

RESEARCH ARTICLES

Comparison of Dental Follicle Stem Cells and Dental Pulp Stem Cells in a Translational Bone Tissue Engineering Protocol
Translasyonel Kemik Doku Mühendisliği Protokolünde Diş Folikül Kök Hücreleri ve Diş Pulpa Kök Hücrelerinin Karşılaştırılması

Elif M. Ozcan, Yucel Erbilgin, Selin Yıldırım, Noushin Zibandeh, Tunc Akkoc, Kamil Goker

Comparison of Marginal Bone Loss Between Titanium and Titanium-Zirconium Implants at Least 5 Years at Function
En Az 5 Yıllık Fonksiyon Süresiyle Titanyum ve Titanyum-Zirkonyum İmplantlar Arasındaki Marjinal Kemik Kaybının Karşılaştırılması

Selin Görgündür, Esra Beyler

İliak Kemik Grefti ile Ogmente Edilmiş Kretlere Yerleştirilen İmplantların Uzun Dönem Sağkalımlarının Değerlendirilmesi
Long-Term Survival Assessment of Dental Implants Placed in Atrophic Ridges Augmented with Iliac Bone Grafts

Yusuf Tamer, Seçil Çubuk

CASE REPORTS

Gunshot Injury to The Mandible of a Patient Due to Domestic Violence With Tough Treatment Process: A Case Report
Aile İçi Şiddet Sebebiyle Bir Hastanın Mandibulasına Oluşan Ateşli Silah Yaralanmasının Zorlu Tedavi Süreci: Bir Olgu Raporu

Nihat Akbulut, Aseel Halawa



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



e-ISSN: 2687-5497

EDITORIAL BOARD

Editor-in Chief

M. Ercüment Önder, PhD, Lokman Hekim University, Ankara

Co-Editors

Burcu Baş Akkor, PhD, Ondokuz Mayıs University, Samsun
Nükhet Kütük, PhD, Bezmialem Vakıf University, İstanbul

Section Editors

Craniofacial Deformity Section Editor

Hakan H. Tuz, PhD
Hacettepe University, Ankara

Dentoalveolar Surgery Section Editor

Fatih Mehmet Coşkunes, PhD
İstanbul Sağlık ve Teknoloji University (İSTÜN), İstanbul

Oral Medicine and Pathology Section Editor

Kıvanç Bektaş, PhD
İstanbul University, İstanbul

Tissue and Bioengineering Section Editor

Doğan Dolanmaz, PhD
Bezmialem Vakıf University, İstanbul

TMJ/Facial Pain Section Editor

Umut Tekin, PhD
Sağlık Bilimleri University, Ankara

Trauma Section Editor

Doruk Koçyiğit, PhD
Kırıkkale University, Kırıkkale

Implant Surgery Section Editor

Fethi Atıl, PhD
Mersin University, Mersin

Language Editor

Z. Özgür Pektaş, PhD
Private Practice, Adana

Anesthesia Section Editor

Gözde Nur Erkan, MD
Kırıkkale University, Kırıkkale



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)

EurAsian Journal of
Oral and Maxillofacial Surgery



ADVISORY BOARD

Gülsüm Ak
Cemal Akay
Nihat Akbulut
Sıdıka Sinem Akdeniz
Alper Aktaş
Hilal Alen
Alper Alkan
Serpil Altundoğan
Ayşegül Apaydın
Kenan Araz
Ahmet Hamdi Arslan
Hanife Ataoğlu
Yavuz Sinan Aydıntuğ
Selçuk Basa
Burcu Baş
Gürkan Raşit Bayer
Burak Bayram
Emel Bulut
Mine Cambazoğlu
Ülkem Cilasun
Figen Çizmeci Şenel
Ezher Hamza Dayısoylu
Ömür Dereci
Gühan Dergin
Ercan Durmuş
Özgür Erdoğan
Sertan Ergun
Behçet Erol
Alparslan Esen
Emin Esen
Yavuz Fındık
Hasan Garip
Mehmet Kamil Göker

Onur Gönül
Nurhan Güler
Belgin Gülsün
Sevtap Günbay
Ahmet Muhtar Gürol
Kubilay Işık
Onur İçten
Abdullah Kalaycı
Beyza Kaya
Kıvanç Bektaş Kayhan
Erdem Kılıç
Adnan Kılınç
Reha Kişnişçi
Hülya Koçak Berberoğlu
Gülperi Koçer
Mahmut Koparal
Meltem Koray
Mehmet Kürkçü
Nükhet Kütük
Deniz Gökçe Meral
Asriye Mocan
Mehtap Muğlalı
Bora Özden
Nedim Özer
Özkan Özgül
Aydın Özkan
Nilüfer Özkan
Yaşar Özkan
Alper Pampu
Mustafa Ramazanoğlu
Alp Saruhanoğlu
Turgay Seçkin
Fırat Selvi

Bahar Sezer
Berkay Tolga Süer
Metin Şençimen
Göksel Şimşek Kaya
Hakkı Tanyeri
Fatih Taşkesen
Emre Tosun
Ufuk Toygar Memikoğlu
Funda Tuğcu
Ayşegül Mine Tüzüner
Sina Uçkan
Cem Üngör
Altan Varol
Mehmet Yaltırık
Gülsün Yıldırım Öz
Nergiz Yılmaz
Nuray Yılmaz Altıntaş
Ümit Yolcu

*Board members above are listed alphabetically by surname.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



RESEARCH ARTICLE

Comparison of Dental Follicle Stem Cells and Dental Pulp Stem Cells in a Translational Bone Tissue Engineering Protocol

Translasyonel Kemik Doku Mühendisliği Protokolünde Diş Folikül Kök Hücreleri ve Diş Pulpa Kök Hücrelerinin Karşılaştırılması

Elif M. Ozcan¹ , Yucel Erbilgin² , Selin Yıldırım³ , Noushin Zibandeh⁴ , Tunc Akkoc⁵ , Kamil Goker⁶

¹DDS, PhD, Private Practice, Istanbul, Turkey

²PhD, Department of Genetics, Institute of Experimental Medicine (DETAE) Istanbul University

³MSc, Private Practice, Istanbul, Turkey

⁴MSc, PhD, CAMS Oxford Institute, Nuffield Department of Medicine, University of Oxford

⁵PhD, Prof.Division of Pediatric Allergy-Immunology Marmara University Faculty of Medicine

⁶PhD, Prof. Private Practice, Istanbul, Turkey

ABSTRACT

Purpose: This study aims to establish and refine a translational protocol and compare the osteogenic potential of dental pulp stem cells (DPSC) and dental follicle stem cells (DFSC) on nano mesh containing polycaprolactone (nmPCL) and plain polycaprolactone (m) scaffolds in vitro and contribute the translational medicine protocols in bone regeneration.

Materials and Methods: DPSCs and DFSCs were osteogenically differentiated on plain polycaprolactone (m) and nano mesh containing polycaprolactone (nm) scaffolds and four groups were examined for cell proliferation and type I collagen formation rates after two weeks of culture. Following immunofluorescence labeling, nonparametric (Kruskal Wallis) and multiple comparison tests were used to compare the groups.

Results: Among all groups, mean cell counts on scaffolds ranged from 30.8 to 82.6 cells/0.0915 mm², and total collagen formation ranged from 2.79% to 17.9%. DFSC and nmPCL complex showed significantly higher cell counts ($p<0.01$) and collagen formation rates ($p<0.01$) in comparison to other groups.

Conclusion: The DFSCnm group is found to show superior properties on cell proliferation and bone matrix formation. This complex is a promising tool for maxillofacial tissue engineering applications.

Keywords: Dental Follicle Stem Cell, Dental Pulp Stem Cell, Polycaprolactone, Tissue Engineering, Bone, Translational Medicine

ÖZET

Amaç: Bu çalışmada, DPSC ve DFSC'lerin nanomesh içeren (nmPCL) ve düz (PCL) polikaprolakton iskeleler üzerindeki osteojenik potansiyellerini in vitro olarak karşılaştırmak ve kemik rejenerasyonunda translasyonel tıp protokollerine katkıda bulunmak amaçlanmıştır.

Gereç ve Yöntemler: DPSC'ler ve DFSC'ler PCL ve nmPCL iskeleleri üzerinde osteojenik olarak farklılaştırılmış ve iki haftalık kültür sonrasında dört grup hücre proliferasyonu ve tip I kollajen oluşum oranları açısından incelenmiştir. İmmüno Floresan etiketlemenin ardından, dört grubu karşılaştırmak için parametrik olmayan (Kruskal Wallis) ve çoklu karşılaştırma testleri kullanılmıştır.

Sonuçlar: Tüm gruplar arasında, iskelelerdeki ortalama hücre sayıları 30.8 ila 82.6 hücre/0.0915 mm² arasında ve toplam kollajen oluşumu %2.79 ila %17.9 arasında değişmektedir. DFSC ve nmPCL kompleksi diğer gruplara kıyasla anlamlı olarak daha yüksek hücre sayısı ($p<0.01$) ve kollajen oluşum oranları ($p<0.01$) göstermiştir.

Sonuç: DFSC/nmPCL grubunun hücre proliferasyonu ve kemik matriksi oluşumu üzerinde üstün özellikler gösterdiği bulunmuştur. Bu kompleks maksillofasial doku mühendisliği uygulamaları için umut verici bir araçtır.

Anahtar Kelimeler: Diş Folikülü Kök Hücresi, Diş Pulpası Kök Hücresi, Polikaprolakton, Doku Mühendisliği, Kemik, Translasyonel Tıp

Submission Date: April 7, 2025
Acceptance Date: May 16, 2025
Corresponding author: Elif Ozcan
Address: Private Practice, Istanbul, Turkey
Phone:
Email: elifmozcan@gmail.com

Elif M. Ozcan 0000-0002-4435-5124
Yucel Erbilgin 0009-0005-3691-188X
Selin Yıldırım 0009-0003-3096-2825
Noushin Zibandeh, 0000-0002-4078-8029
Tunc Akkoc 0000-0001-9179-2805
Kamil Goker 0009-0005-4600-0721



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



Repair and regeneration of bone defects is one of the main research areas of maxillofacial surgery. Bone tissue diseases, injuries and congenital malformations often require treatment with grafting approaches. Allogenic, xenogenic, alloplastic and synthetic materials, as well as autogenous bone grafts, are currently used for reconstruction of maxillofacial complex. Providing the most efficient vascularization and regeneration of bone tissue within the defect area is one of the main objectives for craniofacial tissue engineering. Although autologous bone grafts are accepted as the gold standard for reconstruction of bone defects, a limited amount can be considered as an important problem. Since autologous bone graft harvesting also has the disadvantage of donor site morbidity and all the other materials lack osteogenic potential, tissue engineering methods need to be studied as an alternative to conventional grafting.¹ With the aim of efficient reconstruction of large defects, studies on tissue scaffolds have gained importance in recent years.^{2,3}

Polycaprolactone (PCL) is a biodegradable and biocompatible synthetic material that is reported to have suitable chemical and physical properties in osteogenic differentiation and bone tissue engineering research.⁴ With the goal of autologous bone regeneration, allogeneic and xenogeneic stem cell transplantation, have been studied and obtained successful results.⁵⁻⁷ However the success of treatment also varies according to the characteristics of the tissue scaffolds and type and source of transplanted stem cells^{8,9}. Evaluation of the osteogenic differentiation and regeneration capacity of stem cell colony types is an important criterion in the specific reconstruction of bone defects.¹⁰

Stem cells have been identified as clonogenic cells which have the ability to self-renewal, differentiating into various cell types and forming new cell lines.¹¹ In the area of bone tissue engineering, bone-marrow derived stem cells are one of the most reported sources in the literature.^{3,12,13} Due to the lack of donor site morbidity and the presence of strong osteogenic potential, investigations on dental pulp (DPSC) and dental follicle (DFSC) derived stem cells are recently increased and satisfactory results on bone regeneration are reported.¹⁴⁻¹⁹

In order to apply successful clinical translation of the tissue engineering techniques, tissue specific differentiation potential of stem cell types is needed to be carefully evaluated. The studies that directly compare the results of osteogenic potential of different mesenchymal stem cell colony types are

rare in the literature.

Instead of bone marrow-derived cell lines, mesenchymal stem cell (MSC) colonies that are cultured in this study can be obtained from tissues that will be discarded as medical waste at the end of an oral surgical operation. Hence, clinical translation of DPSC and DFSC studies will result in improved patient comfort by eliminating donor site morbidity and complications.

