyeri* in sea bass has been examined in various studies (Tokşen, 2007; Er&Şevki, 2015; Özak et al., 2016; Yavuzcan Yildiz & Korkmaz, 2021), however, this parasite has not been studied as a welfare biomarker till now. Yet, the easy visibility of *Lernanthropus* to the naked eye may provide the basis for its potential use as a biomarker for European sea bass.

The development and implementation of non-invasive fish welfare biomarkers are essential for ensuring the ethical treatment of fish, promoting sustainable practices in the aquaculture industry, and enhancing our understanding of fish welfare in various settings. Despite the extensive attempts made towards the authentication of welfare indicators that are specific to each species, the identification of suitable combinations of these measures still proves to be limited for the majority of farmed species as stated by Magalhães et al. (2020).

In this study, the aim was to validate the potential use of visible gill parasites (*L. kroyeri*) as the non-invasive biomarker of fish welfare by correlating them with key stress parameters such as hematocrit, blood glucose, and lactate.

MATERIAL AND METHODS

The fish examination was done immediately after harvesting on a commercial farm in the Aegean Sea. Fish samples ($N=37$) were randomly selected among the harvested batch from one cage in the autumn. The weight of European sea bass (*D. labrax*) was around 350 g. During the harvesting period, the water temperature was measured as 23 °C and the dissolved oxygen level as 6.5 mg/L.

Blood samples were collected promptly upon the taking of the fish out. Prior to blood sampling, the fish underwent a brief anesthetic procedure involving a clove oil solution (5 mg/L) for a duration of 5 minutes. Heparinized syringes were utilized to collect blood via the cardiac puncture. Hematocrit measurements were conducted immediately following blood sampling. Plasma was obtained by centrifugation of heparinized blood at 3000g for 10

min and pooled where necessary. The plasma was then preserved at a temperature of -18°C until further analysis. Commercial kits (Cayman Chemicals, USA) and a Shimadzu UV-1210V spectrophotometer were used to measure plasma glucose and lactate, in accordance with the manufacturer's instructions.

The copepod parasite *L. kroyeri* was carefully plucked from the gills by using forceps in the newly harvested fish (Figure 1). Parasites on both sides of the gills were counted on the gills with the naked eye (Figure 2). The sex of the parasite was not considered.

Statistical analyses: The variation in stress parameters was examined using an ANOVA test based on parasite counts. Regression analyses were conducted to test whether there is a significant relationship between parasite counts on the gills and secondary stress parameters (hematocrit, plasma glucose and lactate).

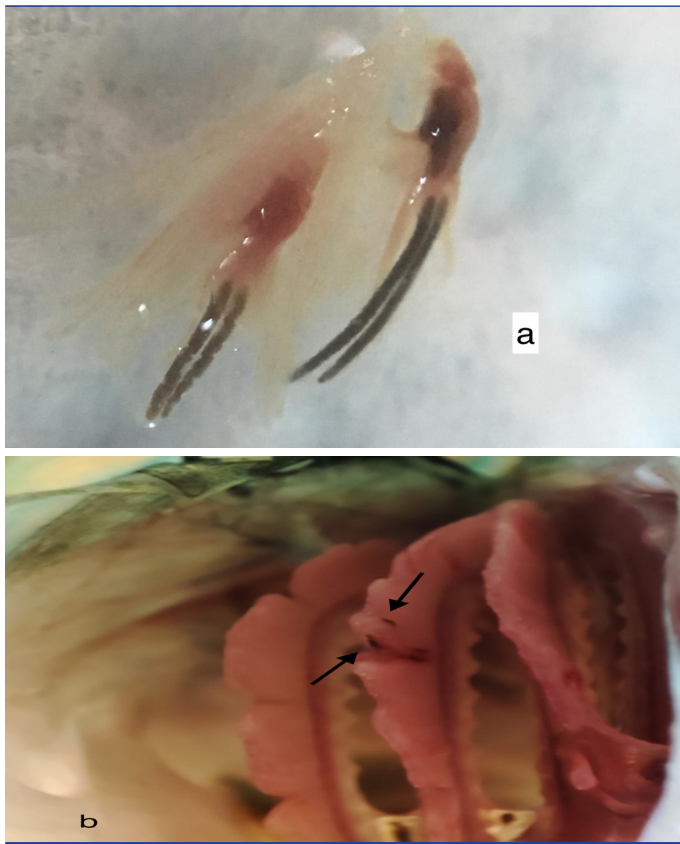


Figure 1. a) Copepod parasite, *Lernantropus kroyeri*
b) *L. kroyeri* on the gills of European sea bass.

RESULTS AND DISCUSSION

In this study, the validation of visible ectoparasites (crustacean parasite, *L. kroyeri*) on the fish gills as a non-invasive fish welfare biomarker has been evaluated. The welfare status of the fish was assessed using the stress indicators as the concept of stress is closely connected to that of fish well-being (Galhardo & Oliveira, 2009). When an organism's coping mechanisms fail to adapt to its environment adequately, it leads to poor welfare (Broom, 2008). Stress can negatively impact fish health, growth, reproduction, and overall well-being of fish.

The prevalence of *L. kroyeri* on sea bass gills was 78.37%. Stress indicators (hematocrit, plasma glucose, and lactate) showed significant differences by the parasite *L. kroyeri* number in the gills of the fish ($p < 0.05$). The variation in the stress indicators was parasite number-dependent (Table 1).

Wells & Pankhurst (1999) noted that the persistence of altered states of glucose and lactate as well as hematological changes following stress are clearly-defined indicators of the stress response although the variations in these indicators may be considered high to observe chronic stress (Magalhães et al., 2020). The increased hematocrit is an indication of the increase in the oxygen requirement of the fish body. Under stressful circumstances, there exists a possibility for hematocrit values to exhibit an elevation, which is a compensatory mechanism aimed at augmenting the supply of oxygen to the organs as a response to the escalated metabolic demand (Fazio et al., 2015). Increased hematocrit levels have been reported for the hypoxic conditions in sea bream (Bermejo-Nogales et al 2014). Thus, here, the higher levels of hematocrit in sea bass with higher numbers of *L. kroyeri* can be explained by the disrupted oxygen balance in the gills. The increase in glucose and lactic acid levels in fish during stress reflects their physiological response to challenging conditions (Levy de Carvalho Gomes, 2007). These metabolic changes help the fish adapt to the stressor, providing the necessary energy for survival, but prolonged or severe stress can have negative effects on fish health and welfare (Wells & Pankhurst, 1999). Blood glucose serves as the primary energy fuel in stressful conditions (Wendelaar-Bonga, 1997) and blood lactate is produced as a metabolic byproduct during certain physiological conditions, including stress (Schreck & Tort, 2016). However, high blood lactate can limit oxygen transport to the tissues in fish (Olsen et al., 1992). Plasma glucose and lactate increased by the parasite numbers on the gills in a manner that is parasite number-dependent in this study. Here, the increase in parasite numbers being associated with an increase in hematocrit, plasma glucose, and lactate levels is an important finding of this study.

Table 1. The changes of stress indicators in sea bass by the *L. kroyeri* intensity.

<i>L. kroyeri</i> intensity in the gills	Frequency of parasitized fish (N _{total} =37)	Hematocrit (%)	Plasma glucose (mg/dL)	Plasma lactate (mg/dL)
0 (uninfested fish)	8	30.62*	67*	10.25*
1-3	9	33	83.88	15.33
4-10	11	35	91.72	17.18
>11	9	36.11	129.11	17.44

*refers the difference for the values in the column

The regression analysis resulted in a significant overall effect of the independent variables (parasite intensity on the gills) on the dependent variables; stress indicators including hematocrit, plasma glucose, and plasma lactate (Table 2).

The regression results for hematocrit provides a moderate-to-good explanation of the hematocrit variability ($R^2=0.56$ and $F=44.75$), however, the individual regression coefficient is statistically significant ($p<0.01$), suggesting that parasite number has some predictive power for hematocrit values of sea bass (Figure 2).

For plasma glucose, the regression model appeared to be highly significant ($R^2=0.85$ and $F=200.45$), indicating that the ectoparasite numbers have a substantial impact on the plasma glucose of sea bass (Figure 3).

The regression model was statistically significant ($F = 36.41$) and explains a moderate amount of the variability in the plasma lactate ($R^2=0.50$), indicating that the model has identified significance between parasite intensity and plasma lactate (Figure 4).

Regarding the potential use of visible ectoparasite presence as a non-invasive biomarker, regression analyses have shown a strong relationship between changes in stress indicators and parasite numbers in the present study. The regression results

provide strong evidence that the parasite numbers on the gills have a significant impact on the secondary stress parameters of hematocrit, plasma glucose, and plasma lactate. Based on these regression results, the presence and intensity of parasites can be utilized as a non-invasive biomarker for fish welfare assessment. Furthermore, to strengthen the argument for the usability of parasites as biomarkers, it should be emphasized that parasites serve as a biomarker indicating both fish welfare and environmental quality in aquaculture. The rise in parasite numbers in fish can serve as a warning sign of an unsuitable environment. Thus, a comprehensive approach has been implemented for addressing sea lice in salmon farming in Norway. Sea lice are regarded as indicators of regional environmental sustainability, and certain limitations on biomass production are imposed based on the recorded parasite levels on farms within a specific area. Farms that consistently maintain low parasite abundance on fish are permitted to produce larger biomasses (Bui et al., 2019). It is also essential to highlight their consistent presence and relevance in different ecological contexts. Copepod parasites such as *L. kroyeri* and their copepodites are available throughout the year, resulting in constant infection pressure in the sea cages (Yavuzcan Yildiz & Korkmaz 2021). The crustacean parasite's continuous presence provides suitability for using these parasites as non-invasive biomarkers.

Table 2. The key parametric in regression analysis for stress indicators.

Dependent variable*	R-squared	F value	P value
Hematocrit	0.56	44.75	<0.01
Plasma glucose	0.85	200.45	<0.01
Plasma lactate	0.50	36.41	<0.01

* The independent variable was the parasite number on the gills

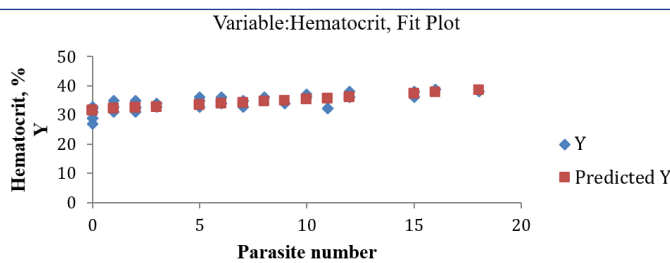


Figure 2. Hematocrit.

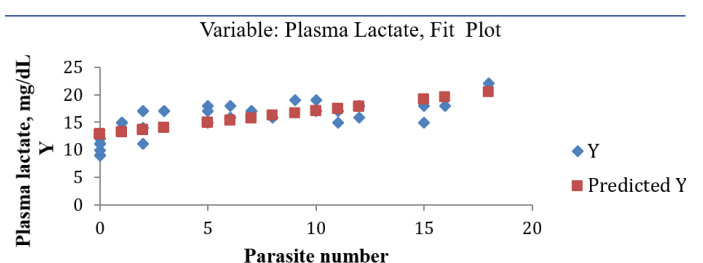


Figure 4. Plasma lactate.

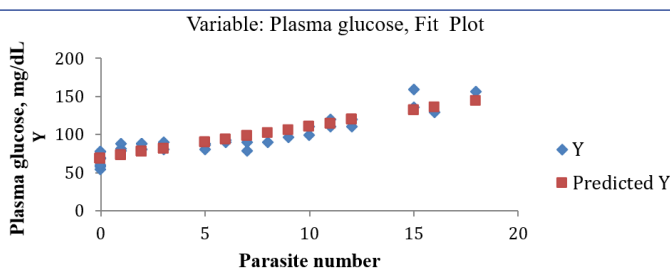


Figure 3. Plasma glucose.

CONCLUSION

While varying between moderate and strong, a statistically significant relationship exists between the numbers of copepod parasites (*L. kroyeri*) on the gills and secondary stress indicators (hematocrit, blood glucose, and lactate) in sea-caged European sea bass. In the regression analysis, R-squared values (approximately 50%) for the hematocrit and plasma lactate indicate that the independent variables in the model explain half of the vari-

ance in the dependent variable, necessitating cautious interpretation. An R-squared value of 0.85 is relatively high and suggests a strong positive linear relationship between the plasma glucose and the parasite number on the gills in the regression. Nevertheless, with R-squared values of each biomarker consistently above 50%, it is considered that the results of this study can support the utilization of parasite count as a non-invasive biomarker for fish welfare assessment. Therefore, the visible ectoparasite presence on the surface of the fish and their intensity can be suggested as a non-invasive biomarker for assessing fish welfare in aquaculture and an effective tool in explaining the welfare status of the fish. The validation process typically included defining requirements, establishing testing protocols, conducting tests, analyzing results, and documenting the findings. The goal was to provide confidence that the presence of visible parasites is fit to welfare assessment as a non-invasive biomarker, however, both validation and verification would be essential steps in the integration of visible parasites on fish gills in operational welfare indicators as the non-invasive biomarker.

It's worth noting that the development and validation of non-invasive biomarkers for fish welfare are ongoing research areas. Exploring new techniques and biomarkers to improve the assessment of fish welfare while minimizing the impact on the fish is significant for sustainable aquaculture.

Conflict of interest: The author declares no conflict of interest.

Ethics committee approval: Ethical approval for this study was obtained from ANKARA UNIVERSITY (2018-16-101). The present study followed international, national, and/or institutional guidelines for humane animal treatment.

Financial disclosure: None

Acknowledgments: The author thanks Agro-Mey aquaculture company for providing fish.

REFERENCES

- Barreto, M.O., Rey Planellas, S., Yang, Y., Phillips, C., Descovich, K. (2021). Emerging indicators of fish welfare in aquaculture. *Reviews in Aquaculture*. <https://doi.org/10.1111/raq.12601>
- Bermejo-Nogales, A., Calduch-Giner, J.A., Pérez-Sánchez, J. (2014) Tissue-specific gene expression and functional regulation of uncoupling protein 2 (UCP2) by hypoxia and nutrient availability in gilthead sea bream (*Sparus aurata*): implications on the physiological significance of UCP1-3 variants. *Fish Physiology & Biochemistry*, 40, 751–762.
- Browning, H. (2023). Improving welfare assessment in aquaculture. *Frontiers in Veterinary Science*. <https://doi.org/10.3389/fvets.2023.1060720>
- Bui, S., Oppedal, F., Sievers, M. and Dempster, T. (2019). Behaviour in the toolbox to outsmart parasites and improve fish welfare in aquaculture. *Reviews in Aquaculture*, 11: 168-186.
- Carbajal, A., Soler, P., Tallo-Parra, O., Isasa, M., Echevarria, C., Lopez-Bejar, M., Vinyoles, D. (2019). Towards Non-Invasive Methods in Measuring Fish Welfare: The Measurement of Cortisol Concentrations in Fish Skin Mucus as a Biomarker of Habitat Quality. *Animals*, <https://doi.org/10.3390/ani9110939>
- Ellis, T., Yildiz, H. Y., López-Olmeda, J. F., Spedicato, M. T., Tort, L., Øverli, Ø., Martins, C. I. M., & Martins, C. I. M. (2012). Cortisol and finfish welfare. *Fish Physiology & Biochemistry*, <https://doi.org/10.1007/S10695-011-9568-Y>
- Er, A. & Kayış, Ş. (2015). Intensity and prevalence of some crustacean fish parasites in Turkey and their molecular identification. *Turkish Journal of Zoology*, 39 (6), <https://doi.org/10.3906/zoo-1409-35>
- FAO (2022). The state of World Fisheries and Aquaculture. Rome. ISBN 978-92-5-136364-5
- Fazio, F., Ferrantelli, V., Fortino, G., Arfuso, F., Giangrosso, G., Faggio, C. (2015). The Influence of Acute Handling Stress on Some Blood Parameters in Cultured Sea Bream (*Sparus aurata* Linnaeus, 1758). *Italian Journal of Food Safety*. 11, 4(1):4174.
- Galhardo, L., & Oliveira, R. F. (2009). Psychological Stress and Welfare in Fish. *Annual Review of Biomedical Sciences*, <https://doi.org/10.5016/1806-8774.2009V11P1>
- Garratt, J. K., McCulloch, S. (2022). Wild Fish Welfare in UK Commercial Sea Fisheries: Qualitative Analysis Of Stakeholder Views. *Animals*, <https://doi.org/10.3390/ani12202756>
- Guardiola, F. A., Cuesta, A., & Esteban, M. Á. (2016). Using skin mucus to evaluate stress in gilthead seabream (*Sparus aurata* L.). *Fish & Shellfish Immunology*, <https://doi.org/10.1016/J.FSI.2016.11.005>
- Henry, M., Alexis, M., Fountoulaki, E., & Nengas, I. (2009). Effects of a natural parasitological infection (*Lernanthropus kroyeri*) on the immune system of European sea bass, *Dicentrarchus labrax* L. *Parasite Immunology*, <https://doi.org/10.1111/j.1365-3024.2009.01150.x>
- Iwama, G. K. (1998). *Fish stress and health in aquaculture*. 21(3). Cambridge University Press. <https://doi.org/10.2307/1352849>
- Kulczykowska, E. (2019). Stress Response System in the Fish Skin-Welfare Measures. *Frontiers in Physiology*, 10. <https://doi.org/10.3389/FPHYS.2019.00072>
- Levy de Carvalho Gomes, L. de C. G. (2007). Physiological responses of pirarucu (*Arapaima gigas*) to acute handling stress. *Acta Amazonica*, <https://doi.org/10.1590/S0044-59672007000400019>
- Magalhães, C. R. de, Schrama, D., Farinha, A. P., Revets, D., Kuehn, A., Planchon, S., Rodrigues, P. M., & Cerqueira, M. (2020). Protein changes as robust signatures of fish chronic stress: a proteomics approach to fish welfare research. *BMC Genomics*, <https://doi.org/10.1186/S12864-020-6728-4>
- Martins, C. I. M., Martins, C. I. M., Galhardo, L., Noble, C., Damsgård, B., Spedicato, M. T., Zupa, W., Beauchaud, M., Kulczykowska, E., Massabuau, J.-C., Carter, T., Rey Planellas, S., & Kristiansen, T. S. (2012). Behavioural indicators of welfare in farmed fish. 38(1). *Fish Physiology & Biochemistry*, <https://doi.org/10.1007/S10695-011-9518-8>.
- Olsen, Y., Falk, K., & Reite, O. (1992). Cortisol and lactate levels in Atlantic salmon *Salmo salar* developing infectious anaemia (ISA). 14. *Diseases of Aquatic Organisms*, <https://doi.org/10.3354/DAO014099>
- Özak, A. A., Demirkale, I., & Yanar, A. (2016). Lernanthropid copepods parasitic on marine fishes in Turkish waters, including two new records. *Zootaxa*, 4174(1), 161-175.
- Øverli, Ø., Nordgreen, J., Mejdell, C. M., Janczak, A. M., Kittilsen, S., Johansen, I. B., & Horsberg, T. E. (2014). Ectoparasitic sea lice (*Lepeophtheirus salmonis*) affect behavior and brain serotonergic activity in Atlantic salmon (*Salmo salar* L.): Perspectives on animal welfare. *Physiology and Behaviour*, <https://doi.org/10.1016/J.PHYSBEH.2014.04.031>
- Schreck, C. B., & Tort, L. (2016). 1 - The Concept of Stress in Fish. In C. B. Schreck, L. Tort, A. P. Farrell, & C. J. Brauner (Eds.), *Biology of Stress in Fish* (Vol. 35, pp. 1–34). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-802728-8.00001-1>
- Segner, H., Sundh, H., Buchmann, K., Douxfils, J., Sundell, K.S., Mathieu, C., Ruane, N., Jutfelt, F., Toften, H., Vaughan, L. (2012). Health of farmed fish: its relation to fish welfare and its utility as welfare indicator. *Fish Physiology & Biochemistry*, 38(1), 85–105.
- Segner, H., Reiser, S., Ruane, N., Rösch, R., Steinhagen, D., Vehanen, T. (2019). Welfare of fishes in aquaculture. FAO Fisheries and Aquaculture Circular No. 1189. Budapest: FAO.

- Seibel, H., Baßmann, B., & Rebl, A. (2021). Blood Will Tell: What Hematological Analyses Can Reveal About Fish Welfare. *Frontiers in Veterinary Science*, <https://doi.org/10.3389/fvets.2021.616955>
- Stien, L. H., Bracke, M. B. M., Folkedal, O., Nilsson, J., Oppedal, F., Torgersen, T., Kittilsen, S., Midtlyng, P. J., Vindas, M. A., Øverli, Ø., & Kristiansen, T. S. (2013). *Salmon Welfare Index Model (SWIM 1.0): a semantic model for overall welfare assessment of caged Atlantic salmon: review of the selected welfare indicators and model presentation*. 5(1). *Reviews in Aquaculture*, <https://doi.org/10.1111/J.1753-5131.2012.01083.X>
- Tokşen, E. (2007). *Lernanthropus kroyeri* van Beneden, 1851 (Crustacea: Copepoda) infections of cultured sea bass (*Dicentrarchus labrax* L.). *Bulletin of the European Association of Fish Pathologists*, 27 (2), 49.
- Wells, R. M. G., & Pankhurst, N. W. (1999). Evaluation of Simple Instruments for the Measurement of Blood Glucose and Lactate, and Plasma Protein as Stress Indicators in Fish. 30(2). *World Aquaculture Society*, <https://doi.org/10.1111/J.1749-7345.1999.TB00876.X>
- Wendelaar Bonga, S. E. (1997). The stress response in fish. *Physiological Reviews*, 77(3). <https://doi.org/10.1152/PHYSREV.1997.77.3.591>
- Yavuzcan Yildiz, H., Chatzifotis, S., Anastasiadis, P., Parisi, G., & Papandroulakis, N. (2021). Testing of the Salmon Welfare Index Model (SWIM 1.0) as a computational welfare assessment for sea-caged European sea bass. *Italian Journal of Animal Science*, <https://doi.org/10.1080/1828051X.2021.1961106>
- Yavuzcan Yildiz, H., & Korkmaz, A. S. (2021). Parasitic copepod (*Lernanthropus kroyeri*) on caged sea bass (*Dicentrarchus labrax*): An estimation of abundance and internal infestation pressure. *Journal of Fish Diseases*, <https://doi:10.1111/jfd.13504>

Length – Weight Relationships, Meat Yield and Morphometric Indices of Five Commercial Bivalve Species Collected from the Çanakkale Strait (Türkiye)

Serhat Çolakoğlu¹ , Fatma Çolakoğlu² , İbrahim Ender Künili³ 

Cite this article as: Colakoglu, S., Colakoglu F., Kunili, I.E. (2023). Length – weight relationships, meat yield and morphometric indices of five commercial bivalve species collected from the Canakkale Strait (Türkiye). *Aquatic Sciences and Engineering*, 39(1), 36-42. DOI: <https://doi.org/10.26650/ASE20241371586>

ABSTRACT

This study was conducted to determine the meat yield, morphometric characteristics, length–weight relationships (LWRs) and their correlations with environmental variables of five commercial bivalve species collected monthly between 2014 and 2015 from the coastal waters of the Çanakkale Strait. A total of 8588 individuals were examined, and different ranges for both shell length (9.00–108.50 mm) and total weight (0.30–234.20 g) were determined according to species. The highest meat yields from *Donax trunculus* (16.40–23.34%), *Mytilus galloprovincialis* (14.89–34.35%) and *Ostrea edulis* (5.91–26.24%) were determined in spring, while *Ruditapes philippinarum* (10.80–29.53%) and *Chamelea gallina* (12.26–18.92%) had maximum yield in late summer and early autumn ($p<0.05$). Elongation index (SH/SL), compactness index (SW/SL), convexity index (SW/SH), and density indexes (TW/SL) were significant ($p<0.05$) and had high correlation coefficients ($r=0.806$ – 0.975). The mean value of the allometry coefficient (b) was 3.257 ± 0.168 , ranging from 2.291 to 4.058. Four species had negative allometries, namely *D. trunculus* (2.738), *C. gallina* (2.889), *M. galloprovincialis* (2.597) and *O. edulis* (2.728), while *R. philippinarum* (3.137) displayed positive allometry. The morphometric indices show high morphological resemblances. As a result, it is thought that the data obtained in this study can both provide data in the fields of biology and ecology for current scientific studies on these species, and can be used as a resource for the sustainable production of these commercial species.

Keywords: Meat yield, morphometric indices, bivalve species, Çanakkale Strait

ORCID IDs of the author:
S.Ç. 0000-0003-3526-6477;
F.Ç. 0000-0002-2211-8371;
İ.E.K. 0000-0003-2830-6979

¹Çanakkale Onsekiz Mart University,
Çanakkale Vocational of Technical
Sciences, Çanakkale, Türkiye

²Çanakkale Onsekiz Mart University,
School of Applied Sciences,
Çanakkale, Türkiye

³Çanakkale Onsekiz Mart University,
Faculty of Marine Sciences and
Technology, Çanakkale, Türkiye

Submitted:
05.10.2023

Revision Requested:
30.10.2023

Last Revision Received:
13.11.2023

Accepted:
28.11.2023

Online Published:
00.00.0000

Correspondence:
Serhat Çolakoğlu
E-mail:
serhat_colakoglu@comu.edu.tr

INTRODUCTION

Bivalves are unique organisms in terms of ecological impact in the marine environment and as a nutritional food in many countries (Wijsman et al., 2019). Their economic value is directly proportional to consumer demand, and demand can cause significant population reductions in regions where stock-supporting activities such as aquaculture are not carried out (Wijsman et al., 2019). Türkiye has significant bivalve production areas from the Aegean Sea to the Black Sea, where the primary production

method is based on fishing.

Bivalve fishing has been performed using various methods (hydraulic and mechanical dredges, hand dredges, SCUBA diving) since the 1970s in Türkiye, and socio-economically contributes to coastal communities (Çolakoğlu & Palaz, 2015). In the coastal waters of the Marmara Sea and the Çanakkale Strait (0–20 m), there are various bivalve species with high economic value and extensive stocks, such as wedge clam (*Donax trunculus* Linnaeus, 1758), striped venus (*Chamelea gallina* Linnaeus,



1758), venerid clam (*Ruditapes philippinarum* Adams and Reeve, 1850), Mediterranean mussel (*Mytilus galloprovincialis* Lamarck, 1819) and flat oyster (*Ostrea edulis*). Among them, the most produced (fished) bivalve species along the coasts of Türkiye in 2020 were reported to be *Mytilus galloprovincialis* (~4000 tons) and *Chamelea gallina* (~30 000 tons) (Turkstat, 2021).

The length–weight relationship (LWRs) and morphometric characteristics are important for biology and fisheries in population dynamics (Gaspar et al., 2002). This information is useful especially for predicting current conditions and stock assessment, as well as morphological comparisons of bivalve species (Gaspar, Santos & Vasconcelos, 2001; Vasconcelos et al., 2018). In relative studies conducted worldwide, LWRs, relative growth and shell morphometric relationships were assessed for different bivalve species (Charef, Langar & Gharsallah, 2012; Gaspar et al., 2001; 2002; Petetta et al., 2019; Vasconcelos et al., 2018). In Türkiye, different bivalve species caught along the coastal areas between the Aegean and Black Seas were analysed in terms of morphometric and population characteristics (Çolakoğlu & Palaz, 2014; Çolakoğlu & Tokaç, 2014; Dalgıç, 2006; Deval, 2009). Also, several studies in the southern Marmara Sea and the Çanakkale Strait focused on diverse subjects such as fishing, population dynamics, aquaculture, chemical and biological contaminants (Çolakoğlu et al., 2011; Künili, Çolakoğlu & Çolakoğlu, 2021a; Künili et al., 2021b). However, there is a limited number of studies to determine the meat yields and morphometric characteristics of these commercial bivalve species and to compare them with environmental parameters in the South Marmara Sea and Çanakkale Strait, where one of the densest populations is found (Çolakoğlu, 2011). The present study was performed to determine and compare the meat yield, morphometric relationships (between shell length, height, width, and total weight), morphometric indices (elongation, compactness, convexity and density) and relative growth (isometry vs allometry) of *D. trunculus*, *M. galloprovincialis*, *O. edulis*, *R. philippinarum*, and *C. gallina* collected along the Çanakkale Strait coast in Türkiye.

