

## Bulletin of Biotechnology

# Computational study on anti-inflammatory and anti-hypertensive drug molecules interaction with base pairs

Işıl Öztürk<sup>1</sup>, Armağan Kınal<sup>2</sup>, Toomas Tamm<sup>3</sup>

<sup>1</sup> Department of Chemistry, Faculty of Science, Ege University, Turkey

<sup>2</sup> Department of Chemistry, Faculty of Science, Ege University, Turkey

<sup>3</sup> Department of Chemistry and Biotechnology, School of Science, Tallinn University of Technology, Estonia

\*Corresponding author : isilayozturk@gmail.com  
Orcid No: <https://orcid.org/0000-0002-9134-6917>

Received : 09/05/2020  
Accepted : 16/08/2020

**Abstract:** Non-steroidal anti-inflammatory drugs (NSAIDs) and anti-hypertensive drugs have been in use for a long time for the treatment of inflammation, pain, hypertension. Besides these functions, they also show different types of other activities. Many of them exhibit critical side effects in different types of cancer such as colon, lung, and breast cancer. In the present study, we computationally investigated the interactions of some nonsteroidal anti-inflammatory and anti-hypertensive drugs (acebutolol, naproxen, diflunisal, bisoprolol) with nucleobases and interaction of drugs with nucleobase pairs by optimized at the B3LYP/6-31+G (d), B3LYP-D2/6-31+G (d) and  $\omega$ B97X-D/6-31+G (d) levels of DFT. The main purpose of this study is was to determine the strengths of drug-DNA-base interactions and drug-DNA-base pair interactions that can provide insights about the side effects of the drugs. The calculations were produced the following results. Acebutolol has the highest interaction between adenine in single base-drug complexes. However, acebutolol has the strongest interaction between the guanine-cytosine base pair. The  $\omega$ B97X-D method, which accounts for dispersion interaction properly, gives better results than the B3LYP and B3LYP-D2 methods.

**Keywords:** NSAIDs; beta blockers; nucleobase ; nucleobase pairs; binding energies; DFT methods.

© All rights reserved.

### 1 Introduction

The DNA - drug interaction has a major role in pharmacology and this interaction has a vital feature for the determination of the mechanisms of drug action. Non-covalent interactions are important for biological systems (Toupanloo and Rahmani 2018). Non-covalent DNA- interacting agents can cause a conformational change in DNA. Non-covalent interactions are weaker than a chemical bond and they have a vital role in controlling the structure and function of DNA and RNA in understanding of replication and origin of life (Ghosh et al. 2010). Therefore, interactions between drugs and nucleobases play an critical role in determining side effects (Abbas 2017-Kennard 1993). Non-steroidal anti-inflammatory drugs (NSAIDs) are a significant class of compounds to reduce inflammation and pain associated with infection or injury. Nevertheless, when NSAIDs are used for a long time, they cause various side-effects such as kidney failure, gastrointestinal problems, colon, lung, and breast cancer (Azam F et al. 2018). The  $\beta$ -blockers belong to the antihypertensives class of medications and they are mainly used to manage high blood pressure (hypertension), treat

arrhythmia and decrease the risk of heart complications after a heart attack.

The studies performed in the past few years aimed to provide an understanding of the fact that how drugs interact with the nucleobases of DNA/RNA. In 2003, Baik et al. studied how two possible hydrolysis products of cis-platinum complexes bind to adenine and guanine nucleobases. Their findings indicate that guanine is the preferred reactant for platination, and guanine is more suitable both thermodynamically and kinetically. In 2018, Toupanloo and Rahmani investigated some bicyclic fragments, which may have possible genotoxic effects, interacting with nucleobases. Hence, they focused on the  $\pi$ - $\pi$  stacking interactions and this interaction leads to both an increase and a decrease in hydrogen bond lengths in the drug-nucleobase interactions. The main objective of this study is to quantitatively determine and scale the interactions of some nonsteroidal anti-inflammatory and anti-hypertensive drugs with nucleobases with quantum chemical methods, especially with DFT methods.

