Investigation of Serum and Wool Levels of Cobalt, Manganese, Selenium and Zinc in Liver-Trematode-Infected Sheep

Selçuk Seçkin TUNCER*, Vural DENİZHAN2, Süleyman KOZAT3
1Van Yuzuncu Yıl University, Agricultural Faculty, Department of Animal Sciences, Van, Turkey
2Van Yuzuncu Yıl University, Özalp Vocational School, Department of Medical Laboratory Technician, Van, Turkey
3Van Yuzuncu Yıl University, Faculty of Veterinary Medicine, Department of Internal Medicine, Van, Turkey

ABSTRACT
In this study, the hematological and biochemical blood values and the differences in serum and wool trace elements (cobalt [Co], manganese [Mn], zinc [Zn], and selenium [Se]) were compared in healthy and liver-trematode-infected sheep. A total of 100 ovines (80 trematode-infected and 20 healthy sheep) were included. The trematode-infected sheep had significantly greater \((P<0.01)\) leucocyte (WBC), neutrophil (Neu), and eosinophil (Eo) values and significantly lower \((P<0.01)\) erythrocyte (RBC), hematocrit (Hct), and hemoglobin (Hb) values when compared with the healthy control group. The trematode-infected sheep had significantly higher serum alanine aminotransferase (ALT), aspartate aminotransferase (AST), and gamma glutamyl transferase (GGT) enzyme activity values when compared with the healthy control group \((P<0.01, P<0.05, \text{ and } P<0.01, \text{ respectively})\). Serum and wool Co, Mn, Zn, and Se levels were significantly lower \((P<0.01)\) in the trematode-infected sheep than in the healthy control group. The effect of liver infections on trace element concentrations was determined to be similar in measurements in wool and serum. Decreases in trace element concentrations were mostly attributed to changes in the biotransformation of trace elements induced by pathologic disorders in the liver.

Keywords: Liver-trematode, Serum, Sheep, Trace element, Wool.

To cite this article: Tuncer S.S. Denizhan V. Kozat S. Investigation of Serum and Wool Levels of Cobalt, Manganese, Selenium and Zinc in Liver-Trematode-Infected Sheep. Kocatepe Vet J. (2020) 13(2):185-191

Submission: 02.03.2020 Accepted: 20.05.2020 Published Online: 24.05.2020

ORCID ID; SST: 0000-0001-8252-8009, VD: 0000-0002-0531-9550, SK: 0000-0001-5089-2623

*Corresponding author e-mail: selcukseckintuncer@gmail.com
INTRODUCTION

Liver trematode infections decrease animal growth, reduce yields, suppress the immune system, and ultimately cause death in cases of severe infection. Consequently, these infections generate significant economic losses every year (Levieux et al. 1992, Matanovic et al. 2007). Trematodes migrating to the liver may deplete potential glycogen reservoirs, as the liver is the primary glycogen depot of animals, so these parasites can cause severe destruction of hepatocytes (Phiri et al. 2007, Kozat and Denizhan 2010). The most precise indicators of the level of liver cell damage in liver diseases, including trematode infections, are the activities of glutamate dehydrogenase (GLDH) and gamma glutamyl transferase (GGT) (Kozat et al. 2006, Mert et al. 2006, Kozat and Denizhan 2010). Plasma GLDH and GGT activities are therefore considered better and more sensitive indicators of liver cell damage, such as that caused by sub-clinical and chronic fascioliasis, when compared to aspartate aminotransferase (AST) activity (Sykes et al. 1980). In general, GGT might be more appropriate for the diagnosis of liver cell damage due to its greater stability.