MATERIALS AND METHODS

Material, study area and sampling

The research materials were *D. trunculus*, *M. galloprovincialis*, *O. edulis*, *R. philippinarum*, and *C. gallina* collected from coastal areas containing both hard substrate and sandy bottoms along the Çanakkale Strait in the west of the Marmara Sea (Figure 1). The sampling locations were selected according to current information on bivalve harvesting and from local fishermen. Due to the living habits of research materials, two sampling methods were used for the two predetermined sampling groups. In the first group, *D. trunculus*, *C. gallina*, and *R. philippinarum* samples were collected from 1–8 m depths by using a mechanical dredge towed parallel to the shoreline for 5 min at a constant speed of 1–2 knots (length of dredge width and height: 55 and 30 cm; the number of teeth and length: 25 and 16 cm; mesh size: 5 mm). For the second group, *M. galloprovincialis* and *O. edulis* samples found at 1–12 m depths around sampling locations were collected by hand during SCUBA dives. All samples were collected between September 2014 and August 2015. Samples were firmly packed with wire meshes and transported to the laboratory via an ice-cooled insulated box within 1–2 hours.

Environmental parameters of the sampling areas, such as Sea Surface Temperature (SST) (°C), salinity (ppt), dissolved oxygen ($\text{mg}\cdot\text{L}^{-1}$), and pH, were measured *in situ* using a YSI 650 MDS multi-parameter water quality meter. Chlorophyll-*a* (Chl-*a*) concentrations in seawater samples obtained from the locations were determined according to the method described by the American Public Health Association (APHA, 1995).

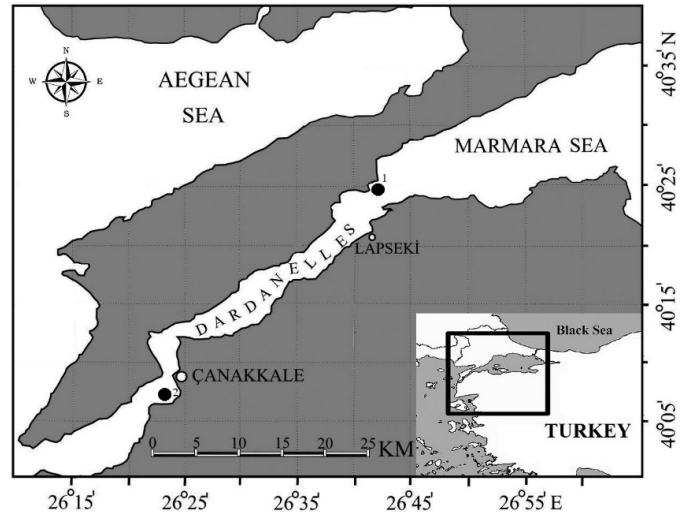


Figure 1. Map showing the study area along the Çanakkale Strait (Türkiye).

Data treatment and statistical analyses

The shell length (SL), shell height (SH), and shell width (SW) of individual specimens were measured using a digital vernier caliper (0.01 mm accuracy). The total wet weight (TW) and wet meat weight (MW) of each bivalve were measured using an electronic balance (d: 0.01 g, Max: 3100, Acculab, ALC-3100). Meat yield (MY) was calculated with the following equation: $\text{MY} (\%) = [(\text{MW}/\text{TW}) \times 100]$. Morphometric relationships were determined according to the allometric equation of Ricker (1973) $Y = aX^b$, where Y is SH, SW, or TW, X is the shell length (SL), a is the intercept, and b is the slope. The allometry coefficient is expressed by the exponent b in the linear regression equations. In these equations, in correlations between different types of variables and/or between different measuring units, the weight–length relationship reflects isometric growth when the exponent $b=3$ (Gaspar et al., 2002). To confirm whether the values of b obtained from linear regressions were significantly different from the isometric value ($b=3$) and described a negative ($b<3$) or positive ($b>3$) allometric relationship (Huxley & Teissier, 1936), the student t-test was applied with a confidence level of $\pm 95\%$ (Sokal & Rohlf, 1987).

Moreover, to characterize the morphology and growth shapes of bivalves, diverse morphometric indices including elongation index (SH/SL), compactness index (SW/SL), convexity index (SW/SH) and density index (TW/SL) were used (Vasconcelos et al., 2018; Caill–Milly et al., 2012, 2014). The significance for all statistical analyses was set at $P<0.05$ (Zar 1999).

RESULTS AND DISCUSSION

The present study evaluated the stock status of five commercial bivalve species from two stations in the Çanakkale Strait. Major environmental variables were measured and correlated with meat yield, as one of the most important economic properties, for complementary evaluation of the results. The results of the environmental parameters measured during the study period are summarized in Fig 2. The minimum and maximum values for SST (°C), salinity (ppt), DO (mg·L⁻¹), pH, and Chl-a (µg·L⁻¹) during the study period were determined as follows; 7.85 (Feb '15) – 23.85 (Jul '15) for SST, 22.00 (Aug '15)–26.00 (Mar '15) for salinity, 6.51 (Jul '15)–8.52 (Feb '15) for DO, 8.05 (Dec '14) – 8.40 (Mar '15) for pH, and 0.95 (Mar '15)–4.10 (Sep '14) for Chl-a, respectively.

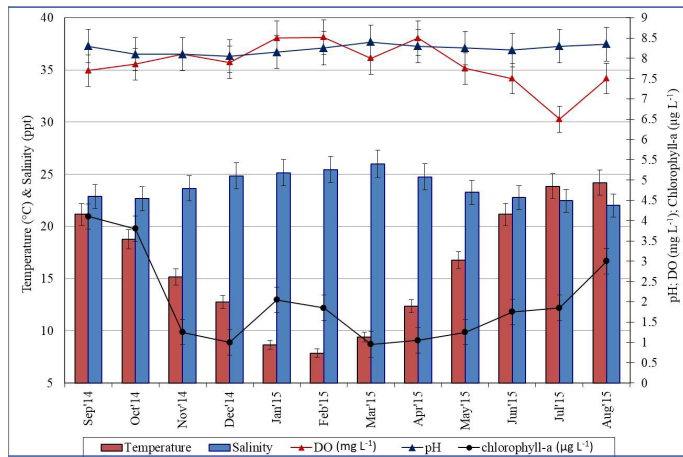


Figure 2. Summary of environmental variable data by months.

During the research period, 3394 individuals of *D. trunculus*, 2433 *C. gallina*, 1869 *R. philippinarum*, 580 *M. galloprovincialis* and 312 *O. edulis* were collected. The descriptive statistics and LWRs parameters of the samples are summarized in Table 1. The allometry coefficients (*b*) varied among species: *D. trunculus* =2.738, *C. gallina* =2.889, *R. philippinarum* =3.137; *M. galloprovincialis* =2.597 and *O. edulis* =2.728 ($P<0.05$). In general, regression analysis demonstrated a significant linear relationship for SH (range=0.481–1.128) ($P<0.05$), SW (range=0.197–0.891) ($P<0.05$), and TW

(range=0.323–2.159) ($P<0.01$) with SL. There were negative allometries (TW/SL) for four species *C. gallina* ($b=2.889$), *D. trunculus* ($b=2.738$), *M. galloprovincialis* ($b=2.597$) and *O. edulis* ($b=2.728$), while one bivalve species *R. philippinarum* ($b=3.137$) was observed to have positive allometries.

The morphometric indices, elongation (SH/SL), compactness (SW/SL), convexity (SW/SH), and density index (TW/SL) of the bivalve species studied are shown in Table 2. The morphometric indices of species were in the range 0.254–1.938 for SH/SL, 0.197–1.500 for SW/SL, 0.222–2.124 SW/SH, and 0.020–2.159 for TW/SL, respectively. In this study, four species had negative allometries, namely *D. trunculus* (2.738), *C. gallina* (2.889), *M. galloprovincialis* (2.597) and *O. edulis* (2.728), while *R. philippinarum* (3.137) displayed positive allometry.

According to the results, the lowest values were present for SH/SL in *M. galloprovincialis* and *O. edulis*; SW/SL in *O. edulis* and *D. trunculus*; SW/SH in *O. edulis* and *D. trunculus*, and TW/SL in *M. galloprovincialis* and *D. trunculus*. These values were found to be similar to those reported from Italian coasts for *M. galloprovincialis* (Orban et al., 2002), southern Black Sea for *D. trunculus* (Aydın, Tunca & Ersoy, 2020) and slightly different from the values from the Algarve coast, Portugal (Vasconcelos et al., 2018). In this study, mean values of SH/SL, SW/SL, SW/SH, and TW/SL were strongly correlated with SST and Chl-a ($P<0.05$). The morphology and physiology of bivalves are strongly influenced by fisheries and biomass (Gaspar et al., 2001, 2002), along with environmental conditions (Lucas et al., 1981) such as space competition for some species (Caill-Milly et al., 2014), differences in nutritional conditions and defence against predators (Caill-Milly et al., 2012; Tokeshi, Ota & Kawai, 2000; Watanabe & Katayama, 2010).

In comparison with previous reports, the allometric coefficient *b* (2.89) for *C. gallina* in this study was similar to values obtained from the Tyrrhenian Sea (2.74) and the Adriatic Sea (2.69) (Petetta et al., 2019), the West Marmara Sea (2.89) (Çolakoğlu & Tokaç, 2014), the North Sea (2.87) (Robinson et al., 2010), and the Algarve coast (Southern Portugal) (2.80) (Gaspar et al., 2001; Rufino et al., 2006), but higher than from the Mediterranean Sea (2.37) (Kasapoğlu & Düzgüneş, 2013). The *b* coefficient (2.74) value obtained for *D. trunculus* was similar to the values for the same species studied in the Tyrrhenian Sea (2.77) (Petetta et al., 2019) and

Table 1. Descriptive statistics, length–weight relationships and type of growth for five economic bivalves collected from the Çanakkale Strait (Türkiye).

Species	N	Length (mm)			Weight (g)			Morphometric relationship				Type of growth
		Min	Max	Mean±SD	Min	Max.	Mean±SD	a	b	SE (b)	r	
Dt	3394	13.00	40.50	28.78±0.07	0.36	7.69	2.93±0.02	0.0003	2.738	0.003	0.951	-A
Mg	580	15.05	84.00	59.02±0.39	0.30	50.17	16.06±0.28	0.0004	2.597	0.006	0.912	-A
Oe	312	53.00	108.50	75.73±0.54	17.30	234.20	72.52±1.78	0.0005	2.728	0.008	0.806	-A
Rp	1869	26.50	62.00	42.66±0.14	3.70	63.90	20.47±0.24	0.0001	3.137	0.003	0.932	+A
Cg	2433	9.00	38.00	23.65±0.11	0.30	17.76	4.67±0.07	0.0005	2.889	0.003	0.975	-A

Dt: Donax trunculus, Mg: Mytilus galloprovincialis, Oe: Ostrea edulis, Rp: Ruditapes philippinarum, Cg: Chamelea gallina, N: Number, SD: Standart Deviation, SE: Standart Error, a: Intercept, b: Slope, r: Correlation Coefficient; -A: Negative Allometry, +A: Positive Allometry

the West Marmara Sea (2.69) (Çolakoğlu, 2014), and higher than in the Adriatic Sea (2.48) (Petetta et al., 2019) and the Algarve coast (Southern Portugal) (2.57) (Gaspar et al., 2001). The b value (3.14) for *R. philippinarum* was similar to exponential values obtained on the southern coast of the Marmara Sea (3.14) (Çolakoğlu & Palaz, 2014) and the Taehwa River, Ulsan (3.04) (Choi et al., 2011), but higher than for the coast of Yeongi in Tongyeong, Korea (2.99) (Cho, Jeong & Lee, 2008) and the Amurshy Bay, Sea of Japan (2.95) (Ponurovsky, 2008). The b (2.73) for *O. edulis* was lower than findings obtained in Mersin Bay, Aegean Sea (3.15) (Acarlı et al., 2011), but higher than in the Black Sea (2.46) (Aydın & Biltekin, 2020). The b value (2.60) for *M. galloprovincialis* was similar to the findings obtained in the Istanbul Bosphorus (2.63) (Balçioğlu & Gönülal, 2017) and Gökçeada Island, North Aegean Sea (2.73) (Keskin, Ekici & Serdar, 2020), but higher than in the Çanakkale Strait (2.33) (Balçioğlu and Gönülal, 2017). Discrepancies in the value of b in LWRs could be affected by the fishing gear used and selectivity, and variations in environmental conditions such as ecological differences (water temperature, salinity, etc.) in the sampling areas, type of bottom and type of sediment, the intensity of predation, and lack or abundance of food (Gaspar et al., 2001; 2002; Çolakoğlu, 2020).

The meat yields of species were in the range of 16.40–23.34% for *D. trunculus*, 14.89–34.35% for *M. galloprovincialis*, 5.91–26.24% for *O. edulis*, 10.80–29.53% for *R. philippinarum*, and 12.26–18.92% *C. gallina*, respectively (Figure 3a). The highest meat yields for *D. trunculus*, *M. galloprovincialis* and *O. edulis* were determined in spring, while *C. gallina* and *R. philippinarum* had maximum yield in late summer and early autumn ($P < 0.05$). The meat yields were found to be affected significantly by seasonal and species differences ($P < 0.05$) (Figure 3a). The correlation of the meat yield with environmental variables and indexes is summarized in Figure 3b. The highest positive and negative correlations among meat yields of species and variables were observed as follows; *D. trunculus* had positive moderate correlations with salinity ($r = 0.598$) and pH ($r = 0.630$); *M. galloprovincialis* and *O. edulis* had strong–moderate positive correlations with Chl- a ($r: 0.827$, $r: 0.597$); and *C. gallina* and *R. philippinarum* had strong–moderate positive correlations with pH ($r: 0.545$, $r: 0.731$) ($P < 0.05$). The highest negative strong correlations were observed as follows; *D. trunculus* with SST ($r: -0.489$); *M. galloprovincialis* with

compactness index (C-I) ($r: 0.512$), *O. edulis* with elongation index (E-I) ($r: -0.518$), *R. philippinarum* with DO ($r: -0.356$); and *C. gallina* with DO ($r: -0.387$) ($P > 0.05$). The weakest correlations observed among species were determined as follows; *D. trunculus* with elongation index (E-I) ($r: -0.069$); *M. galloprovincialis* with density index (D-I) ($r: 0.014$); *O. edulis* with density index ($r: -0.077$); *R. philippinarum* with salinity ($r: 0.062$); and *C. gallina* with elongation index (E-I) ($r: 0.163$, $P > 0.05$).

In general, meat yield is an indicator of the condition status of bivalves which can change with seasons, reproduction period and food accessibility (Okumuş & Stirling, 1998; Orban et al., 2002). The highest meat yield of *D. trunculus*, *M. galloprovincialis*, and *O. edulis* were observed in the samples from the spring months, while it was at the highest level in the summer months for *R. philippinarum* and in the autumn months for *C. gallina*. Although the highest and lowest levels of meat yield slightly varied, the means of findings between months are similar to those reported in previous studies (Çolakoğlu & Tokaç, 2014; Vernocchi et al., 2007; Chen et al., 2020). Salinity was positively correlated with only *D. trunculus*, while pH was correlated with *D. trunculus*, *R. philippinarum* and *C. gallina* ($P < 0.05$). A significant positive correlation for the most important parameter, Chl- a , was determined only for *M. galloprovincialis* and *O. edulis* ($r: 0.545$ – 0.731 ; $P < 0.05$). The living habits of both these bivalves (*M. galloprovincialis* and *O. edulis*) differs from other species (*D. trunculus*, *R. philippinarum*, and *C. gallina*) due to the requirements of hard substrates to adhere to in water columns (Wilcox & Jeffs, 2017; Potet et al., 2021). In seawater, the planktonic mass first increases in warmer months and then sedimentation and distribution occur to benthic and lower depths of the water column throughout seasons (Graf et al., 1982; Benedetti et al., 2019). This case may be an indicator that initially, meat yield increases in parallel with the planktonic bloom in the water column for *M. galloprovincialis* and *O. edulis*, then with the precipitation of the planktonic mass in late summer, other sub-benthic clams achieve higher meat yields. This is also most probably related to active feeding on these species from the spring to autumn as the Çanakkale Strait is characterized by SST and Chl- a level increases causing increased planktonic activity (Turkoglu, 2010), which is an important factor affecting food accessibility and growth of bivalves (Robinson et al., 2010). The information about meat yields in dif-

Table 2. Morphometric indices of five economic bivalves collected from the Çanakkale Strait (Türkiye)

Species	N	Elongation index (SH/SL)			Compactness index (SW/SL)			Convexity index (SW/SH)			Density index (TW/SL)		
		Min	Max	Mean±SD	Min	Max	Mean±SD	Min	Max	Mean±SD	Min	Max	Mean±SD
Dt	3394	0.71	0.50	0.59±0.001	0.42	0.27	0.33±0.001	0.78	0.36	0.56±0.001	0.20	0.03	0.10±0.001
Mg	580	1.94	0.25	0.37±0.003	1.50	0.37	0.52±0.003	2.12	0.42	1.45±0.006	0.60	0.02	0.26±0.004
Oe	312	1.13	0.48	0.81±0.006	0.89	0.20	0.34±0.004	0.96	0.22	0.43±0.006	3.99	3.14	3.58±0.008
Rp	1869	1.03	0.79	0.90±0.001	1.03	0.50	0.60±0.001	1.32	0.36	0.69±0.004	1.05	0.14	0.46±0.004
Cg	2433	1.07	0.77	0.92±0.001	0.59	0.39	0.51±0.001	0.67	0.42	0.56±0.001	0.48	0.03	0.19±0.002

Dt: Donax trunculus, Mg: Mytilus galloprovincialis, Oe: Ostrea edulis, Rp: Ruditapes philippinarum, Cg: Chamelea gallina, N: Number, SD: Standard Deviation, SL: Shell Length, SH: Shell Height, SW: Shell Width; TW: Total weight

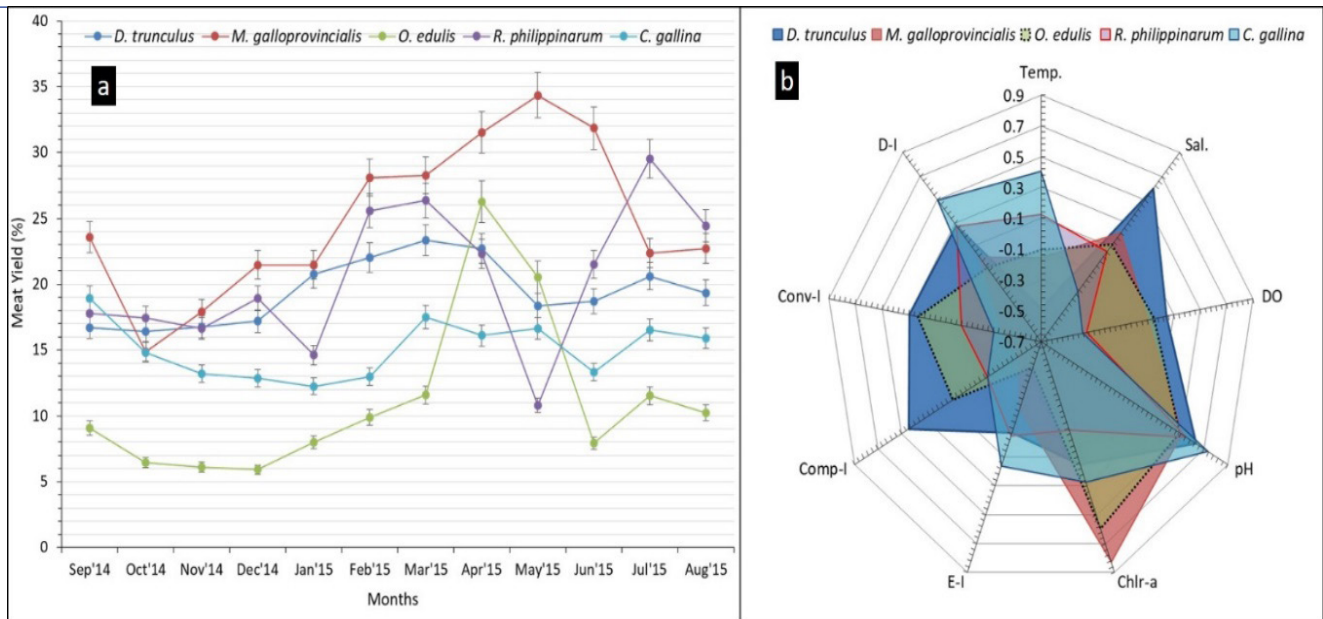


Figure 3. (a) Meat yields of bivalve species during study period (mean±SD); (b) correlations of meat yields with indexes and environmental variables.

Temp: SST, Sal: Salinity, DO: dissolved oxygen, Chl-a: Chlorophyll-a, E-I: Elongation Index (SH/SL), Comp-I: Compactness Index (SW/SL), Conv-I: Convexity Index (SW/SH), D-I: Density Index (TW/SL).

ferent bivalves could be useful for maximizing catch and controlling the exploitation of bivalves in their natural beds in different geographical areas.

CONCLUSION

This study showed that condition, growth, and morphometric indices of *D. trunculus*, *C. gallina*, *R. philippinarum*, *M. galloprovincialis* and *O. edulis* could vary based on seasonal and environmental conditions. Although the LWRs, coefficient factors and morphometric indices showed good growth for the five species, the future of the populations of these species may be negatively affected since meat yield is one of the most important reasons for fishing due to consumer demand. The growth of species varied at a high rate with changing environmental factors, especially warmer seawater and planktonic abundance (Chl-a) by season. In further studies, commercial bivalve species should be monitored by including morphometric characteristics, growth, conditions indexes, as well as meat yield and environmental parameters, sustainability, and conservation of natural stocks in terms of fisheries management.

Conflict of interest: The authors declare that they have no conflict of interests for this article.

Ethics committee approval: There is not necessity for ethical approval for this research.

Financial disclosure: This research did not receive any specific grant.

REFERENCES

Acarlı, S., Lök, A., Küçükdermenci, A., Yıldız, H. & Serdar, S. (2011). Comparative growth, survival and condition index of flat oyster,

Ostrea edulis (Linnaeus 1758) in Mersin Bay, Aegean Sea, Türkiye. *Journal of the Faculty of Veterinary Medicine, Kafkas University*, 17: 203–210.

American Public Health Association (APHA), (1995). Standard Methods for Examination of Water and Wastewater. 19th Edition, American Public Health Association, New York.

Aydın, M. & Biltekin, D. (2020). First morphometric aspects and growth parameters of the European flat oyster (*Ostrea edulis* Linnaeus, 1758) for the Black Sea, Türkiye. *Natural and Engineering Sciences*, 5: 101–109. <https://doi.org/10.28978/nesciences.756736>

Aydın, M., Tunca, E. & Ersoy, N. E. (2020). Morphometric Aspects and Growth Parameters of the Wedge Clam (*Donax trunculus*) of the Black Sea, Türkiye. *Journal of Anatolian Environmental and Animal Sciences*, 1: 11–18. <https://doi.org/10.35229/jaes.637729>

Balçioğlu, B. E. & Gönülal, O. (2017). A study on biometry of mussels (*Mytilus galloprovincialis*, Lamarck, 1819) collected from various regions of Marmara Sea. *Süleyman Demirel University Journal of Natural and Applied Sciences*, 21: 397–400. <https://doi.org/10.19113/sdufbed.56809>

Benedetti, F., Jalabert, L., Sourisseau, M., Becker, B., Cailliau C., Desnos, C., Elineau, A., Irison, J. O., Lombard, F., Picheral, M., Stemmann, L. & Pouline, P. (2019). The Seasonal and Inter-Annual Fluctuations of Plankton Abundance and Community Structure in a North Atlantic Marine Protected Area. *Frontiers in Marine Science*, 6: 214. <https://doi.org/10.3389/fmars.2019.00214>

Caill-Milly, N., Bru, N., Barranger, M., Gallon, L., & D'Amico, F. (2014). Morphological trends of four Manila clam populations (*Venerupis philippinarum*) on the French Atlantic coast: identified spatial patterns and their relationship to environmental variability. *Journal of Shellfish Research*, 33: 355–372. <https://doi.org/10.2983/035.033.0205>

Caill-milly, N., Bru, N., Mahe, K., Borie, C., & D'Amico, F. (2012). Shell shape analysis and spatial allometry patterns of Manila clam (*Ruditapes philippinarum*) in a mesotidal coastal lagoon. *Journal of Marine Biology*, 281206. <https://doi.org/10.1155/2012/281206>