## 2 Materials and Method

This study consists of two main parts: a) interaction of drugs with nucleobases b) interaction of drugs with nucleobase pairs. All structures have been optimized at the B3LYP/6-31+G (d), B3LYP-D2/6-31+G (d) and  $\omega$ B97X-D /6-31+G (d) levels of DFT. That each optimized geometry corresponds to a local minimum on the potential energy surface was identified via harmonic vibrational analyses. The effect of implicit water solvation on complex formation was calculated by employing integral equation formalism of the Polarized Continuum Model (IEF-PCM). All calculations were performed using the Gaussian09 program (<https://gaussian.com/g09citation/>)

## 3 Results and Discussion

### 3.1 Binding Enthalpies of Drugs to Nucleobases

In this part, we scrutinized the effects of each drug on a single nucleobase. In order to determine drug-nucleobase interactions, all drugs-nucleobase complexes were optimized at the B3LYP/6-31 G(d), B3LYP-D2/6-31 G(d) and  $\omega$ B97X-D/6-31 G(d) levels of DFT. The binding energies were calculated exploiting the following equation.

$$\Delta H_{bind} = H_{298}(complex) - (H_{298}(drug) + H_{298}(base)) \quad (1)$$

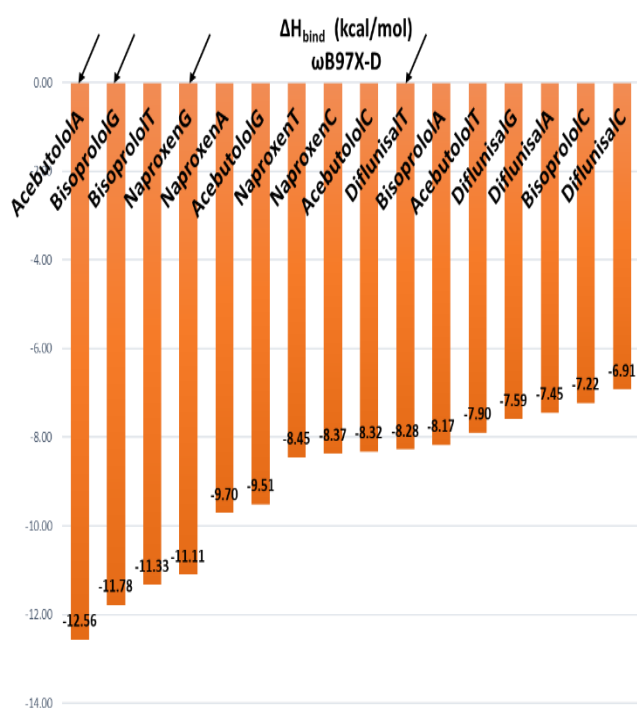
**Table 1** Binding enthalpy (kcal/mol) values of the drug–nucleobase complexes optimized with the B3LYP, B3LYP-D2 and  $\omega$ B97X-D methods.

Drug name	Base	B3LYP	B3LYP-D2	$\omega$ B97X-D
Bisoprolol	A	2.02	8.3	-8.17
	T	1.7	3.33	-11.33
	G	-2.39	5.72	-11.78
	C	0.17	5.25	-7.22
Acebutolol	A	1.44	9.03	-12.56
	T	1.02	7.74	-7.9
	G	1.57	8.29	-9.51
	C	-2.83	*	-8.32
Naproxen	A	0.91	6.24	-9.7
	T	1.03	7.07	-8.45
	G	*	3.86	-11.11
Naproxen	C	0.94	7.36	-8.37
Diflunisal	A	-5.4	7.62	-7.45
	T	-6.05	7.22	-8.28
	G	-8.59	6.15	-7.59
	C	*	6.37	-6.91

\* Calculations did not converge to a stable geometry.