A healthy liver is essential for the preservation of animal health and growth performance (Chalabis-Mazurek and Walkuska 2014). In particular, the liver organ plays a central role in the use and distribution of macro and micro elements absorbed from the intestines, and animal health depends on sufficient and well-balanced trace element concentrations. The serum cobalt (Co) and zinc (Zn) levels in ruminant animals are directly related to the Co and Zn concentrations of the animals diets (Jacob 1987, Oldfield 1987). These two trace elements play significant roles in hair follicle maturation, ovulation, and the estrus cycle (Unal 1987), so their deficiencies can generate significant animal yield losses and consequent economic losses for growers (Şendil et al. 1975, Jacob 1987, Aytuğ et al. 1990). Zn, in particular, serves as a cofactor for more than 300 enzymes and plays a substantial role in growth, DNA synthesis, immune system performance, neuro-sensory functions, and several other cellular process in both humans and animals (Kozat 2007). Zn deficiency reduces cell division and appetite, decreases growth and development, and generates parakeratosis lesions over the skin (Kaneko et al. 1997). Zn is present in all animal tissues and particularly in muscles, bones, blood, glands, genital organs, skin, hair, wool, and nails (Gabor 1991).

Cobalt is an essential trace element in ruminants, as it is required for vitamin B<sub>12</sub> synthesis in the rumen. Unlike the case in other domesticated animals, this vitamin is synthesized by the microorganisms of the rumen proventriculus, so sufficient quantities of cobalt are required in the diet (Stangl et al. 1999). Co absorption is quite low in ruminants, and only 3% of the Co taken up is synthesized into vitamin B<sub>12</sub> and only 3% of the synthesized vitamin B<sub>12</sub> is absorbed (NCR 1987). Vitamin B<sub>12</sub> synthesis by microorganisms is totally inhibited when the Co content of the rumen fluid is less than 0.5 ng/ml (Underwood 1977). The trace element concentrations are significantly lower in sheep with liver cystic echinococcosis than in healthy sheep (P <0.01) (Taşçı et al. 1995). Decreases in serum trace element levels also occur in animals afflicted with parasitic diseases, mostly due to malnutrition, transfer of parasites to other sections of the body, or substantial uptake of these elements by the parasites (Seyrek et al. 2009).

Manganese is also an essential trace element for animals (Kozat 2007). It is required for the activity of glucosidases enzymes that form the mucopolysaccharide chondroitin that is involved in cartilage, cartilage activity, and bone formation (Miranda et al. 2006). Absorption of Mn in the gastrointestinal tract depends on its chemical form. Absorbed Mn is initially transported to the liver and then secreted into the intestines with the gall. The Mn secreted into intestines is reabsorbed through the enterohepatic cycle. The absorbed Mn is initially sequestered in the mitochondria of the liver, kidneys, and pancreas. About 40% of the body Mn is preserved in the bone marrow (Kozat 2007).

Selenium is another important biochemical component, as it forms part of the structure of glutathione peroxidase enzyme. Together with vitamin E, this enzyme inhibits oxidative processes that destroy cells and tissues (Değer et al. 2008).

The trace element contents of animal tissues are determined from serum, wool, hair, and liver and kidney tissues (Baysu et al. 1984). Determination of mineral levels can aid in determining preventive health measures, thereby maintaining high animal yields (Spears 2003), so several studies have investigated the relationships of trace element deficiencies with these yields (White et al. 1994). In sheep, trace element deficiencies result in yield losses, fleece abnormalities, and various malnutritional problems (Kozat 2007). Minerals taken up by the animal, together with vitamins, play critical roles in the healthy development of fetuses and young, in the improvement of yield and resistance, in the sustainability of production, and in the performance of various metabolic functions. Trace elements, and especially those with co-factor roles, are of significant importance for the functions of metalloenzymes (Şendil et al. 1975, Jacob 1987).

The deficiency or abundance of trace elements can therefore result in a number of functional disorders (Aytuğ et al. 1990). In the present study, changes in serum and wool trace element concentrations were
investigated in sheep with disorders in the liver induced by liver trematode infection. According to the results of the research, it is thought that the determination of trace element in wool instead of serum in animals with liver infection will be beneficial for animal welfare.

**MATERIAL and METHODS**

**Animals and Sampling**
The final report of this research study was approved (2017/12) by Van Yuzuncu Yil University Animal Researches Local Ethic Committee. This study was conducted on 8 sheep farms with an average size of 200-280 ewes. Animal feces samples were analyzed to select 80 sheep with trematode infection and 20 healthy sheep. This selection was conducted by collecting 30-50 g feces samples from the rectums into sample cups and analyzing them in the laboratory using the Benedek Sedimentation method (Toparlağ and Tüzer 1994). After taking feces and blood samples, 10 mg / kg triclabendazole + 7.5 mg / kg levamisole were administered orally to sheep infected with trematodes.