The purpose of our study is to compare the osteogenic potential of DPSC and DFSC colonies which are cultured on polycaprolactone mesh and polycaprolactone nanomesh scaffolds.

MATERIALS AND METHODS

1. Study Groups

For 10 scaffolds for each of four different groups (DPSCm, DPSCnm, DFSCm, DFSCnm), a total of 40 scaffolds were examined. For each group, two additional scaffolds were cultured as negative controls.

2. Isolation and Expansion of DPSCs and DFSCs

This comparative experimental study was approved by Medipol University Research Ethics Committee (number of approval: 10840098-153). The authors have read the Helsinki Declaration and have followed the guidelines in this investigation. A fully impacted wisdom tooth with its follicle was extracted in aseptic conditions and cracked in sterile conditions with an osteotome. Removed pulp tissue and follicle tissues were finely minced with a scalpel and transported into a 15 ml falcon tube. 2 ml of collagenase type I was added (1:500, ab 34710; Abcam, Cambridge, UK) to each sample and mixture was incubated at 37°C for 1 hour. Samples were then filtered through a 70 µm cell strainer and washed two times with equal volumes of phosphate buffered saline (PBS). After washing, supernatant was removed and 1 ml of culture medium was added to the pellet. Then, cells were seeded into the cell culture flasks and incubated. Culture media was changed every three days and cells were passaged when 80% of confluency was achieved.

3. Characterization of DPSCs and DFSCs

Analyses were performed in every sub-culture from passage 1 to passage 5 using flow cytometry. Characterization of the DPSCs and DFSCs was done with regard to described



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



characteristic MSC markers including CD45, CD14, CD34, CD25, CD28, CD105, CD146, CD90, CD73 and CD29.

For osteogenic differentiation, culture medium was supplemented by 50 µg/ml ascorbic acid (Sigma, USA), 10 mm β-glycerophosphate (Sigma, USA) 100 nmol/L dexamethasone (Sigma, USA). In the fourth week of culture, samples were stained with Alizarin red to examine the formation of mineralized nodules within the culture. For differentiation into adipocytes, cells were cultured in adipogenic differentiation media. Adipocytes were identified by inverted microscopy examination and Oil red O (Sigma, USA) staining. For differentiation into chondrocytes, culture medium was supplemented with chondrogenic differentiation media. Chondrocytes were characterized by Alican blue staining.

4. Scaffolds, Cell Seeding and Culture

Mesenchymal stem cells were seeded on sterile polycaprolactone scaffolds (3D Biotek, USA). Before the cell seeding procedure, cell colonies were washed with PBS (phosphate buffer saline), and incubated with alpha-MEM with 10% fetal calf serum. Then, DPSC and DFSC suspensions were seeded into the 5x1.5 mm PCL mesh (m) and 5x1 mm PCL nanomesh (nm) scaffolds in 96 well plates. For 10 scaffolds for each of four different groups (DPSCm, DPSCnm, DFSCm, DFSCnm), total of 40 scaffolds were examined. For each group, two additional scaffolds were cultured as negative controls. A number of 2.0×10^5 cells in 25 µl of suspension were transferred into each well. For higher seeding efficiency, careful manipulation was applied in order to avoid the contact of the pipette tip with the walls of the wells. After three hours of incubation in 5% CO₂ and 37° C, 175 µl of medium containing 10% FCS and 1% Penstrep was added to the wells. After examining the cell morphology by microscopic examination, cells were taken into the incubator. After adhesion of the cells to the tissue scaffolds for two days, cell-scaffold complexes were supplemented with alpha-MEM, gentamycin (50 µg/ml) and 15% FCS containing 50 µg/ml ascorbic acid (Sigma, USA), 10 mm β-glycerophosphate (Sigma, USA) 100 nmol/L dexamethasone (Sigma, USA) for osteogenic differentiation. Culture medium was changed in every 2 or 3 day of intervals. Cell-scaffold complexes were incubated in 5% CO₂ and 37° C. On 14th day of culture, scaffolds were fixed for immunofluorescence staining.

5. Type I Collagen Formation and Cell Count Analysis

Scaffolds were fixed in 0.05% PFA + 4° C for overnight, then washed with PBS for 2 minutes. After incubation with 70% cold ethanol for a period of 15 minutes, permeabilization was performed for 15 minutes with 0.1% PBS Tween. Cells were blocked with 10% goat serum for 1 hour. Scaffolds were incubated with primary antibody (1: 500 by rabbit anti-collagen I, EU 34710; Abcam, Cambridge, UK) for overnight at + 4° C. A conjugated goat anti-rabbit IgH antibody (DyLight488, Abcam, Cambridge, UK) was used as the secondary antibody. Nuclear staining was performed with 4',6-diamidino-2-phenylindole (DAPI, Sigma) for 5 minutes. Examples were maintained at 4° C until examination. For total cell count, seven representative images were captured using a phase contrast fluorescent microscope at 20x and 40x magnification (Leica, Germany). For each scaffold, seven random representative sections were obtained by confocal laser scanning microscopy (CLSM) (Leica, Germany). The total area of the collagen formation was quantified and total cell counts were calculated for each slide with ImageJ software (National Institutes of Health, Bethesda, MD) and also confirmed manually by the same observer.

6. Statistical Analysis

Datasets were analyzed by using GraphPad Prism 5 software (GraphPad Software, Inc., CA, USA). Comparisons of the multiple groups were performed with nonparametric Kruskal-Wallis test and Bonferroni correction was used when comparing the groups. For all analyses, a P value less than 0.05 was considered statistically significant.

RESULTS

1. Characterization of DPSCs and DFSCs

Characterization of DPSCs was performed using flow cytometry analysis and differentiation potentials of the cell colonies. Flow cytometric analysis demonstrated that DPSCs express stem cell markers CD73, CD90, CD105, CD146 and CD29 and do not express hematopoietic cell markers CD45, CD34, CD25, CD28 and CD14 on their cell surface. With supplementation of the culture by the according differentiation media, isolated DPSC colonies were shown to be capable of in vitro adipogenic, chondrogenic and osteogenic differentiation.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



2. Cell Counts

At the conclusion of the culture period, no deformation was observed in the scaffold structures. Immediately following the seeding process, the cells exhibited a round morphology.

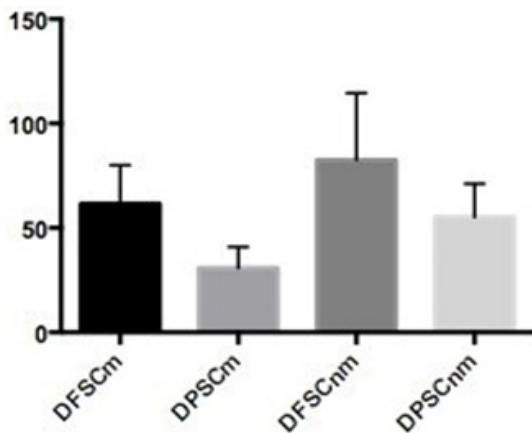


Fig. 1. Cell counts were significantly higher in DFSCnm group.

However, as the culture progressed, a transition to a spindle-like morphology was observed. This change occurred in response to the initiation of adhesion and tissue bridge organization on the PCL fibers.

The mean cell count values for the group DPSCm, DFSCm, DPSCnm, and DFSCnm were 30.8 cells/ 0.0915 mm², 61.6 cells/ 0.0915 mm², 55.2 cells/ 0.0915 mm², and 82.6 cells/ 0.0915 mm², respectively (Fig. 1-3).

The results of the statistical analysis indicated that the DFSCnm group exhibited a statistically significant increase ($p<.001$) in cell count compared to the other groups. Furthermore, observations of DAPI-stained samples revealed the most frequent and uniform cell organization throughout the DFSCnm group. A comparative analysis of the cell counts in the DPSCnm/DFSCnm, DPSCm/DPSCnm, and DPSCm/DFSCm groups was performed. The analysis revealed statistically significantly higher cell counts in the latter groups ($p<.05$). An analysis of the data yielded no statistically significant differences between the DFSCm and DFSCnm groups.

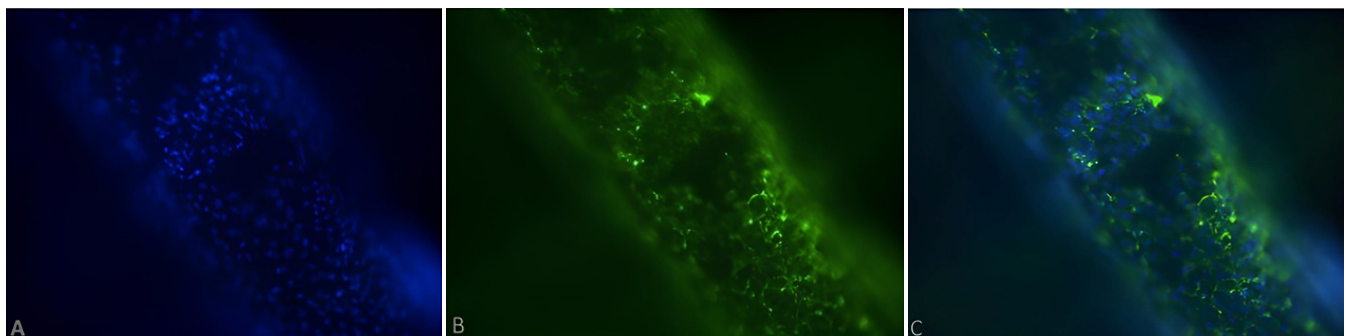


Fig. 2. DAPI (A), Type I Collagen (B) and merged (C) images of DFSCs on PCL fiber structure (X20).

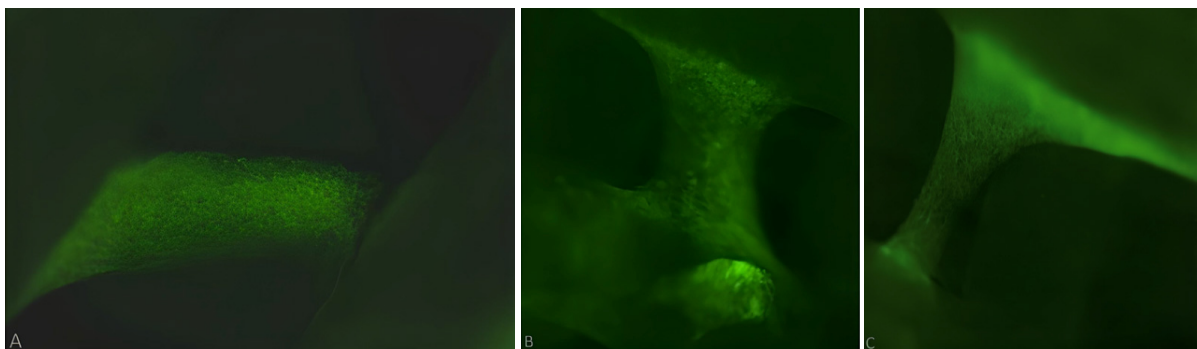


Fig. 3. A-C: Tissue bridging formations of DFSCs on nanomesh scaffold structures.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



3. Type I Collagen Formation

The mean values of collagen formation ratio for the DPSCm, DFSCm, DPSCnm, and DFSCnm groups were 2.79%, 3.93%, 12.7%, and 17.9%, respectively. In the statistical analysis, the rate of type I collagen formation in the DFSCnm group was

found to be significantly higher than in the other groups ($p < .001$). Evaluation via confocal microscopic analysis revealed that the DFSCnm group exhibited the most profound and uniform distribution of collagen organization (Fig. 4-6). A comparison of type I collagen formation rates in DPSCm/DPSCnm and DFSCm/DFSCnm groups revealed statistically significantly higher rates in the latter groups ($p < 0.05$). The comparison of the groups, DFSCnm/DPSCnm and DFSCm/DPSCm, revealed no significant statistical difference.