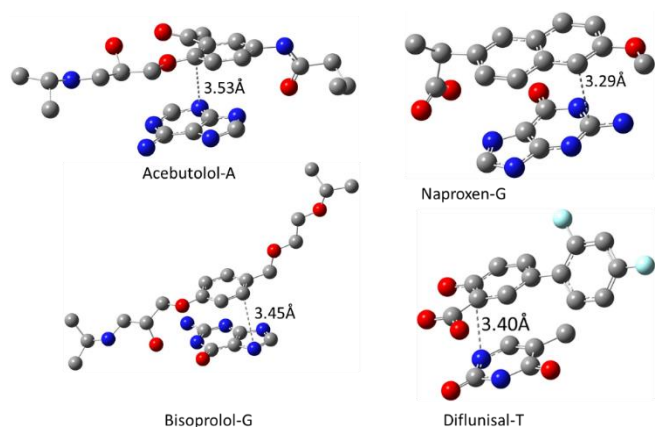
Table 1 shows the binding enthalpy values of the drug–nucleobase complexes optimized with the B3LYP, B3LYP-D2 and  $\omega$ B97X-D methods.

As seen from the table, the most negative binding energy is between adenine and acebutolol while the lowest one is between diflunisal and guanine. The  $\omega$ B97X-D method predicted that all drug-nucleobase complexes are rather stable. Our expectation was that the B3LYP-D2 method would produce similar results since the D2 part includes the contribution of dispersion interactions that are not incorporated in the B3LYP method. However, this is not the case. Although the B3LYP methods produce somewhat reasonable results, the D2 correction made them worse. Hence, it can be said that the  $\omega$ B97X-D method gives more reasonable results than the B3LYP-based methods.



**Figure 1** Highly interacting drug-base structures

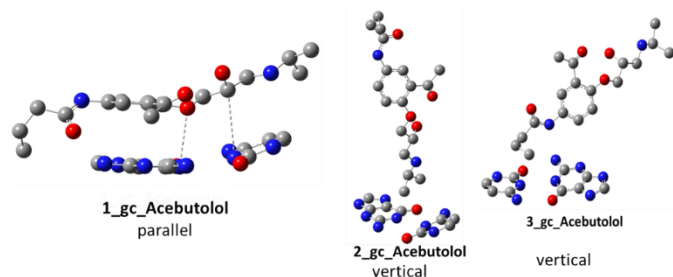
The figure shows the highly-interacting-drug-nucleobase structures obtained with the  $\omega$ B97X-D method. As previously stated, it is more likely that this method gives better results than the other two since it includes long-range interactions and it is claimed to give good results in the weakly interacting systems. Therefore, the ranking obtained with this method is presented here. Accordingly, acebutolol and adenine have the most negative binding energy among all single nucleobase–drug complexes. The second most interacting complex is the bisoprolol-guanine complex. The next two complexes are naproxen-guanine and diflunisal-thymine complexes, too.



**Figure 2**  $\omega$ B97X-D optimized structures of drug–nucleobase complexes

### 3.2 Drug-Nucleobase-pair Binding Enthalpies

In this part, we investigated the effects of each drug on nucleobase -pairs. To determine these interactions, all drugs-nucleobase pair complexes were optimized at the B3LYP/6-31G(d), and  $\omega$ B97X-D/6-31G(d) levels of theory. The binding energies were calculated using equation (1) as in the previous section.



**Figure 3** Orientations of drug–nucleobase pair complexes optimized by  $\omega$ B97X-D

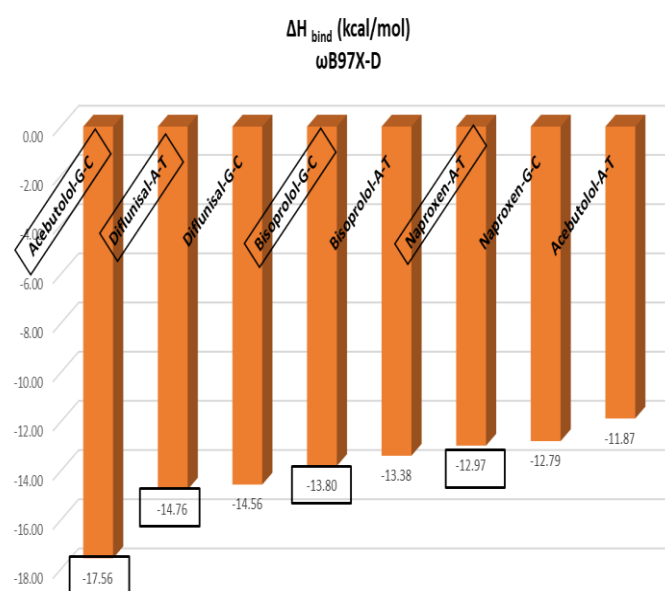
Drug molecules were interacted with nucleobase pairs in the orientations shown in Figure 3, and Table 2 shows the binding enthalpy values for all drugs that interact with the nucleobase pairs in these three orientations. The calculations predict that mostly the parallel oriented drug complexes have more negative interaction interaction than the vertical interacted ones.