Charef, A., Langar, N. Z., & Gharsallah, I. H. (2012). Stock size assessment

- and spatial distribution of bivalve species in the Gulf of Tunis. *Journal of the Marine Biological Association of the United Kingdom*, 92: 179–186. <https://doi.org/10.1017/S0025315411000403>
- Chen, L., Yu, F., Sun, S., Liu, X., Sun, Z., Cao, W., Liu, S., Li, Z., & Xue, C. (2020). Evaluation indicators of *Ruditapes philippinarum* nutritional quality. *Journal of Food Science and Technology*, 58: 2943–2951. <https://doi.org/10.1007/s13197-020-04796-6>
- Cho, S.M., Jeong, W.G. & Lee, S.J. 2008. Ecologically sustainable management of short-necked clam, *Ruditapes philippinarum*, on the coast of Yeongi at Tongyeong, Korea. *The Korean Journal of Malacology*, 24:189–197.
- Choi, Y.M., Yoon, S.C., Lee, S.I., Kim, J.B., Yang, J.H., Yoon, B.S. & Park, J.H. (2011). The study of stock assessment and management implications of the Manila clam, *Ruditapes philippinarum* in Taehwa river of Ulsan. *The Korean Journal of Malacology*, 27:107–114. <https://doi.org/10.9710/KJM.2011.27.2.107>
- Çolakoğlu, F. A., Ormanci, H. B., Berik, N., Künili, I.E. Çolakoğlu, S. (2011). Proximate and elemental composition of *Chamelea gallina* from the southern coast of the Marmara Sea (Türkiye). *Biological Trace Element Research*, 143: 983–991. <https://doi.org/10.1007/s12011-010-8943-3>
- Çolakoğlu, S. & Palaz, M. (2014). Some population parameters of *Ruditapes philippinarum* (Bivalvia, Veneridae) on the southern coast of the Marmara Sea, Türkiye. *Helgoland Marine Research*, 68: 539–548. <https://doi.org/10.1007/s10152-014-0410-7>
- Çolakoğlu, S. & Palaz, M. (2015). Population structure and dynamics of warty venus, *Venus verrucosa* (Bivalvia, Veneridae), in the North Aegean Sea, Türkiye. *Journal of Shellfish Research*, 34: 347–354. <https://doi.org/10.2983/035.034.0217>
- Çolakoğlu, S. & Tokaç, A. (2014). Properties growth of populations the striped venus (*Chamelea gallina* L., 1758) and the wedge clam (*Donax trunculus* L., 1758) in the West Marmara Sea. *Journal of FisheriesSciences.com*, 8: 27–41. <https://doi.org/10.3153/jfsc.com.2014004>
- Çolakoğlu, S. (2011). Stock assessment of striped venus (*Chamelea gallina* L., 1758) and wedge clam (*Donax trunculus* L., 1758) in the Çanakkale Strait with The West Marmara Sea. PhD thesis, Institute of Sciences, Ege University (in Turkish).
- Çolakoğlu, S. (2014). Population structure, growth and production of the wedge clam *Donax trunculus* (Bivalvia, Donacidae) in the West Marmara Sea, Türkiye. *Turkish Journal of Fisheries and Aquatic Sciences*, 14: 221–230. https://doi.org/10.4194/1303-2712-v14_1_24
- Çolakoğlu, S. (2020). Bycatch and discards from two types of bivalve dredges targeting *Donax trunculus* and *Chamelea gallina* used in the southern coast of the Marmara Sea, Türkiye. *Fisheries Science*, 86: 995–1004. <https://doi.org/10.1007/s12562-020-01473-7>
- Dalgıç, G. (2006). Determination of the spawning period and growing performance of the Black Sea Striped Venus *Chamelea gallina* (L., 1758) population (in Turkish). PhD thesis; Karadeniz Teknik Üniversitesi, Fen Bilimleri Enstitüsü, Trabzon.
- Deval, M.C. (2009). Growth and reproduction of the wedge clam (*Donax trunculus*) in the Sea of Marmara, Türkiye. *Journal of Applied Ichthyology*, 25: 551–558. <https://doi.org/10.1111/j.1439-0426.2009.01258.x>
- Gaspar, M.B., Santos, M.N., & Vasconcelos, P. (2001). Weight-length relationships of 25 bivalve species (Mollusca: Bivalvia) from the Algarve coast (southern Portugal). *Journal of the Marine Biological Association of the United Kingdom*, 81: 805–807. <https://doi.org/10.1017/S0025315401004623>
- Gaspar, M.B., Santos, M.N., Vasconcelos, P. & Monteiro, C.C. (2002). Shell morphometric relationships of the most common bivalve species (Mollusca: Bivalvia) of the Algarve coast (southern Portugal). *Hydrobiologia*, 477: 73–80. <https://doi.org/10.1023/A:1021009031717>
- Graf, G., Bengtsson, W., Diesner, U., Schulz, R. & Theede, H. (1982). Benthic Response to Sedimentation of a Spring Phytoplankton Bloom: Process and Budget. *Marine Biology*, 201–208. <https://doi.org/10.1007/BF00401286>
- Huxley, J. S. & Teissier, G. (1936). Terminology of relative growth. *Nature*, 137: 780–781. <https://doi.org/10.1038/137780b0>
- Kasapoglu, N. & Düzgüneş, E. (2013). Length-weight relationships of marine species caught by five gears from the Black Sea. *Mediterranean Marine Science*, 15: 95–100. <https://doi.org/10.12681/mms.463>
- Keskin, I., Ekici, A. & Serdar, S. (2020). Determination of the growth performance of *Mytilus galloprovincialis* in nets at Gökçeada Island. *The European Zoological Journal*, 87: 559–570. <https://doi.org/10.1080/24750263.2020.1818856>
- Künili, I.E., Çolakoğlu, S. & Çolakoğlu, F. (2021a). Levels of PAHs, PCBs, and toxic metals in *Ruditapes philippinarum* and *Donax trunculus* in Marmara Sea, Türkiye. *Journal of the Science of Food and Agriculture*, 101: 1167–1173. <https://doi.org/10.1002/jsfa.10728>
- Künili, I.E., Ertürk-Gürkan, S., Aksu, A., Turgay, E., Çakır, F., Gürkan, M. & Altınağaç, U. (2021b). Mass mortality in endangered fan mussels *Pinna nobilis* (Linnaeus 1758) caused by co-infection of *Haplosporidium pinnae* and multiple *Vibrio* infection in Çanakkale Strait, Türkiye. *Biomarkers*, 5: 450–461. <https://doi.org/10.1080/1354750X.2021.1910344>
- Lucas, A. (1981). Adaptations écopysiologiques des bivalves aux conditions de culture. *Bulletin de la Société d'Ecophysiologie*, 6: 27–35.
- Okumuş, İ., & Stirling, H. P. (1998). Seasonal variations in the meat weight, condition index and biochemical composition of mussels (*Mytilus edulis* L.) in suspended culture in two Scottish sea lochs. *Aquaculture*, 159(3-4), 249-261.
- Orban, E., Di Lena, G., Nevigato, T., Casini, I., Marzetti, A. & Caproni, R. (2002). Seasonal changes in meat content, condition index and chemical composition of mussels (*Mytilus galloprovincialis*) cultured in two different Italian sites. *Food Chemistry*, 77: 57–65. [https://doi.org/10.1016/S0308-8146\(01\)00322-3](https://doi.org/10.1016/S0308-8146(01)00322-3)
- Petetta, A., Bargione, G., Vasapollo, C., Virgili, M. & Lucchetti, A. (2019). Length-weight relationships of bivalve species in Italian razor clam *Ensis minor* (Chenu, 1843) (Mollusca: Bivalvia) fishery. *The European Zoological Journal*, 86: 363–369. <https://doi.org/10.1080/24750263.2019.1668066>
- Ponurovsky, S.K. (2008). Population structure and growth of the Japanese littleneck clam, *Ruditapes philippinarum* in Amursky Bay, Sea of Japan. *Russian Journal of Marine Biology*, 34:329–332. <https://doi.org/10.1134/S1063074008050106>
- Potet, M., Fabien, A., Chaudemanche, S., Sebaibi, N., Guillet, T., Gachelin, S., Cochet, H., Boutouil, M. & Pouvreau, S. (2021). Which concrete substrate suits you? *Ostrea edulis* larval preferences and implications for shellfish restoration in Europe. *Ecological Engineering*, 162: 106159. <https://doi.org/10.1016/j.ecoleng.2021.10615>
- Ricker, W.E. (1973). Linear regressions in fishery research. *Journal of the Fisheries Research Board of Canada*, 30: 409–434. <https://doi.org/10.1139/f73-072>
- Robinson, L.A., Greenstreet, S.P.R., Reiss, H., Callaway, R., Craeymeersch, J., DE Boois, I., Degraer, S., Ehrich, S., Fraser, H.M. Goffin, A., Kröncke, I., Jorgenson, L.L., Robertson, M.R. & Lancaster, J. (2010). Length-weight relationships of 216 North Sea benthic invertebrates and fish. *Journal of the Marine Biological Association of the United Kingdom*, 90: 95–104. <https://doi.org/10.1017/S0025315409991408>
- Rufino, M.M., Gaspar, M.B., Pereira, A.M. & Vasconcelos, P. (2006). Use of shape to distinguish *Chamelea gallina* and *Chamelea striatula* (Bivalvia: Veneridae): Linear and geometric morphometric methods. *Journal of Morphology*, 267: 1433–1440. <https://doi.org/10.1002/jmor.10489>
- Sokal, R.R. & Rohlf, F.J. (1987). Introduction to biostatistics. 2nd edition.

- New York, NY: Freeman.
- Tokeshi, M., Ota, N. & Kawai, T.A. (2000). Comparative study of morphometry in shell-bearing molluscs. *Journal of Zoology*, 251: 31–38. <https://doi.org/10.1017/S0952836900005057>
- Turkoglu, M. (2010). Temporal variations of surface phytoplankton, nutrients and chlorophyll a in the Dardanelles (Turkish Straits System): a coastal station sample in weekly time intervals. *Turkish Journal of Biology*, 34: 319–333. <https://doi.org/10.3906/biy-0810-17>
- Turkstat, (2021). *Fishery statistics* [online]. Turkish Statistical Institute. Ankara. Database of Seafood Production. <https://data.tuik.gov.tr/Bulten/Index?p=Fishery-Products-2020-37252>.
- Vasconcelos, P., Moura, P., Pereira, F., Pereira, A. & Gaspar, M. (2018). Morphometric relationships and relative growth of 20 uncommon bivalve species from the Algarve coast (southern Portugal). *Journal of the Marine Biological Association of the United Kingdom*, 98: 463–474. <https://doi.org/10.1017/S002531541600165X>
- Vernocchi, P., Maffei, M., Lanciotti, R., Suzzi, G. & Gardini, F. (2007). Characterization of Mediterranean mussels (*Mytilus galloprovincialis*) harvested in Adriatic Sea (Italy). *Food Control*, 18: 1575–1583. <https://doi.org/10.1016/j.foodcont.2006.12.009>
- Watanabe, S. & Katayama, S. (2010). Relationships among shell shape, shell growth rate, and nutritional condition in the manila clam (*Ruditapes philippinarum*) in Japan. *Journal of Shellfish Research*, 29: 353–359. <https://doi.org/10.2983/035.029.0210>
- Wijsman, J.W.M., Troost, K., Fang, J. & Roncarati, A. (2019). Goods and Services of Marine Bivalves. Global production of Marine Bivalves. Trends and Challenges. (Chapter), Springer, 598 pp.
- Wilcox, M. & Jefss, A. (2017). Is attachment substrate a prerequisite for mussels to establish on soft sediment substrate? *Journal of Experimental Marine Biology and Ecology*, 495: 83-88. <https://doi.org/10.1016/j.jembe.2017.07.004>
- Zar, J.H. (1999). *Biostatistical Analysis*. 4th edition. Prentice Hall: Englewood Cliffs, New Jersey. 929 pp.

Records of Three Immature Gelatinous Specimens for the Turkish Mediterranean Coast with an Emphasis on Alternative Pathways

Erhan Mutlu¹ , Dođukan Karaca² 

Cite this article as: Mutlu, E., & Karaca, D. (2023). Records of three immature gelatinous specimens for the Turkish Mediterranean coast with an emphasis on alternative pathways. *Aquatic Sciences and Engineering*, 39(1), 43-53. DOI: <https://doi.org/10.26650/ASE20241326004>

ABSTRACT

From samples for the phytoplankton collected from the sea surface water of 67 stations along the entire Turkish Mediterranean coast during June-July 2019, three juvenile gelatinous organisms were recorded. Two cnidarian-hydrozoan species (*Podocorynoides minima* and *Gastroblasta raffaelei*) and one ctenophore (*Bolinopsis cf. vitrea*) occurred near Mersin Bay. The specimens were determined at the juvenile or eumedusoid stages. Referring to the literature, *Gastroblasta raffaelei* was presumably about 4-5-day old (1.05 x 1.56 mm in elliptical diameter), and *Podocorynoides minima* about a stage of liberated eumedusoid (0.327 x 0.316 mm in bell diameter x height). The specimen of *Bolinopsis cf. vitrea* was measured to be 11 mm in lobate (total) length and 8.6 mm in body width. Interestingly, all species were found at different locations close to each other on the set of the water rim current speeding easterly up to about 0.5 m s⁻¹. *Gastroblasta raffaelei* and *Podocorynoides minima* were first recorded on the Turkish Mediterranean coast. Early-staged specimens of these three species were described and discussed for their diagnostic structures with their occurrence in the Turkish Mediterranean Sea after the hydrozoans were reported in the Sea of Marmara, the ctenophore in the Black Sea, and other seas of the Mediterranean basin. The present study also discussed possible and presumable pathways of recent increased Turkish Mediterranean records of specimens that have been observed in the West Mediterranean Sea and the Adriatic Sea.

Keywords: New records, gelatinous species, Turkish Mediterranean waters

ORCID IDs of the author:
E.M. 0000-0002-6825-3587;
D.K. 0000-0002-1140-2342

¹Akdeniz University, Fisheries Faculty,
Main Campus, Antalya, Turkiye

²Akdeniz University, Medical Faculty,
Main Campus, Antalya, Turkiye

Submitted:
11.07.2023

Revision Requested:
19.09.2023

Last Revision Received:
20.09.2023

Accepted:
26.10.2023

Online Published:
25.12.2023

Correspondence:
Erhan Mutlu
E-mail:
emutlu@akdeniz.edu.tr

INTRODUCTION

The Mediterranean Sea is specifically biologically well-diversified and significantly researched for its marine life (Vasilakopoulos et al., 2017). Although the Mediterranean Sea is recognized as a biodiversity hotspot (Coll et al., 2010), it has been threatened by pollution, over-exploitation, and global warming (Cuttelod et al., 2009). The eastern Mediterranean Sea, particularly the Levantine Sea is well open to the new records of the gelatinous organisms (Ctenophora and Cnidaria) besides other taxa. Most of them are invasive alien species (Galil, 2007). Many recent records (32 records since 2006, most of them since 2015) of zooplankton,

especially gelatinous organisms, most inhabiting the West Mediterranean, and Adriatic Sea, to the Turkish marine system were noticed (Table 1). Furthermore, recent records have increased for other taxa (e.g., Patania & Mutlu, 2021; Garuti & Mutlu, 2021; Mutlu et al., 2023a). For all increased records there could be a vector for their transportation, such as water current among the other vectors, the straits, aquaculture, ship ballast water, etc.

Early staged-medusoid and juvenile ctenophore specimens are often encountered in the water samples for zooplankton and even phytoplankton. Their occurrences suggested that they could be established in the seas or trans-



Table 1. Recent records and sampling year of gelatinous zooplankton in the Turkish marine coasts. * is not a new record but is, at one location, extremely abundant specimens and ** is open water species but abundantly found at locations in Figure 1, which is found in the West Mediterranean Sea and recorded in the area symbolized in Figure 1.

Taxa	Regions	Year	Citations
Published specimen			
One ctenophore	Bosporus exit to Black Sea	2007	Öztürk, Mihneva, & Shiganova, 2011
Sixteen hydrozoans, One scyphozoan	Sea of Marmara, Aegean Sea	2006-2013	İşinibilir et al., 2015a; İsinibilir, Yılmaz, & Demirel, 2015b; İşinibilir, Ulucam, & Yüksel, 2019
One hydrozoan	Sea of Marmara	2015	Yılmaz et al., 2017
Two scyphozoans, One hydrozoan, One thaliacean	Sea of Marmara	2019-2020	İşinibilir et al., 2022
Two hydrozoans	Aegean Sea	2012, 2015-2016	Gülşahin, Tarkan, & Bilge, 2013; Gülşahin et al., 2016
One ctenophore	Aegean Sea	2015-2018	Killi, Abyzova, & Shiganova, 2019
One hydrozoan	Northernmost Aegean Sea	2021	İsinibilir, Yüksel, & Guresen, 2021
One lobat ctenophore	Turkish water	2015	Gülşahin and Türker, 2017
One new scyphozoan	Levant Sea	2018	Mutlu et al., 2020
One lobat ctenophore	Levant Sea	2020	Gokoglu and Galil, 2020
One cydippid ctenophore	Levant Sea	2019	Mutlu and Özvarol, 2022a
One hydrozoan	Levant Sea	2019	Mutlu and Ozvarol, 2022b
Unpublished specimens noticed			
One calanoid copepod (24 inds)	Levant Sea	2019	Duman, 2023
Four phytoplankton*	Levant Sea	2019	Karaca, 2023
One cheatognath**	Levant Sea	2019	Duman, 2023

ferred from one sea to another sea depending on their life longevity. In many cases, the literature is needed to identify immature gelatinous species as needed for the other taxa. To overcome such difficulties in the identification, laboratory studies were conducted to follow growth through different stages of the gelatinous species. This stage of growth could be differentiated by their diagnostic morphometries under different environmental conditions (Mayer, 1912; Oliveira et al., 2007; Gravili et al., 2007; Shiganova, 2020; Fabien Lombard pers. comm.).

Mediterranean plankton is easily drifted and moved from intra-seas of the Mediterranean Sea and inter-seas by water current induced by a variety of mechanisms of atmosphere-sea interactions over the Mediterranean Sea besides a variety of introduction vectors such as Suez Canal, ship ballast water, and aquaculture (Zenetos et al., 2012). Such events could change the temporal structure of ecosystems, fisheries, and surface water currents. Globally, the Mediterranean Sea is influenced by atmospheric actions, mainly North Atlantic Oscillation (NAO) (Raitso et al., 2011) and slightly North Pacific Oscillation (NPO), Pacific Decadal Oscillation (PDO) and El Niño (Báez et al., 2022). Locally, the Mediterranean Sea is influenced by the Bimodal Oscillation System (BiOS) decadal-occurring around the Adriatic Sea and global warming (Poulain et al., 2013; Civitarese et al., 2023). The study area of the present study is one of the regions mostly affected by global warming and undergoes a process of tropicalization (Encarnação et al., 2019). All these events induced new aspects of the water current in the Mediterranean system and introduced new records of plankton among the Mediterranean

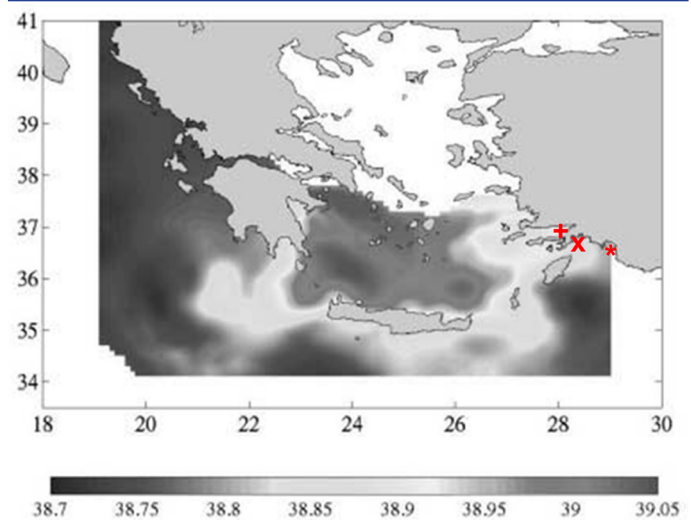


Figure 1. Water salinity at a deep layer having $\sigma_t=29.18$ (LIW) to show depth-wise changes of the layer along the different seas' interaction in 1991 (from Briand, Lascaratos, & Klein, 2000). + denotes records by Gülşahin, Tarkan, & Bilge, 2013; Gülşahin et al., 2016, x denotes records by Duman, 2023, Karaca 2023, and * denotes records by Mutlu and Özvarol, 2022a, b (see Table 1 for taxa observation and sampling dates).

seas, and ocean to the Mediterranean Sea. New records of the organisms particularly in the Turkish marine waters increased as the research was performed, or presumable new pathways could be developed in the Mediterranean Sea. Such introduction signaled the impulse of new records in the Turkish waters (Table 1).

Besides the endemic gelatinous species including medusoids of the hydrozoans of the Mediterranean Sea, the gelatinous alien species have altered the Mediterranean ecosystem (Coll et al., 2010; Dragičević et al., 2019); for instance, a new sea medusa, *Chrysaora pseudoocellata* Mutlu, Tulay, Olguner & Yılmaz 2020 was established in the eastern Mediterranean Sea (Dragičević et al., 2019; Mutlu et al., 2020; Douek et al., 2020). The records particularly the recent records coincided with decadal atmospheric and oceanic events, especially with a decadal period of BiOS which was first noticed in 1988. Furthermore, Duman (2023) and Karaca (2023) determined a new record of a calanoid copepod (specific to the Adriatic Sea) at one location and extreme-abundantly (more than at least 100-fold-higher at one location than other sampling stations of the present study) occurred phytoplankton species (three of four common in Adriatic Sea) at one location, respectively (Table 1, Figure 1). A delayed record could occur in the northern Turkish waters referring to southern Turkish waters. All species recorded in Table 1 originated from Atlantic-Mediterranean waters. Most of them were found in the western waters of the Levantine Sea. Depth-wise rise of a deep layer characterized by a water density of $\sigma_t=29.18$ (LIW) was observed along the Ionian Sea through the Aegean Sea to a part of the Levant Sea in 1991 (Briand et al., 2000). This event however coincided with the new records of some species for the Turkish waters (Table 1, Figure 1). Decadal events and this example (Figure 1) alert transportation of westerly originated species to the western coasts of Turkish waters beside the BiOS effect. Nevertheless, the BiOS induced acceleration of water velocity in the present study area (Figure 2).

A model that was recently conducted to simulate jellyfish dispersion in space showed a pathway of jellyfish distributional extension in time from the Egyptian (around the Suez Canal) and Israeli coast to the Turkish water on the set of the Atlantic water current (Edelist et al., 2022). Alternatively, BiOS is a water circulation process switching from cyclonic to anticyclonic or vice versa on decadal intervals via the North Ionian Gyre (NIG) (Civitarese et

al., 2010) and leads to changes in the thermohaline current structure in the Southern Adriatic (Civitarese et al., 2010), followed effect on Surface Water (LSW) and Intermediate water (LIW) in the Levant Sea (Ozer et al., 2017) via Ionian Jet (Figure 2). The Ionian jet then pumped the waters to the Levantine Sea (Poulain et al., 2013) when the NIG became cyclonic circulation. Studies have started to understand the effect of the NIG on the plankton communities, for instance on the phytoplankton in the Mediterranean Sea (Jasprica et al., 2022), and spatiotemporal benthic-pelagic coupling changes (Ricci et al., 2022) as the severity of the winter condition has changed globally yielding BiOS (Poulain et al., 2013).

Gelatinous biodiversity has increased in the Turkish marine environment. Öztürk, Mihneva, & Shiganova, (2011) contributed occurrence of one ctenophore, İşinibilir et al., 2015a, İsinibilir, Yılmaz, & Demirel, 2015b and İşinibilir, Ulucam, & Yüksel (2019) 16 hydrozoans and one scyphozoan, Yılmaz et al. (2017) one hydrozoan, Gülşahin, Tarkan, & Bilge (2013) and Gülşahin et al. (2016) two hydrozoans, and Gokoglu & Galil (2020) one ctenophore to the gelatinous biodiversity of the Turkish marine waters, followed by species records reported by Mutlu & Özvarol (2022a, b) and the present study (Table 1).

Therefore, the present study aims to mark the evidence of the occurrence of three gelatinous invertebrates in jelly form from a different region characterized with comprehensive environmental variables rather than other regions by the published studies and to discuss possible introductory pathways of western species-level intrusion undergoing the Mediterranean marine basin, particularly the Levantine Sea, and the Turkish coast.

MATERIAL AND METHODS

During a summer survey (June-July 2019) of acoustical studies on the vegetation along the entire Turkish Mediterranean coast (Taşucu Bay, the Mediterranean region, is the easternmost end – Datça Bay, the Aegean region is the westernmost end of the study area), water samples for a study of phytoplankton were collected from the surface waters of 67 stations (Figure 3). The locations of the sampling stations and the study area were described in detail in a study published by Mutlu et al. (2023b). The bottom depth of the stations varied between 5 m and 1000 m. One-third of the stations were in offshore waters and the rest were in coastal waters. Three major rivers (Goksu, Seyhan, and Ceyhan) flow into the eastern part, the coastal zone of which is eutrophic. On the contrary, Antalya and Muğla Bays are oligotrophic due to the absence of rivers compared to Mersin Bay. Sea surface temperature was at maxima (28 C in the Aegean part - 31 °C in the Mediterranean part), and salinity varied between 38 PSU and ~40 PSU in summer, respectively. Three of the stations were included in the present study because the species considered in the study were recorded only in the three stations (Table 1, Figure 1).

On board R/V "Akdeniz Su", 100 ml of the water sample was taken from the sea surface at each station using a 5-l Nansen bottle. The water was then fixed in a dark bottle using a 1% glutaraldehyde solution. During the survey, samples for physicochemical and optical parameters were collected from the surface (prefix S)

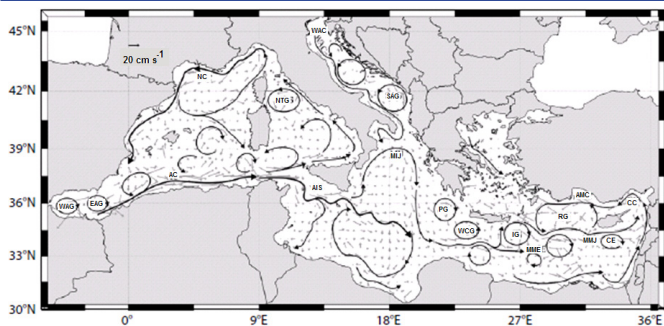


Figure 2. Mediterranean Sea surface water circulations affected by BiOS (Poulain et al., 2013).

and near-bottom (prefix N) waters some of them were measured on board, and the rest were frozen for lab analyses. One board and one liter of the water were filtered through CF/C for the nutrients and total suspended matter, and another one liter through CF/F filters for chl a. The samples were then frozen at -20 °C on board. These parameters were physical (T; temperature in °C, pH; S; salinity in PSU) using a multi-parameter probe (YSI, Hi-Tech), optical (Secchi disk depth), and chemical parameters (nutrients; NO₂+NO₃, NH₄, and PO₄, SiO₂, chl a and TSM; total suspended matter).

Three immature and early-staged gelatinous species were observed during the lab microscopic study. Each specimen was recorded at the different stations of the three stations (Figure 3). These three stations were sampled on July 02, 2019. All species were found at their early stages. Diagnostic terms of hydrozoan followed the description made by Bouillon et al. (2006) The staged species were identified using the descriptions ascribed by Mayer (1910, 1912), Madin (1991), Oliveira et al. (2007). Gravili et al. (2007) determined the stage development of *Gastroblasta raffaelei* Lang, 1886, regarding the different lab conditions with water temperature, and Schuchert (2007) of *Podocorynoides minima* (Trinci, 1903) and species of the ctenophores (Chun, 1880; Mayer, 1912; Oliveira et al., 2007; Shiganova, 2020). Photos of the specimens were taken, and their size was measured under the microscope; elliptical diameters of *G. raffaelei* and maximum bell width and height of *P. minima*.

The nutrients were measured following the standard UV-spectrophotometric procedures described by APHA (1999). The values of the nutrients were then converted to a unit µM. Total suspended solids (the material was dried in an oven at 60 °C for 24 h, and then weighed before the weight of the dried membrane was subtracted from the total dry weight), and chlorophyll a (chl-a) were determined following Lorenzen's method (1967).

RESULTS AND DISCUSSION

Three gelatinous organisms were determined along the Turkish Mediterranean coast. These are *G. raffaelei* found at 22E, *P. minima* at 25E (hydrozoans), and *Bolinopsis cf. vitrea* at 23E (the ctenophore) (Figure 3).

Environment of the study area

The 22E, 23E, and 25E were the coastal stations within inshore waters (Figure 3). The coastal stations had a bottom depth of 49 m, 48 m, and 52 m, respectively (Table 2). The Secchi disk depth at all stations varied between 13 m and 15 m. The sea surface temperature was measured in a similar range of 26-27 °C. The surface and near-bottom water salinity varied between 36.4 and 37.6 (PSU) (Table 2).

The concentration of the sea surface chl-a was 2-fold lower at two coastal stations (23E and 25E) than that of another station (22E). However, the total suspended matter was measured in a similar range at all the stations (Table 3). Sea surface SiO₂ was minima at 22E and maxima at 23E, like NH₄, and contrasted to NO₂+NO₃. Sea surface water PO₄ varied between 1.41 µM at 23E and 3.39 µM at 25E (Table 2).

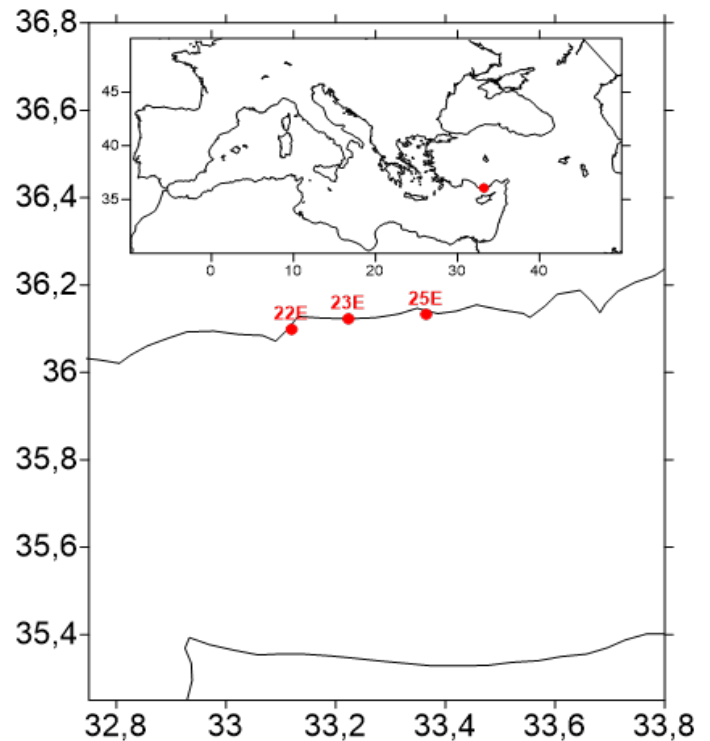


Figure 3. The study area (rectangle in color cyan) of the summer survey, and the area where the identified gelatinous specimens were found (red dot) at the sampling stations of the specimens (red dots labeled with station codes).

The stations were located between Turkey and the northern coast of Cyprus, which caused the derivation of the jet of the rim current. The current velocity was faster than 20 cm s⁻¹ which is an average value for the Mediterranean Sea, and a maximum velocity of >50 cm s⁻¹ was also measured along the Turkish Mediterranean coast (Poulain et al., 2013). These gelatinous planktons could be converged to the location by the rim current circumstance (Figures 2, 3).