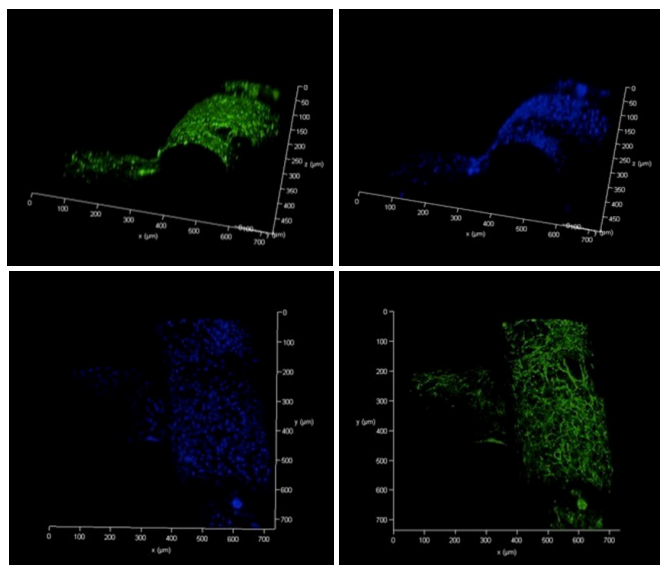


Fig. 4. DAPI (B, C) and Type I Collagen (A, D) confocal microscopy images of tissue bridge forming DFSC lines on nanomesh PCL fibers.

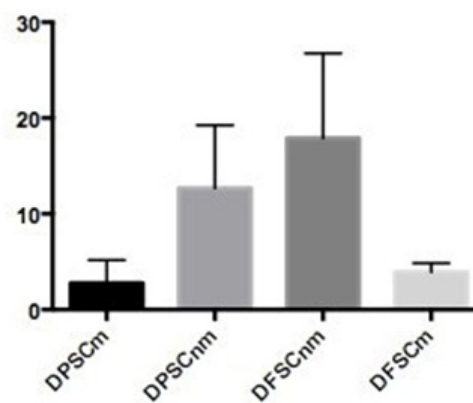


Fig. 5. Type I collagen formation rates were significantly higher in DFSCnm group.

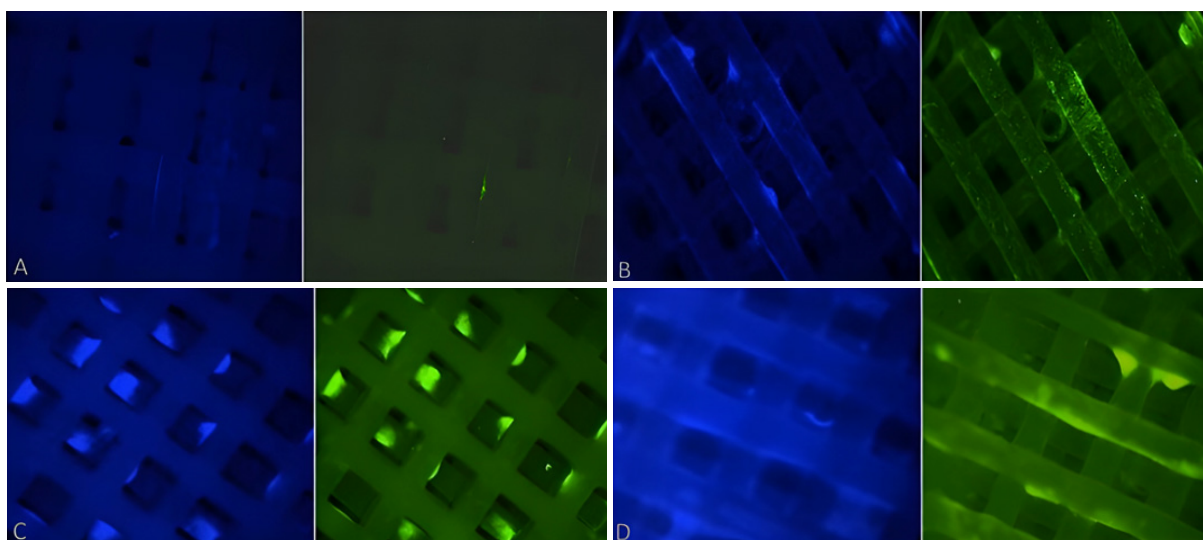


Fig. 6. Immunofluorescence microscopy images of (A) DPSC, (B) DFSC, (C) DPSCnm and (D) DFSCnm groups (X4). DAPI (left) and Type I Collagen (right).



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



DISCUSSION

In the context of bone tissue engineering, the successful organization of bone tissue in qualitative and quantitative aspects is critically dependent on the utilization of scaffold materials to provide support for cells and expressed extracellular matrix. Also, these materials are able to provide guidance to the geometric shape of the tissue growth. The most important task of the material to be used as scaffold is to help fulfill tissue function by providing a temporary support to the cell colonies and an environment that will create the biological orientation of the cells.^{4,20-23} Abukawa et al. studied pig mandibular condyle reconstruction by culturing mesenchymal stem cell colonies on PLGA (polylactic-co-glycolic acid) scaffolds, and reported that the bone formation was observed only on the surface of the construct²⁴. In a subsequent study, Abukawa et al. cultivated porcine bone marrow stem cells on a channeled PLGA scaffold for a period of 10 days. The cells were then implanted into a porcine mandibular critical-sized defect. In the second, fourth, and sixth weeks of healing, histologic sections were obtained, and there was observation of more intensive, uniform, and highly vascularized bone formation on channeled-PLGA scaffolds. In the case of non-channeled scaffolds, the formation of bone tissue was observed exclusively on the surface area. The addition of channels and micropores to the scaffold structure was found to enhance the permeability and transport of the culture medium, cell number and cell distribution, thereby facilitating the formation of bone tissue. In addition, the maximum cell count was documented to occur during the second week of the culture, and a substantial decline in cell counts after this period was reported.²⁵

Scaglione et al. indicated the significance of total porosity, fully connected interior structure, and chemical composition of a scaffold. They further suggested a new "open-pore" tissue scaffold architecture. The formation of tissue and vascular infiltration in both in vitro and in vivo models was found to be satisfactory when using mesh-formed, calcium phosphate-coated hydroxyapatite polymer structures.²⁶

Polycaprolactone structures can be made by three-dimensional fabricating technologies without exposure to chemical solvent materials. Porter et al. evaluated short term biocompatibility and long-term bioactivity assays of PCL nanomesh structures that were produced with three-dimensional printing methods.

In this study, the cultivation of rat bone marrow-derived mesenchymal stem cells on PCL structures was conducted. The results indicated that there was an enhancement in cell adhesion, viability, and elevated levels of bone tissue biochemical markers on nano surfaces during the first, second, and third weeks in comparison to the control groups. In this study, the findings suggest that three-dimensionally printed, solvent-free PCL scaffolds have a positive impact on the biological performance of mesenchymal stem cells and can be utilized as an effective form of tissue scaffolds for bone regeneration.²⁷

Binulal et al. evaluated the adhesion and proliferation potential of human mesenchymal stem cells on nanofibrous and microfibrous electro-spun PCL scaffolds. Adhesion, organization, proliferation and osteoblastic differentiation features of the stem cells were observed to be superior on nanofibrous structures.²⁸

After the implantation of a cultured cell-scaffold complex into a defect area, viability and successful fusion depends on the angiogenesis activity within the first three days. In in-vitro conditions, mineralization of the extracellular matrix and formation of bone nodules is observed from the beginning of the fourth week of culture. In the studies on bone tissue engineering that have been documented, the cell-scaffold complex is predominantly indicated to be implanted subsequent to the observation of mineralization in the tissues. In the present study, the interconnected multilayer mesh scaffold design and culture duration were selected to ensure optimal nutrient and oxygen diffusion, vascular penetration, and uniform bone formation, thereby facilitating the clinical translation of the technique.

Jensen et al. compared three scaffold models for osteogenic differentiation of DPSCs on the 1st, 7th, 14th and 21st days. Cell proliferation, migration, osteoblastic activity and calcium deposition were observed to be increased at day 21 in nano-structure hyaluronic acid / TCP modified scaffold group when compared to control group. According to the results, DPSC/PCL scaffold complexes were stated to be a suitable implementation method for in vivo bone regeneration studies²⁹.

Studies that directly compare the osteogenic potential of DPSCs and DFSCs are rare in the literature. Shoi et al. evaluated the cell proliferation, colony forming capacity, gene expression,



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



cell surface markers and differentiation capacity of DPSCs and DFSCs isolated from supernumerary incisors. Due to the increased obtainable tissue amount, DFSCs are indicated to be a more accessible stem cell source for isolation protocols. The rate of cell proliferation and colony forming capacity of DFSCs were found to be significantly higher in comparison to the DPSCs. In the appropriate culture medium, osteogenic differentiation potential of both cell lines was shown. Despite the similar stem cell characteristics of the DPSCs and DFSCs, due to easier access and higher proliferation rate of DFSC, it is indicated that DFSCs are a more favorable source of stem cells in regenerative applications.³⁰⁻³³

Surface topography is one of the main factors in determination of the differentiation of mesenchymal stem cell lines. This process is based on cell-cell, cell-extracellular matrix and cell-biomaterial interactions via signaling mechanisms.³⁴ Osteogenic differentiation is reported to be more effective on fibrillar nanostructured constructs.³⁵⁻³⁷

In our study, the highest cell count and type I collagen formation rates were observed in DFSCnm group. The effect of the electro-spun nanomesh base membrane structure was found to have a positive impact on cell spreading, adhesion, and proliferation. The observed variations in morphology, cell counts, and type I collagen expression rates may have resulted from the distinct osteogenic differentiation potentials of the cell types, as well as the asynchronous differentiation of cell lines.

In conclusion, using nanomesh PCL scaffold and DFSC complexes is found to be a suitable and promising method for bone tissue engineering applications. In vitro characteristics of stem cell-tissue scaffold complexes are needed to be correlated with in vivo bone regeneration studies. Based on these results, an experimental orthotopic critical size defect model should be studied in order to elucidate the impact of the technique on in situ osteogenesis.

FUNDING

The authors declare that no funds, grants, or other support were received during the preparation of this manuscript.

COMPETING INTERESTS

The authors have no relevant financial or non-financial interests to disclose.

ETHICAL APPROVAL

This study was performed in line with the principles of the Declaration of Helsinki. This study was approved by Medipol University Research Ethics Committee (number of approval: 10840098-153).

CONSENT TO PARTICIPATE

Informed consent was obtained from individual participants included in the study.

CONSENT TO PUBLISH

Non applicable.

ACKNOWLEDGEMENTS

This article is derived from Elif M. Ozcan's PhD dissertation entitled "Experimental evaluation of the effects of mesenchymal stem cell types on bone regeneration", with registration number 388594, conducted under the supervision of M. Kamil Goker and Co-Supervisor Tunc Akkoc.

REFERENCES

1. Chaliserry EP, Nam SY, Park SH, A. S. Therapeutic potential of dental stem cells. *J Tissue Eng.* 2017 May 23;8:2041731417702531. doi: 10.1177/2041731417702531.
2. Arinze TL, Peter SJ, Archambault MP, van den Bos C, Gordon S, Kraus K et al. Allogeneic mesenchymal stem cells regenerate bone in a critical-sized canine segmental defect. *J. Bone Joint Surg. Am.* 2003 Oct;85(10):1927-35. doi: 10.2106/00004623-200310000-00010.
3. Panetta, N. J., Gupta, D. M., Quarto, N. Longaker, M. T. Mesenchymal cells for skeletal tissue engineering. *Panminerva Med.* 2009 Mar;51(1):25-41.
4. Yang X, Yang F, Walboomers XF, Bian Z, Fan M, Jansen JA. The performance of dental pulp stem cells on nanofibrous PCL/gelatin/nHA scaffolds. *J. Biomed. Mater. Res. A.* 2010 Apr;93(1):247-57. doi: 10.1002/jbm.a.32535.
5. Rezai-Rad M, Bova JF, Orooji M, Pepping J, Qureshi A, Del Piero