As seen from the table, the strongest interaction is between Acebutolol and the G-C base pair. The  $\omega$ B97X-D method predicted that all drug-nucleobase complexes be more stable than the ones predicted with B3LYP. However, in only one case, the B3LYP method predicts the  $\pi$ - $\pi$  stacking interaction of the acebutolol drug with the A-T base pair approximately four times stronger than the one predicted by  $\omega$ B97X-D. This is most probably spurious stability originating from the inability of the B3LYP method in this calculation. Because this high binding energy (-45 kcal/mol) is larger than the covalent bond in the  $F_2$  molecule whose bond dissociation energy is about 37 kcal/mol [Ref F2\_BDE]. This clearly shows that B3LYP does not work for this particular case. Figure 4 displays the binding enthalpy order of the drug-nucleobases pairs calculated with the  $\omega$ B97X-D method.

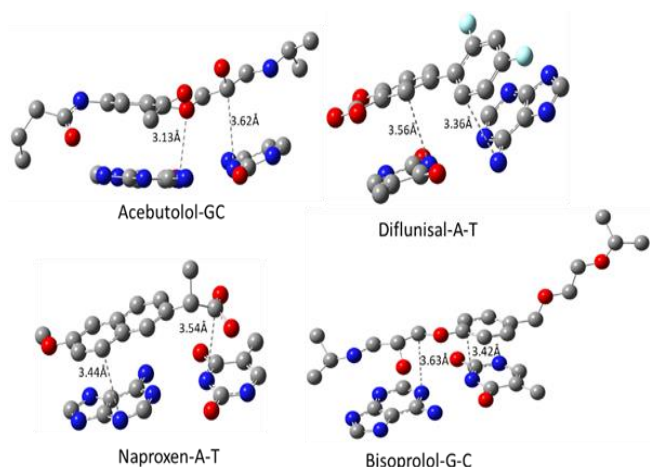
**Table 2**  $\Delta H_{\text{bind}}$  (kcal/mol) values of B3LYP and  $\omega$ B97X-D optimized drug–nucleobase pair complexes

Interreaction Site	Drug Name	Basepair	B3LYP	$\omega$ B97X-D
1_gc	Bisoprolol	G-C	1.26	-13.8
2_gc			1.74	-4.67
3_gc			1.66	-4.82
1_at	Bisoprolol	A-T	*	-13.38
2_at			1.67	-4.56
3_at			1.68	-4.66
1_gc	Acebutolol	G-C	0.63	-17.56
2_gc			0.59	-3.73
3_gc			1.13	-9.62
1_at	Acebutolol	A-T	-45.02	-11.87
2_at			-44.59	-3.67
3_at			-45.34	-6.63
1_gc	Naproxen	G-C	0.19	-9.99
2_gc			0.85	-12.79
3_gc			-1.83	-12.69
1_at	Naproxen	A-T	0.74	-10.31
2_at			1.02	-11.3
3_at			-2.29	-12.97
1_gc	Diflunisal	G-C	-6.15	-14.56
2_gc			-5.9	-11.52
3_gc			*	*
1_at	Diflunisal	A-T	-6.74	-14
2_at			-5.9	-14.76
3_at			-5.87	-10.94

\* Calculations did not converge to a stable geometry.