Gelatinous species

Two species belonging to the phylum Cnidaria and one species to phylum Ctenophora were recorded for the first time along the Turkish Mediterranean coast. All three species were staged at a level of juvenile.

Species *Gastroblasta raffaelei* Lang, 1886

Phylum: Cnidaria
Class: Hydrozoa
Order: Leptothecata
Family: Campanulariidae
Genus: *Gastroblasta*

The material examined was an immature medusoid specimen collected from Tekeli (36° 05' 91" N, 33° 07' 18" E, station 22E, Mersin coast, Turkey, Eastern Mediterranean Sea) on 02 July 2019. Collection of the material was taken using a Nansen bottle from surface water at a seafloor depth of 49 m by Doğukan Kara-

Table 2. Distribution of the environmental parameters; bottom depth (depth), Secchi depth (SDD), surface (prefix S) and near-bottom (prefix N) water temperature (T), salinity (S), pH (pH), chl-a, total suspended matter (TSM), and the nutrients at stations of the present study. nm; not measured.

	22E	23E	25E
Depth (m)	49	48	52
SDD (m)	13	15	12.5
ST (°C)	26	27.1	26.5
NT (°C)	nm	nm	22.5
SS (PSU)	37.4	37.4	37.6
NS (PSU)	nm	nm	36.4
SpH	9.2	9.1	9.34
NpH	nm	nm	9.35
SChl-a (µg/l)	0.473	0.215	0.216
STSM (mg/l)	0.045	0.048	0.039
NTSM (mg/l)	nm	nm	0.054
SSiO ₂ (µM)	8.83	10.51	9.39
NSiO ₂ (µM)	12.18	nm	27.85
SNO ₂ +NO ₃ (µM)	0.30	0.51	0.47
NNO ₂ +NO ₃ (µM)	0.20	nm	0.28
SNH ₄ (µM)	43.91	67.24	64.65
NNH ₄ (µM)	422.36	nm	90.57
SPO ₄ (µM)	2.07	1.41	3.39
NPO ₄ (µM)	3.72	nm	3.72

ca. The specimen was measured to be 1.05 x 1.56 mm in the elliptical umbrella diameter.

Description: Umbrella (u) is much flatter, and elliptical and has a shorter axis of 1.05 mm and a longer axis of 1.56 mm (Figure 4A). There are eight tentacles (t) of which half of the completed tentacle is curly in length and the other half is straight with each originating from an incomplete pear-shaped tentacular bulb (TB) (Figure 4A, B). Manubria (m) were multiple urn-shaped and the central manubrium (m) was fully formed (Fig. 4) and the other two manubria were smaller (one just formed with two lips) than the central one. Developed two manubria had four lips (l) which were more pronounced in the central manubrium, developing manubrium has two lips (Figure 4). Two opposite weak radial canals (rc) were hardly observed crossing diagonally three manubria and a centripetal (circular) canal (cc) was developed. The velar opening (vo, velum) is rather wide (Figure 4).

Remarks: *Gastroblasta timida* resembles *G. raffaelei*, both having multiple manubria and centripetal radial canals, and differentiated in bell shape; *G. timida* in circular, and *G. raffaelei* in elliptical umbrella (Gravili et al., 2007). The specimen was about 4-5 days old medusa according to the description diagnosed by Gravili et al. (2007). *G. raffaelei* had different diagnostics under different temperature conditions (15 °C and 18 °C). Morphometrically, our specimen resembles a specimen of 4-5 days old *G. raffaelei* reared at 18 °C by Gravili et al. (2007), but it is like stage reared at 15 °C considering the size of the specimen.

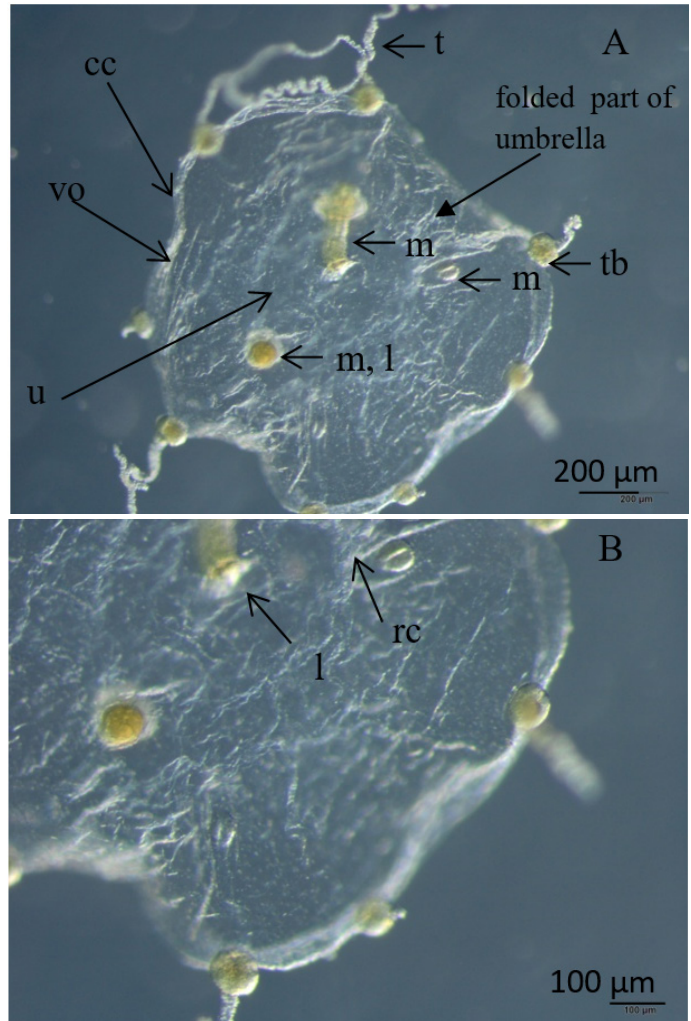


Figure 4. Oral view of *Gastroblasta raffaelei*: entire specimen (A), central manubrium, and newly occurred manubria at either side of the central manubrium (B). Umbrella (u), tentacles (t), tentacular bulb (tb), manubria (m), central manubrium (m), lips (l), radial canals (rc), centripetal (circular) canal (cc), and velar opening (vo, velum).

Species *Podocorynoides minima* (Trinci, 1903)

Phylum: Cnidaria

Class: Hydrozoa

Order: Anthoathecata

Family: Rathkeidae

Genus: *Podocorynoides*

The material examined specimen was just released medusoid after the spawning and collected from surface water at a seafloor depth of 52 m off Akkuyu, Aydıncık (36° 07' 98" N, 33° 21' 90" E, station 25E, eastern Mediterranean Sea, Mersin coast, Turkey) on 02 July 2019, using a Nansen bottle by Doğukan Karaca. The size of the specimen was 0.327 x 0.316 mm in bell diameter x height.

Description: The body of the medusoid specimen is characterized by an incomplete sphere bell and untapered global pole of

Table 3. Previous occurrence, sampling year and location of three species in the Mediterranean Sea and adjacent seas.

Species	Region	Year	Location	Citation
Gastroblasta raffaelei				
	Tyrrhenian sea	1886	Gulf of Naples	Lang, 1886
	Ligurian Sea	1980	Portofino and Pon-tetto	Boero, 1980; Boero & Fresi 1986
	Balearic Sea	2003	Catalonian waters	Guerrero et al., 2018
	Ionian Sea, Adriatic Sea	2005	Otranto	Gravili et al., 2007
	Adriatic Sea		Northern coast	REGIONE DEL VENETO, SHAPE, 2013
	Marmara Sea	2008	İzmit Bay	Isinibilir, Yilmaz, & Demirel, 2015b
Podocorynoides minima				
	Ligurian Sea	1963-1964	Riviera-Corsica	Goy, 1972
	Red Sea	?		Schmidt, 1973
	Eastern Mediterranean		Lebanon water	Goy, Lakkis, & Zeidane, 1991
	Tyrrhenian Sea		Naples	Brinckmann-Voss, 1987
	Western Mediterranean	2004-2005	Tunisia	Touzri et al., 2012
	Marmara Sea	2006	Istanbul Bay	Isinibilir, Yilmaz, & Demirel, 2015b
	Adriatic Sea		Northern coast	REGIONE DEL VENETO, SHAPE, 2013
	Western Mediterranean	2014	Algeria	Kherchouche & Hafferssas, 2019
	Black Sea	2020	Romania	Mureşan, Teacă, & Begun, 2021
Bolinopsis vitrea				
	Aegean Sea	?	Turkey	Ergen, 1967
	Adriatic Sea	2003-2006	Italy	Shiganova & Malej, 2009.
	Black Sea	2007	Turkey	Öztürk, Mihneva, & Shiganova, 2011
	Adriatic Sea	2009	Montenegro	Lucic et al., 2012
	Adriatic Sea	2009-2010	Montenegro -Croatia	Branka et al., 2014
	Black Sea	2010	Bulgaria	Öztürk, Mihneva, & Shiganova, 2011; Öztürk, 2021
	Adriatic Sea		Northern coast	REGIONE DEL VENETO, SHAPE, 2013
	Levant Sea	?	Turkey	Çinar et al., 2014
	Adriatic Sea	2010-2019	Adriatic eastern coasts	Pestic et al., 2021

the oral side and is shaped in a dome having thin global jelly and thicker apical jelly (Figure 5A). The subumbrella has a long gastric peduncle, cylindrical manubrium (m), elongated perradial lip (l) margins, and gonads (g) surrounding the manubrium (Figure 5A, B). The oral part has four radial canals (rc) extending to the apical pole, a narrow ring canal (rrc), four perradial tentacle bulbs (tb) pad-like, and tentacles (t) are very contractile (Figure 5A). Tentacles are first perpendicularly positioned (pp) to the umbrella or velum and then extended outward (Figure 5A).

Remarks: *P. minima* is distinguished by having a distinct, round trunk, and oral tentacles. The species resembles members of the family Rathkeidae as compared to that of the Hydractiniidae (Schuchert 2007). The initial parts of oral tentacles are positioned perpendicular to the oral velum (Figure 5A) unlike the oblique tentacles of *Lizzia blondina* Forbes, 1848 (Schuchert 2007).

Species *Bolinopsis cf. vitrea*

Phylum: Ctenophora

Class: Tentaculata

Order: Lobata

The material examined was a specimen at the stage of immature and collected from surface water at a seafloor depth of 48 m off Aydıncık, (36° 07' 37" N, 33° 13' 39" E, station 23E, Mersin coast, Turkey, eastern Mediterranean Sea) on 02 July 2019 using a Nansen bottle by Doğukan Karaca. The size of the specimen was 11.1 x 8.5 mm in the total (lobate) length x maximal body width. The specimen had also an aboral length of 9.6 mm, and an auricle length of 8.6 mm (Figure 6).

Description: Body shape from the tentacular plane is semi-ellipsoid oval, and laterally compressed. It has a blunt-aboral apex and wide oral lobes terminated between the apical location and

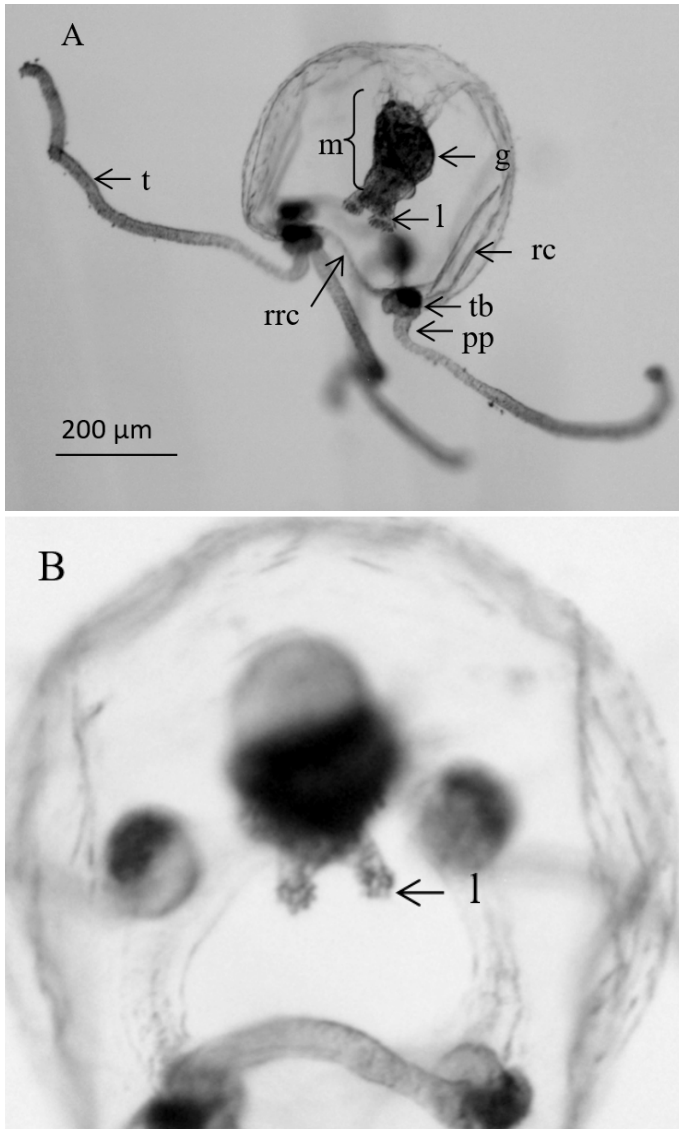


Figure 5. Liberated medusoid specimen of *Podocorynoides minima*; lateral view (A) and oral view focusing on the mouth lips (B). Manubrium (m), perradial lip (l), gonads (g), radial canals (RC), ring canal (rrc), tentacle bulbs (tb), tentacles (t), and tentacle in perpendicular position (pp) to velum.

mouth. Two poles of the specimen are about equal bluntness and lateral parts around the apical pole are characterized by multi-corners on the lateral of the body. The body surface has small granulation. Auricle (a) is developed and edged by ciliary combs (cc) and extends to just above the mouth. Tentacles (t) are very short and simple, initiated from tentacle bulbs (tb) located on each side of the mouth by extending to the bases of the auricles. Stomodaeum (s) is simple, long, and very narrow, has fine caliber chymiferous tubes, very simple meridional ventral tubes joined with the circumoral vessel, and two contrasted directional canals in "V" shape on each of two poles by joining to stomodaeum (Figure 6). Lobe (l) is initiated (li) in the middle part between mouth (m) and infundibulum (i)

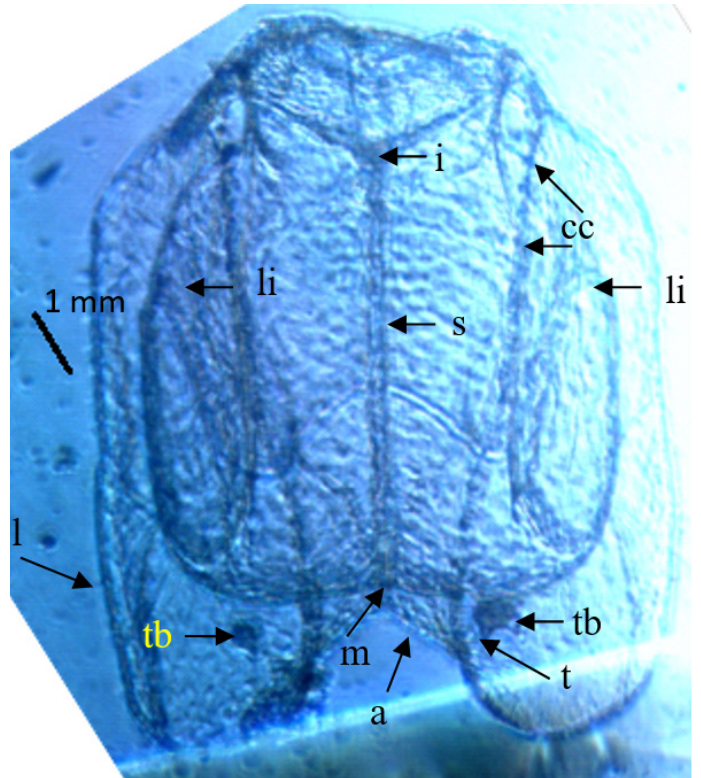


Figure 6. *Bolinopsis cf. vitrea*; view of the tentacular plane of its juvenile specimen. Auricle (a), ciliary combs (cc), tentacles (t), tentacle bulbs (tb), stomodaeum (s), lobe (l), initial region of the lobe (li), mouth (m), and infundibulum (i)

Remarks: According to descriptions performed by Mayer (1912), Oliveira et al. (2007), and Oliveira & Migotto (2006) for the adult specimen of *Bolinopsis vitrea* (L. Agassiz, 1860), all characters of a very simple and narrow stomodaeum, position of large tentacle bulbs, tip position of just developed auricle and oral lobe and mouth shape with reverse "V" (Fig. 4) support that the specimen seems to be *B. vitrea*. Immature and mature *Mnemiopsis* specimens are distinguished as having auricle furrows (Oliveira et al., 2007), small tentacle bulbs positioned at the middle of the body and not acute and blunt aboral apex (Shiganova, 2020), and the specimen of the present study lacks the auricle furrows but having blunt aboral apex. Specimens of genus *Leucothea* pass to the stage of genus *Bolinopsis* which is closely related to *Leucothea* covered with small papillae on the whole outer surface (Mayer, 1912) as occurred in the present specimen with the very small pits-like granulates on the outer surface of the body. However, the present specimen lacks 2 remarkable blindly-ending sacs and the tentacle bulbs nearly to the level of the funnel. The tentacle bulbs are positioned at a level of mouth in the present specimen as differentiated by Oliveira et al. (2007). It is hereby remarked that the species could be *Bolinopsis vitrea* but was recognized as *Bolinopsis cf. vitrea* due to the following remarks: "I once got a juvenile like this and grown it in the lab (thinking it was either *Bolinopsis* or *Mnemiopsis*)... to turn out it was *Leucothea multicornis* once grown" (Fabien Lombard, pers. comm.) and "It is difficult with the small ones as the lobe morphology about sta-

toicyt change. What I can say for sure is that it does look like neither *Mnemiopsis* nor *Bolinopsis infundibulum*" (Cornelia Jaspers, pers. comm.).

Early stages of gelatinous specimens are very passive swimmers at movement and could be much more drifted by water currents than the adult specimens. Eumedusoid hydrozoan had a short stage passage to become adult (Gravili et al., 2007). This occurrence of such newcomers to a sea suggested that the specimen could be established in the region where they are found and could be passively moved from one sea to another sea via the water current depending on its velocity or via ship ballast water. This could induce a broad-scale distribution of the organisms in the Mediterranean Sea (Table 3).

Body morphometry and structure of gelatinous organisms were differentiated by the environmental parameters under lab conditions or from sea to sea worldwide (Gravili et al., 2007; Shiganova, 2020). Such differences led to the importance of local morphometry of the organisms reported to widen their descriptive knowledge. There are some studies published on such structural differences; Gravili et al. (2007) reared *G. raffaelei* under lab conditions in two different seasons and temperature conditions; 15 and 18 °C. The size of specimens at the same age is larger at 18 °C than at 15 °C. Our specimen fits with both sizes (4-5 days old) of specimen reared at 15 °C and the diagnostic structure of specimen reared at 18 °C rather lower than our sampling temperature of the water with higher chl-a concentration coincided with higher ammonium concentration than the other two stations (Table 2). Isinibilir, Yilmaz, & Demirel, 2015b showed a similar diagnostic structure of *G. raffaelei* in the Sea of Marmara, which is colder than the present study area, to the structure of specimen reared at 15 C (Gravili et al., 2007). Nutritional conditions as well as temperature could induce such differences between different Turkish and Italian waters. Near-bottom temperature was measured at around 22 C (Table 2). Similar structural differences were observed for a ctenophore species, *M. leidy* in the European seas (Shiganova, 2020).

Gastroblasta raffaelei which endemic species to the Mediterranean Sea overspread the Mediterranean Sea, particularly the Adriatic, Tyrrhenian and Ligurian Seas (Bouillon et al., 2004; Gravili et al., 2007) and was first reported occurring in the Sea of Marmara for the Turkish waters (Isinibilir, Yilmaz, & Demirel, 2015b). Up to now, the species was however not reported for the Turkish Mediterranean coast (Çinar et al., 2014). *G. raffaelei* was recently recorded in the Catalan Sea (Guerrero et al., 2018) (Table 3). *Podocorynoides minima* has been distributed globally in the temperate waters of the world; Mediterranean, Atlantic Ocean, Indo-Pacific Ocean (Schuchert, 2007), Sea of Marmara (Isinibilir, Yilmaz, & Demirel, 2015b) and Black Sea (Table 3).

Regarding ctenophoran lobate distribution only in the Turkish waters, *M. leidy* A. Agassiz, 1865 which was introduced first to the Russian waters of the Black Sea in 1987 (Vinogradov et al., 1989) was reported in the Turkish Black Sea in 1991 (Mutlu et al., 1994; Mutlu, 1999) and in the Mediterranean Sea, Mersin Bay in 1992 (Uysal & Mutlu, 1993; Kideys & Niermann 1993). *Bolinopsis vitrea* which is a cosmopolitan species in the world was reported in the

Black Sea (Öztürk, Mihneva, & Shiganova, 2011), and *Leucothea multicornis* (Quoy & Gaimard, 1824), which is a cosmopolitan species in the world, in the Turkish Mediterranean Sea, Antalya Bay from a sample taken in 2020 (Gokoğlu & Galil, 2020). However, *B. vitrea* was generally observed in the eastern Mediterranean Sea and adjacent seas (Table 3). Recently, the occurrence of *Beroe mitrata* (Moser, 1907) was reported in the Turkish waters. (Killi, Ayzova, & Shiganova, 2019). However, Çinar et al. (2014) previously reported the species, *B. vitrea* from the Turkish Levant water. Nevertheless, *L. multicornis* was reported for the first observation in the eastern Mediterranean (Galil et al., 2014), followed by occurrences in the Syrian waters (Mamish, Durgham, & Ikhtiyar, 2019), and Greek waters (Digenis & Gerovasileiou, 2020).

Most of all three species records were performed from the western Mediterranean Sea and Adriatic Sea (Table 3). Regarding their Mediterranean-wide occurrence, a common location is Adriatic Sea with occurrence in our one sampling conducted in the summer of 2019. Furthermore, increased records have occurred in the Turkish marine waters (Table 1). This brings about speculation about the coincidence of the decadal formation of BiOS (Gacic et al., 2010) with three species found in the present study and other records (Table 1). The BiOS has the power to create the high-level variability of the Mediterranean Sea and the strong interconnection of its sub-basins by affecting the vectors of the saltier LIW, the LSW, and the fresher Atlantic Water (Civitaresse et al., 2023). The BiOS which changed the cycle pattern of the Northern Ionian Gyre affected the Levant hydrographs in decadal periods. The Mid-Ionian Jet pumped the water toward the Turkish Mediterranean waters via two vectors: one directly to Rhodes gyre (RG) and the other to easternmost Levant water by the Atlantic current (CC) (Figure 2). The water of the study area had fewer saline waters and relatively high primary production (Table 2) as compared to the expected value (39 PSU) of the summer salinity in the Levant Sea (Lascaratos, Williams, & Tragou, 1993; Poulos, Drakopoulos, & Collins, 1997).

In comparison to the western vector introducing specimens to the Turkish Levant waters, Civitaresse et al. (2023) monitored the decadal formation of the NIG circulation from 1996 to 2018. Regarding the records given in Table 1 and the present study, all physical data suggested an alternative way of water transportation by the BiOS or LIW derived by BiOS toward the Turkish waters (Figures 1, 2). Compared to regular summer measures existence of less saline and warmer water indicated that there was physical water transportation from somewhere to the present study area during the summer of 2019 as inferred from decadal surface water currents (Table 2, Figure 3).

In comparison to the eastern (Red Sea) vector introducing specimens to the Turkish Levant waters, Edelist et al. (2022) conducted a particle dispersal model to simulate jellyfish dispersion in space and showed a pathway of jellyfish distributional extension in time starting from the Egyptian (around Suez Canal) and Israeli coasts to the Turkish water on the set of the Atlantic water current path during 2017-2018 based on the sea surface current. Consequently, the particle arrived at İskenderun Bay which was far away from the record locations of the present study if the particles were released from the Egyptian coast. Subsequently, the particles arrived at An-

talya Gulf which was on the set of record locations of the present study but far away from the record locations given in Figure 1 and locations of unpublished records given in Table 1 if the particles were released from the Israeli coast (Edelist et al., 2022).

Recently, Mutlu & Özvarol (2022a, b) and Uttieri et al. (2023) discussed possible introductory pathways of the zooplankton to the eastern Mediterranean Sea regarding the effect of the BiOS; three species (*Hormiphora plumosa* M. Sars, 1859, *Oceania armata* Kölliker, 1853 and *Pseudodiptomus marinus*, Sato, 1913, respectively) are also observed in the Adriatic Sea and adjacent seas. Jasprica et al. (2022) related the BiOS caused by severe winter conditions to phytoplankton composition and distribution in the southern Adriatic Sea.

CONCLUSION

Early-staged specimens of hydrozoans and ctenophores could be morphometrically different under different environmental conditions and sea-to-sea. Description of these three immature gelatinous organisms was characterized for the Turkish Levant coast. However, records of the gelatinous organisms have increased in the Turkish waters. Discussing the possible vectors of their introduction to the Turkish coasts, water pumped by BiOS is bifurcated at Rhodes gyre. One branch was entrapped in Rhodes gyre, and another branch was joined to the Atlantic current, both affecting the Turkish Mediterranean coast (Figure 2) (Poulain et al., 2013). Therefore, it is possible that the species could have entered the eastern Mediterranean by the Atlantic and met BiOS through the western Mediterranean.

Acknowledgments: The present study was conducted in a work of the project (grant no: 117Y133) funded by The Scientific and Technological Research Council of Turkey (TUBITAK). We thank Peter Schuchert (Muséum d'histoire Naturelle, Switzerland) for confirming the identification of *Podocorynoides minima*, Lombard Fabien (Sorbonne Université, France) and Cornelia Jaspers (DTU Aqua, Denmark) for their remarks on juvenile ctenophore. We also thank crew of R/V "Akdeniz Su".

Compliance with Ethical Standard: The authors declare that all applicable guidelines for sampling, care, and experimental use of animals in the study have been followed.

Conflict of interest: The authors have no conflicts of interest to declare that are relevant to the content of this article.

Ethical approval: The authors declare that all applicable guidelines for sampling, care, and experimental use of animals in the study have been followed.

Financial disclosure: The present study was funded by The Scientific and Technological Research Council of Turkey (TUBITAK). Project no: 117Y133.

REFERENCES

APHA. (1999). *Standard Methods for the Examination of Water and Wastewater*. American Public Health Association, Washington DC.

Báez, J. C., Pennino, M. G., Czerwinski, I. A., Coll, M., Bellido, J. M., Sánchez-Laulhé, J. M., García, A., Giraldez, A., García-Soto, C. (2022).

Long term oscillations of Mediterranean sardine and anchovy explained by the combined effect of multiple regional and global climatic indices. *Regional Studies in Marine Science*, 56, 102709. <https://doi.org/10.1016/j.rsma.2022.102709>.

Boero, F. & Fresi, E. (1986). Zonation and evolution of a rocky bottom hydroid community. *P. S. Z. N. Marine Ecology*, 7, 123–150.

Boero, F. (1980). Life cycles of hydroids and hydromedusae: Some cases of difficult interpretation. *Memorie di Biologia Marina e di Oceanografia*, 10, 141–147.

Bouillon, J., Medel, M. D., Pagès, F., Gili, J. M., Boero, F., Gravili, C. (2004). Fauna of the Mediterranean Hydrozoa. *Scientia Marina*, 68, 1–454.