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



- F, et al. Evaluation of bone regeneration potential of dental follicle stem cells for treatment of craniofacial defects. *Cytherapy*. 2015 Nov;17(11):1572-81. doi: 10.1016/j.jcyt.2015.07.013.
6. Wongsupa N, Nuntanaranont T, Kamolmattayakul S, Thuaksuban N. Assessment of bone regeneration of a tissue-engineered bone complex using human dental pulp stem cells/poly(ϵ -caprolactone)-biphasic calcium phosphate scaffold constructs in rabbit calvarial defects. *J Mater Sci Mater Med*. 2017 May;28(5):77. doi: 10.1007/s10856-017-5883-x.
7. Jang JY, Park SH, Park JH, Lee BK, Yun JH, Lee B et al. In Vivo Osteogenic Differentiation of Human Dental Pulp Stem Cells Embedded in an Injectable In Vivo-Forming Hydrogel. *Macromol. Biosci*. 2016 Aug;16(8):1158-69. doi: 10.1002/mabi.201600001.
8. Niveditha Sundaram M, Sowmya S, Deepthi S, Bumgardener JD, Jayakumar R. Bilayered construct for simultaneous regeneration of alveolar bone and periodontal ligament. *J Biomed Mater Res B Appl Biomater*. 2016 May;104(4):761-70. doi: 10.1002/jbm.b.33480.
9. Wongsupa N, Nuntanaranont T, Kamolmattayakul S, Thuaksuban N. Biological characteristic effects of human dental pulp stem cells on poly- ϵ caprolactone-biphasic calcium phosphate fabricated scaffolds using modified melt stretching and multilayer deposition. *J. Mater. Sci. Mater. Med*. 2017 Feb;28(2):25. doi: 10.1007/s10856-016-5833-z.
10. Matsubara T, Suardita K, Ishii M, Sugiyama M, Igarashi A, Oda R et al. Alveolar bone marrow as a cell source for regenerative medicine: differences between alveolar and iliac bone marrow stromal cells. *J. Bone Miner. Res*. 2005 Mar;20(3):399-409. doi: 10.1359/JBMR.041117.
11. Rimondini, L, Mele, S. Stem cell technologies for tissue regeneration in dentistry. *Minerva Stomatol*. 2009 Oct;58(10):483-500.
12. Kwan MD, Slater BJ, Wan DC, Longaker MT. Cell-based therapies for skeletal regenerative medicine. *Hum. Mol. Genet*. 2008 Apr 15;17(R1):R93-8. doi: 10.1093/hmg/ddn071.
13. Goel A, Sangwan SS, Siwach RC, Ali AM. Percutaneous bone marrow grafting for the treatment of tibial non-union. *Injury* 2005 Jan;36(1):203-6. doi: 10.1016/j.injury.2004.01.009.
14. Pisciotta A, Riccio M, Carnevale G, Beretti F, Gibellini L, Maraldi T et al. Human serum promotes osteogenic differentiation of human dental pulp stem cells in vitro and in vivo. *PLoS One* 2012;7(11):e50542. doi: 10.1371/journal.pone.0050542.
15. Mori G, Ballini A, Carbone C, Oranger A, Brunetti G, Di Benedetto A et al. Osteogenic differentiation of dental follicle stem cells. *Int. J. Med. Sci*. 2012;9(6):480-7. doi: 10.7150/ijms.4583.
16. Yao S, He H, Gutierrez DL, Rad MR, Liu D, Li C et al. Expression of bone morphogenetic protein-6 in dental follicle stem cells and its effect on osteogenic differentiation. *Cells. Tissues. Organs* 2013;198(6):438-47. doi: 10.1159/000360275.
17. Laino G, Carinci F, Graziano A, d'Aquino R, Lanza V, De Rosa A et al. In vitro bone production using stem cells derived from human dental pulp. *J. Craniofac. Surg*. 2006 May;17(3):511-5. doi: 10.1097/00001665-200605000-00021.
18. Todorovic V, Markovic D, Milošević-Jovčić N, Petakov M, Balint B, Čolić M et al. Dental pulp stem cells: Potential significance in regenerative medicine. *Stomatol. Glas. Srb*. 2008 55(3):170-179. doi: 10.2298/SGS0803170T
19. Chuenjitkuntaworn B, Osathanon T, Nowwarote N, Supaphol P, Pavasant P. The efficacy of polycaprolactone/hydroxyapatite scaffold in combination with mesenchymal stem cells for bone tissue engineering. *J Biomed Mater Res A*. 2016 Jan;104(1):264-71. doi: 10.1002/jbm.a.35558.
20. Nakashima M, Reddi AH. The application of bone morphogenetic proteins to dental tissue engineering. *Nat. Biotechnol*. 2003 Sep;21(9):1025-32. doi: 10.1038/nbt864.
21. Batorsky A, Liao J, Lund AW, Plopper GE, Stegemann JP. Encapsulation of adult human mesenchymal stem cells within collagen-agarose microenvironments. *Biotechnol. Bioeng*. 2005 Nov 20;92(4):492-500. doi: 10.1002/bit.20614.
22. Ravichandran A, Lim J, Chong MSK, Wen F, Liu Y, Pillay YT et al. In vitro cyclic compressive loads potentiate early osteogenic events in engineered bone tissue. *J. Biomed. Mater. Res. - Part B Appl. Biomater*. 2017 Nov;105(8):2366-2375. doi: 10.1002/jbm.b.33772.
23. Flores-Cedillo ML, Alvarado-Estrada KN, Pozos-Guillén AJ, Murguía-Ibarra JS, Vidal MA, Cervantes-Uc JM et al. Multiwall carbon nanotubes/polycaprolactone scaffolds seeded with human dental pulp stem cells for bone tissue regeneration. *J Mater Sci Mater Med*. 2016 Feb;27(2):35. doi: 10.1007/s10856-015-5640-y.
24. Abukawa H, Terai H, Hannouche D, Vacanti JP, Kaban LB, Troulis MJ. Formation of a mandibular condyle in vitro by tissue engineering. *J. Oral Maxillofac. Surg*. 2003 Jan;61(1):94-100. doi: 10.1053/joms.2003.50015.
25. Abukawa H, Shin M, Williams WB, Vacanti JP, Kaban LB, Troulis MJ. Reconstruction of mandibular defects with autologous tissue-engineered bone. *J. Oral Maxillofac. Surg*. 2004 May;62(5):601-6. doi: 10.1016/j.joms.2003.11.010.
26. Scaglione S, Ilengo C, Fato M, Quarto R. Hydroxyapatite-coated polycaprolactone wide mesh as a model of open structure for bone regeneration. *Tissue Eng. Part A* 2009 Jan;15(1):155-63. doi: 10.1089/ten.tea.2007.0410.
27. Porter JR, Henson A, Popat KC. Biodegradable poly(ϵ -caprolactone) nanowires for bone tissue engineering applications. *Biomaterials*. 2009 Feb;30(5):780-8. doi: 10.1016/j.biomaterials.2008.10.022.
28. Binulal NS, Deepthy M, Selvamurugan N, Shalumon KT, Suja S, Mony U et al. Role of nanofibrous poly(caprolactone) scaffolds



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



- in human mesenchymal stem cell attachment and spreading for in vitro bone tissue engineering--response to osteogenic regulators. *Tissue Eng. Part A* 2010 Feb;16(2):393-404. doi: 10.1089/ten.TEA.2009.0242.
29. Jensen J, Kraft DC, Lysdahl H, Foldager CB, Chen M, Kristiansen AA et al. Functionalization of polycaprolactone scaffolds with hyaluronic acid and β -TCP facilitates migration and osteogenic differentiation of human dental pulp stem cells in vitro. *Tissue Eng. Part A*. 2015 Feb;21(3-4):729-39. doi: 10.1089/ten.TEA.2014.0177.
 30. Shoi K, Aoki K, Ohya K, Takagi Y, Shimokawa H. Characterization of pulp and follicle stem cells from impacted supernumerary maxillary incisors. *Pediatr. Dent.* 2014 May-Jun;36(3):79-84.
 31. Zhang Y, Xing Y, Jia L, Ji Y, Zhao B, Wen Y et al. An In Vitro Comparative Study of Multisource Derived Human Mesenchymal Stem Cells for Bone Tissue Engineering. *Stem Cells Dev.* Dec 1;27(23):1634-1645.
 32. Salgado CL, Barrias CC, Monteiro FJM. Clarifying the Tooth-Derived Stem Cells Behavior in a 3D Biomimetic Scaffold for Bone Tissue Engineering Applications. *Front Bioeng Biotechnol.* 2020 Jun 26;8:724. doi: 10.3389/fbioe.2020.00724.
 33. Li Z, Wang D, Li J, Liu H, Nie L, Li C. Bone Regeneration Facilitated by Autologous Bioscaffold Material: Liquid Phase of Concentrated Growth Factor with Dental Follicle Stem Cell Loading. *ACS Biomater Sci Eng.* 2024 May 13;10(5):3173-3187
 34. Potier E, Ferreira E, Andriamanalijaona R, Pujol JP, Oudina K, Logeart-Avramoglou D et al. Hypoxia affects mesenchymal stromal cell osteogenic differentiation and angiogenic factor expression. *Bone* 2007 Apr;40(4):1078-87. doi: 10.1016/j.bone.2006.11.024.
 35. Lee JH, Rim NG, Jung HS, Shin H. Control of osteogenic differentiation and mineralization of human mesenchymal stem cells on composite nanofibers containing poly(lactic-co-[glycolic acid]) and hydroxyapatite. *Macromol. Biosci.* 2010 Feb 11;10(2):173-82. doi: 10.1002/mabi.200900169.
 36. Martins A, Pinho ED, Correlo VM, Faria S, Marques AP, Reis RL et al. Biodegradable nanofibers-reinforced microfibrillar composite scaffolds for bone tissue engineering. *Tissue Eng. Part A* 2010 Dec;16(12):3599-609. doi: 10.1089/ten.TEA.2009.0779.
 37. Sadeghzadeh H, Mehdipour A, Dianat-Moghadam H, Salehi R, Khoshfetrat AB, Hassani A et al. PCL/Col I-based magnetic nanocomposite scaffold provides an osteoinductive environment for ADSCs in osteogenic cues-free media conditions. *Stem Cell Res Ther* 2022 Apr 4;13(1):143. doi: 10.1186/s13287-022-02816-0.



RESEARCH ARTICLE

Comparison of Marginal Bone Loss Between Titanium and Titanium-Zirconium Implants at Least 5 Years at Function

En Az 5 Yıllık Fonksiyon Süresiyle Titanyum ve Titanyum-Zirkonyum İmplantlar Arasındaki Marjinal Kemik Kaybının Karşılaştırılması

Selin Görgündür¹ , Esra Beyler² 

¹DDS; Başkent University faculty of dentistry oral and maxillofacial surgery

²DDS PhD; Başkent University faculty of dentistry oral and maxillofacial surgery

ABSTRACT

Titanium (Ti) is the leading material in implant dentistry for the treatment of partial or full edentulism.. More recently, Titanium- Zirconium (TiZr) alloy has been developed for more demanding clinical conditions, as it shows greater mechanical and biological features than commercially pure Ti Grade 4. Survival of dental implants is based on the relationship between implant and oral tissues. Marginal bone level stability around implants has been used as one of the main criteria for implant success. Implant failures are often associated with implant mobility due to marginal bone loss. The aim of this study was to compare Titanium and Titanium-Zirconium implants marginal bone loss values which were at function more than 5 years. Titanium and 13-17% Zr containing TiZr alloy implants included in this study. Marginal bone loss measurements were performed digitally in computer software programme. TiZr alloy implants showed less marginal bone loss compared to traditional titanium implants but this difference statistically was not significant. TiZr alloy may become the dominant material in implant material choice in order to increase clinical implant success.

Keywords: dental implants, marginal bone loss, titanium zirconium implants

ÖZET

Titanyum (Ti), kısmi veya tam dişsizlik tedavisinde implant diş hekimliğinde en yaygın kullanılan materyaldir. Son zamanlarda zorlu klinik koşullar için, ticari olarak saf Ti Grade 4'e kıyasla daha üstün mekanik ve biyolojik özellikler sergileyen Titanyum-Zirkonyum (TiZr) alaşımı geliştirilmiştir. Dental implantların başarısı, implant ile ağız dokuları arasındaki ilişkiye dayanmaktadır. İmplant çevresindeki marjinal kemik seviyesi stabilitesi, implant başarısının temel kriterlerinden biri olarak kabul edilmektedir. İmplant kayıpları sıklıkla, marjinal kemik kaybına bağlı implant mobilitesi ile ilişkilidir. Bu çalışmanın amacı, 5 yıldan daha uzun süredir fonksiyonda olan Titanyum ve Titanyum-Zirkonyum implantlarının marjinal kemik kaybı değerlerini karşılaştırmaktır. Çalışmaya Titanyum ve %13-17 oranında Zr içeren TiZr alaşımlı implantlar dahil edilmiştir. Marjinal kemik kaybı ölçümleri bilgisayar destekli yazılım programı kullanılarak dijital olarak gerçekleştirilmiştir. TiZr alaşımlı implantlar, geleneksel Titanyum implantlara kıyasla daha az marjinal kemik kaybı göstermiştir, ancak bu fark istatistiksel olarak anlamlı bulunmamıştır. Klinik implant başarısını artırmak amacıyla TiZr alaşımının ilerleyen dönemlerde implant materyali seçiminde dominant hale gelebileceği düşünülmektedir.