**Figure 4** The highly interacting drug-base pair structures



**Figure 5**  $\omega$ B97X-D optimized structures of drug–nucleobase pair complexes

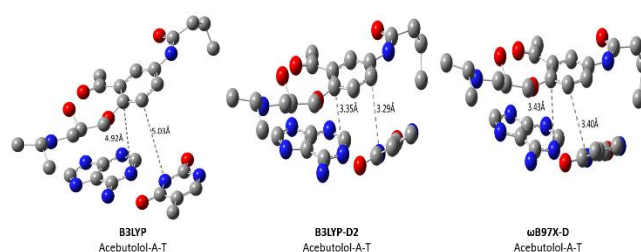
Figure 5 shows the highly-interacting -drug-nucleobase structures obtained with the  $\omega$ B97X-D method. Acebutolol and Bisoprolol, which are antihypertensive drugs, show the largest interaction with guanine-cytosine base pair. Besides, Diflunisal and Naproxen which are Non-steroidal anti-inflammatory drugs, give the largest interaction with the adenine-thymine base pair.

**Table 3.**  $\Delta H_{\text{bind}}$  (kcal/mol) values of  $\omega$ B97X-D and B3LYP, B3LYP-D2 optimized the parallel-oriented drug–nucleobase pair complexes

Interreaction site	B3LYP	B3LYP-D2	$\omega$ B97X-D
Bisoprolol-G-C	*	11.39	-13.8
Bisoprolol-A-T	1.26	11.77	-13.38
Acebutolol-A-T	-45.02	11.35	-11.87
Acebutolol-G-C	0.63	*	-17.56
Naproxen-A-T	-2.29	3.68	-12.97
Naproxen-G-C	0.85	13.29	-12.79
Diflunisal-A-T	-5.9	13.39	-14.76
Diflunisal-G-C	-6.15	12.6	-14.56

\* Calculations did not converge to a stable geometry.

Table 3 shows the parallel-oriented drug-nucleobase pair complexes that were optimized at the B3LYP/6-31G(d), B3LYP-D2/6-31G(d) and  $\omega$ B97X-D/6-31G(d) levels of DFT. All of these complexes are  $\pi$ - $\pi$  stacking complexes. Here, it is seen that the B3LYP underestimates the stability of the complexes as expected except for acebutolol A-T as explained above. However, B3LYP-D2 results are very surprising. Although we performed D2 calculations to improve the results of B3LYP method, B3LYP-D2 made them worse as in the single base-drug interaction case.



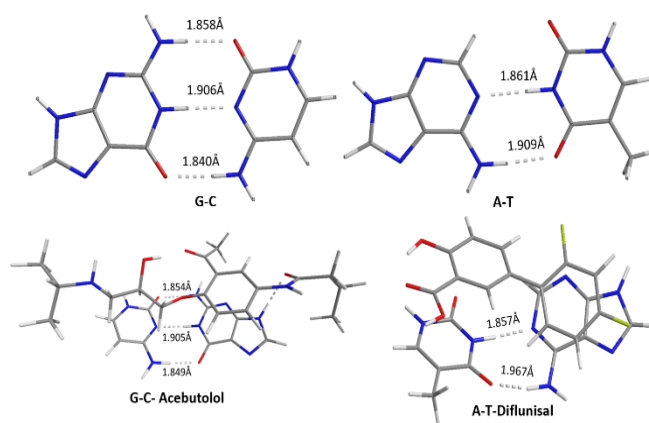
**Figure 6** Optimized structures of drug–nucleobase pair complexes

**Table 4** Distances between hydrogen bonds of  $\omega$ B97X-D optimized drug–G-C basepair complexes

Angstrom	O-H	N-H	O-H	difference	difference	difference
G-C	1.840	1.906	1.858	0.000	0.000	0.000
G-C- Acebutolol	1.849	1.905	1.854	0.009	-0.001	-0.004
G-C-Bisoprolol	1.857	1.899	1.866	0.018	-0.007	0.009

**Table 5** Distances between hydrogen bonds of  $\omega$ B97X-D optimized drug– A-T basepair complexes

Angstrom	O-H	N-H	difference	difference
A-T	1.909	1.861	0.000	0.000
A-T-Diflunisal	1.967	1.857	0.058	-0.004
A-T-Naproxen	1.863	1.896	-0.045	0.034



**Figure 7** Intermolecular interaction of drug–nucleobase pair complexes

Table 4 shows that the acebutolol affects the bond lengths of the hydrogen bonds between the G-C base pair. On the other hand, the diflunisal affects the bond lengths of the hydrogen bonds between the adenine-thymine base pairs, shown in Table 5 Accordingly, it is likely that these drugs may cause DNA breakage in long-term usage.