From blood-sampled sheep, wool samples (about 5 g) were also taken from the occipital area with steel scissors. While taking the wool samples, the wool was cut only from the bottom to eliminate potential differences in Co, Mn, Se, and Zn concentrations in the top, middle, and bottom sections of the wool. Samples were placed into polyethylene bags and kept sealed until analysis.

**Laboratory Analyses**
Blood samples were taken from trematode-infected and healthy sheep in accordance with the relevant techniques for hematologic and biochemical analyses. Samples were collected into anticoagulant tubes and analyzed for hematocrit value (Hct), hemoglobin concentration (Hb), and leukocyte (WBC), erythrocyte (RBC), eosinophil (Eo), and neutrophil (Neu) ratios with a veterinary hemogram device (Veterinary MS4-s-Melet Schloesing Laboratories in France) in the Animal Hospital laboratory.

**Biochemical Analyses**
Blood samples collected into non-anticoagulant tubes (biochemistry tubes) were centrifuged at 3000 rate per minute (rpm) for 10 minutes to obtain blood sera, which were then stored in serum preservation tubes in a fridge at -20 °C until biochemical analysis (Özdemir et al. 2014). Serum AST, alanine aminotransferase (ALT), and GGT levels were measured with autoanalyzer device (BS-120 Vet-Mindray). Serum Co, Mn, Zn, and Se concentrations were measured with an Inductively Coupled Plasma – Mass Spectrometer (ICP-MS) instrument in the Scientific Research Center of University.

**Analysis of Wool Samples**
Wool samples were initially washed with a 1% Triton-X 100 solution four times and then rinsed with double distilled deionized water. The washed samples were then dried in a sterilizer at 100 °C for two hours. About 100 mg of the dried wool samples were placed into tubes, 1 ml of a 1.5 nitric acid + perchloric acid mixture was added, and the samples were allowed to dissolve for four hours. The dissolved mixtures were made up to 10 ml with distilled water. From this mixture, 1 ml was removed and diluted with 2 ml distilled water. The prepared and diluted (1:50) wool samples were then subjected to Co, Mn, Zn, and Se analyses with an ICP-MS instrument in the Scientific Research Center of University (Kozat 2007).

**Statistical Analysis**
Statistical analyses were performed using the GLM sub-procedure of the SAS 9.4 statistical software (SAS 2018). Data from serum and wool samples were subjected to variance analysis, and significant means were compared with Duncan’s multiple range test.

**RESULTS**

**Clinical Results**
Inspections of the feces revealed *Fasciola hepatica* and *Dicrocoelium dendriticum* eggs. The trematode-infected sheep showed inappetence, weight loss, poor performance, reductions in milk yield and fleece quality, and digestive disorders; some sheep had diarrhea and anemia.

**Hematological Results**
Data for hematological parameters of the trematode-infected and healthy sheep are provided in Table 1. The WBC, Neu, and Eo levels were significantly higher in the trematode-infected sheep (13.91, 9.28, and 0.73, respectively) than in the control group (8.60, 5.80, and 0.18, respectively) (P<0.01), but the RBC (×10<sup>6</sup> µL), Hct (%), and Hb (g/dL) values were lower in the infected sheep (9.80, 25.90, and 9.75, respectively) than in control group (11.80, 33.00, and 13.41, respectively) (P<0.01).

**Biochemical Results**
Statistical analysis results for blood serum biochemical parameters of infected and healthy sheep are provided in Table 2. The ALT (30.60 U/L), AST (148.25 U/L), and GGT (80.44 U/L) values were significantly greater in the trematode-infected sheep than in the healthy sheep (17.70 U/L, 94.20 U/L, and 54.20 U/L, respectively; P<0.01, P<0.05, and P<0.01, respectively).