Anahtar Kelimeler: dental implantoloji, marjinal kemik kaybı, titanyum zirkonyum

Submission Date: April 29, 2025

Acceptance Date: May 22, 2025

Corresponding author: Selin Görgündür

Address: Başkent University faculty of dentistry oral and maxillofacial surgery

Phone: +90 5068326070

Email: selingrgndr@gmail.com

Selin Görgündür

Esra Beyler

0000-0002-5809-018X

0000-0003-0824-1629



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



INTRODUCTION

Since Branemark introduced the concept of osseointegration in 1977, dental implants have become a popular method for the treatment of total or partial edentulism.¹ According the AAID (American Academy of Implant Dentistry) implant treatment have been shown almost %95 success rate.² In a systematic review and meta-analysis including 23 studies performed in 2015, the success rate of implants was 94.6% in a total of 7711 implants (mean follow-up year 13 years).³

Titanium is a biocompatible material that causes little or no reaction in the tissues in which it is loaded⁴ It has also been found that titanium is a material that is resistant to corrosion and allows osseointegration.⁵ Branemark proved that titanium forms a permanent connection with the bone due to the titanium oxide layer and this created the concept of osseointegration today¹. Titanium have been recognized as the gold standard in implant materials.⁶ However, the stress/strength ratio of narrow diameter titanium implants were found higher than regular diameter implants, indicating higher risk of fatigue failure.⁷ Implants made of titanium alloys containing zirconium (TiZr; commercially known as Roxolid; Institut Straumann AG, Switzerland) have been introduced to the market showing better mechanical properties than pure titanium while maintaining its biocompatibility.⁸ This particular TiZr alloy dental implant manufactured from mixing Ti with 13%-17% of Zr.

The ICOI's (The International Congress of Oral Implantologists) Pisa Consensus defined implant success criteria as pain, mobilization, radiographic marginal bone loss and pocket depth and peri-implant disease.⁹ Regarding of bone loss, Albrektsson et al. recommended that a successful implant should have a marginal bone loss of less than 1 mm in the first year and less than 0.2 mm in the following years.¹⁰ The aim of this study was to compare the marginal bone loss between conventional titanium and TiZr implants under function at least 5 years

MATERIALS AND METHOD

This study was approved by Baskent University Institutional Review Board (Project no: D-KA24/07) and supported by Baskent University Research Fund. This retrospective study was conducted using clinical and radiographic records of patients who received dental implants between 2015 and 2018

at the Department of Oral and Maxillofacial Surgery, Başkent University Faculty of Dentistry.

Patients with posterior single tooth, same brand (Institut Straumann AG, Switzerland) tissue level implants in 4.1 mm diameter and 10 mm length were included. Patients divided into two groups with 40 implants in titanium and 40 implants in Ti-Zr group. Of the 80 implants included in the study, 44 were located in the mandible and 36 in the maxilla. Patients with uncontrolled systemic diseases, active periodontal diseases, keratinized mucosa less than 2 mm, implants required bone augmentation were excluded from the study.

Panoramic radiographs were taken via Veraviewepocs 2D (J. Morita Corp., Japan) device. Digital panoramic radiographs which were taken after the implant placement (T1) and after minimum 5 years in function (T2) were used in peri-implant bone loss evaluations. Measurements were performed digitally in ImageJ software programme (ImageJ, USA). Measurements of one of the digital panoramic radiography are shown in Figure 1. The values were obtained by measuring the distance from the apical end of the implant to the marginal bone level separately on the mesial and distal sides, and then subtracting these values from the total implant length. Twenty percent of the radiographs were randomly selected and remeasured in order to ensure intra-observer reliability and the the Intraclass Correlation Coefficient (ICC) was calculated.

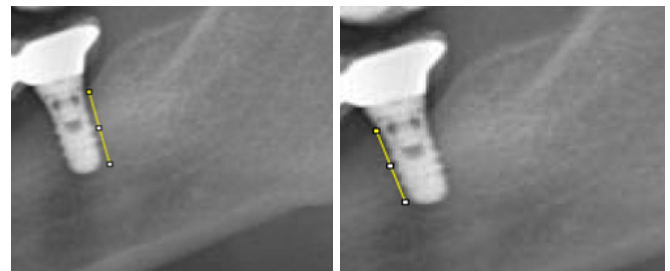


Fig 1: Distally and mesially measurements

Statistical analyses were performed with SPSS version 25.0 program. The conformity of the variables to normal distribution was examined by Shapiro-Wilk test. Mean, standard deviation, median, minimum and maximum values were used to present descriptive analyses. Mann Whitney U Test was used to evaluate the variables that were not normally distributed between Ti and Ti-Zr groups. p-values below 0.05 were considered as statistically significant results.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



RESULTS

Thirty nine patients with a total of eighty implant included in this study. Patient demographic characteristics were summarized as; 16 male (45 %) and 20 female (55 %), with an average age of 41.75 (ranging between 24 and 60 years).

The mean marginal bone loss was 1.46 ± 1.038 mm in the Ti group and 1.11 ± 0.698 mm in the Ti-Zr group. There was no statistically significant difference between the Ti group and the Ti-Zr group ($p=0.101$) regarding of peri-implant bone loss values. Ti and Ti-Zr implants did not show different results in the follow-up of marginal bone loss over 5 years (Table 1).

Table 1: No statistical difference between Titanium and Titanium Zirconium Group

	Titanium		Titanium - Zirconium		p
	Mean \pm sd	Median(Min-Max)	Mean \pm sd	Median(Min-Max)	
Bone loss	1,464 \pm 1,038	1,212 [0-6,289]	1,113 \pm ,698	1,11 [0-2,377]	0,101

DISCUSSION

The marginal bone around the implant is considered as a significant indicator of implant health. In the literature the most common method to measure bone loss is by radiographic evaluation. However, conventional two dimension radiographics allow to monitor only the mesial and distal aspect of bone loss. It was suggested that the peri-implant bone level measurements should be related to the original marginal bone level which was measured at implant insertion, rather than to a previous measurement.⁹

The use of Ti-Zr dental implants is well published in several in vitro and animal studies, showing similar results to Grade IV titanium dental implants.¹¹

In a clinical trial by Quirynen et al., Ti-Zr alloy implants were compared to titanium grade IV implants. The marginal bone level values were found similar in the two groups, which is compatible with the results of this study.¹²

In our study, slightly less marginal bone loss was observed in titanium-zirconium implants compared to titanium implants. In their in vitro study, Sista et al. showed that more osteoblasts adhered to the 50% Ti-zr surface compared to the titanium surface and more alkaline phosphatase and osteocalcin were released. This may lead to better osseointegration and thus indirectly to less marginal bone loss.¹³

A study by Ghazal et al. in 2019, the 5-year mean bone loss of ti-zr standard implants was measured as -0.48 ± 0.67 mm, which was less than the average marginal bone loss in the ti-zr group of our study. This may be due to the fact that Ghazal et al. included only single tooth implants or excluded heavy smokers (more than 10 per day).¹⁴

Carlson et al, measured the 10-year periimplant marginal bone loss of conventional titanium implants as 0.9 mm .This result was close to the marginal bone loss of titanium implants in our study, but slightly less than our result.¹⁵

CONCLUSION

The relatively new alloy implants appear as a reliable treatment option. Ti-Zr implants showed slightly less bone loss compared to Ti group. Although this difference clinically is not significant, Ti-Zr alloy may be the dominant material in implant material choice in order to increase clinical implant success. Due to the fact that peri-implant bone loss depends on many factors, multiparameter studies with larger sample size are needed.

REFERENCES

1. Brånemark PI. Osseointegrated implants in the treatment of the edentulous jaw: experience from a 10-year period. Scand J Plast Reconstr Surg. 1977;16:1-132.
2. What is the Success Rate of Dental Implants? [Internet]. [cited 2025 Apr 16]. Available from: <https://connect.aaid-implant.org/blog/success-rate-dental-implants>
3. Moraschini V, Poubel LADC, Ferreira VF, Barboza EDSP. Evaluation of survival and success rates of dental implants reported in longitudinal studies with a follow-up period of at least 10 years: a systematic review. Int J Oral Maxillofac Surg. 2015;44(3):377-88.
4. Silva RCS, Agrelli A, Andrade AN, et al. Titanium dental implants: an overview of applied nanobiotechnology to improve biocompatibility and prevent infections. Materials. 2022;15(9):1-23.
5. Steinemann SG. Titanium – the material of choice? Periodontology 2000. 1998;17(1):7-21.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



6. Karoussis IK, Salvi GE, Heitz-Mayfield LJA, Brägger U, Hämmerle CHF, Lang NP. Long-term implant prognosis in patients with and without a history of chronic periodontitis: a 10-year prospective cohort study of the ITI® Dental Implant System. Clin Oral Implants Res. 2003;14(3):329–39.
7. Neto HT, Tuzita AS, Gehrke SA, et al. A comparative analysis of implants presenting different diameters: extra-narrow, narrow and conventional. Materials. 2020;13(8):1888.
8. Kobayashi E, Matsumoto S, Doi H, Yoneyama T, Hamanaka H. Mechanical properties of the binary titanium-zirconium alloys and their potential for biomedical materials. J Biomed Mater Res. 1995;29(8):943–50.
9. Misch CE, Perel ML, Wang HL, et al. Implant success, survival, and failure: the International Congress of Oral Implantologists (ICOI) Pisa Consensus Conference. Implant Dent. 2008;17(1):5–15.
10. Albrektsson T, Zarb G, Worthington P, Eriksson AR. The long-term efficacy of currently used dental implants: a review and proposed criteria of success. Int J Oral Maxillofac Implants. 1986;1(1):11–25.
11. Clinical evidence on titanium–zirconium dental implants: a systematic review and meta-analysis. International Journal of Oral and Maxillofacial Surgery, 45(7), 842–850 | 10.1016/j.ijom.2016.01.004.
12. Müller F, Al-Nawas B, Storelli S, et al. Small-diameter titanium grade IV and titanium-zirconium implants in edentulous mandibles: five-year results from a double-blind, randomized controlled trial. BMC Oral Health. 2015;15:1–10.
13. Sista S, Wen C, Hodgson PD, Pande G. The influence of surface energy of titanium-zirconium alloy on osteoblast cell functions in vitro. J Biomed Mater Res A. 2011;97A(1):27–36.
14. Ghazal SS, Huynh-Ba G, Aghaloo T, et al. Randomized controlled multicenter clinical study evaluating crestal bone level change of narrow-diameter versus standard-diameter Ti-Zr implants for single tooth replacement in anterior and premolar region. Int J Oral Maxillofac Implants. 2019;34(3):708–15.
15. Carlsson GE, Lindquist LW, Jemt T. Long-term marginal periimplant bone loss in edentulous patients. Int J Prosthodont. 2000 Jul-Aug;13(4):295–302. PMID: 11203645.



RESEARCH ARTICLE

İliak Kemik Grefti ile Ogmente Edilmiş Kretlere Yerleştirilen İmplantların Uzun Dönem Sağkalımlarının Değerlendirilmesi

Long-Term Survival Assessment of Dental Implants Placed in Atrophic Ridges Augmented with Iliac Bone Grafts

Yusuf Tamer¹ , Seçil Çubuk² 

¹Dr. Öğr. Üyesi, Baskent Üniversitesi, Diş Hekimliği Fakültesi, Ağız, Diş ve Çene Cerrahisi Bölümü, Adana

²Doç.Dr., Baskent Üniversitesi, Diş Hekimliği Fakültesi, Ağız, Diş ve Çene Cerrahisi Bölümü, Ankara

ÖZET

Bu retrospektif çalışma, atrofik çenelerin rekonstrüksiyonunda iliak kemik grefti ile ogmente edilen kretlere yerleştirilen dental implantların uzun dönemli sağkalımını ve komplikasyon profilini değerlendirmeyi amaçlamaktadır. 2008-2018 yılları arasında Baskent Üniversitesi Adana Turgut Noyan Uygulama ve Araştırma Hastanesi'nde tedavi edilen, en az 5 yıllık takibi tamamlanmış 12 hasta çalışmaya dahil edilmiştir. Toplam 67 implant yerleştirilmiş, hastaların ortalama yaşı 62,8 ve takip süresi 83,5 ay olarak hesaplanmıştır. Greft başarı oranı %100 olup, alıcı bölgede cerrahi sonrası komplikasyon izlenmemiştir. Ancak donör bölgede 5 hastada çeşitli komplikasyonlar gözlenmiştir. Bu komplikasyonlar arasında en sık karşılaşılanlar yürüme güçlüğü ve hematomdur. Toplam 6 implant (%9,2) kaybedilmiş olup, bunların 4'ü osseointegrasyon başarısızlığı nedeniyle, 2'si ise protetik tedavi sonrasında kaybedilmiştir. Bulgular, iliak kemik greftleriyle ogmente edilen alanlara yerleştirilen implantların uzun vadeli sağkalım oranlarının literatürle uyumlu olduğunu göstermektedir. Ancak iliak kemiğin yüksek rezorpsiyon oranı ve embriyolojik köken farkı nedeniyle bazı implant kayıpları yaşanmıştır. Greft stabilitesini artırmaya yönelik tekniklerin değerlendirilmesi, gelecekteki çalışmalarda klinik başarının artırılmasına katkı sağlayabilir.