#### 4 Conclusion

In this study, we calculated drug-nucleobase and drug nucleobase base-pair interactions with various DFT methods, and we aimed to quantitatively determine the strengths of drug-DNA-base interactions that may improve our understandings about the side effects of the drugs. The most negative binding energy among the drug-single base complexes is between acebutolol and adenine. In the base pair drug complexes, however, acebutolol has the strongest attractive interaction between the guanine-cytosine base pair. Furthermore, we can conclude that the  $\omega$ B97X-D method gives better results for such non-covalently interacting systems than both the B3LYP and B3LYP-D2 methods since this method accounts for dispersion interaction more accurately.

#### 5 Acknowledgements

The authors thank the all calculations reported in this paper were performed at High Performance and Grid Computing Center (TRUBA resources). I.Ö. wishes to thank Armağan Kınal and Toomas Tamm.

#### References

- Abbas H (2017) First-principles study of interaction of serine with nucleobases of DNA and RNA. *J Biol Phys*, 43(1):105–111
- Azam F, Alabdullah NH, Ehmedat HM (2018) NSAIDs as potential treatment option for preventing amyloid  $\beta$  toxicity in Alzheimer's disease: an investigation by docking, molecular dynamics and DFT studies. *Journal of Biomolecular Structure and Dynamics* 36(8):2099–2117
- Baik MH, Richard AF and Stephen JL (2003) Theoretical Study of Cisplatin Binding to Purine Bases: Why Does Cisplatin Prefer Guanine over Adenine? *JACS* 125(46): 14082–92.
- Foresman JB and Frisch A (1996) *Exploring Chemistry with Electronic Structure Methods*, 2nd Edition, Gaussian, Inc, Pittsburgh
- Frisch E, Hratchian HP, Dennington RD, Keith TA, Millam J, Nielsen AB, Holder AJ, Hiscocks J (2009) *GaussView 5 Reference*, Gaussian, Inc, Wallingford
- Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Mennucci B, Petersson GA, Nakatsuji H, Caricato MLX, Hratchian HP, Izmaylov AF, Bloino J, Zheng G, Sonnenberg JL, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery JA, Peralta JE, Ogliaro F, Bearpark M, Heyd JJ, Brothers E, Kudin KN, Staroverov VN, Keith TA, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant JC, Iyengar SS, Tomasi J, Cossi M, Rega N, Millam JM, Klene M, Knox JE, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Martin RL, Morokuma K, Zakrzewski VG, Voth GA, Salvador P, Dannenberg JJ, Dapprich S, Daniels AD, Farkas O, Foresman JB, Ortiz JV, Cioslowski J and Fox DJ (2010) *Gaussian 09, Revision B.01*, Gaussian, Inc, Wallingford
- Gaussian 09 Citation <https://gaussian.com/g09citation/> Accessed 4 Feb 2014
- Ghosh D, Kosenkov D, Vanovschi V, Williams C F, Herbert J M, Gordon M S, Krylov A I (2010) Noncovalent Interactions in Extended Systems Described by the Effective Fragment Potential Method: Theory and Application to Nucleobase Oligomers. *J. Phys. Chem.* 114(48):12739–12754.
- Kennard O (1993) DNA-drug interactions. *Pure and App Chem* 65(6):1213–1222
- Toupanloo HA and Rahmani Z (2018) An in-depth study on noncovalent stacking interactions between DNA bases and aromatic drug fragments using DFT method and AIM analysis: conformers, binding energies, and charge transfer. *Appl Biol Chem* 61(2):209–226
- Wenthold PG, Squires RR (2005) Bond Dissociation Energies of F<sub>2</sub> and HF<sub>2</sub> A Gas-Phase Experimental and G2 Theoretical Study. *Phys Chem* 9(7):2002–2005