Bouillon, J., Gravili, C., Pagès, F., Gili, J.-M., Boero, F. (2006). *An introduction to Hydrozoa*. Publications Scientifiques du Muséum, Paris, p. 591

Branka, P., Marijana, M., Dragana, D., Davor, L. (2014). Chaetognaths in Boka Kotorska Bay. *Studia Marina*, 27(1), 109–130. doi:UDC559.135(262.3)

Briand F., Lascaratos, A., Klein, B. (2000). *The Eastern Mediterranean climatic transient : its origin, evolution and impact on the ecosystem*. Trieste, 29 March - 1st April 2000 CIESM Workshop Series no 10, p 86

Brinckmann-Voss, A. (1987). *Seasonal distribution of hydromedusae (Cnidaria, Hydrozoa) from the Gulf of Naples and vicinity, with observations on sexual and asexual reproduction in some species*. In: Bouillon J, Boero F, Cicogna F, Cornelius PFS (eds). *Modern trends in the systematics, ecology and evolution of hydroids and hydromedusae*. Oxford: Clarendon Press. pp. 133–141.

Chun, C. (1880). *Die Ctenophoren des Golfes von Neapel*. Fauna und Flora des Golfes von Neapel 1, 1–313.

Çinar, M. E., Yokeş, M. B., Açıık, Ş., Bakır, A. K. (2014). Checklist of Cnidaria and Ctenophora from the coasts of Turkey. *Turkish Journal of Zoology*, 38, 677–697. doi:10.3906/zoo-1405-68

Civitarese, G., Gacic, M., Lipizer, M., Borzelli, G. L. E. (2010). On the impact of the Bimodal Oscillating System (BiOS) on the biogeochemistry and biology of the Adriatic and Ionian Seas (Eastern Mediterranean). *Biogeosciences*, 7, 3987–3997. <http://doi.org/10.5194/bg-7-3987-2010>

Civitarese, G., Gacic, M., Batistic, Mirna., Bensi, M., Cardin, V., Dulcic, J., Garic, R., Menna, M. (2023). The BiOS mechanism: History, theory, implications. *Progress in Oceanography*, 216, 103056. <https://doi.org/10.1016/j.pocean.2023.103056>

Coll, M., Piroddi, C., Steenbeek, J., Kaschner, K., Ben Rais Lasram, F., Nike Bianchi, C., Corbera, J., Dailianis, T., Danovaro, R., Estrada, M., Frogli, C., Galil, B. S., Gasol, J. M., Gertwagen, R., Gil, J., Guilhaumon, F., Kesner-Reyes, K., Kitsos, M. S., Koukouras, A., Lampadariou, N., Laxamana, E., Lopez-Fe de la Cuadra, C. M., Lotze, H. K., Martin, D., Mouillot, D., Oro, D., Raicevich, S., Rius-Barile, J., Saiz-Salinas, J. I., San Vicente, C., Somot, S., Templado, J., Turon, X., Vafidis, D., Villanueva, R., Voultsiadou, E. (2010). The biodiversity of the Mediterranean sea: estimates, patterns, and threats. *PLoS One*, 5 (8), e11842. <https://doi.org/10.1371/journal.pone.0011842>

Cuttelod, A., García, N., Malak, D.A., Temple, H.J., Katariya, V. (2009). *The Mediterranean: a biodiversity hotspot under threat*. Wildlife in a Changing World—an analysis of the 2008 IUCN Red List of Threatened Species 89, 1–2.

Digenis, M. & Gerovasileiou, V. (2020). First Record of *Leucothea multicornis* (Quoy & Gaimard, 1824) (Ctenophora: Leucotheidae) in Greek Waters. *Acta Zoologica Bulgarica*, 72(3), 499–500.

Douek, J., Paz, G., Rinkevich, B., Gevili, R., Galil, B. S. (2020). First record of a non-native pelagiid jellyfish (Scyphozoa: Pelagiidae: Chrysaora) in the easternmost Mediterranean Sea. *BioInvasions Record*, 9(3), 482–489. doi:10.3391/bir.2020.9.3.04

Dragičević, B., Anadolı, O., Angel, D., Benabdi, M., Bitar, G., Castriota, L., Crocetta, F., Deidun, A., Dulčić, J., Edelist, D., Gerovasileiou, V.,

- Giacobbe, S., Goruppi, A., Guy-Haim, T., Konstantinidis, E., Kuplik, Z., Langeneck, J., Macali, A., Manitaras, I., Michailidis, N., Michaloudi, E., Ovalis, P., Perdikaris, C., Pillon, R., Piraino, S., Renda, W., Rizgalla, J., Spinelli, A., Tempesti, J., Tiralongo, F., Tirelli, V., Tsiamis, K., Turan, C., Uygur, N., Zava, B., Zenetos, A. (2019). New Mediterranean Biodiversity Records 2019. *Mediterranean Marine Science*, 20, 645–656. doi:10.12681/mms.20913
- Duman, G. S. (2023). Türkiye Akdeniz kıyılarının Eepipelajik zonunda bulunan mesozooplanktonik Oorganizmaların dağılımı. "Distribution of Epipelagic Mesozooplankton in Mediterranean Coast of Turkey". Akdeniz University, MSc Thesis, 98 p
- Edelist, D., Knutsen, Ø., Ellingsen, I., Majaneva, S., Aberle, N., Dror, H., Angel, D. L. (2022). Tracking jellyfish swarm origins using a combined oceanographic-genetic-citizen Science approach. *Frontiers in Marine Science*, 9, 869619. https://doi.org/10.3389/fmars.2022.869619
- Encarnaçao, J., Morais, P., Baptista, V., Cruz, J., Teodósio, M. A. (2019). New Evidence of Marine Fauna Tropicalization off the Southwestern Iberian Peninsula (Southwest Europe). *Diversity*, 11, 48. doi:10.3390/d11040048
- Ergen, Z. (1967). İzmir Korfezi'nde tespit edilen başlıca planktonik organizmalar. *Sci Rep Fac Sci Ege Univ* 47, 1–47 (in Turkish).
- Galil, B. (2007). Seeing Red: Alien species along the Mediterranean coast of Israel. *Aquatic Invasions*, 2(4), 281-312. doi:10.3391/ai.2007.2.4.2
- Galil, B. S., Rothman, S., Gevili, R., Shiganova, T. (2014). First record of *Leucothea multicornis* (Quoy & Gaimard, 1825) (Ctenophora; Lobata; Leucothidae) in the Eastern Mediterranean. *Marine Biodiversity Record*, 7, e89.
- Garuti, A. & Mutlu, E. (2021). Spatiotemporal and ecological distribution of megabenthic non-crustacean invertebrates in an ultraoligotrophic gulf, the eastern Mediterranean Sea. *Journal of Marine Systems*, 224, 103644.
- Gokoglu, M. & Galil, B. S. (2020). New records of siphonophores and ctenophores on the Levant Sea. *Journal of the Black Sea/ Mediterranean Environment*, 26(2), 190-202.
- Goy, J., Lakkis S., Zeidane, R. (1991). Les Meduses (Cnidaria) des eaux Libanaises. *Annales de l'Institut océanographique*, 67, 99-128.
- Goy, J. (1972). Les Hydroméduses de la mer Ligure. *Bulletin du Muséum national d'histoire Naturelle*, 83(62), 965–1008.
- Gravili, C., Bouillon, J., D'Elia, A., Boero, F. (2007). The life cycle of *Gastroblasta raffaelei* (Cnidaria, Hydrozoa, Leptomedusae, Campanulariidae) and a review of the genus *Gastroblasta*. *Italian Journal of Zoology*, 74, 395–403. doi:10.1080/11250000701650768
- Guerrero, E., Gili, J.-M., Maynou, F., Sabatés, A. (2018). Diversity and mesoscale spatial changes in the planktonic cnidarian community under extreme warm summer conditions. *Journal of Plankton Research*, 40(2), 178–196. doi:10.1093/plankt/fby001
- Gülşahin, N., Çelik, M., Turan, C., Ateş, C. (2016). First record of the hydrozoan *Oceania armata* Kölliker, 1853 from Turkey. *Zoology in the Middle East*, 62(2), 187-188. doi:10.1080/09397140.2016.1182782
- Gülşahin, N., Tarkan, A.N., Bilge, G. (2013) The hydrozoan *Geryonia proboscoidalis* (Forskål, 1775), new for Turkey (Hydrozoa). *Zoology in the Middle East*, 59(1), 93-94. doi:10.1080/09397140.2013.795077
- Gülşahin, N. & Türker, A. (2017). A New Ctenophora Species for Turkey – *Leucothea multicornis* (Quoy & Gaimard, 1824) (Order: Lobata). 2. Workshop on Invasive Species, Global meeting on invasion ecology, 27-29 September 2017, Bodrum, Muğla, TURKEY.
- Isinibilir, M., Martell, L., Topçu, E.N., Yılmaz, I.N., Piraino, S. (2015a). First inventory of the shallow-water benthic hydrozoan assemblages of Gökçeada Island (northern Aegean Sea). *Italian Journal of Zoology*, 82(2), 281-290. doi:10.1080/11250003.2014.977970
- İsinibilir, M., Ulucam, K., Yüksel, E. (2019). First record of *Physophora hydrostatica* Forskål, 1775 (Cnidaria, Hydrozoa) for the Turkish seas. *Journal of the Black Sea/Mediterranean Environment*, 25(2), 160-165.
- Isinibilir, M., Yılmaz, I.N., Demirel, N., (2015b). New records of jellyfish species in the Marmara Sea. *Italian Journal of Zoology*, 82(3), 425-429. doi:10.1080/11250003.2015.1040858
- Isinibilir, M., Yüksel, E., Guresen, O. (2021). The first record of *Geryonia proboscoidalis* (Forskål, 1775), (Cnidaria: Hydrozoa) on the coasts of Gökçeada, the Aegean Sea, Turkey. *Aquatic Sciences and Engineering*, 36(4), 215-217
- Isinibilir, M., Yüksel, E., Turkeri, E.E., Dogan, O., Karakulak, F.S., Uzer, U., Dalyan, C., Furfaro, G., Piraino, S. (2022). New Additions to the Jellyfish Fauna of the Sea of Marmara. *Aquatic Sciences and Engineering*, 37(1), 53-57.
- Jasprica, N., Čalić, M., Kovačević, V., Bensi, M., Radić, I. D., Garić, R., Batistić, M. (2022). Phytoplankton distribution related to different winter conditions in 2016 and 2017 in the open southern Adriatic Sea (eastern Mediterranean). *Journal of Marine Systems*, 226, 103665. https://doi.org/10.1016/j.jmarsys.2021.103665
- Karaca, D. (2023). Distribution of phytoplankton found along Turkish coasts of the Mediterranean Sea. MSc Thesis, Akdeniz University, 115 p.
- Kherchouche, A. & Hafferssas, A. (2019). Species composition and distribution of Medusae (Cnidaria: medusozoa) in the Algerian coast between 2°E and 7°E (SW Mediterranean Sea). *Mediterranean Marine Science*, 21(1), 52–61. https://doi.org/10.12681/mms.20849
- Kideys, A.E. & Niermann, U. (1993). Intrusion of *Mnemiopsis mcgradyi* (Ctenophora: Lobata) into the Mediterranean Sea. *Senckenbergiana Maritima*, 23(1), 43-47.
- Killi, N., Abyzova, G., Shiganova, T. (2019). Occurrence *Beroe mitrata* (Beroidae) associated with its prey aggressive invader ctenophore *Mnemiopsis leidyi* A. Agassiz from Gökova Bay. In Özcan, T. (Ed.). 2019. International Biodiversity & Ecology Sciences Symposium Proceeding (Bioeco2019), 26-28 September 2019, İstanbul/Turkey. Palas Academic Organization and Trade Corporation, Publication number: 1, İskenderun-Hatay, Turkey, 259-259.
- Lang, A. (1886). *Gastroblasta raffaelei* Eine durch eine Art unvollständiger Theilung entstehende Medusen-Kolonie. *Jenaische Zeitschrift für Naturwissenschaft*, 19, 735–763.
- Lascaratos, A., Williams, R. G., Tragou, E. (1993). A Mixed-Layer Study of the Formation of Levantine Intermediate Water. *Journal of Geophysical Research*, 98, 739-749.
- Lorenzen, C. J. (1967). Determination of chlorophyll and phaeopigments: spectrophotometric equations. *Limnology and Oceanography*, 12(2), 343-346. doi:10.4319/lo.1967.12.2.0343
- Lucic, D., Pestoric, B., Malej, A., Lopez-Lopez, L., Drakulovic, D., Onofri, V., Miloslavic, M., Gangai, B., Onofri, I., Benovic, A. (2012). Mass occurrence of the ctenophore *Bolinopsis vitrea* (L. Agassiz, 1860) in the nearshore southern Adriatic Sea (Kotor Bay, Montenegro). *Environmental Monitoring and Assessment*, 184, 4777–4785. DOI 10.1007/s10661-011-2301-6
- Madin, L. P. (1991). *Distribution and taxonomy of zooplankton in the Alboran Sea and adjacent western Mediterranean: A literature survey and field guide*. Woods Hole Oceanographic Institution, Technical Report, WHOI-91-26, USA.
- Mamish, S., Durgham, H., Ikhtiyar, S. (2019). First record of *Leucothea multicornis* (Quoy & Gaimard, 1824) off the Syrian coastal water (eastern Mediterranean Sea). *SSRG International Journal of Agriculture & Environmental Science*, 6(4), 1-4.
- Mayer, A. G. (1910). *Medusae of the world; volume 1: Hydromedusae*. The Carnegie Institution of Washington. Washington, D. C., 230 p.
- Mayer, A. G. (1912) *Ctenophores of the Atlantic Coast of North America*. The Carnegie Institution of Washington. Washington, D. C., 162, 1-58.
- Mureşan, M., Teacă, A., Begun, T. (2021). First record of the hydrozoan *Podocorynoides minima* (Trinci, 1903) in the Romanian Black Sea waters. *Biologia*, 77, 1819–1828, https://doi.org/10.1007/s11756-022-01051-5
- Mutlu, E. (1999). Distribution and abundance of ctenophores and their zooplankton food in the Black Sea. II. *Mnemiopsis leidyi*. *Marine Biology*, 135, 603–613. doi:10.1007/s002270050661

- Mutlu, E., Bingel, F., Gücü, A. C., Melnikov, V. V., Niermann, U., Ostr, N. A., Zaika, V. E. (1994). Distribution of the new invader *Mnemiopsis* sp. and the resident *Aurelia aurita* and *Pleurobrachia pileus* populations in the Black Sea in the years 1991–1993. *ICES Journal of Marine Science*, 51(4), 407–421. doi:10.1006/jmsc.1994.1042.
- Mutlu, E., Çağatay, I. T., Olguner, M. T., Yılmaz, H. E. (2020). A new sea-nettle from the Eastern Mediterranean Sea: *Chrysaora pseudoocellata* sp. nov. (Scyphozoa: Pelagiidae). *Zootaxa*, 4790(2), 229–244. doi:10.11646/zootaxa.4790.2.2
- Mutlu, E. & Özvarol, Y. (2022a). First record of *Hormiphora plumosa* (Ctenophora: Pleurobrachiidae) from the Turkish coast of the Mediterranean, 438–439. In Kousteni, V., Anastasiadis, A., Bariche, M., Battaglia, P., Bonifazi, A., Četković, I., Chimienti, G., Colombo, M., Constantinou, C., Corsini-Foka, M., Dalyan, C., Dogrammatz, A., Domenichetti, F., El Zrelli, R., Fernández-Álías, A., Kampouris, T. E., Kesici, N. B., Küpper, F. C., Lipej, L., Mancini, E., Manunza, B., Marcos, C., Mavrič, B., Mavruk, S., Mutlu, E., Özvarol, Y., Papadimitriou, E., Pešić, A., Pérez-Ruzafa, A., Pey, A., Poursanidis, D., Rizgalla, J., Samaha, Z., Stipa, M. G., Trkov, D., Türel, C., Ventura, P., Yacoubi, L., Zacchetti, L., Zava, B. (co-authors), New records of rare species in the Mediterranean Sea (May 2022). *Mediterranean Marine Science*, 23, 417–446. https://doi.org/10.12681/mms.28372
- Mutlu, E., Özvarol, Y. (2022b). Recent record of *Oceania armata* and Near-past records of other gelatinous organisms in the Turkish waters presumably derived by basin-scale current. *COMU Journal of Marine Science and Fisheries*, 5(1), 48–55. https://doi.org/10.46384/jmsf.1083023
- Mutlu, E., Meo, I. d., Miglietta, C., Deval, M. C. (2023a). Ecological indicative stressors of native vs non-native fish in an ultra-oligotrophic region of the Mediterranean Sea. *Sustainability*, 15, 2726. https://doi.org/10.3390/su15032726.
- Mutlu, E., Karaca, D., Duman, G. S., Şahin, A., Özvarol, Y., Olguner, C. (2023b). Seasonality and phenology of an epiphytic calcareous red alga, *Hydrolython boreale*, on the leaves of *Posidonia oceanica* (L) Delile in the Turkish water. *Environmental Science and Pollution Research*, 30, 17193–17213. https://doi.org/10.1007/s11356-022-23333-w
- Oliveira, O. M. P., Mianzan, H., Migotto, A. E., Marques, A. C. (2007). Chave de identiicação dos Ctenophora da costa Brasileira. *Biota Neotropica*, 7(3), 341–350. doi:10.1590/S1676-06032007000300034
- Oliveira, O. M. P. & Migotto, A. E. (2006). Pelagic ctenophores from the São Sebastião Channel, southeastern Brazil. *Zootaxa*, 1183, 1–26.
- Ozer, T., Gertman, I., Kress, N., Silverman, J., Herut, B. (2017). Interannual thermohaline (1979–2014) and nutrient (2002–2014) dynamics in the Levantine surface and intermediate water masses, SE Mediterranean Sea. *Global and Planetary Change*, 151, 60–67. https://doi.org/10.1016/j.gloplacha.2016.04.001
- Öztürk, B., Mihneva, V., Shiganova, T. (2011). First records of *Bolinopsis vitrea* (L. Agassiz, 1860) (Ctenophora: Lobata) in the Black Sea. *Aquatic Invasions*, 6(3), 355–360. doi: 10.3391/ai.2011.6.3.12
- Öztürk, B. (2021). Non-indigenous species in the Mediterranean and the Black Sea. Studies and Reviews No. 87 (General Fisheries Commission for the Mediterranean). Rome, FAO. https://doi.org/10.4060/cb5949en
- Patania, A. & Mutlu, E. (2021). Spatiotemporal and ecological distribution of megabenthic crustaceans on the shelf-shelf break of Antalya Gulf, the eastern Mediterranean Sea. *Mediterranean Marine Science*, 22, 446–465.
- Pesticor, B., Lucic, D., Bojanic, N., Vodopivec, M., Kogovšek, T., Violic, I., Paliaga, P., Malej, A. (2021). Scyphomedusae and Ctenophora of the eastern Adriatic: Historical overview and new data. *Diversity*, 13, 186. https://doi.org/10.3390/d13050186
- Poulain, P.-M., Bussani, A., Gerin, R., Jungwirth, R., Mauri, E., Menna, M., Notarstefano, G. (2013). Mediterranean surface currents measured with drifters: From basin to subinertial scales. *Oceanography*, 26(1), 38–47. doi:10.5670/oceanog.2013.03.
- Poulos, S. E., Drakopoulos, P.G., Collins, M. B. (1997). Seasonal variability in sea surface oceanographic conditions in the Aegean Sea (Eastern Mediterranean): an overview. *Journal of Marine Systems*, 13(1–4), 225–244. https://doi.org/10.1016/S0924-7963(96)00113-3.
- Raitsos, D. E., Kassis, D., Sarantopoulos, A. (2011). The summer North Atlantic Oscillation influence on the Eastern Mediterranean. *Journal of Climate*, 24, 5584–5596. DOI: 10.1175/2011JCLI3839.1
- Ricci, P., Carlucci, R., Capezuto, F., Carluccio, A., Cipriano, G., D’Onghia, G., Maiorano, P., Sion, L., Tursi, A., Libralato, S. (2022). Contribution of intermediate and high trophic level species to benthic-pelagic coupling: Insights from modelling analysis. *Frontiers in Marine Science*, 9, 887464. https://doi.org/10.3389/fmars.2022.887464
- Schmidt, H. E. (1973). Die Hydromedusen (Hydrozoa: Coelenterata) des Roten Meeres und seiner angrenzenden Gebiete. *Meteor Forschungsergebnisse Reihe D – Biologie*, 15, 1–35.
- Schuchert, P. (2007). The European athecate hydroids and their medusae (Hydrozoa, Cnidaria): Filifera Part 2. *Revue suisse de Zoologie*, 114(2), 195–396. doi:10.5962/bhl.part.80395
- Shiganova, T. A. (2020). Adaptive strategies of *Mnemiopsis leidyi* A. Agassiz 1865 in different environments of the Eurasian seas. *Marine Pollution Bulletin*, 161, 111737. doi:10.1016/j.marpolbul.2020.111737
- Shiganova, T. & Malej, A. (2009). Native and non-native ctenophores in the Gulf of Trieste, Northern Adriatic Sea. *Journal of Plankton Research*, 31(1), 61–71. doi:10.1093/plankt/fbn102.
- Regione Del Veneto, Shape (2013). *Definition of the Adriatic ecosystem quality as basis for Maritime Spatial planning: Contribution to the initial assessment of marine Adriatic waters according to Directive 2008/56/CE Action 4.2. Action 4.2 Final Report*. 21863-REL-T-003.2. Thetis S.p.a., pp. 322. 15 October 2013
- Touzri, C., Hamdi, H., Goy, J., Yahia, M. N. D. (2012). Diversity and distribution of gelatinous zooplankton in the Southwestern Mediterranean Sea. *Marine Ecology*, 33, 393–406. doi:10.1111/j.1439-0485.2012.00510.x
- Uttieri, M., Anadoli, O., Banchi, E., Battuello, M., Besiktepe, S., Carotenuto, Y., Marques, S. C., de Olazabal, A., Di Capua, I., Engell-Sørensen, K., Goruppi, A., Guy-Haim, T., Hure, M., Kourkoutmani, P., Lucic, D., Mazzocchi, M. G., Michaloudi, E., Morov, A. R., Terbiyik Kurt, T., Tirelli, V., Vannini, J., Velasquez, X., Vidjak, O., Wootton, M. (2023). The Distribution of *Pseudodiaptomus marinus* in European and Neighbouring Waters—A Rolling Review. *Journal of Marine Science and Engineering*, 11, 1238. https://doi.org/10.3390/jmse11061238
- Uysal, Z. & Mutlu, E. (1993). Preliminary note on the occurrence and biometry of Ctephoran *Mnemiopsis leidyi* finally invaded Mersin Bay. *Turkish Journal of Zoology*, 17(2), 229–236
- Vasilakopoulos, P., Raitsos, D. E., Tzanatos, E., Maravelias, C. D. (2017). Resilience and regime shifts in a marine biodiversity hotspot. *Scientific Reports*, 7, 13647. https://doi.org/10.1038/s41598-017-13852-9
- Vinogradov, M. Ye., Shushkina, E. A., Musayeva, E. I., Sorokin, F. Yu. (1989). A newly acclimated species in the Black Sea The ctenophore *M. leidyi* (Ctenophora Lobata). *Oceanology*, 29, 220–224.
- Yılmaz, I. N., Isinibilir, M., Vardar, D., Dursun, F. (2017). First record of *Aequorea vitrina* Gosse, 1853 (Hydrozoa) from the Sea of Marmara: a potential invader for the Mediterranean Sea. *Zoology in the Middle East*, 63(2), 1–3. doi:10.1080/09397140.2017.1299334
- Zenetos, A., Gofas, S., Morri, C., Rosso, A., Violanti, D., Garcia Raso, J., Cinar, M., Almogi-Labin, A., Ates, A., Azzurro, E., Ballesteros, E., Bianchi, C., Bilecenoglu, M., Gambi, M., Giangrande, A., Gravili, C., Hyams-Kaphzan, O., Karachle, P. K., Katsanevakis, S., Lipej, L., Mastrototaro, F., Mineur, F., Pancucci-Papadopoulou, M., Ramos Espla, A., Salas, C., San Martin, G., Sfriso, A., Streftaris, N., Verlaque, M. (2012). Alien species in the Mediterranean Sea by 2012. A contribution to the application of European Union’s Marine Strategy Framework Directive (MSFD). Part 2. Introduction trends and pathways. *Mediterranean Marine Science*, 13(2), 328–352. https://doi.org/10.12681/mms.327

Lakes of Turkey: Comprehensive Review of Lake Çıldır

Özgür Eren Zariç¹ , Abuzer Çelekli¹ , Sidar Yaygır¹ 

Cite this article as: Zariç, Ö.E., Çelekli, A, & Yaygır, S. (2024). Lakes of Turkey: comprehensive review of Lake Çıldır. *Aquatic Sciences and Engineering*, 39(1), 54-63. DOI: <https://doi.org/10.26650/ASE20241353730>

ABSTRACT

Lake Çıldır, situated in eastern Turkey, is one of the most noteworthy freshwater lakes in the region, distinguished for its aesthetic allure and its paramount ecological and socio-economic roles. This review thoroughly examines Lake Çıldır, beginning with an introduction that contextualizes its importance. The climatic conditions influencing the lake's dynamics, its hydrographic characteristics, and the physicochemical variables determining its water quality have been specified. Our review synthesizes findings about the lake's biological attributes, including its varied flora and fauna. Detailed sections are dedicated to the fish species, macrophytes, benthic macroinvertebrates, and phytoplankton that inhabit the lake. Using land cover data, temporal changes in the lake's environmental structure have been evaluated. The lake's trophic state and overall ecological health are critically discussed, shedding light on its current status. Furthermore, the potential and existing ecotourism practices around Lake Çıldır are explored, emphasizing its value as a tourist destination. In conclusion, the challenges faced by the lake are addressed, advocating for robust conservation and management initiatives. This comprehensive review of Lake Çıldır's complex ecosystem is a foundation for forthcoming research.