Anahtar Kelimeler: İliak Kemik Greftleme, Dental İmplant, Osteointegrasyon

ABSTRACT

This retrospective study aimed to evaluate the long-term survival and complication profile of dental implants placed in alveolar ridges augmented with iliac bone grafts for the reconstruction of atrophic jaws. Twelve patients treated at Baskent University Adana Turgut Noyan Application and Research Hospital between 2008 and 2018, with a minimum follow-up period of 5 years, were included. A total of 67 implants were placed, with a mean patient age of 62.8 years and an average follow-up duration of 83.5 months. The graft success rate was 100%, with no postoperative complications observed at the recipient sites. However, donor site complications were noted in five patients, including gait disturbances and hematomas. A total of six implants (9.2%) failed—four due to lack of osseointegration before prosthetic loading and two after prosthetic use. The findings indicate that the long-term survival rates of implants placed in iliac bone-grafted areas are consistent with existing literature. Nevertheless, some implant losses were attributed to the high resorption potential and embryological origin differences of iliac bone. Future studies assessing techniques to enhance graft stability may contribute to improved clinical outcomes.

Keywords: Iliac bone graft, Dental implant, Osseointegration

Submission Date: May 2, 2025

Acceptance Date: May 20, 2025

Corresponding author: Yusuf Tamer

Address: Baskent Üniversitesi, Adana Uygulama ve Araştırma Merkezi, Kazım Karabekir cad.

59. Sok. No. 91. 01250 Yüreğir, Adana, Türkiye

Phone: +90 5426285654

Email: dryusuftamer@gmail.com

Yusuf Tamer

Seçil Çubuk

0000-0003-4338-7180

0000-0003-2065-7555



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



Son yıllarda diş implantları kısmen veya tamamen dişsiz olan hastaların rehabilitasyonu için rutin bir tedavi yöntemi haline gelmiştir.¹ Birçok vakada dental implantlar herhangi bir zorluk veya komplikasyon olmadan yerleştirilebilmektedir. Ancak bazı durumlarda, şiddetli periodontal hastalıklar, uzun süreli dişsizliğe bağlı alveolar kemik atrofi, travma, enfeksiyon ya da kemik patolojileri kaynaklı kemikte ileri dereceli defektler oluşması implantın doğru aç ve uzunlukta yerleştirilmesini imkansız hale getirebilir.^{2,3}

Atrofik kretlerin rekonstrüksiyonunda otojen kemik greftleri kullanılarak implantın ideal pozisyonunda ve biomekanik açıdan uygun şekilde yerleştirilmesi sağlanabilir. Otojen kemik greftleri, osteokondüktif, osteoindüktif ve osteojenik özellikleri bir arada barındırması nedeniyle kemik rejenerasyonunda altın standart olarak kabul edilmektedir.⁴

Otojen kemik greftleri intraoral ve ekstraoral bölgelerden elde edilebilir. İntraoral kemik greftleri düşük rezorpsiyon oranlarına sahip olmakla beraber sağlayabilecekleri kemik hacmi sınırlıdır. Geniş kemik defektlerinin rekonstrüksiyonunda kalvarial kemik, kosta, tibia ve iliak kemikler gibi ekstraoral donör bölgelerden greft temin etmek gerekebilir.⁵

İliak kemik, yüksek osteojenik potansiyele sahip olması, vaskülarizasyonunun hızlı gerçekleşmesi ve ortalama 50 cm³ kortikokansellöz greft materyali temin edilebilmesi nedeniyle, greftleme işlemleri için optimal donör sahalardan biridir.⁶

İliak kemikle yapılan rekonstrüksiyonda greftin uzun vadeli sağkalım oranı ve stabilitesi, özellikle kemik rezorpsiyon oranlarına ilişkin belirsizlikler nedeniyle halen tartışmalıdır.³ Bu çalışmanın amacı, kliniğimizde atrofik çenelerde gerçekleştirilen iliak kemik grefti augmentasyonu sonrasında uygulanan dental implantlar ile implant üstü protetik rehabilitasyonun en az 5 yıllık kümülatif başarı oranlarını, erken ve geç dönem komplikasyon profillerini, ayrıca radyografik ve klinik takip bulgularını değerlendirmektir.

MATERYAL METOT

Bu çalışma, Baskent Üniversitesi Adana Turgut Noyan Uygulama ve Arastırma Hastanesi Kurumsal Etik Kurulu tarafından onaylanmıştır (Protokol No: KA25/14). Çalışma protokolü, Dünya Tıp Birliği'nin Helsinki Bildirgesi etik ilkelerine uygun olarak tasarlanmış ve tüm katılımcılardan yazılı bilgilendirilmiş onam alınmıştır.

Bu retrospektif çalışmaya Baskent Üniversitesi Adana Turgut Noyan Uygulama ve Arastırma Hastanesi Ağız, Diş ve Çene Cerrahisi kliniğinde 2008-2018 yılları arasında iliak greft ile alveolar augmentasyon uygulanarak dental implant yerleştirilmiş ve implant üstü protetik rehabilitasyonu tamamlanmasının ardından en az 5 yıl süre geçmiş, tüm tedavi süreci ile takip ziyaretlerine ait klinik muayene bulguları ve radyografik görüntüleri hasta kayıtlarında eksiksiz olarak bulunan 18 yaşından büyük bireyler dahil edilmiştir.

Sistemik hastalık öyküsü olan, kemik metabolizmasını etkileyen ilaç kullanan, greft bölgesinde postoperatif komplikasyon (enfeksiyon vb.) gelişen, düzenli takipleri bulunmayan, klinik veya radyografik verileri eksik olan, tedavi süreci tamamlanamayan, parafonksiyonel alışkanlık (ör. brüksizm) öyküsü olan ve 18 yaş altı bireyler çalışma dışı bırakılmıştır.

Greft Başarısı ve İmplant Sağkalımının Değerlendirilmesi: Greftin başarılı kabul edilmesi için gerekli görülen kriterler greftleme işleminden sonra ve implant yerleştirilme aşamasında greft bölgesinde enfeksiyon bulunmaması, radyografik incelemelerde greftlenen sahada radyolüseni gözlenmemesi, greftin alıcı bölgede entegre ve hareketsiz olması, implant yerleştirilmesi sırasında greftten kanama gözlenmesi ve implant yerleşimi için yeterli miktarda kemik dokusu bulunmasıdır.⁷

İmplant sağkalımı; implantın ağızda tutulabiliyor olması, hareketli olmaması, fonksiyon sırasında ağrı oluşturmaması kriterlerine göre değerlendirildi.⁸

Veriler tanımlayıcı istatistiklerle değerlendirildi. Sürekli veriler ortalama ± standart sapma (SD), kategorik veriler sayı ve yüzde (%) olarak sunuldu.

SONUÇLAR

Bu çalışmaya toplam 67 adet implant uygulanmış olan 12 hasta (6 kadın, 6 erkek) dahil edildi (Tablo 1). Hastaların yaş ortalaması 62,8± 4,9 yıl (54-72 yıl) ve takip süresi ortalama 83.5 ±12.1 ay olarak hesaplandı.

Yedi hasta tam dişsizlik ve 5 hasta kısmi dişsizlik nedeniyle tedavi edildi. Mandibulaya toplam 11 adet, maksillaya 54 adet implant yerleştirildi.

Greft başarı oranı %100'dü. Ameliyat sonrası alıcı bölgede herhangi bir komplikasyon görülmedi. Çalışmada incelenen 12 hastanın donör bölge komplikasyonlarına ilişkin



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)

EurAsian Journal of
Oral and Maxillofacial Surgery



değerlendirmesinde, toplam 5 hastada çeşitli komplikasyonlar gözlenmiştir. En sık rastlanan komplikasyon yürüme güçlüğü olup 4 hastada bildirilmiştir. Hematom ise 2 hastada izlenmiştir. Geriye kalan 7 hastada ise herhangi bir donör bölge komplikasyonu gözlenmemiştir (Tablo 2).

Toplam 6 implant (%9,2) kaybedilmiştir. Bu implantlardan 4'ü, protez için ölçü alınmadan önce osseointegrasyon başarısızlığı nedeniyle; 2'si ise protez kullanımı başladıktan sonra kaybedilmiştir.

Tablo 1: Hastaların Yaş, Cinsiyet, Uygulanan Protez Türü, İmplant Sayısı ve Takip Süresi Dağılımı

Yaş	Cinsiyet	Protez Türü	Uygulandığı Çene	İmplant Sayısı	Süresi(ay)
72	Kadın	All on Four	mandibula	4	84
56	Erkek	Hibrit Protez	maksilla	6	92
64	Erkek	All on Four/Hibrit	maksilla/mandibula	6	68
63	Kadın	All on Six	maksilla	6	86
63	Kadın	Hibrit Protez	maksilla	4	110
61	Kadın	All on Six	maksilla	6	74
67	Erkek	All on Four	mandibula	4	79
58	Erkek	All on Six	maksilla	6	69
61	Kadın	All on Six/Hibrit	maksilla/mandibula	8	82
64	Erkek	All on Six	mandibula	6	76
61	Kadın	Hibrit Protez	mandibula	5	81
68	Erkek	All on Six	maksilla	6	105

Tablo2: İmplant Kaybı Nedenleri ve Donör Bölge Komplikasyonlarının Sıklığı

Hastalar	İmplant Kaybı		Donör Bölge
	Protez Öncesi	Protez Sonrası	
1	0	0	Yok
2	0	0	Yok
3	0	0	Yok
4	2	1	hematom, yürüme güçlüğü
5	0	0	Yok
6	0	0	yürüme güçlüğü
7	1	0	Yok
8	1	0	Yok
9	0	1	hematom
10	0	0	Yok
11	0	0	yürüme güçlüğü
12	0	0	yürüme güçlüğü



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



TARTIŞMA

Dental implantlar, eksik dişlerin tedavisinde sabit veya hareketli protez uygulamalarını kolaylaştıran etkili bir tedavi seçeneğidir. Ancak implant yerleştirilecek bölgedeki alveol kemiğinin aşırı rezorbe olması, implantın ideal açıda ve yeterli uzunlukta yerleştirilmesini zorlaştırabilir. Bu gibi durumlarda, uzun vadede fonksiyonel ve estetik açıdan başarılı bir sonuç elde etmek için yetersiz kemik hacminin artırılması gerekir. Böylece implantın stabilitesi ve dayanıklılığı sağlanarak hastanın hem işlevsel hem de estetik beklentileri karşılanabilir.⁹

Yakın zamanda yayınlanmış olan bir sistematik derlemede iliak kemikten elde edilen greftler ile rehabilite edilen kretlere uygulanan implantların sağkalım oranlarının, greft uygulanmamış bölgelere veya intraoral kaynaklı kemik greftleri ile ogmente edilen kretlere uygulanan implantların sağkalımına kıyasla belirgin şekilde daha düşük olduğu gösterilmiştir. Bu çalışmada iliak kemik bölgesine yapılan implantların 5 yıllık sağkalım oranı % 90 olduğu rapor edilmiştir.¹⁰ Çalışmamızın sonuçları iliak greft ile ogmente kretlere yerleştirilen implantların sağkalım oranları bakımından literatür ile benzerlik göstermektedir.