Data for the serum mineral levels of trematode-infected and healthy sheep are provided in Table 3. The serum Co, Mn, Se and Zn levels were significantly lower (P<0.01) in the infected sheep (2.50 µg/dL, 31.00 µg/dL, 2.73 µg/dL, 31.12 µg/dL, respectively) than in the control group (2.50 µg/dL, 31.00 µg/dL, 2.73 µg/dL, 31.12 µg/dL, respectively).
respectively) than in the healthy controls (4.90 µg/dl, 58.00 µg/dl, 4.69 µg/dl, 83.76 µg/dl, respectively).

Data for the wool mineral levels of trematode-infected and healthy sheep are provided in Table 4. The mineral (Co, Mn, Se and Zn) levels were significantly lower (P<0.01) in the infected sheep (0.03 µg/g, 4.22 µg/g, 1.35 µg/g and 26.71 µg/g, respectively) than in the healthy controls (0.06 µg/g, 10.61 µg/g, 4.66 µg/g and 45.96 µg/g, respectively).

The correlations between serum trace element levels of the healthy sheep are provided in Table 5. Significant positive correlations were observed between Se and Co (0.948; P<0.01) and significant negative correlations were observed between Se and Zn (-0.647; P<0.05).

The correlations between serum trace element levels of the trematode-infected sheep are provided in Table 6. Significant positive correlations were observed between Se and Zn (0.340; P<0.01), and significant negative correlations were observed between Zn and Mn (-0.624; P<0.01) and between Se and Mn (-0.227; P<0.05). No significant correlations were noted between the wool trace element levels in healthy sheep (Table 7). The correlations between wool trace element levels of liver trematode-infected sheep are provided in Table 8. Significant positive correlations were noted between Zn and Mn (0.06); between Se and Zn (0.521; P<0.01).

**Table 1.** Hematologic blood parameters of healthy control sheep and sheep infected with liver trematodes

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control (n=20)</th>
<th>Trematode infected (n=80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBC (x10^3/µl)</td>
<td>8.60±0.60&lt;sup&gt;a&lt;/sup&gt;</td>
<td>13.91±2.27&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Neu (%)</td>
<td>5.80±0.34&lt;sup&gt;a&lt;/sup&gt;</td>
<td>9.28±1.58&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Eo (%)</td>
<td>0.18±0.03&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.73±0.06&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>RBC (x10^6 µl)</td>
<td>11.80±0.50&lt;sup&gt;a&lt;/sup&gt;</td>
<td>9.80±0.48&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Hct (%)</td>
<td>33.00±2.27&lt;sup&gt;a&lt;/sup&gt;</td>
<td>25.90±2.33&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Hb (g/dl)</td>
<td>13.41±1.33&lt;sup&gt;a&lt;/sup&gt;</td>
<td>9.75±1.76&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>a, b</sup>: Means with different lower case letters in the same row are significantly different

<sup>: P<0.01</sup>


**Table 2.** Biochemical parameters of healthy control sheep and sheep infected with liver trematodes

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control (n=20)</th>
<th>Trematode infected (n=80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT (U/L)</td>
<td>17.70±2.32&lt;sup&gt;a&lt;/sup&gt;</td>
<td>30.60±6.15&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>94.20±18.79&lt;sup&gt;a&lt;/sup&gt;</td>
<td>148.25±60.14&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>GGT (U/L)</td>
<td>54.20±5.55&lt;sup&gt;a&lt;/sup&gt;</td>
<td>80.44±16.12&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>a, b</sup>: Means with different lower case letters in the same row are significantly different

<sup>: P<0.05; **P<0.01</sup>

ALT: alanine aminotransferase, AST: aspartate aminotransferase, GGT: gamma glutamyl transferase.

**Table 3.** Serum trace element levels in healthy control sheep and sheep infected with liver trematodes

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control (n=20)</th>
<th>Trematode infected (n=80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co (µg/dl)</td>
<td>4.90±0.80&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.50±0.50&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Mn (µg/dl)</td>
<td>58.00±5.10&lt;sup&gt;b&lt;/sup&gt;</td>
<td>31.00±10.10&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Se (µg/dl)</td>
<td>4.69±1.20&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.73±1.29&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Zn (µg/dl)</td>
<td>83.76±43.64&lt;sup&gt;b&lt;/sup&gt;</td>
<td>31.12±10.32&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>a, b</sup>: Means with different lower case letters in the same row are significantly different