Keywords: Ecology, Ecotourism, Environmental structure, Lake Çıldır, Land cover, Trophic state

INTRODUCTION

Situated in the northern region of East Anatolia, Lake Çıldır is located between the cities of Kars and Ardahan at 41° 00' north latitude and 43° 12' longitude in Turkey. Nestled within the provinces of Ardahan and Kars, Lake Çıldır, with its expanse of 123 km², stands as the second-largest lake in Eastern Anatolia. Positioned at an altitude of 1959 meters above sea level, it reaches a depth of 42 meters, encircled by a vast stretch of approximately 60 kilometers. The lake's genesis can be attributed to a confluence of a lava flow (Aykir & Fıçıcı, 2022). Lake Çıldır's hydrography is an intricate web of inflows and outflows. While myriad streams and springs replenish it, its sole drainage point is the Telek Creek, branching out to the Arpaçay near the Armenian frontier. Adjacent to the Akçakale ruins lies an island, further enhancing the lake's topography (Şimşek, 2019). Despite

its limited vegetation, the lake's vicinities burgeon with pastures, underscoring the socio-economic dependencies on animal husbandry (Yerli, 1993). The regional economy is substantially bolstered by fishing, a year-round activity that persists even during icy winters. Predominantly, the mirror carp (*Cyprinus carpio*) is the prized catch (Yerli, 1997). Ecologically, the lake confronts adversities. Declining water levels in arid spells imperil habitats essential for fish reproduction, while unsustainable fishing practices exacerbate the strains on fish populations (Başçınar et al., 2009). Notably, the lake's northwestern fringes, adorned with wetlands and marshes, amplify its ecological heterogeneity. More than a mere geographic landmark, Lake Çıldır embodies a vibrant ecosystem, an economic linchpin, and a vault of regional heritage. Figure 1 offers a cartographic representation of its location, providing a spatial context;

ORCID IDs of the author:
Ö.E.Z. 0000-0001-5293-871X;
A.Ç. 0000-0002-2448-4957;
S.Y. 0009-0005-0878-3366

¹Gaziantep University, Environmental
Research Center (GÜÇAMER),
Gaziantep, Türkiye

Submitted:
01.09.2023

Revision Requested:
10.10.2023

Last Revision Received:
13.10.2023

Accepted:
13.10.2023

Online Published:
25.12.2023

Correspondence:
Abuzer Çelekli
E-mail:
celekli.a@gmail.com



according to the Turkish General Directorate of Mapping Data, maps were prepared using *ESRI ArcMap 10.7* software. This paper holistically examines Lake Çıldır, advocating its preservation while spotlighting its multifaceted challenges in our evolving global context.

the connection between the Lake Çıldır basin and the Çıldır Plain, eventually fostering the formation of the lake, nurtured by streams originating from the encompassing terrains (Atalay et al., 2018). The prevalent soil types within the basin are chestnut-colored and basaltic (Karaoğlu & Çelim, 2018). The basaltic soils,

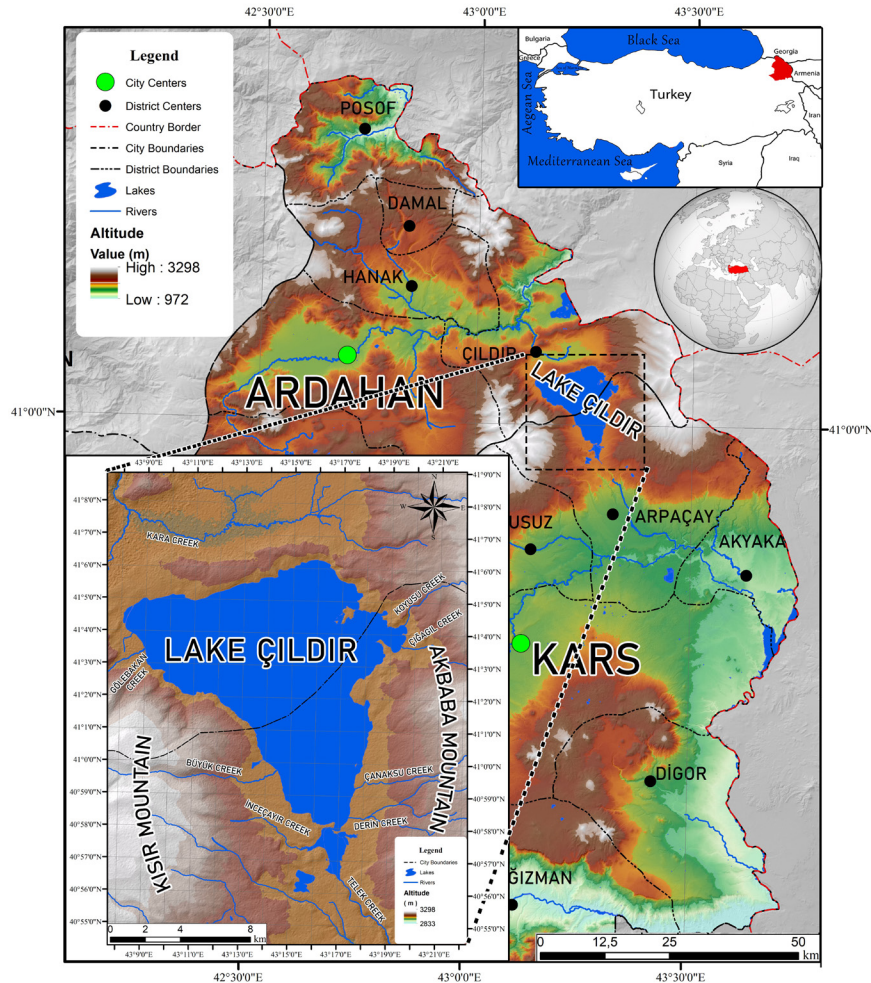


Figure 1. Geographical Position of Lake Çıldır in Turkey.

Geographical features

The geologic foundation of the Lake Çıldır basin is predominantly composed of volcanic tuff, block tuff, andesitic, and basaltic flows, supplemented by younger marl and conglomerates from the post-Oligocene period. During the Pliocene era, the region experienced significant vertical tectonic activities, precipitating block descents along the fault lines (Atalay et al., 2018). Historically, the Lake Çıldır basin and the adjacent Çıldır Plain existed as a unified depression, channeling its hydrologic flow towards the Kura River via the Çıldır Stream (N. E. Akbulut et al., 2022). However, subsequent lava flows partitioned this expansive basin into two distinct sections, creating an intermediary plain between the lava flow's forefront and the Kısır Mountain. Over time, the debris deposited from the Kısır Mountain culminated in the total occlusion of this corridor. This geologic evolution effectively severed

emerging from the volcanic bedrock, are characterized by their proficient drainage capabilities and clayey surface strata (Öztürk et al., 2015).

Climatic features

Situated in the northeastern part of the Eastern Anatolia Region, Lake Çıldır exemplifies the pronounced continental climate characteristic of the area. Due to its distinctive geographical position, the region is one of Turkey's most intensely affected areas by continental climate. This results in short, cool summers and bitterly cold winters (Şimşek, 2019).

The climatic characteristics of a region, encompassing temperature variations and precipitation patterns, are fundamental to comprehending its hydrological and ecological dynamics. Such

knowledge is pivotal for formulating strategies for water resource management, agricultural activities, and the preservation of ecosystems. In this context, an analysis was conducted utilizing meteorological data collated by the Turkish State Meteorological Service from 1931 to 2023 for Kars and Ardahan. Figure 2 shows the precipitation and temperature averages for 2000-2023. As the temperature rises, the decrease in precipitation may pose a problem for species diversity and lake health. The findings indicate that the mean annual temperature for Kars is 4.8°C, reaching a peak in August at 17.8°C and a nadir in January at -10.8°C. Ardahan, on the other hand, exhibits a marginally lower mean annual temperature of 3.7°C, with the highest temperatures recorded in July and August at 16.0°C and the lowest in January at -11.2°C. Concerning precipitation, Kars records an annual average of 506.0 mm, while Ardahan experiences a slightly elevated average of 555.3 mm (Turkish State Meteorological Service, 2018). Notably, May is the month with the highest precipitation in both regions. This characteristic diverges from the typical precipitation patterns observed in other parts of Turkey, where summer is identified as the season with the highest rainfall. Conversely, the winter season is associated with the lowest precipitation levels in both Kars and Ardahan. This climatic information is crucial for developing effective strategies for water resource management, agricultural practices, and ecosystem conservation initiatives.

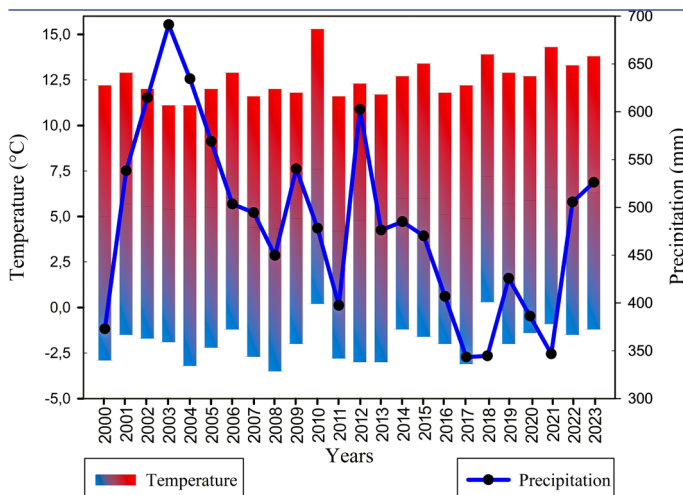


Figure 2. Temperature and precipitation with averages of Lake Çıldır.

Hydrographic features

Lake Çıldır, located in the catchment areas of the Aras and Kura rivers, holds a crucial hydrological role in the region, ultimately discharging into the Caspian Sea via these intersecting rivers in Azerbaijan's Sabirabad. The lake is predominantly replenished by snowmelt and several seasonal and perennial streams, originating from the Kısır Mountains in the west and the Akbaba Mountains in the east. Major tributaries on the western side include the Gölbelen, Kindırğa, and Gölebakan rivers. In contrast, the eastern side receives contributions from the Gülyüzü River and smaller streams passing through the Göldalı and Çanaksu

villages (Lahn, 1947). Lake Çıldır, characterized by its freshwater, has an overflow management mechanism that directs excess water through a narrow corridor near Taşbaşı Village to the Telek Creek and eventually to the Arpaçay. Additionally, the natural barrier between the Çıldır Plain and the lake basin facilitates the lake's assimilation of waters from various sources and streams originating from the plain. As a critical hydrological entity nourished by diverse geographical sources and feeding major river systems, Lake Çıldır's recognition is vital for informed water management and local ecosystem protection (Alkan et al., 2016).

Physicochemical variables

The health and function of lake ecosystems are intrinsically linked to their physicochemical properties, necessitating a comprehensive understanding of the interactions between these factors and their broader ecological consequences (Langdon et al., 2010). Parameters such as temperature, pH, dissolved oxygen, nutrient concentrations, and pollutants are diagnostic tools for assessing water quality and ecological status. These factors influence the metabolic rates of aquatic organisms, their life cycles, ecological interactions, nutrient solubility, productivity, and biodiversity (Brown et al., 2004). Furthermore, nutrient dynamics, particularly nitrogen and phosphorus, play a crucial role in lake ecology. Excessive anthropogenic inputs can lead to eutrophication, which reduces light and oxygen availability, adversely affecting aquatic systems; pollutants, heavy metals, and organic compounds pose significant risks, potentially leading to bioaccumulation and biomagnification in the food chain (Ali & Khan, 2019).

By integrating physicochemical data with environmental and biological indicators, we can comprehensively evaluate lake health and identify potential ecological disturbances (Beck & Hatch, 2009). Such insights are crucial for developing effective management strategies to restore and maintain the balance of lake ecosystems. Therefore, understanding and analyzing the interrelationships between physicochemical properties and biological dynamics is essential for preserving our precious freshwater resources. Table 1 indicates the physicochemical data of Lake Çıldır.

Lake Çıldır has been reported to show no signs of sewage and eutrophication, and it has been characterized as oligotrophic based on physicochemical parameters. However, due to population growth and the effects of pollution elements originating from residential areas and agricultural and livestock enterprises in its vicinity, Lake Çıldır is gradually becoming polluted (Çakır & Çiçek, 2015).

The heavy metal levels in the surface sediments of the northern littoral zone of Lake Çıldır have been studied, and it has been found that the lake is affected by the ecological effects of heavy metals. Additionally, the lake has been found to contain heavy metal levels, including Cu, Pb, Zn, Ni, Mn, Fe, As, Cd, Cr, and Hg, which may have ecological effects (Kükreker et al., 2015).

Biological feature

From a botanical standpoint, Turkey's Central and Eastern Anatolia regions are characterized by the presence of Iran-Turan flora, with a notable distribution in Northeast Anatolia. In contrast, the area surrounding Lake Çıldır dominates the European-Siberian

Table 1. Physicochemical variables in Lake Çıldır (Çelekli et al., 2020).

Variables	unit	Mean±SD	Minimum	Maximum
Temperature	°C	16.9±2.7	14.80	19.90
pH		7.94±1.29	6.47	8.86
EC	µS/cm	134±16	117.00	149.00
Salinity	ppt	0.10±0.05	0.07	0.15
DO	mg/l	8.86±0.29	8.53	9.05
TSS	mg/l	19.9±26.1	2.80	50.00
BOD ₅	mg/l	4.7±1.4	3.70	6.30
COD	mg/l	19.6±5.0	14.50	24.40
TOC	mg/l	2.5±0.4	2.10	2.80
TN	mg/l	0.40±0.10	0.32	0.51
N-NH ₄	mg/l	0.11±0.02	0.10	0.14
N-NO ₂	mg/l	0.01±0.01	0.00	0.01
N-NO ₃	mg/l	0.14±0.05	0.10	0.20
TP	mg/l	0.14±0.12	0.05	0.28
P-PO ₄	mg/l	0.11±0.16	0.01	0.30
Secchi depth	m	3.00±1.32	2.50	4.00

The average and standard deviation (SD) of Lake Çıldır's environmental variables and their minimum and maximum values in brackets are provided. These variables include DO (dissolved oxygen), EC (electrical conductivity), TSS (total suspended solids), BOD5 (biological oxygen demand), TOC (total organic carbon), TN (total nitrogen), N-NO₂ (nitrite), N-NO₃ (nitrate), N-NH₄ (ammonium), P-PO₄ (orthophosphate), and TP (total phosphorus).

phytogeographic elements, reflecting the lake's unique geographic positioning (Atalay et al., 2018). Identifies four distinct vegetation types around the lake: steppe, meadow, rock, and tree-shrub. Salient species include willow variants (*Salix caucasica* and *Salix caprea*), shrubs (*Spiraea hypericifolia*, *Padus avium*), and wild plum (*Prunus spinosa*). The vegetation's diversity can be attributed to the area's short 4-5 months period and distinct geographical conditions. At Lake Çıldır, according to the International Union for Conservation of Nature, endemic plants are in danger. Some endemic plants found in the flora of Kars-Ardahan and Lake Çıldır are *Senecio integrifolius* subsp. *karsianus*, *Veronica thymoides*, *Chaerophyllum karsianum*, *Veronica gentianoides*, *Lathyrus karsianus* (Figure 3), *Festuca karsianum*, *Allium karsianum*, *Nonea karsensis* (Akgül & Aytac, 2008; Armağan et al., 2017).

The aquatic ecosystem of Lake Çıldır is remarkably diverse. The lake is a habitat for native fish species such as the common carp (*Cyprinus carpio*) and the Caspian trout (*Salmo trutta caspius*). Additionally, the Prussian carp - (*Carassius gibelio*) and the crayfish (*Pontastacus leptodactylus*) are shown in Figure 4. There are also rainbow trout populations (*Oncorhynchus mykiss*) within the lake (Çiçek et al., 2022; Çiçek & Birecikligil, 2016; Koçyiğit & Önder, 2018; Zengin et al., 2012).

Avifauna around Lake Çıldır is also noteworthy. Regional data indicates the presence of 86 bird species, including significant ones like velvet ducks (*Melanitta fusca*) and pintails (*Anas acuta*). Two "bird islands" within the lake are hubs for dense bird populations, mainly gulls, providing a visually snow-like spectacle. Historically, pelican species nested here, with recent sightings suggesting potential breeding activity linked to neighboring Lake Aktaş (Kartzahi). The Dalmatian pelican (*Pelecanus crispus*), ruddy shelduck (*Tadorna ferruginea*), common crane (*Grus grus*,

and Armenian gull (*Larus armenicus*) are bird species that meet global and/or, regional Important Bird and Biodiversity Area criteria. The area is a critical stopover for avian migration during spring and autumn. Notably, from mid-September onward, significant numbers of common buzzards (*Buteo buteo*) and lesser spotted eagles (*Aquila pomarina*) can be observed in the north-eastern and southwestern parts of the area. It is possible to monitor with the Bird Observation Tower in Lake Çıldır (Figure 5). However, there is no available data regarding the exact population sizes of these species (Azizoğlu & Adizel, 2020; Crivelli & Vizi, 1981; Özkoç, 2020; Şimşek, 2019; Yılmaz et al., 2018).



Figure 3. *Lathyrus karsianus* a. live plant (Savran, 2021) b. herbarium sample (POWO, 2023).

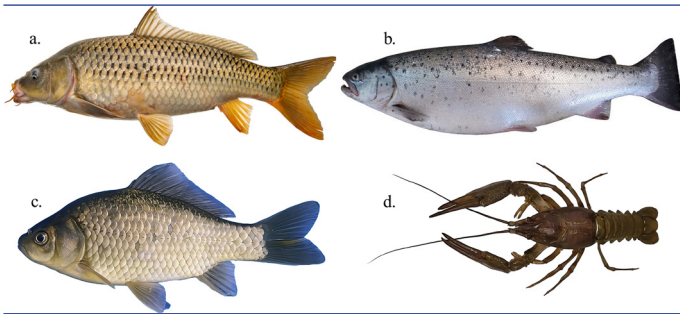


Figure 4. a. *Cyprinus carpio* (The Fishes of North Carolina, 2023), b. *Salmo trutta caspius* (Jouladeh-Roudbar et al., 2020), c. *Carassius gibelio* (Kalous et al., 2012), d. *Pontastacus leptodactylus*.



Figure 5. Photos from the Bird Observation Tower in Lake Çıldır (Çıldır District Governorate, 2023).

Lake Çıldır showcases a diverse macrozoobenthic community, with notable species such as *Armiger crista* and *Paratanytarsus lauterborni*. Recent research has highlighted the lake's rich aquatic oligochaete diversity, identifying 22 species, including new records like *Pristina synclites* and *Bratislavia palmeni*. Furthermore, a comprehensive analysis revealed that the lake's macrozoobenthic fauna encompasses 47 taxa, two unprecedented records for Turkey's Oligochaeta fauna (Arslan & Mercan, 2020; Mercan et al., 2022).

Lake Çıldır is characterized by a rich and varied phytoplankton community, reflecting its ecological complexity and significance in freshwater ecosystems. Phytoplankton play a crucial role in the global carbon cycle and are responsible for producing a significant amount of oxygen. Problems related to the carbon cycle are the causes of global warming and can cause various problems, up to human migrations (Çelekli et al., 2023). Their species diversity and abundance act as water quality indicators due to their different responses to environmental condi-

tions (Çelekli et al., 2020). In addition, phytoplankton are of significant biotechnological importance; some are the removal of harmful dyes. (Zariç et al., 2022). As part of the research, 74 phytoplankton species have been identified from five lakes in the Aras River basin, including Lake Çıldır. *Cocconeis placentula*, *Ulnaria ulna*, *Cymbella affinis*, *Lindavia bodanica*, *Pseudopediatrum boryanum*, and *Ankistrodesmus arcuatus* were among the commonly found species; some of them indicated in Figure 6. Bacillariophyta was the dominant group in the lake based on their values of phytoplankton biovolume in all sampling times. The diatoms contributed (89%) to the total phytoplankton biovolume in Lake Çıldır (Çelekli et al., 2020). As mentioned above, the common diatom species of Lake Çıldır are also the most widespread and abundant in other lakes in Turkey. However, some species were found to be rare, such as *Coscinodiscus* sp., *Stauroneis marlyi*, *Didymosphenia geminata*, *Stauroneis acute*, *Diatoma hiemale*, *Aulacoseira ambigua*, and *Ellerbeckia arenaria* (Akbulut & Yıldız, 2002).

In light of the above findings, it is evident that Lake Çıldır represents a complex nexus of biogeographic, ecological, and anthropogenic influences. Given its environmental significance, devising and implementing holistic conservation and management strategies is imperative.

Trophic state and ecological status of Lake Çıldır

Phytoplankton biomass, chlorophyll-a, total phosphorus (TP), and Secchi disk depth are essential indicators of lakes' trophic state and ecological status (Poniewozik & Lenard, 2022). Changes in water color intensity can affect a lake's physical, chemical, and biological parameters, including the concentration of TP and nitrogen and the biomass and composition of phytoplankton (Lenard et al., 2018). Secchi disk depth and TP values obtained during the study were used to determine the trophic status of ecosystems according to OECD criteria. Results of Carlson's trophic state index (Carlson, 1977) based on Secchi depth (TSI_{SD}) indicated that Lake Çıldır was mesotrophic characteristic (Çelekli et al., 2020). The modified phytoplankton trophic index-MPTI (Çelekli, 2016) and the Mediterranean phytoplankton trophic index-Med-PTI (Marchetto et al., 2009) were used to estimate the ecological status of the lake. The high ecological status in the Aras River basin was only found in Lake Çıldır (Çelekli et al., 2020). Although the lake's condition appears better than others, it could be due to its larger surface area compared to others. Continued conservation efforts are necessary because of the significant human impact on the lake.

The land cover changes Lake Çıldır

Lake Çıldır has undergone significant land cover transformations from 2000 to 2018. This was analyzed using CORINE Land Cover (CLC) (EAA,2018) data from the Copernicus Land Monitoring Service and visualized using ESRI ArcMap 10.7 software, as demonstrated in Figure 7 and their results are given in Table 2. This amalgamation of satellite, sensor network data, and land cover data allowed for a thorough examination and intuitive comprehension of the spatial distribution and transformations of various land cover types over the studied period.

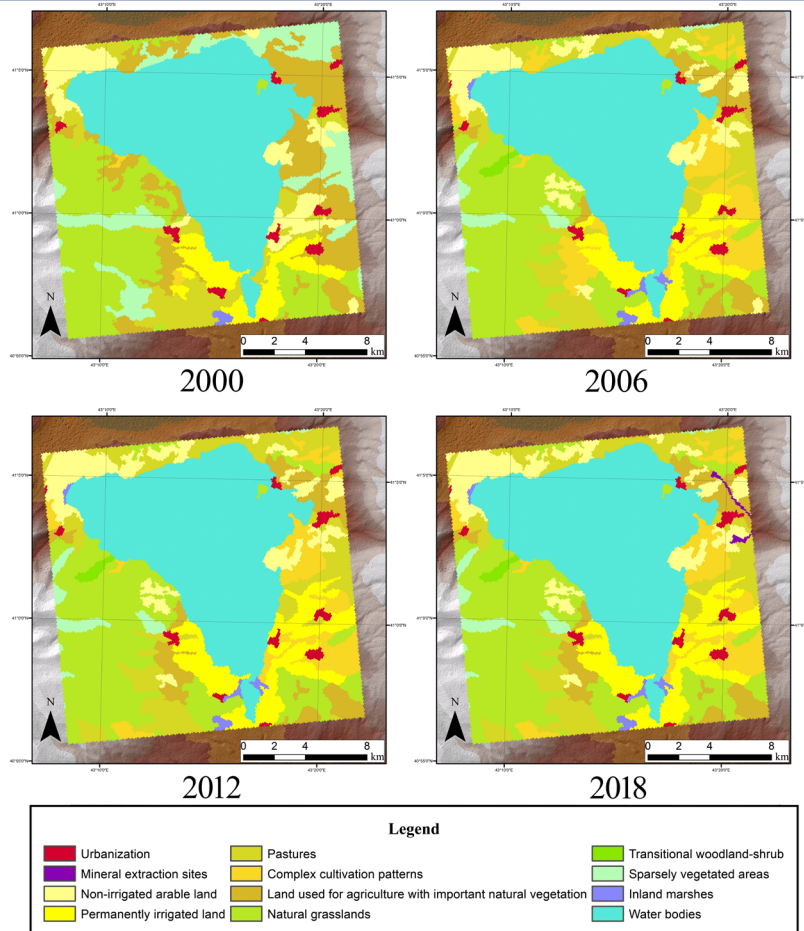
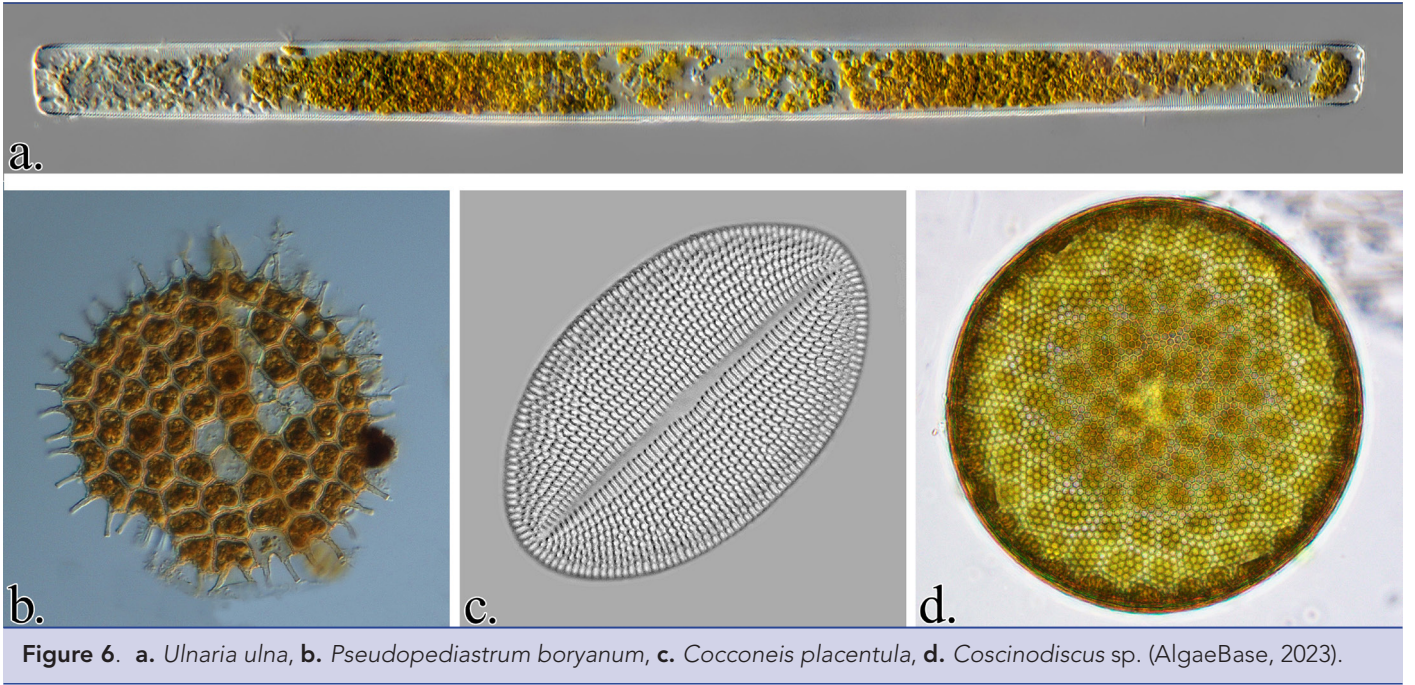


Figure 7. Lake Çıldır CLC Map (2000-2018).

Table 2. Land cover changes around Lake Çıldır from 2000 to 2018 (CLC, 2018).

Landcover (%)	2000	2006	2012	2018
Urbanization	0.49	0.45	0.55	0.56
Mineral extraction sites	0.00	0.00	0.00	0.14
Permanently irrigated land	2.39	3.22	3.36	3.42
Non-irrigated arable land	3.54	3.90	5.06	5.35
Pastures	2.90	20.31	24.26	22.46
Complex cultivation patterns	0.09	5.01	3.30	3.79
LUAINV	6.33	2.75	6.21	6.49
Natural grasslands	39.43	50.61	40.67	40.99
Transitional woodland-shrub	0.00	0.16	0.19	0.19
Sparsely vegetated areas	33.48	1.28	1.79	1.81
Inland marshes	0.10	0.24	0.22	0.23
Water bodies	11.24	12.06	14.39	14.58

LUAINV: Land used for agriculture with important natural vegetation

Lake Çıldır experienced considerable alterations in its land cover between 2000 and 2018, underscoring a significant anthropogenic impact on the region. This period saw an uptick in urbanization, indicating steady urban development near the lake. Concurrently, mineral extraction sites, which were absent until 2006, accounted for a notable proportion of the land use by 2018. Moreover, a marginal increase was observed in irrigated and non-irrigated agricultural lands, hinting at a broader expansion of agriculture in the region. Similarly, the expansion of pastures was observed, which is likely attributable to intensified livestock farming. Conversely, natural grasslands experienced a slight increment, and there was a discernible increase in water bodies, potentially attributable to land use modifications impacting water runoff. These transformations carry profound implications for the ecosystem of Lake Çıldır. The enlargement of agricultural lands and pastures may escalate nutrient runoff into the lake, thereby heightening the risk of eutrophication, a phenomenon detrimental to aquatic life due to the resultant depletion of oxygen levels in the water. Additionally, the advent of mineral extraction sites could adversely affect water quality, contingent on the extraction techniques employed. The augmentation of water bodies might precipitate modifications to the lake's hydrological regime, thereby influencing its water level and, subsequently, its ecological attributes. Furthermore, the marginal alterations in natural grasslands and the reduction in sparsely vegetated areas may engender shifts in the region's flora and fauna, impacting the lake's ecosystem.

In light of these findings, it is imperative to institute robust conservation and management measures to ameliorate the potential adverse repercussions on the Lake Çıldır ecosystem. Such measures should encompass implementing sustainable agricultural practices to curtail nutrient runoff, oversee and regulate mineral extraction operations, and execute land use planning and management strategies to safeguard natural habitats and biodiversity. Additionally, it is crucial to regularly monitor the lake's water quality and ecological attributes to identify and address any untoward alterations expeditiously.