Çene kemikleri ile iliak kemik greftleri arasında embriyolojik köken açısından belirgin farklılıklar bulunmaktadır. Çene kemikleri intramembranöz ossifikasyon yoluyla, ilium ise endokondral ossifikasyon ile gelişir. Bu gelişimsel farklılık, greftlerin rezorpsiyon oranları ile greft ve implant sağkalımı üzerinde önemli bir etkiye sahip olabilir. Özellikle, iliak kemik greftlerinin en önemli dezavantajlarından biri, yüksek rezorpsiyon oranlarına sahip olmalarıdır. Yapılan çalışmalarda, iliak greftlerde altı ay sonrasında hacimsel olarak %24 ila %49 oranında azalma meydana geldiği bildirilmiştir.^{11,12,13}

Literatürde, erken dönem kemik rezorpsiyon miktarını azaltmak amacıyla greft ve implantın aynı zamanda yerleştirildiği tek aşamalı tedavi ve bariyer membran kullanımı önerilmektedir.^{14,15} Tek aşamalı yöntemin işlem sayısı ve tedavi süresini kısaltma gibi avantajlarının yanı sıra bu yöntemde implantların primer stabilitesinin sağlanmasında zorluk yaşanabilir. Çalışmamızda değerlendirilen hiçbir olguda greftleme ile eş zamanlı implant yerleştirilmemiş veya iliak greft üzerine bariyer membran uygulanmamıştır.

Bu çalışmanın bulguları, iliak kemik greftleri ile ogmente edilen kretlere yerleştirilen dental implantların uzun dönemli sağkalım oranlarının literatürle uyumlu olduğunu

göstermektedir. Her ne kadar greft başarı oranı yüksek olsa da, iliak greft kaynaklı implantların bir kısmında erken dönemde osseointegrasyon başarısızlığı ve ileri derecede periimplant kemik rezorpsiyonu nedeniyle implant kayıpları gözlenmiştir. Bu sonuçlar, greftin embriyolojik kökeni ve yüksek rezorpsiyon potansiyelinin implant başarısı üzerinde etkili olabileceğini düşündürmektedir. İleriye dönük çalışmalarda, greft stabilitesini artırmaya yönelik tekniklerin değerlendirilmesi klinik başarıyı artırmada faydalı olabilir.

Bu çalışmamızın bazı sınırlamaları bulunmaktadır. Çalışmanın retrospektif olması, verilerin hasta kayıtlarından elde edilmesi nedeniyle bazı parametrelerin standardizasyonunda sınırlamalar bulunmaktadır. Ayrıca, çalışmaya dahil edilen hasta sayısının ve implant sayısının nispeten düşük olması, istatistiksel gücü ve sonuçların genellenebilirliğini etkileyebilir. Ayrıca donör bölge komplikasyonları (yürüme güçlüğü, hematoma) hasta bazında değerlendirilmiş olsa da, komplikasyonların şiddeti ve süresine ilişkin nicel verilerin hasta epikriz notlarından ve hastalara geçmişe dair sorulan sorulardan elde edilmiştir. Bu durum bulguların objektif olarak karşılaştırılmasını kısıtlamaktadır.

Bu kısıtlılıklara rağmen çalışmanın bulguları, iliak kemik greftleri ile ogmente edilen atrofik kretlerde yerleştirilen dental implantların uzun dönemli sağkalım oranlarının klinik açıdan kabul edilebilir düzeyde olduğunu göstermektedir.

KAYNAKLAR

1. Moraschini V, Poubel LA, Ferreira VF, Barboza ES. Evaluation of survival and success rates of dental implants reported in longitudinal studies with a follow-up period of at least 10 years: a systematic review. *Int J Oral Maxillofac Surg*. 2015;44(3):377-88.
2. Stellingsma C, Vissink A, Meijer HJ, Kuiper C, Raghoobar GM. Implantology and the severely resorbed edentulous mandible. *Crit Rev Oral Biol Med* 2004; 15(4): 240-8.
3. Nguyen TTH, Eo MY, Kuk TS, Myoung H, Kim SM. Rehabilitation of atrophic jaw using iliac onlay bone graft combined with dental implants. *Int J Implant Dent*. 2019 Mar 19;5(1):11.
4. Rodham PL, Giannoudis VP, Kanakaris NK, Giannoudis PV. Biological aspects to enhance fracture healing. *EFORT Open Rev*. 2023 May 9;8(5):264-82.
5. Sjöström M, Sennerby L, Nilson H, Lundgren S. Reconstruction of the Atrophic Edentulous Maxilla with Free Iliac Crest Grafts and Implants: A 3-Year Report of a Prospective Clinical Study. *Clin Implant Dent Relat Res* 2007; 9(1): 46-59.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



6. Dahlin C, Johansson A. Iliac crest ABG versus alloplastic graft and guided bone regeneration in the reconstruction of atrophic maxillae: a 5-year retrospective study on cost-effectiveness and clinical outcome. *Clin Implant Dent Relat Res*. 2011;13:305-10.
7. Barone A, Covani U. Maxillary alveolar ridge reconstruction with nonvascularized autogenous block bone: clinical results. *J Oral Maxillofac Surg*. 2007;65(10):2039-46.
8. Misch CE, Perel ML, Wang HL, et al. Implant success, survival, and failure: the International Congress of Oral Implantologists (ICOI) Pisa Consensus Conference. *Implant Dent* 2008;17:5-15.
9. Roccuzzo M, Roccuzzo A, Marruganti C, Fickl S. The importance of soft tissue condition in bone regenerative procedures to ensure long-term peri-implant health. *Periodontol* 2000. 2023 Oct;93(1):129-138.
10. McKenna GJ, Gjengedal H, Harkin J, Holland N, Moore C, Srinivasan M. Effect of autogenous bone graft site on dental implant survival and donor site complications: a systematic review and meta-analysis. *J Evid Based Dent Pract*. 2022 Sep;22(3):101731.
11. Mertens C, Decker C, Seeberger R, Hoffmann J, Sander A, Freier K. Early bone resorption after vertical bone augmentation--a comparison of calvarial and iliac grafts. *Clin Oral Implants Res*. 2013 Jul;24(7):820-5.
12. Johansson, B., Grepe, A., Wannfors, K. & Hirsch, J. M. (2001) A clinical study of changes in the volume of bone grafts in the atrophic maxilla. *Dentomaxillofacial Radiology* 30: 157-161.
13. Chiapasco M, Zaniboni M, Boisco M. Augmentation procedures for the rehabilitation of deficient edentulous ridges with oral implants. *Clin Oral Implants Res*. 2006;17(Suppl 2):136-159.
14. Kang YH, Kim HM, Byun JH, Kim UK, Sung IY, Cho YC, Park BW: Stability of simultaneously placed dental implants with autologous bone grafts harvested from the iliac crest or intraoral jaw bone. *BMC Oral Health*. 2015, 15:172. 10.1186/s12903-015-0156-x
15. Vermeeren JI, Wismeijer D, van Waas MA. One-step reconstruction of the severely resorbed mandible with onlay bone grafts and endosteal implants. A 5-year follow-up. *Int J Oral Maxillofac Surg*. 1996 Apr;25(2):112-5.



CASE REPORT

Gunshot Injury to The Mandible of a Patient Due to Domestic Violence With Tough Treatment Process: A Case Report

Aile İçi Şiddet Sebebiyle Bir Hastanın Mandibulasına Oluşan Ateşli Silah Yaralanmasının Zorlu Tedavi Süreci: Bir Olgu Raporu

Nihat Akbulut¹ , Aseel Halawa²

¹Professor, Ondokuz Mayıs University, Faculty of Dentistry, Oral and Maxillofacial Department, Samsun, Türkiye

²Research Asistant, Ondokuz Mayıs University, Faculty of Dentistry, Oral and Maxillofacial Department, Samsun, Türkiye

ABSTRACT

Gunshot wounds (GSWs) are a significant cause of trauma in the maxillofacial region, particularly in the mandible. Following the establishment of a treatment plan, patients may experience various complications, which often necessitate a secondary surgical intervention to correct the patient's concerns. This case study presents the details of a 36-year-old female patient who experienced a gunshot injury six years ago, that resulted in a fracture of the mandibular ramus. As a consequence of the injury, the mandibular condyle and coronoid process migrated into the infraorbital fossa, causing difficulty with mastication and mouth opening. To correct this issue, a surgical procedure was performed that included submandibular and preauricular incisions. The condyle was replaced and fixed with rigid internal fixation and maxillomandibular fixation techniques. Additionally, an abdominal fat graft was harvested and transplanted into the mandible. Postoperative follow-up was carried out; however, two weeks later, the patient developed an infection, requiring transfer to the infectious diseases department. In the follow-up period, the patient has acceptable function and well-being in the other health condition.

Keywords: Gunshot, wound, trauma, mandible, infection, fat graft.

ÖZET

Ateşli silah yaralanmaları (ASY), özellikle mandibula bölgesinde, maksillofasiyal bölgede önemli bir travma nedenidir. Tedavi planının belirlenmesini takiben, hastalarda genellikle çeşitli komplikasyonlar gelişebilir ve bu durum hastanın şikayetlerini gidermek amacıyla ikincil cerrahi müdahaleleri gerekli kılar. Bu olgu sunumunda, altı yıl önce ASY geçirmiş 36 yaşındaki bir kadın hastanın durumu ele alınmaktadır. Yaralanma sonucunda mandibular ramusta bir kırık oluşmuş ve mandibular kondil ile koronoid çıkıntı infraorbital fossaya göç etmiştir. Bu durum hastada çiğneme ve ağız açmada zorluklara neden olmuştur. Bu sorunu düzeltmek amacıyla submandibular ve preauriküler insizyonları içeren bir cerrahi işlem gerçekleştirilmiştir. Kondil, yerine yerleştirilmiş ve rijid internal fiksasyon ile maksillomandibular fiksasyon teknikleri kullanılarak sabitlenmiştir. Ayrıca, karın bölgesinden yağ grefti alınarak mandibula bölgesine transplant edilmiştir. Ameliyat sonrası takip süreci yürütülmüş; ancak iki hafta sonra hastada enfeksiyon gelişmiş ve enfeksiyon servisinde tedavi görmesi gerekmiştir. Enfeksiyon bir ay içerisinde başarıyla tedavi edilmiş ve hasta sağlıklı bir şekilde taburcu edilmiştir. Takip sürecinde, hastanın genel sağlık durumu iyi olup fonksiyon açısından kabul edilebilir düzeyde iyileşme gözlemlenmiştir.

Anahtar Kelimeler: Ateşli silah, yara, travma, mandibula, enfeksiyon, yağ grefti

Submission Date: May 14, 2025

Acceptance Date: May 22, 2025

Corresponding author: Nihat Akbulut

Address: Ondokuz Mayıs University, Faculty of Dentistry, Oral and Maxillofacial Department, Samsun, Türkiye.

Phone: +90 5054489263

Email: drnihatakbulut@yahoo.com

Nihat Akbulut

Aseel Halawa

0000-0001-6950-8214

0009-0001-5663-362X



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



INTRODUCTION

Gunshot wounds (GSWs) frequently result in traumatic injuries to the maxillofacial region, with the mandible being particularly vulnerable. Patients may experience various complications, including limited mouth opening, fibrous or bony ankyloses, scarring, trismus, paresthesia, and facial asymmetry.¹⁻⁴ These complications often necessitate a secondary surgical intervention to correct the patient's concerns. This case aims to illustrate the surgical procedures employed for the replacement and reconstruction of a migrated condyle resulting from a GSW. Additionally, it highlights the technique of harvesting a fat graft from the abdominal dermis to replenish the incision site. The case also sheds light on the topic of domestic violence and its impacts.

CASE REPORT

A 36-year-old female with no known systemic illnesses applied to Ondokuz Mayıs University. Six years ago, the patient suffered a gunshot to her right mandible, which was inflicted by a family member. She had several operations in other clinics with unacceptable outcomes. She reported experiencing pain while opening her mouth and expressed difficulty in chewing food. After the examination of the patient; pain while opening mouth, trismus, and facial asymmetry were noted.

At the radiological exam computed tomography (CT), panoramic and posteroanterior x-rays were obtained. The radiographs clearly showed that the mandibular condyle and coronoid process had migrated into the infratemporal fossa after gunshot injury (GSI).

The fracture was unfavorable, so masseter muscle's movement helped in the displacement. As a firearm wound, the fracture was in multi-piece which requires rigid fixation in the treatment plan.

A comprehensive operation under general anesthesia was planned to reconstruct the condyle. The potential complications associated with the surgery were explained to the patient, and all the necessary preparations were made.

Preauricular and submandibular incisions were made during the surgical procedure. The mandibular condyle was separated from the coronoid process, inserted into the glenoid fossa, and fixed with a reconstruction plate. In addition, an intraoral closed reduction fixation of the mandible (MMF) was performed to provide stability to the dental arch (Figure 1).

To close the gap in the incision area, a dermis fat graft was harvested from the patient's abdomen. The harvested fat was then used to fill the mandibular area in the same surgical session. Subsequently, the wound was sutured to facilitate proper healing.