<sup>: P<0.05; **P<0.01</sup>


**Table 4.** Wool trace element levels in healthy control sheep and sheep infected with liver trematodes

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control (n=20)</th>
<th>Trematode infected (n=80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co (µg/g)</td>
<td>0.06±0.01&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.03±0.01&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Mn (µg/g)</td>
<td>10.61±3.89&lt;sup&gt;b&lt;/sup&gt;</td>
<td>4.22±1.78&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Se (µg/g)</td>
<td>4.66±2.17&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.35±0.50&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Zn (µg/g)</td>
<td>45.96±16.62&lt;sup&gt;b&lt;/sup&gt;</td>
<td>26.71±12.80&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>a, b</sup>: Means indicated with different small letters in the same row are significantly different

<sup>: P<0.05; **P<0.01</sup>

**Table 5.** Correlations between serum trace element levels of healthy sheep

<table>
<thead>
<tr>
<th>Co</th>
<th>Mn</th>
<th>Se</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn</td>
<td>-0.123</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Se</td>
<td>0.948&lt;sup&gt;**&lt;/sup&gt;</td>
<td>-0.297</td>
<td>1</td>
</tr>
<tr>
<td>Zn</td>
<td>-0.534</td>
<td>0.362</td>
<td>-0.647&lt;sup&gt;*&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>: P<0.05; **P<0.01</sup>

**Table 6.** Correlations between serum trace element levels of trematode-infected sheep

<table>
<thead>
<tr>
<th>Co</th>
<th>Mn</th>
<th>Se</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn</td>
<td>0.164</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Se</td>
<td>0.019</td>
<td>-0.227&lt;sup&gt;*&lt;/sup&gt;</td>
<td>1</td>
</tr>
<tr>
<td>Zn</td>
<td>-0.113</td>
<td>-0.624&lt;sup&gt;**&lt;/sup&gt;</td>
<td>0.340&lt;sup&gt;**&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>: P<0.05; **P<0.01</sup>

**Table 7.** Correlations between wool trace element levels in healthy sheep

<table>
<thead>
<tr>
<th>Co</th>
<th>Mn</th>
<th>Se</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn</td>
<td>-0.128</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Se</td>
<td>-0.236</td>
<td>0.493</td>
<td>1</td>
</tr>
<tr>
<td>Zn</td>
<td>-0.090</td>
<td>0.169</td>
<td>0.496</td>
</tr>
</tbody>
</table>

**Table 8.** Correlations between wool trace element levels in liver trematode-infected sheep

<table>
<thead>
<tr>
<th>Co</th>
<th>Mn</th>
<th>Se</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn</td>
<td>-0.120</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Se</td>
<td>0.094</td>
<td>0.069</td>
<td>1</td>
</tr>
<tr>
<td>Zn</td>
<td>-0.210</td>
<td>0.449&lt;sup&gt;**&lt;/sup&gt;</td>
<td>0.521&lt;sup&gt;**&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>: P<0.01</sup>
DISCUSSION

The RBC, Hct, and Hb values were lower in the trematode-infected sheep than in the healthy sheep (Table 1), in agreement with the literature (Vengust et al. 2003; Samadieh et al. 2017). Of the hematological parameters of healthy and trematode-infected sheep, the WBC, RBC, Hct, and Hb levels were in compliance with the reference values specified for sheep, whereas the Neu and Eo levels were lower than the reference values (Babeker and Elmansoury 2013).

The higher ALT, AST, and GGT levels in the trematode-infected sheep than in the healthy sheep (Table 2) can be attributed to enzyme leakage from destroyed organs (Gerber 1969). Increasing AST and ALT activities have been reported previously in lambs infected with 1000 and 3000 metacercariae of D. dendriticum (Manga-González et al. 2004). The ALT, AST, and GGT values of the trematode-infected sheep in the present study were similar to those reported in the literature for liver parasite infections (Mert et al. 2006, Kozat and Denizhan 2010). The ALT, AST, and GGT values of the healthy sheep were also in compliance with the values reported in similar studies (Kozat and Denizhan 2010, Samadieh et al. 2017). The mean GGT value for the trematode-infected sheep (80.44 U/L) was greater than the specified reference values (20–52 U/L), but the GGT value for the healthy sheep (54.20 U/L) was similar to the reference values. The ALT (17.70 U/L) and 30.60 U/L) and AST (94.20 U/L and 148.25 U/L) values of the healthy and infected animals were in compliance with the normal reference values (26-34 U/L and 60-280 U/L, respectively) (Kahn and Line 2011).