Ecotourism of Lake Çıldır

Lake Çıldır offers significant potential for ecotourism, with its pristine natural beauty, diverse wildlife, and unique ecological characteristics attracting nature enthusiasts and adventure seekers. The establishment of nature trails around the lake allows visitors to explore the surrounding ecosystems, providing a close observation of its rich flora and fauna. The lake and its vicinity host diverse wildlife, including mammals, reptiles, and amphibians. Observation points facilitate the study and appreciation of these local species. Beyond its natural wonders, Lake Çıldır's ecotourism encompasses cultural experiences. Visitors can engage with local communities, learn about their traditional practices and the cultural significance of the lake, and experience artisanal showcases and traditional food tastings. One of the most unique experiences is walking on the frozen expanse of the lake during winter. However, visitors should be mindful of the conditions and prioritize safety. During the peak of winter, when temperatures plummet, the lake freezes over with ice reaching up to 80 cm in thickness (Çimen & Erginal, 2015). This provides a unique opportunity for ice fishing in an "Eskimo-style." The fish caught are then prepared and served, offering the region an authentic taste. Another popular winter activity is taking horse-drawn sleigh tours on the frozen lake, providing a traditional and delightful experience. Organized every February, the Lake Çıldır Festival sees participation from neighboring countries like Georgia, Azerbaijan, Turkmenistan, and Armenia. The Minstrel Festival coincides with this, making it a prime time for visitors to experience winter tourism and cultural festivities. This attraction resembles a frozen city, featuring an ice tower, green caves of pine leaves, a fish museum representing the Lake Çıldır fish, a fruit museum, a crystal cave, and a ghost house. Photographs from the lake Çıldır are shown in Figure 8. Visiting the Ice City, especially when illuminated at night, allows visitors to explore polarly (Çıldır District Governorate, 2023; Sezer, 2022).

Lake Çıldır is a testament to nature's splendor, rich cultural heritage, and diverse ecosystems. It presents a unique blend of ecotourism activities, from walking on its frozen expanse to delving into local traditions, making it an invaluable destination for na-



Figure 8. Lake Çıldır a-c. (Çıldır District Governorate, 2023), d. (Zariç, 2023).

ture aficionados and cultural enthusiasts. As the push for sustainable and immersive travel experiences continues to grow, Lake Çıldır undeniably offers an authentic and enriching journey that resonates with the core values of ecotourism.

CHALLENGES FACING LAKE ÇILDIR AND RECOMMENDATIONS FOR SUSTAINABLE MANAGEMENT

Lake Çıldır a pristine aquatic gem in eastern Turkey. Lake Çıldır has faced ecological challenges since the 1990s. Introducing invasive species, specifically the silver Prussian carp (*Carassius gibelio*) and the narrow-clawed crayfish (*Pontastacus leptodactylus*), has threatened the lake's natural balance. Moreover, attempts at introducing aquaculture, like rainbow trout (*Oncorhynchus mykiss*) farming, have further strained native species and raised sustainability issues. The situation has been aggravated by the operational expansion of the Arpaçay Hydroelectric Plant, which has negatively impacted the lake's shallow coasts, essential breeding grounds for native carp (Zengin et al., 2012). As tourism activities burgeon and exotic species make their mark, an exigent need exists to address these issues with sustainable and academically sound solutions. The following section delineates the challenges faced by the lake and provides recommendations for its sustainable management:

Preservation of current state and ecotourism enhancements:

Recommendation: Ecotourism activities around Lake Çıldır should underscore the importance of environmental education and conservation. It is crucial to instill in visitors the essence of ecosystem preservation, minimizing anthropogenic impacts, and ardently supporting local conservation endeavors. In tandem with this, developing eco-friendly accommodations, such as sustainable eco-lodges and camping sites, can significantly boost the sustainability quotient of ecotourism in the region. Infrastructure developments should be meticulously planned and executed, ensuring a minimal ecological footprint and a relentless commitment to conserving the natural landscape.

Exotic and invasive species

The lake has witnessed the incursion of invasive species, such as the Prussian carp and freshwater crayfish, corroborated by local accounts and other studies (Zengin et al., 2012). Prussian carp are recognized for their aggressive colonization of new environments and their ability to establish dominance, often outcompeting the majority of native species in the process. (Ruppert et al., 2017). Nonetheless, environmental, historical, and anthropogenic processes closely affect the alterations in the native fish population in the ecosystems (Anas & Mandrak, 2021). It is imperative to introduce stringent monitoring systems to track the population dynamics of these exotic species. Furthermore, awareness campaigns should be launched to educate the local

populace about the detrimental impacts of introducing non-native species into the lake.

Environmental hygiene and cleanliness:

Robust sanitation and waste management protocols should be implemented. Regular underwater and on the shores, standard clean-up drives will help maintain the lake's pristine nature. Efforts should also be geared towards sensitizing visitors about the importance of leaving no trace.

Promotion of ecotourism activities

The winter charm of Lake Çıldır, characterized by its frozen expanse, offers many sports and recreational activities. The region can garner significant economic benefits by promoting ice tourism, which appeals to adventure enthusiasts and tourists driven by novelty-seeking motivations. This includes activities like sleigh rides, the Golden Horse Festival, ice skating, ice hockey, and ATV tours. However, it is paramount that these activities are conducted with environmental sensitivity to avoid undue strain on the lake's ecosystem.

Regulation and oversight of local enterprises

Stringent guidelines and regulations should be established for local businesses near the lake. Regular audits and checks can ensure compliance with environmental standards, ensuring the lake's health is not compromised for commercial gains.

Educating and empowering the local community

The local community plays a pivotal role in conserving Lake Çıldır. Organizing workshops, seminars, and interactive sessions can impart knowledge and skills to locals, enabling them to champion the cause of lake conservation. We can foster a collective responsibility toward its well-being by ensuring its stake in the lake's future.

In conclusion, while Lake Çıldır faces multifaceted challenges, with strategic interventions and a commitment to sustainability, it can continue to thrive as both a natural wonder and a hub of ecotourism.

CONCLUSION

Lake Çıldır, situated in Eastern Turkey, presents a significant confluence of ecological and cultural richness. The empirical evaluations underscore its superior environmental status within the Aras River basin. However, its pristine conditions have been incrementally perturbed due to anthropogenic intrusions, notably the introduction of invasive species and burgeoning tourism activities. The repercussions of these interventions are manifested not only in the fluctuating trophic state of the lake but also in its altered phytoplankton composition and biomass.

The juxtaposition of ecological challenges and the robust potential for ecotourism necessitates a paradigm shift in the lake's management strategies. To this end, a suite of recommendations has been advanced, pivoting on sustainable eco-tourism, vigilant monitoring of invasive species, enhanced environmental hygiene protocols, judicious oversight of local enterprises, and a pivotal emphasis on community education and empowerment.

It is paramount to accentuate that the sustainable management of Lake Çıldır not only bears implications for its intrinsic ecological integrity but also resonates with the broader tenets of sustainable ecotourism. Embracing a holistic approach that harmoniously integrates conservation with socio-economic aspirations is, thus, imperative for the lake's future trajectory.

Author Contribution Statement: Özgür Eren Zariç, Abuzer Çelekli, and Sidar Yaygır designed the overall review work.

Ethical Approval: There is no need for ethical approval for this study.

Funding Statement: The authors do not declare any funds.

Conflict Of Interest: The authors declare that they have no conflict of interest.

Data Availability Statement: Data used in this study are available from the corresponding author upon reasonable request.

REFERENCES

- Akbulut, A., & Yıldız, K. (2002). The Planktonic Diatoms of Lake Çıldır (Ardahan-Turkey). *Turkish Journal of Botany*, 26(2), 55–75.
- Akbulut, N. E., Bayarı, S., Akbulut, A., Özyurt, N. N., & Sahin, Y. (2022). Rivers of Turkey. In *Rivers of Europe* (pp. 853–882). Elsevier. <https://doi.org/10.1016/B978-0-08-102612-0.00021-3>
- Akgül, G., & Aytaç, Z. (2008). Çıldır ve Aktaş (Ardahan) Gölleri Arasında Kalan Bölgenin Florası. *The Herb Journal of Systematic Botany*, 15, 37–70.
- AlgaeBase. (2023). *AlgaeBase*. Retrieved from <https://www.algaebase.org/> (accessed 16.06.23)
- Ali, H., & Khan, E. (2019). Trophic transfer, bioaccumulation, and biomagnification of non-essential hazardous heavy metals and metalloids in food chains/webs—Concepts and implications for wildlife and human health. *Human and Ecological Risk Assessment*, 25(6), 1353–1376. <https://doi.org/10.1080/10807039.2018.1469398>
- Alkan, A., Gökçek, Ç., Akbas, U., & Alkan, N. (2016). Spatial Distributions of Heavy Metals in the Water and Sediments of Lake Çıldır, Turkey. *Ekoloji*, 25(98), 9–16. <https://doi.org/10.5053/ekoloji.2015.23>
- Anas, M. M., & Mandrak, N. E. (2021). Drivers of native and non-native freshwater fish richness across North America: Disentangling the roles of environmental, historical and anthropogenic factors. *Global Ecology and Biogeography*, 30(6), 1232–1244.
- Armağan, M., Özgökçe, F., & Çelik, A. (2017). Notes on the genus *Gypsophila* (Caryophyllaceae) in Turkey, with a description of *G. guvengorkii* sp. nov. *Phytotaxa*, 295(3), 271–279. <https://doi.org/10.11646/phytotaxa.295.3.8>
- Arslan, N., & Mercan, D. (2020). The aquatic oligochaete fauna of Lake Çıldır, Ardahan-Kars, Turkey, including an updated checklist of freshwater annelids known to occur in the country. *Zoosymposia*, 17(1), 53–76. <https://doi.org/10.11646/zoosymposia.17.1.8>
- Atalay, İ., Saydam, C., Kadir, S., & Eren, M. (2018). *Pedogeomorphology* (S. Kapur, E. Akça, & H. Günel (eds.); pp. 75–103). Springer International Publishing. https://doi.org/10.1007/978-3-319-64392-2_6
- Aykır, D., & Fiçıcı, M. (2022). Çıldır Gölü Havzasında Erozyon Risk Analizi. *Jeomorfolojik Araştırmalar Dergisi*, 9, 38–49. <https://doi.org/10.46453/jader.1144699>
- Azizoğlu, E., & Adizel, Ö. (2020). Ornithofauna of Ardahan Province (Turkey). *Bitlis Eren Üniversitesi Fen Bilimleri Dergisi*, 9(2), 711–718. <https://doi.org/10.17798/bitlisfen.684942>
- Başçınar, N. S., Düzgüneş, E., Mısır, D. S., Polat, H., & Zengin, B. (2009). Growth and flesh yield of the swan mussel [*Anodonta cygnea*

- (Linnaeus, 1758)] (*Bivalvia*: Unionidae) in Lake Çıldır (Kars, Turkey). *Turkish Journal of Fisheries and Aquatic Sciences*, 9(2), 127–132. <https://doi.org/10.4194/trjfas.2009.0201>
- Beck, M. W., & Hatch, L. K. (2009). A review of research on the development of lake indices of biotic integrity. *Environmental Reviews*, 17(NA), 21–44. <https://doi.org/10.1139/A09-001>
- Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., & West, G. B. (2004). Toward a metabolic theory of ecology. *Ecology*, 85(7), 1771–1789. <https://doi.org/10.1890/03-9000>
- Çakır, N., & Çiçek, E. (2015). Çıldır Gölü *Acanthobrama microlepis* (De filippi, 1863) popülasyonu için bazı popülasyon dinamiği parametrelerinin belirlenmesi [Master's Thesis]. Nevşehir Hacı Bektaş Veli University.
- Carlson, R. E. (1977). A trophic state index for lakes. *Limnology and Oceanography*, 22(2), 361–369. <https://doi.org/10.4319/lo.1977.22.2.0361>
- Çelekli, A. (2016). Guide Document of Phytoplankton Indexes. Directorate General for Water Management of the Ministry of Forestry and Water Affairs Republic of Turkey, Ankara.
- Çelekli, A., Kayhan, S., & Çetin, T. (2020). First assessment of lakes' water quality in Aras River catchment (Turkey); Application of phytoplankton metrics and multivariate approach. *Ecological Indicators*, 117, 106706. <https://doi.org/10.1016/j.ecolind.2020.106706>
- Çelekli, A., Yaygır, S., & Zariç, Ö. E. (2023). A review of climate change-induced migration. *Acta Biologica Turcica*, 36(2), 1–3. <https://doi.org/10.5281/zenodo.8190755>
- Çiçek, E., & Bircikligil, S. S. (2016). Ichthyofauna of the Turkish parts of Kura-Aras River Basin. *FishTaxa*, 1(1), 14–26.
- Çiçek, E., Seçer, B., Sungur, S., Eagderi, S., & Bahçeci, H. (2022). Length-Weight Relationships and Condition Factors of Eight Exotic Fish Species from Turkey. *Turkish Journal of Water Science and Management*, 6(2), 260–274. <https://doi.org/10.31807/tjwsm.1067360>
- Çimen, H., & Erginal, G. (2015). Donan Göllerin Turizm Potansiyeli: Çıldır Gölü Örneği. *Doğu Karadeniz Bölgesi Sürdürülebilir Turizm Kongresi*, 82.
- Çıldır District Governorate. (2023). *Bird Observation Tower in Lake Çıldır*. Retrieved from <http://cildir.gov.tr/> (accessed 10.08.23)
- Crivelli, A., & Vizi, O. (1981). The Dalmatian pelican, *Pelecanus crispus* Bruch 1832, a recently world-endangered bird species. *Biological Conservation*, 20(4), 297–310. [https://doi.org/10.1016/0006-3207\(81\)90016-1](https://doi.org/10.1016/0006-3207(81)90016-1)
- Huys, R., Oidtmann, B., Pond, M., Goodman, H., & Clark, P. F. (2014). Invasive crayfish and their symbionts in the Greater London area: New data and the fate of *Astacus leptodactylus* in the Serpentine and Long Water Lakes. *Ethology Ecology and Evolution*, 26(2–3), 320–347. <https://doi.org/10.1080/03949370.2014.903433>
- Jouladeh-Roudbar, A., Ghanavi, H. R., & Doadrio, I. (2020). Ichthyofauna from iranian freshwater: Annotated checklist, diagnosis, taxonomy, distribution and conservation assessment. *Zoological Studies*, 59, 1–303. <https://doi.org/10.6620/ZS.2020.59-21>
- Kalous, L., Bohlen, J., Rylková, K., & Petrý, M. (2012). Hidden diversity within the Prussian carp and designation of a neotype for *Carassius gibelio* (Teleostei: Cyprinidae). *Ichthyological Exploration of Freshwaters*, 23(1), 11–18.
- Karaoğlu, M., & Çelim, Ş. (2018). Doğu Anadolu Bölgesi ve İçdir'in Jeolojisi ve Toprak Özellikleri. *Journal of Agriculture*, 1(1), 14–26.
- Koçyiğit, A., & Önder, A. (2018). Çıldır Gölü'ndeki Kerevit (*Astacus leptodactylus* Eschscholtz, 1823) Avcılığının Sosyal, Yapısal Ve Ekonomik Yönden Değerlendirilmesi. *International Journal of Veterinary and Animal Research*, 1(3), 8–12.
- Kükrer, S., Erginal, A. E., Şeker, S., & Karabıyıköğlu, M. (2015). Distribution and environmental risk evaluation of heavy metal in core sediments from Lake Çıldır (NE Turkey). *Environmental Monitoring and Assessment*, 187(7). <https://doi.org/10.1007/s10661-015-4685-1>
- Lahn, E. (1947). Çıldır Gölü ve Hazapın Gölünün jeolojisi hakkında (Kars ili). *Türkiye Jeoloji Bülteni*, 2(1), 113–117.
- Langdon, P. G., Ruiz, Z., Wynne, S., Sayer, C. D., & Davidson, T. A. (2010). Ecological influences on larval chironomid communities in shallow lakes: Implications for palaeolimnological interpretations. *Freshwater Biology*, 55(3), 531–545. <https://doi.org/10.1111/j.1365-2427.2009.02345.x>
- Lenard, T., Ejankowski, W., & Poniewozik, M. (2018). Does an increase in water color intensity affect the lake trophic status and phytoplankton metrics. *Knowledge and Management of Aquatic Ecosystems*, 2018-Janua(419). <https://doi.org/10.1051/kmae/2018035>
- Marchetto, A., Padedda, B. M., Mariani, M. A., Lugliè, A., & Sechi, N. (2009). A numerical index for evaluating phytoplankton response to changes in nutrient levels in deep mediterranean reservoirs. *Journal of Limnology*, 68(1), 106–121. <https://doi.org/10.4081/jlimnol.2009.106>
- Mercan, D., Arslan, N., Çamur Elipek, B., Ertorun, N., & Odabaşı, D. A. (2022). First evaluation of three lakes' (Çıldır, Aktaş and Yaygır) macrozoobenthic community structure in Aras River basin (North Eastern Türkiye). *Aquatic Research*, 5(4), 307–318. <https://doi.org/10.3153/ar22030>
- Özkoç, Ö. Ü. (2020). Kars ilinde üreyen ve geçit yapan kuş türleri. *Turkish Journal of Forestry*, 21(2), 179–187.
- Öztürk, M., Tatlı, Â., Özçelik, H., & Behçet, L. (2015). General Characteristics of Flora and Vegetation Formations of Eastern Anatolia Region and Its Environs (Türkiye). *SDU Journal of Science (E-Journal)*, 10(1), 23–48.
- Poniewozik, M., & Lenard, T. (2022). Phytoplankton Composition and Ecological Status of Lakes with Cyanobacteria Dominance. *International Journal of Environmental Research and Public Health*, 19(7). <https://doi.org/10.3390/ijerph19073832>
- POWO. (2023). *Plants of the World Online (POWO)*. Plants of the World Online. Facilitated by the Royal Botanic Gardens, Kew. Retrieved from <http://www.plantsoftheworldonline.org> (accessed 10.08.23)
- Ruppert, J. L. W., Docherty, C., Neufeld, K., Hamilton, K., MacPherson, L., & Poesch, M. S. (2017). Native freshwater species get out of the way: Prussian carp (*Carassius gibelio*) impacts both fish and benthic invertebrate communities in North America. *Royal Society Open Science*, 4(10). <https://doi.org/10.1098/rsos.170400>
- Savran, A. (2021). *Lathyrus karsianus* [photograph]. *Türkiyebitkileri*. Retrieved from <https://turkiyebitkileri.com/> (accessed 10.08.23)
- Sezer, B. (2022). *Assessment of Ecotourism Potential in Çıldır District and its Surroundings in Ardahan Province*. Necmettin Erbakan University.
- Şimşek, O. (2019). Some Geographical Observations In Çıldır And Aktaş Lakes. *Belgü*, 4, 103–127.
- The Fishes of North Carolina. (2023). *The Fishes of North Carolina*. The Fishes of North Carolina. <https://doi.org/10.7868/S0042875218020157>
- Turkish State Meteorological Service. (2018). *Meteoroloji Genel Müdürlüğü*. Turkish State Meteorological Service.
- Yerli, S. V. (1993). The assessment of fish stocks in Çıldır Lake. *Aquaculture Studies*, 2005(4).
- Yerli, S. V. (1997). An Investigation on The Growth Criterias of *Cyprinus carpio* Linnaeus 1758 in Çıldır Lake-Ardahan. *Turkish Journal of Zoology*, 21(1), 91–99. <https://doi.org/10.55730/1300-0179.2996>
- Yılmaz, H., Mutlu, E., Mutlu, B., Demir, M., & Irmak, A. (2018). Çıldır Gölü'nün Rekreatyonel Turizm Potansiyelinin Belirlenmesi. *International Symposium on Urbanization and Environmental Problems; Eskişehir, Turkey*, 282-288.
- Zariç, Ö. E., Yeşildağ, İ., Yaygır, S., & Çelekli, A. (2022). Removal of Harmful Dyes Using Some Algae. *3rd International Congress on Plant Biology; Rize, Turkey, 1st Edition*, 173. <https://doi.org/10.5281/zenodo.8190776>
- Zariç, Ö. E. (2023). Lake Çıldır [photograph].
- Zengin, M., Yerli, S. V., Dağtekin, M., Ö Akpınar, İ., Ürünleri Merkez Araştırma Enstitüsü, S., Üniversitesi, H., Bölümü, B., & Ana Bilim Dalı, H. (2012). Çıldır Gölü Balıkçılığında Son Yirmi Yılda Meydana Gelen Değişimler. *Eğirdir Su Ürünleri Fakültesi Dergisi*, 8(2), 10–24.

Instructions to Authors

Aquatic Sciences and Engineering is an international, scientific, open access periodical published in accordance with independent, unbiased, and double-blinded peer-review principles. The journal is the official publication of Istanbul University Faculty of Aquatic Sciences and it is published quarterly on January, April, July, and October. The publication language of the journal is English and continues publication since 1987.

Aquatic Sciences and Engineering aims to contribute to the literature by publishing manuscripts at the highest scientific level on all fields of aquatic sciences. The journal publishes original research and review articles that are prepared in accordance with the ethical guidelines.

The scope of the journal includes but not limited to; aquaculture science, aquaculture diseases, feeds, and genetics, ecological interactions, sustainable systems, fisheries development, fisheries science, fishery hydrography, aquatic ecosystem, fisheries management, fishery biology, wild fisheries, ocean fisheries, biology, taxonomy, stock identification, functional morphology freshwater, brackish and marine environment, marine biology, water conservation and sustainability, inland waters protection and management, seafood technology and safety.

The target audience of the journal includes specialists and professionals working and interested in all disciplines of aquatic sciences.

The editorial and publication processes of the journal are shaped in accordance with the guidelines of the Committee on Publication Ethics (COPE), the European Association of Science Editors (EASE), the International Council of Medical Journal Editors (ICMJE), and National Information Standards Organization (NISO). The journal conforms to the Principles of Transparency and Best Practice in Scholarly Publishing (doaj.org/bestpractice).

Originality, high scientific quality, and citation potential are the most important criteria for a manuscript to be accepted for publication. Manuscripts submitted for evaluation should not have been previously presented or already published in an electronic or printed medium. The journal should be informed of manuscripts that have been submitted to another journal for evaluation and rejected for publication. The submission of previous reviewer reports will expedite the evaluation process. Manuscripts that have been presented in a meeting should be submitted with detailed information on the organization, including the name, date, and location of the organization.

Manuscripts submitted to Aquatic Sciences and Engineering will go through a double-blind peer-review process. Each submission will be reviewed by at least two external, independent peer reviewers who are experts in their fields in order to ensure an unbiased evaluation process. The editorial board will invite an external and independent editor to manage the evaluation processes of manuscripts submitted by editors or by the editorial board members of the journal. The Editor in Chief is the final authority in the decision-making process for all submissions.

An approval of research protocols by the Ethics Committee in accordance with international agreements (World Medical Association Declaration of Helsinki "Ethical Principles for Medical Research Involving Human Subjects," amended in October 2013, www.wma.net) is required for experimental, clinical, and drug studies. If required, ethics committee reports or an equivalent official document will be requested from the authors.

For manuscripts concerning experimental research on humans, a statement should be included that shows the written informed consent of patients and volunteers was obtained following a detailed explanation of the procedures that they may undergo. Information on patient consent, the name of the ethics committee, and the ethics committee approval number should also be stated in the Materials and Methods section of the manuscript. It is the authors' responsibility to carefully protect the patients' anonymity. For photographs that may reveal the identity of the patients, signed releases of the patient or of their legal representative should be enclosed.

Aquatic Sciences and Engineering requires experimental research studies on vertebrates or any regulated invertebrates to comply with relevant institutional, national and/or international guidelines. The journal supports the principles of Basel Declaration (basel-declaration.org) and the guidelines published by International Council for Laboratory Animal Science (ICLAS) (iclas.org). Authors are advised to clearly state their compliance with relevant guidelines.

Aquatic Sciences and Engineering advises authors to comply with IUCN Policy Statement on Research Involving Species at Risk of Extinction and the Convention on the Trade in Endangered Species of Wild Fauna and Flora for research involving plants.

All submissions are screened by a similarity detection software (iThenticate by CrossCheck).

In the event of alleged or suspected research misconduct, e.g., plagiarism, citation manipulation, and data falsification/fabrication, the Editorial Board will follow and act in accordance with COPE guidelines.

Each individual listed as an author should fulfil the authorship criteria recommended by the ICMJE. The ICMJE recommends that authorship be based on the following 4 criteria:

1. Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work; AND
2. Drafting the work or revising it critically for important intellectual content; AND
3. Final approval of the version to be published; AND
4. Agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

In addition to being accountable for the parts of the work he/she has done, an author should be able to identify which co-authors are responsible for specific other parts of the work. In addition, authors should have confidence in the integrity of the contributions of their co-authors.

All those designated as authors should meet all four criteria for authorship, and all who meet the four criteria should be identified as authors. Those who do not meet all four criteria should be acknowledged in the title page of the manuscript.

Aquatic Sciences and Engineering requires corresponding authors to submit a signed and scanned version of the authorship contribution form (available for download through <https://iupress.istanbul.edu.tr/en/journal/ase/home>) during the initial submission process in order to act appropriately on authorship rights and to prevent ghost or honorary authorship. If the editorial board suspects a case of "gift authorship," the submission will be rejected without further review. As part of the submission of the manuscript, the corresponding author should also send a short statement declaring that he/she accepts to undertake all the responsibility for authorship during the submission and review stages of the manuscript.

Conflict of Interest

The journal requires the authors and all individuals taking part in the evaluation process to disclose any existing or potential conflict of interest (such as financial ties, academic commitments, personal relationships, institutional affiliations) that could unduly influence one's responsibilities. To disclose potential conflicts of interest, the ICMJE Potential Conflict of Interest Disclosure Form should be filled in and submitted by authors as explained in the Author Form of the journal. Cases

of a potential conflict of interest are resolved within the scope of COPE Conflict of Interest Flowcharts and ICMJE Conflict of Interest guidelines

Besides conflict of interest, all financial support received to carry out research must be declared while submitting the paper.

The Editorial Board of the journal handles all appeal and complaint cases within the scope of COPE guidelines. In such cases, authors should get in direct contact with the editorial office regarding their appeals and complaints. When needed, an ombudsperson may be assigned to resolve cases that cannot be resolved internally. The Editor in Chief is the final authority in the decision-making process for all appeals and complaints.

Copyright Notice

Authors publishing with the journal retain the copyright to their work licensed under the Creative Commons Attribution-NonCommercial 4.0 International license (CC BY-NC 4.0) (<https://creativecommons.org/licenses/by-nc/4.0/>) and grant the Publisher non-exclusive commercial right to publish the work. CC BY-NC 4.0 license permits unrestricted, non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Open Access Statement

The journal is an open access journal and all content is freely available without charge to the user or his/her institution. Except for commercial purposes, users are allowed to read, download, copy, print, search, or link to the full texts of the articles in this journal without asking prior permission from the publisher or the author. This is in accordance with the BOAI definition of open access.

The open access articles in the journal are licensed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0) license.

PUBLISHERS ETHIC RULES

The Aquatic Sciences and Engineering is committed to upholding the highest standards of publication ethics and takes all possible measures against any publication malpractices. All authors submitting their works to The Aquatic Sciences and Engineering for publication as original articles attest that the submitted works represent their authors' contributions and have not been copied or plagiarized in whole or in part from other works. The authors acknowledge that they have disclosed all and any actual or potential conflicts of interest with their work or partial benefits associated with it. In the same manner, The Aquatic Sciences and Engineering is committed to objective and fair double-blind peer-review of the submitted for publication works and to prevent any actual or potential conflict of interests between the editorial and review personnel and the

reviewed material. is committed to objective and fair double-blind peer-review of the submitted for publication works and to prevent any actual or potential conflict of interests between the editorial and review personnel and the reviewed material.

Further to the above, The Aquatic Sciences and Engineering provides a platform for the open public discussion of the journal contents. To secure accountability and to encourage sincere professional inputs without incivilities the system is set up to require registration and logging for recording of inputs. Some of the website contents will be available without logging but no peer review comments can be posted on the website without the disclosure of the reviewer identity to the journal editors.

It is necessary to agree upon standards of expected ethical behavior for all parties involved in the act of publishing: the author, the journal editor, the peer reviewer and the publisher.

Our ethic statements are based on [COPE's](#) Best Practice Guidelines for Journal Editors.

Publication Decisions

The editor of the TJAS is responsible for deciding which of the articles submitted to the journal should be published. The editor may be guided by the policies of the journal's editorial board and constrained by such legal requirements as shall then be in force regarding libel, copyright infringement and plagiarism. The editor may confer with other editors or reviewers in making this decision.