The patient received prophylactic antibiotics for a duration of two weeks. This included amoxicillin 1000mg taken twice daily, metronidazol 500mg taken twice daily, and cefuroxime 500mg taken once daily.

After two weeks, the patient returned for a follow-up examination and presented with symptoms of swelling, redness, and pain in the surgical site. Therefore, she was

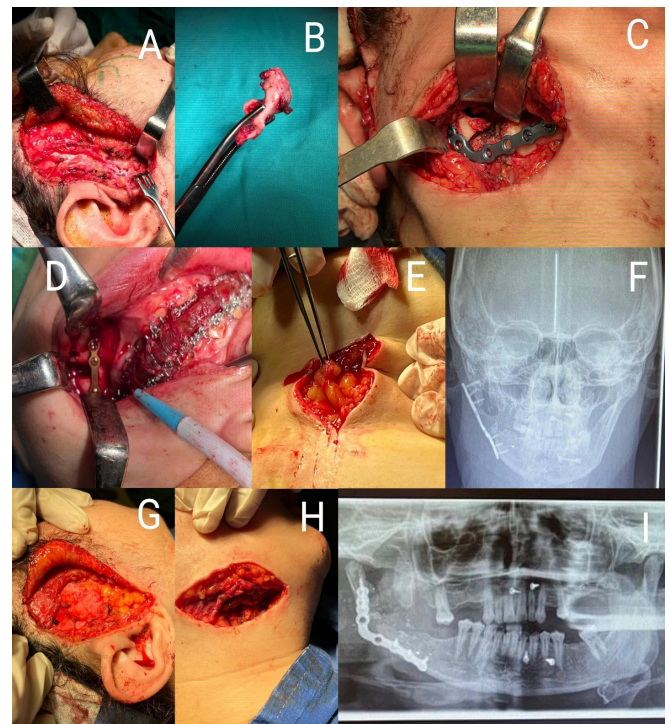


Figure 1 (A) Preauricular incision and access to the infratemporal fossa. (B) The mandible condyle after separating from coronoid process. (C) reconstructing condyle to the mandible corpus using reconstruction palate. (D) closed reduction maxillomandibular fixation. (E) Harvesting fat graft from abdominal dermis. (F) A post-Operative posteroanterior radiography. (G) Filling the preauricular region by the fat autograft. (H) Filling the region of submandibular incision by the fat autograft. (I) A Post-operative panoramic radiography showing the reconstruction plate.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



transferred to the infectious diseases department for further evaluation and treatment. After one month, the infection was successfully eliminated, and the patient was discharged in a good health. The follow-up period is ongoing and there is no discomfort reported so far.

DISCUSSION

GSWs) can affect all parts of the body, with approximately 6% involving the face, and an incidence of 87–91.5% occurring in men.⁵⁻⁸

The mandible is one of the most affected bones in the maxillofacial region following a GSW, with involvement reported in up to 65% of cases.⁹ Mandibular fractures are the most common consequence, occurring in approximately 92% of cases⁷, and these fractures are often comminuted in nature.¹⁰ Fractures can also be classify according to their line and vector of muscle traction into favorable and unfavorable, in favorable (stable) type muscle can support the fracture while unfavorable (unstable) type cause displacement.¹¹ Angle of mandible is one of the weakest regions of the mandible¹², so the percentage of angle fractures in total mandibular fractures is 33.4%.¹³

Recent findings by the World Health Organization (WHO) and its collaborators show that violence against women remains alarmingly widespread and commences at a young age. Data show that approximately one-third of women, or about 736 million individuals, experience violence during their lifetime. Shockingly, this number has remained relatively constant over the past decade.¹⁴

In Turkey, a study has demonstrated that domestic violence is a significant issue. Almost half of the women surveyed reported cases of domestic violence, and the majority of them were victims of violence.¹⁵ In this case, the patient endured violence perpetrated by her own family members, resulting in her being injured by a shotgun.

The structure of the infratemporal fossa is formed by osseous boundaries located inferiorly and medially to the zygomatic arch. It is situated deep to the ramus of the mandible. So when high violent injury such as a gunshot targets the ramus, it's expected to migrate into infratemporal fossa, and that is what happened in this case.¹⁶

Preauricular and submandibular surgical incisions were chosen to access the wound area. The preauricular incision is placed in the preauricular skin fold, which is posterior to the superficial temporal artery and the auriculotemporal nerve. This incision provides maximal lateral and anterior access to the joint. The submandibular incision is a curved incision that is made at least 2 cm inferior to mandible and is very close to the facial vein, facial artery, parotid gland, and marginal mandibular nerve. Therefore, the surgeon must protect these anatomical structures to avoid complications. For example, in the preauricular approach, injury to the auriculotemporal nerve, branch beside parotid gland, can lead to Frey's syndrome, and any pluck to artery can extend surgical time by hours in some cases.¹¹

Open reduction internal fixation (ORIF) and closed reduction MMF are used as treatment method in such situations, both have advantages and disadvantages; for example, ORIF has a higher rate of infection, which can be reduced by antibiotics, careful debridement and surgeon skill and experience. However, fewer complications occur with ORIF, such as non-union, mal-union, and facial asymmetry.^{17, 18, 19, 20}

In this case, an infection developed. Notably, the infection rate of GSWs in the maxillofacial region is significantly higher than that of GSWs in other parts of the body.²¹ And in some cases, it is difficult to predict because it depends on the patient's immunity, environmental factors, and prophylactic medication administered. Therefore, an antibiotic is routinely administered in this type of surgery.

The decision of which antibiotic to use should be based on the area of injury and the degree of wound contamination. Empiric broad-spectrum antibiotics are administered over a period of 7 to 14 days to control potential pathogens.

Additionally, the chosen antibiotic should encompass coverage against staphylococci, *Clostridium perfringens*, and *Acinetobacter baumannii*.^{22,23}

Fat grafting is a highly effective technique for the treatment of maxillofacial deformities resulting from trauma. It has the advantage of providing long-term clinical monitoring and can serve as an alternative to traditional reconstructive procedures. The use of fat grafts reduces complications and healthcare costs.²⁴ Consequently, fat grafting was performed to restore the surgical site and correct facial asymmetry in this case.



EurAsian Journal of Oral and Maxillofacial Surgery

Official publication of AÇBİD
(Association of Oral and Maxillofacial Surgery Society)



"In conclusion, this case study involved gunshot-induced displaced fractures of the mandibular ramus and condyle, which had previously resulted in unsuccessful treatment outcomes in other clinics. However, in an oral and maxillofacial surgery clinic, the case was managed through a challenging yet ultimately successful treatment process, particularly due to recurrent infections. This case may serve as a valuable reference for similar cases in other clinical settings.

REFERENCES

- Erol B, Tanrikulu R, Görgün B. Maxillofacial fractures. Analysis of demographic distribution and treatment in 2901 patients (25-year experience). *J Craniomaxillofac Surg*. 2004;32(5):308-313. doi:10.1016/j.jcms.2004.04.006
- Chrcanovic BR, Abreu MH, Freire-Maia B, Souza LN. 1,454 mandibular fractures: a 3-year study in a hospital in Belo Horizonte, Brazil. *J Craniomaxillofac Surg*. 2012;40(2):116-123. doi:10.1016/j.jcms.2011.03.012
- Grossmann E, Silva AN Jr, Collares MV. Surgical management of a projectile within the temporomandibular joint. *J Craniofac Surg*. 2012;23(2):613-615. doi:10.1097/SCS.0b013e31824cd831
- Pires MS, Giongo CC, Antonello Gde M, Couto RT, Filho Rde O, Junior OL. An interesting case of gunshot injury to the temporomandibular joint. *Craniomaxillofac Trauma Reconstr*. 2015 Mar;8(1):79-82. doi: 10.1055/s-0034-1390244. Epub 2014 Sep 15. PMID: 25709756; PMCID: PMC4329033.
- Demetriades D, Chahwan S, Gomez H, Falabella A, Velmahos G, Yamashita D. Initial evaluation and management of gunshot wounds to the face. *J Trauma*. 1998;45(1):39-41. doi:10.1097/00005373-199807000-00007.
- Lauriti, Leandro et al. "Gunshot injuries in the maxillofacial region: a retrospective analysis and management." *Brazilian Journal of Oral Sciences* 10 (2011): 236-240.
- Newlands SD, Samudrala S, Katzenmeyer WK. Surgical treatment of gunshot injuries to the mandible. *Otolaryngol Head Neck Surg*. 2003;129(3):239-244. doi:10.1016/S0194-5998(03)00481-9
- Guevara C, Pirgousis P, Steinberg B. Maxillofacial Gunshot Injuries: A Comparison of Civilian and Military Data. *J Oral Maxillofac Surg*. 2016;74(4):795.e1-795.e7957. doi:10.1016/j.joms.2015.11.007
- Sali Bukhari SG, Khan I, Pasha B, Ahmad W. Management of facial gunshot wounds. *J Coll Physicians Surg Pak*. 2010;20(6):382-385.
- Çetingül, Erdoğan. Çene ve yüz travmatolojisi. Ege Üniversitesi Diş Hekimliği Fakültesi Yayınları, 2002.
- Fonseca RJ, Marciani RD, Turvey TA. *Oral and Maxillofacial Surgery*. 2nd ed. St. Louis Mo: Saunders/Elsevier; 2009.
- Oral and Maxillofacial Anatomy, The First Affiliated Hospital School of Medicine, Zhejiang University, Prof. Dr. Gu Xinhua, <https://www.slideshare.net/deepak15/part1anatomy>
- Lee KH. Epidemiology of mandibular fractures in a tertiary trauma center. *Emergency Medicine Journal*. 2008;25:565-568
- World World Health Organization. "Devastatingly Pervasive: 1 in 3 Women Globally Experience Violence." *www.who.int*, World Health Organization, 9 Mar. 2021, www.who.int/news/item/09-03-2021-devastatingly-pervasive-1-in-3-women-globally-experience-violence.
- Basar F, Demirci N. Domestic violence against women in Turkey. *Pak J Med Sci*. 2018 May-Jun;34(3):660-665. doi: 10.12669/pjms.343.15139. PMID: 30034434; PMCID: PMC6041515
- Casale J, Bordoni B. *Anatomy, Head and Neck: Infratemporal Fossa*. [Updated 2022 Jul 25]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2023 Jan-.
- Muddassar M, Arshad R, Rabbani S, Qureshi IS, Khattak IK, Rana Z. Management of Gunshot Injuries of Mandible with Open Reduction and Internal Fixation versus Closed Reduction and Maxillo-mandibular Fixation. *Cureus*. 2020;12(4):e7830. Published 2020 Apr 25. doi:10.7759/cureus.7830.
- Siddiqui S-u-d et al., Efficacy of open reduction and internal fixation in achieving bony union of comminuted mandibular fractures caused by civilian gunshot injuries, *The Surgeon*, <https://doi.org/10.1016/j.surge.2019>.
- Rana M, Warraich R, Rashad A, et al. Management of comminuted but continuous mandible defects after gunshot injuries. *Injury*. 2014;45(1):206-211. doi:10.1016/j.injury.2012.09.021.
- Muddassar M, Arshad R, Rabbani S, Qureshi IS, Khattak IK, Rana Z. Management of Gunshot Injuries of Mandible with Open Reduction and Internal Fixation versus Closed Reduction and Maxillo-mandibular Fixation. *Cureus*. 2020;12(4):e7830. Published 2020 Apr 25. doi:10.7759/cureus.7830.
- Fagin AP, Dierks EJ, Bell RB, Cheng AC, Patel AA, Amundson MS. Infection prevalence and patterns in self-inflicted gunshot wounds to the face. *Oral Surg Oral Med Oral Pathol Oral Radiol*. 2019;128(1):9-13. doi:10.1016/j.oooo.2019.02.022.
- Powers DB, Will MJ, Bourgeois SL Jr, Hatt HD. Maxillofacial trauma treatment protocol. *Oral Maxillofac Surg Clin North Am*. 2005;17(3):341-vii. doi:10.1016/j.joms.2005.05.003.
- Sharma, Rohit & Jose, Anson. (2021). Gunshot Injuries of the Maxillofacial Region. 10.1007/978-981-15-1346-6_59.
- Arcuri F, Brucoli M, Baragiotta N, Stellin L, Giarda M, Benech A. The role of fat grafting in the treatment of posttraumatic maxillofacial deformities. *Craniomaxillofac Trauma Reconstr*. 2013 Jun;6(2):121-6. doi: 10.1055/s-0033-1333877. Epub 2013 Mar 8. PMID: 24436747; PMCID: PMC3721009.