Fair Play

An editor at any time evaluate manuscripts for their intellectual content without regard to race, gender, sexual orientation, religious belief, ethnic origin, citizenship, or political philosophy of the authors.

Confidentiality

The editor and any editorial staff must not disclose any information about a submitted manuscript to anyone other than the corresponding author, reviewers, potential reviewers, other editorial advisers, and the publisher, as appropriate.

Disclosure and Conflicts of Interest

Unpublished materials disclosed in a submitted manuscript must not be used in an editor's own research without the express written consent of the author.

Research Ethics

An approval of research protocols by the Ethics Committee in accordance with international agreements (World Medical Association Declaration of Helsinki "Ethical Principles for

Medical Research Involving Human Subjects," amended in October 2013, www.wma.net) is required for experimental, clinical, and drug studies. If required, ethics committee reports or an equivalent official document will be requested from the authors.

For manuscripts concerning experimental research on humans, a statement should be included that shows the written informed consent of patients and volunteers was obtained following a detailed explanation of the procedures that they may undergo. Information on patient consent, the name of the ethics committee, and the ethics committee approval number should also be stated in the Materials and Methods section of the manuscript. It is the authors' responsibility to carefully protect the patients' anonymity. For photographs that may reveal the identity of the patients, signed releases of the patient or of their legal representative should be enclosed.

Aquatic Sciences and Engineering requires experimental research studies on vertebrates or any regulated invertebrates to comply with relevant institutional, national and/or international guidelines. The journal supports the principles of Basel Declaration (basel-declaration.org) and the guidelines published by International Council for Laboratory Animal Science (ICLAS) (iclas.org). Authors are advised to clearly state their compliance with relevant guidelines.

Aquatic Sciences and Engineering advises authors to comply with IUCN Policy Statement on Research Involving Species at Risk of Extinction and the Convention on the Trade in Endangered Species of Wild Fauna and Flora for research involving plants.

Human Subjects and Animal Use in Research, Ethics Committee Approval and Informed Consent

The Journal takes as principle to comply with the ethical standards of [World Medical Association \(WMA\) Declaration of Helsinki – Ethical Principles for Medical Research Involving Human Subjects](#) and [WMA Statement on Animal Use in Biomedical Research](#).

An approval of research protocols by the Ethics Committee in accordance with international standards mentioned above is required for experimental, clinical, and drug studies and for some case reports. If required, ethics committee reports or an equivalent official document will be requested from the authors. For manuscripts concerning experimental research on humans, a statement should be included that shows that written informed consent of patients and volunteers was obtained following a detailed explanation of the procedures that they may undergo. For studies carried out on animals, the measures taken to prevent pain and suffering of the animals should be stated clearly. Information on patient consent, the name of the ethics committee, and the ethics committee approval number

should also be stated in the Materials and Methods section of the manuscript. It is the authors' responsibility to carefully protect the patients' anonymity. For photographs that may reveal the identity of the patients, signed releases of the patient or of their legal representative should be enclosed.

Conflict of Interest

The journal requires the authors and all individuals taking part in the evaluation process to disclose any existing or potential conflict of interest (such as financial ties, academic commitments, personal relationships, institutional affiliations) that could unduly influence one's responsibilities. To disclose potential conflicts of interest, the ICMJE Potential Conflict of Interest Disclosure Form should be filled in and submitted by authors as explained in the Author Form of the journal. Cases of a potential conflict of interest are resolved within the scope of COPE Conflict of Interest Flowcharts and ICMJE Conflict of Interest guidelines

Besides conflict of interest, all financial support received to carry out research must be declared while submitting the paper.

Author's Responsibilities

Reporting Standards

Authors of reports of original research should present an accurate account of the work performed as well as an objective discussion of its significance. Underlying data should be represented accurately in the paper. A paper should contain sufficient detail and references to permit others to replicate the work. Fraudulent or knowingly inaccurate statements constitute unethical behavior and are unacceptable.

Data Access and Retention

Authors are asked to provide the raw data in connection with a paper for editorial review, and should be prepared to provide public access to such data (consistent with the ALPSP-STM Statement on Data and Databases), if practicable, and should in any event be prepared to retain such data for a reasonable time after publication.

Originality and Plagiarism

The authors should ensure that they have written entirely original works, and if the authors have used the work and/or words of others that this has been appropriately cited or quoted.

By submitting articles to Turkish Journal of Aquatic Sciences, the author attest the following:

- None of the part of manuscript is plagiarized from other sources

- Proper reference is provided for all contents extracted from other sources

Strong action will be taken against cases of plagiarism

All the papers submitted have to pass through an initial screening and will be checked through the Advanced Plagiarism Detection Softwares (iThenticate, Copyleaks).

Multiple, Redundant or Concurrent Publication

An author should not in general publish manuscripts describing essentially the same research in more than one journal or primary publication. Submitting the same manuscript to more than one journal concurrently constitutes unethical publishing behaviour and is unacceptable.

Acknowledgement of Sources

Proper acknowledgment of the work of others must always be given. Authors should cite publications that have been influential in determining the nature of the reported work.

Authorship of the Paper

Authorship should be limited to those who have made a significant contribution to the conception, design, execution, or interpretation of the reported study. All those who have made significant contributions should be listed as co-authors. Where there are others who have participated in certain substantive aspects of the research project, they should be acknowledged or listed as contributors.

The corresponding author should ensure that all appropriate co-authors and no inappropriate co-authors are included on the paper, and that all co-authors have seen and approved the final version of the paper and have agreed to its submission for publication.

Hazards and Human or Animal Subjects

If the work involves chemicals, procedures or equipment that have any unusual hazards inherent in their use, the author must clearly identify these in the manuscript.

Disclosure and Conflicts of Interest

All authors should disclose in their manuscript any financial or other substantive conflict of interest that might be construed to influence the results or interpretation of their manuscript. All sources of financial support for the project should be disclosed.

Fundamental Errors in Published Works

When an author discovers a significant error or inaccuracy in his/her own published work, it is the author's obligation to promptly

notify the journal editor or publisher and cooperate with the editor to retract or correct the paper.

Responsibility for the Editor and Reviewers

General duties and responsibilities for editor;

- Actively seek the views of authors, readers, reviewers and editorial board members about ways of improving their journal's processes
- Encourage and be aware of research into peer review and 'journalology' and reassess journal processes in the light of new findings
- Work to persuade their publishers to provide them with appropriate resources, guidance from experts (e.g. designers, lawyers) and adequate training to perform their role in a professional manner and raise the quality of their journal
- Support initiatives designed to reduce academic misconduct
- Support initiatives to educate researchers about publication ethics
- Assess the effects of their journal policies on author and reviewer behaviour and revise policies, as required, to encourage responsible behaviour and discourage misconduct
- Ensure that any press releases issued by the journal reflect the message of the reported article and put it into context

Duties of Reviewers;

- **Contribution to Editorial Decisions:** Peer review assists the editor in making editorial decisions and through the editorial communications with the author may also assist the author in improving the paper.
- **Promptness:** Any selected referee who feels unqualified to review the research reported in a manuscript or knows that its prompt review will be impossible should notify the editor and excuse himself from the review process.
- **Confidentiality:** Any manuscripts received for review must be treated as confidential documents. They must not be shown to or discussed with others except as authorized by the editor.
- **Standards of Objectivity:** Reviews should be conducted objectively. Personal criticism of the author is inappropriate. Referees should express their views clearly with supporting arguments.
- **Acknowledgement of Sources:** Reviewers should identify relevant published work that has not been cited by the authors. Any statement that an observation, derivation, or argument

had been previously reported should be accompanied by the relevant citation. A reviewer should also call to the editor's attention any substantial similarity or overlap between the manuscript under consideration and any other published paper of which they have personal knowledge.

- **Disclosure and Conflict of Interest:** Privileged information or ideas obtained through peer review must be kept confidential and not used for personal advantage. Reviewers should not consider manuscripts in which they have conflicts of interest resulting from competitive, collaborative, or other relationships or connections with any of the authors, companies, or institutions connected to the papers.

PEER REVIEW PROCESS

Peer Review Policies

Only those manuscripts approved by its every individual author and that were not published before in or sent to another journal, are accepted for evaluation.

Submitted manuscripts that pass preliminary control are scanned for plagiarism using iThenticate software. After plagiarism check, the eligible ones are evaluated by editor-in-chief for their originality, methodology, the importance of the subject covered and compliance with the journal scope.

The editor hands over the papers matching the formal rules to at least two national/international referees for double-blind peer review evaluation and gives green light for publication upon modification by the authors in accordance with the referees' claims.

Responsibility for the Editor and Reviewers

Editor-in-Chief evaluates manuscripts for their scientific content without regard to ethnic origin, gender, citizenship, religious belief or political philosophy of the authors. Editor-in-Chief provides a fair double-blind peer review of the submitted articles for publication and ensures that all the information related to submitted manuscripts is kept as confidential before publishing.

Editor-in-Chief is responsible for the contents and overall quality of the publication. He/She must publish errata pages or make corrections when needed.

Editor-in-Chief does not allow any conflicts of interest between the authors, editors and reviewers. Only he has the full authority to assign a reviewer and is responsible for final decision for publication of the manuscripts in the Journal.

Reviewers must have no conflict of interest with respect to the research, the authors and/or the research funders. Their judgments must be objective.

Reviewers must ensure that all the information related to submitted manuscripts is kept as confidential and must report to the editor if they are aware of copyright infringement and plagiarism on the author's side.

A reviewer who feels unqualified to review the topic of a manuscript or knows that its prompt review will be impossible should notify the editor and excuse himself from the review process.

Peer Review Process

Only those manuscripts approved by its every individual author and that were not published before in or sent to another journal, are accepted for evaluation.

Submitted manuscripts that pass preliminary control are scanned for plagiarism using iThenticate software. After plagiarism check, the eligible ones are evaluated by Editor-in-Chief for their originality, methodology, the importance of the subject covered and compliance with the journal scope.

Editor-in-Chief evaluates manuscripts for their scientific content without regard to ethnic origin, gender, citizenship, religious belief or political philosophy of the authors and ensures a fair double-blind peer review of the selected manuscripts.

The selected manuscripts are sent to at least two national/international external referees for evaluation and publication decision is given by Editor-in-Chief upon modification by the authors in accordance with the referees' claims.

Editor-in-Chief does not allow any conflicts of interest between the authors, editors and reviewers and is responsible for final decision for publication of the manuscripts in the Journal.

Reviewers' judgments must be objective. Reviewers' comments on the following aspects are expected while conducting the review.

- Does the manuscript contain new and significant information?
- Does the abstract clearly and accurately describe the content of the manuscript?
- Is the problem significant and concisely stated?
- Are the methods described comprehensively?
- Are the interpretations and conclusions justified by the results?
- Is adequate references made to other Works in the field?
- Is the language acceptable?

Reviewers must ensure that all the information related to submitted manuscripts is kept as confidential and must report to the editor if they are aware of copyright infringement and plagiarism on the author's side.

A reviewer who feels unqualified to review the topic of a manuscript or knows that its prompt review will be impossible should notify the editor and excuse himself from the review process.

MANUSCRIPT PREPARATION

The manuscripts should be prepared in accordance with ICMJE-Recommendations for the Conduct, Reporting, Editing, and Publication of Scholarly Work in Medical Journals (updated in December 2017 - <http://www.icmje.org/icmje-recommendations.pdf>). Authors are required to prepare manuscripts in accordance with the CONSORT guidelines for randomized research studies, STROBE guidelines for observational studies, STARD guidelines for studies on diagnostic accuracy, PRISMA guidelines for systematic reviews and meta-analysis, ARRIVE guidelines for experimental animal studies, TREND guidelines for non-randomized studies, and COREQ guidelines for qualitative studies.

Manuscripts can only be submitted through the journal's online manuscript submission and evaluation system, available at <https://dergipark.org.tr/en/journal/507/submission/step/manuscript/new>. Manuscripts submitted via any other medium will not be processed.

Manuscripts submitted to the journal will first go through a technical evaluation process where the editorial office staff will ensure that the manuscript has been prepared and submitted in accordance with the journal's guidelines. Submissions that do not conform to the journal's guidelines will be returned to the submitting author with technical correction requests.

Authors are required to submit the following forms during the initial submission. These are available for download at istanbul.dergipark.gov.tr/ase

- Copyright Agreement Form,
- Author Contributions Form, and
- ICMJE Potential Conflict of Interest Disclosure Form (should be filled in by all contributing authors)

Preparation of the Manuscript

Title page: A separate title page should be submitted with all submissions and this page should include:

- The full title of the manuscript as well as a short title (running head) of no more than 50 characters,
- Name(s), affiliations, and highest academic degree(s) of the author(s) and ORCID ID (orcid.org)
- Grant information and detailed information on the other sources of financial support,
- Name, address, telephone (including the mobile phone number) and fax numbers, and email address of the corresponding author,

- Acknowledgment of the individuals who contributed to the preparation of the manuscript but who do not fulfil the authorship criteria.

Abstract: A Turkish and an English abstract should be submitted with all submissions except for Letters to the Editor. Submitting a Turkish abstract is not compulsory for international authors. Please check Table 1 below for word count specifications.

Keywords: Each submission must be accompanied by a minimum of three to a maximum of six keywords for subject indexing at the end of the abstract.

Manuscript Types

Original Articles: This is the most important type of article since it provides new information based on original research. The main text should contain Introduction, "Materials and Methods", "Result and Discussion", "Conclusion" and "References" sections.

Statistical analysis to support conclusions is usually necessary. Statistical analyses must be conducted in accordance with international statistical reporting standards. Information on statistical analyses should be provided with a separate subheading under the Materials and Methods section and the statistical software that was used during the process must be specified.

Units should be prepared in accordance with the International System of Units (SI).

After the Conclusion section and before references list, information regarding conflict of interest, financial disclosure, ethics committee approval and acknowledgement are given. These information are to be provided in the author form which must be submitted together with the manuscript.

Conflict of interests: When you (or your employer or sponsor) have a financial, commercial, legal or professional relationship with other organizations or people working with them, a conflict of interest may arise that may affect your research. A full description is required when you submit your article to a journal.

Ethics committee approval: Ethical committee approval is routinely requested from every research article based on experiments on living organisms and humans. Sometimes, studies from different countries may not have the approval of the ethics committee, and the authors may argue that they do not need the approval of their work. In such situations, we consult COPE's "Guidance for Editors: Research, Audit and Service Evaluations" document and evaluate the study at the editorial board and decide whether or not it needs approval.

Financial disclosure: If there is any, the institutions that support the research and the agreements with them should be given here.

Acknowledgment: Acknowledgments allow you to thank people and institutions who assist in conducting the research.

Review Articles: Reviews prepared by authors who have extensive knowledge on a particular field and whose scientific background has been translated into a high volume of publications with a high citation potential are welcomed. These authors may even be invited by the journal. Reviews should describe, discuss, and evaluate the current level of knowledge of a topic in researches and should guide future studies. The main text should start with Introduction and end with "Conclusion" and "References" sections. Authors may choose to use any subheading in between those sections.

After the Conclusion section and before references list, information regarding conflict of interest and acknowledgement are given. These information are to be provided in the author form which must be submitted together with the manuscript.

Conflict of interests: When you (or your employer or sponsor) have a financial, commercial, legal or professional relationship with other organizations or people working with them, a conflict of interest may arise that may affect your research. A full description is required when you submit your article to a journal.

Acknowledgment: Acknowledgments allow you to thank people and institutions who assist in conducting the research.

Short Communication: This type of manuscript discusses important parts, overlooked aspects, or lacking parts of a previously published article. Articles on subjects within the scope of the journal that might attract the readers' attention, particularly educative cases, may also be submitted in the form of a "Short Communication". Readers can also present their comments on the published manuscripts in the form of a "Short Communication". The main text should contain Introduction, "Materials and Methods", "Result and Discussion", "Conclusion" and "References" sections.

After the Conclusion section and before references list, information regarding conflict of interest, financial disclosure, ethics committee approval and acknowledgement are given. These information are to be provided in the author form which must be submitted together with the manuscript.

Conflict of interests: When you (or your employer or sponsor) have a financial, commercial, legal or professional relationship with other organizations or people working with them, a conflict of interest may arise that may affect your research. A full description is required when you submit your article to a journal.

Ethics committee approval: Ethical committee approval is routinely requested from every research article based on experiments on living organisms and humans. Sometimes,

studies from different countries may not have the approval of the ethics committee, and the authors may argue that they do not need the approval of their work. In such situations, we consult COPE's "Guidance for Editors: Research, Audit and Service Evaluations" document and evaluate the study at the editorial board and decide whether or not it needs approval.

Financial disclosure: If there is any, the institutions that support the research and the agreements with them should be given here.

Acknowledgment: Acknowledgments allow you to thank people and institutions who assist in conducting the research.

Tables

Tables should be included in the main document, presented after the reference list, and they should be numbered consecutively in the order they are referred to within the main text. A descriptive title must be placed above the tables. Abbreviations used in the tables should be defined below the tables by footnotes (even if they are defined within the main text). Tables should be created using the "insert table" command of the word processing software and they should be arranged clearly to provide easy reading. Data presented in the tables should not be a repetition of the data presented within the main text but should be supporting the main text.

Table 1. Limitations for each manuscript type

Type of manuscript	Page	Abstract word limit	Reference limit
Original Article	≤20	250	40
Review Article	≤25	250	60
Short Communication	≤5	250	20

Figures and Figure Legends

Figures, graphics, and photographs should be submitted as separate files (in TIFF or JPEG format) through the submission system. The files should not be embedded in a Word document or the main document. When there are figure subunits, the subunits should not be merged to form a single image. Each subunit should be submitted separately through the submission system. Images should not be labeled (a, b, c, etc.) to indicate figure subunits. Thick and thin arrows, arrowheads, stars, asterisks, and similar marks can be used on the images to support figure legends. Like the rest of the submission, the figures too should be blind. Any information within the images that may indicate an individual or institution should be blinded. The minimum resolution of each submitted figure should be 300 DPI. To prevent delays in the evaluation process, all submitted figures should be clear in

resolution and large in size (minimum dimensions: 100 × 100 mm). Figure legends should be listed at the end of the main document.

All acronyms and abbreviations used in the manuscript should be defined at first use, both in the abstract and in the main text. The abbreviation should be provided in parentheses following the definition.

When a drug, product, hardware, or software program is mentioned within the main text, product information, including the name of the product, the producer of the product, and city and the country of the company (including the state if in USA), should be provided in parentheses in the following format: "Discovery St PET/CT scanner (General Electric, Milwaukee, WI, USA)"

All references, tables, and figures should be referred to within the main text, and they should be numbered consecutively in the order they are referred to within the main text.

Limitations, drawbacks, and the shortcomings of original articles should be mentioned in the Discussion section before the conclusion paragraph.

References

While citing publications, preference should be given to the latest, most up-to-date publications. If an ahead-of-print publication is cited, the DOI number should be provided. Authors are responsible for the accuracy of references. List references in alphabetical order. Each listed reference should be cited in text, and each text citation should be listed in the References section. The reference styles for different types of publications are presented in the following examples.

Reference Style and Format

Aquatic Sciences and Engineering complies with APA (American Psychological Association) style 6th Edition for referencing and quoting. For more information:

- American Psychological Association. (2010). Publication manual of the American Psychological Association (6th ed.). Washington, DC: APA.
- <http://www.apastyle.org>

Accuracy of citation is the author's responsibility. All references should be cited in text. Reference list must be in alphabetical order. Type references in the style shown below.

Citations in the Text

Citations must be indicated with the author surname and publication year within the parenthesis.

If more than one citation is made within the same parenthesis, separate them with (;).

Samples:

More than one citation;

(Esin et al., 2002; Karasar, 1995)

Citation with one author;

(Akyolcu, 2007)

Citation with two authors;

(Sayiner & Demirci, 2007)

Citation with three, four, five authors;

First citation in the text: (Ailen, Ciambrune, & Welch, 2000)

Subsequent citations in the text: (Ailen et al., 2000)

Citations with more than six authors;

(Çavdar et al., 2003)

Major Citations for a Reference List

Note: All second and third lines in the APA Bibliography should be indented.

- **A book in print:** Baxter, C. (1997). *Race equality in health care and education*. Philadelphia: Ballière Tindall. ISBN 4546465465
- **A book chapter, print version:** Haybron, D. M. (2008). Philosophy and the science of subjective well-being. In M. Eid & R. J. Larsen (Eds.), *The science of subjective well-being* (pp. 17-43). New York, NY: Guilford Press. ISBN 4546469999
- **An eBook:** Millbower, L. (2003). *Show biz training: Fun and effective business training techniques from the worlds of stage, screen, and song*. Retrieved from <http://www.amacombooks.org/> (accessed 10.10.15)
- **An article in a print journal:** Carter, S. & Dunbar-Odom, D. (2009). The converging literacies center: An integrated model for writing programs. *Kairos: A Journal of Rhetoric, Technology, and Pedagogy*, 14(1), 38-48.
- **An article with DOI:** Gaudio, J. L. & Snowdon, C. T. (2008). Spatial cues more salient than color cues in cotton-top tamarins (*saguinus oedipus*) reversal learning. *Journal of Comparative Psychology*, <https://doi.org/10.1037/0735-7036.122.4.441>
- **Websites - professional or personal sites:** The World Famous Hot Dog Site. (1999, July 7). Retrieved January 5, 2008, from <http://www.xroads.com/~tcs/hotdog/hotdog.html> (accessed 10.10.15)

- **Websites - online government publications:** U.S. Department of Justice. (2006, September 10). Trends in violent victimization by age, 1973-2005. Retrieved from <http://www.ojp.usdoj.gov/bjs/glance/vage.htm> (accessed 10.10.15)
- **Photograph (from book, magazine or webpage):** Close, C. (2002). *Ronald*. [photograph]. Museum of Modern Art, New York, NY. Retrieved from http://www.moma.org/collection/object.php?object_id=108890 (accessed 10.10.15)
- **Artwork - from library database:** Clark, L. (c.a. 1960's). *Man with Baby*. [photograph]. George Eastman House, Rochester, NY. Retrieved from ARTstor
- **Artwork - from website:** Close, C. (2002). *Ronald*. [photograph]. Museum of Modern Art, New York. Retrieved from http://www.moma.org/collection/browse_results.php?object_id=108890 (accessed 10.10.15)

REVISIONS

When submitting a revised version of a paper, the author must submit a detailed "Response to the reviewers" that states point by point how each issue raised by the reviewers has been covered and where it can be found (each reviewer's comment, followed by the author's reply and line numbers where the changes have been made) as well as an annotated copy of the main document. Revised manuscripts must be submitted within 30 days from the date of the decision letter. If the revised version of the manuscript is not submitted within the allocated time, the revision option may be canceled. If the submitting author(s) believe that additional time is required, they should request this extension before the initial 30-day period is over.

Accepted manuscripts are copy-edited for grammar, punctuation, and format. Once the publication process of a manuscript is completed, it is published online on the journal's webpage as an ahead-of-print publication before it is included in its scheduled issue. A PDF proof of the accepted manuscript is sent to the corresponding author and their publication approval is requested within 2 days of their receipt of the proof.

Editor in Chief: Prof. Devrim MEMİŞ

Address: İstanbul Üniversitesi Su Bilimleri Fakültesi Yetiştiricilik Anabilim Dalı Ordu Cad. No:8 34134 Laleli / İstanbul, Türkiye

Phone: +90 212 4555700/16448

Fax: +90 212 5140379

E-mail: mdevrim@istanbul.edu.tr

Copyright Agreement Form / Telif Hakkı Anlaşması Formu

İstanbul University
İstanbul Üniversitesi



Dergi Adı: Aquatic Sciences and Engineering
Journal name: Aquatic Sciences and Engineering

Copyright Agreement Form
Telif Hakkı Anlaşması Formu

Responsible/Corresponding Author Sorumlu Yazar	
Title of Manuscript Makalenin Başlığı	
Acceptance date Kabul Tarihi	
List of authors Yazarların Listesi	

Sıra No	Name - Surname Adı-Soyadı	E-mail E-Posta	Signature İmza	Date Tarih
1				
2				
3				
4				
5				

Manuscript Type (Research Article, Review, etc.) Makalenin türü (Araştırma makalesi, Derleme, v.b.)	
---	--

Responsible/Corresponding Author:
Sorumlu Yazar:

University/company/institution	Çalıştığı kurum	
Address	Posta adresi	
E-mail	E-posta	
Phone; mobile phone	Telefon no; GSM no	

The author(s) agrees that:
The manuscript submitted is his/her/their own original work, and has not been plagiarized from any prior work, all authors participated in the work in a substantive way, and are prepared to take public responsibility for the work, all authors have seen and approved the manuscript as submitted, the manuscript has not been published and is not being submitted or considered for publication elsewhere, the text, illustrations, and any other materials included in the manuscript do not infringe upon any existing copyright or other rights of anyone. İSTANBUL UNIVERSITY will publish the content under Creative Commons Attribution-NonCommercial 4.0 International (CC BY-NC 4.0) license that gives permission to copy and redistribute the material in any medium or format other than commercial purposes as well as remix, transform and build upon the material by providing appropriate credit to the original work. The Contributor(s) or, if applicable the Contributor's Employer, retain(s) all proprietary rights in addition to copyright, patent rights. I/We indemnify İSTANBUL UNIVERSITY and the Editors of the Journals, and hold them harmless from any loss, expense or damage occasioned by a claim or suit by a third party for copyright infringement, or any suit arising out of any breach of the foregoing warranties as a result of publication of my/our article. I/We also warrant that the article contains no libelous or unlawful statements, and does not contain material or instructions that might cause harm or injury. This Copyright Agreement Form must be signed/ratified by all authors. Separate copies of the form (completed in full) may be submitted by authors located at different institutions; however, all signatures must be original and authenticated.

Yazar(lar) aşağıdaki hususları kabul eder
Sunulan makalenin yazar(lar)ın orijinal çalışması olduğunu ve intihal yapmadıklarını,
Tüm yazarların bu çalışmaya asli olarak katılmış olduklarını ve bu çalışma için her türlü sorumluluğu aldıklarını,
Tüm yazarların sunulan makalenin son halini gördüklerini ve onayladıklarını,
Makalenin başka bir yerde basılmadığını veya basılmak için sunulmadığını,
Makalede bulunan metnin, şekillerin ve dokümanların diğer şahıslara ait olan Telif Haklarını ihlal etmediğini kabul ve taahhüt ederler.
İSTANBUL ÜNİVERSİTESİ'nin bu fikri eseri, Creative Commons Atıf-GayriTicari 4.0 Uluslararası (CC BY-NC 4.0) lisansı ile yayınlamasına izin verirler. Creative Commons Atıf-GayriTicari 4.0 Uluslararası (CC BY-NC 4.0) lisansı, eserin ticari kullanım dışında her boyut ve formatta paylaşılmasına, kopyalanmasına, çoğaltılmasına ve orijinal esere uygun şekilde atıfta bulunmak kaydıyla yeniden düzenleme, dönüştürme ve eserin üzerine inşaa etme dâhil adapte edilmesine izin verir.
Yazar(lar)ın veya varsa yazar(lar)ın işverenin telif dâhil patent hakları, fikri mülkiyet hakları saklıdır.
Ben/Biz, telif hakkı ihlali nedeniyle üçüncü şahıslara vuku bulacak hak talebi veya açılacak davalarda İSTANBUL ÜNİVERSİTESİ ve Dergi Editörlerinin hiçbir sorumluluğunun olmadığını, tüm sorumluluğun yazarlara ait olduğunu taahhüt ederim/ederiz.
Ayrıca Ben/Biz makalede hiçbir suç unsuru veya kanuna aykırı ifade bulunmadığını, araştırma yapılırken kanuna aykırı herhangi bir malzeme ve yöntem kullanılmadığını taahhüt ederim/ederiz.
Bu Telif Hakkı Anlaşması Formu tüm yazarlar tarafından imzalanmalıdır/onaylanmalıdır. Form farklı kurumlarda bulunan yazarlar tarafından ayrı kopyalar halinde doldurularak sunulabilir. Ancak, tüm imzaların orijinal veya kanıtlanabilir şekilde onaylı olması gerekir.

Responsible/Corresponding Author; Sorumlu Yazar;	Signature / İmza	Date / Tarih
	/...../.....