Trace elements are micronutrients necessary for the growth and preservation of healthy tissues. Blood is a medium where trace elements are collected and transported. Therefore, serum is usually a suitable sample for determining the trace element status of animals (Özdemir et al. 2014). However, mineral measurements from tissue, hair, or wool may provide more accurate results than serum analyses (Bayşu et al. 1984).

The serum Zn levels in the healthy sheep (83.76 µg/dl) (Table 3) were in compliance with the reference values (80-150 µg/dl) (Kurt et al. 2001, Erdoğan et al. 2002), but the serum Zn levels in the trematode-infected sheep (31.12 µg/dl) (Table 3) were lower than the reference values. Decreasing plasma Zn levels were observed in echinococcosis diseases (Heidarpour et al. 2012), in response to the accelerated metabolism of animals during the progression of disease (Beisel 1991), and in insufficient nutrition, stress, and hyperthermia cases (Heidarpour et al. 2012). Another study reported that serum Zn levels decreased in parasitic diseases (Taşçı et al. 1995). The low Zn levels observed in the trematode-infected sheep in the present study were also attributed to these reasons.

The serum Co concentrations in the healthy control group (4.90 µg/dl) (Table 3) were within the reference value range. The observed decreases in Co concentrations in the trematode-infected sheep (2.50 µg/dl) (Table 3) agreed with the available literature (Taşçı et al. 1995, Seyrek et al. 2009).

The serum Se levels were significantly greater in the healthy sheep (4.69 µg/dl) than in the trematode-infected sheep (2.73 µg/dl) (P<0.05) (Table 3). The serum Se levels of healthy sheep were in compliance with the values specified for sheep, at 5.37µg/dl (Yokus et al. 2004), and for lambs, between 6.0µg/dl (Kozat 2007) and 5.65µg/dl (Değer et al. 2008).

Decreasing plasma Zn levels were reported in Echinococcus-infected camels (Heidarpour et al. 2012), during the disease periods of animals (Beisel 1991), in animals with insufficient Zn nutrition, and in stress and hyperthermia cases (Heidarpour et al. 2012). In the present study, low Zn concentrations were determined in sheep infected with liver trematodes (31.12 µg/dl) (Table 3), in agreement with previous reports (Beisel 1991, Taşçı et al. 1995, Humann–Ziehank et al. 2008, Heidarpour et al. 2012).

CONCLUSIONS

In conclusion, highly significant decreases were observed in blood serum Co and Mn levels (P<0.01) and significant decreases were observed in Se and Zn levels in sheep infected with liver trematodes (P<0.05), with highly significant decreases observed all trace element levels in wool (P<0.01). The effect of liver infections on trace element concentrations was determined to be similar in measurements in wool and serum. This result shows that trace elements can be determined in wool instead of serum for animal welfare. Decreases in trace element concentrations were mostly attributed to changes in the biotransformation of trace elements induced by pathologic disorders in the liver.
ACKNOWLEDGEMENT

This study was supported by Research Fund of the Van Yüzüncü Yıl University. Project number FBA-2018-6823.

The final report of this research project was approved by Van Yüzüncü Yıl University Animal Research Local Ethics Committee, decision number 2017/12.

Conflict of Interest: The authors declare that they have no conflict of interest.

REFERENCES


Mert H, Kożat S, Ekin S, Yoruk İH. Serum sialic acid, lipid-bound sialic acid levels in sheep naturally chronically infected with Fasciola hepatica. YYU Health Sci J. 2006; 2: 40–46.


Toparlar M, Tüzer E. Laboratory Techniques in the Diagnosis of Parasitic Diseases. Istanbul University Faculty of Veterinary Medicine Publication, Istanbul, Turkey. 1994; p. 19.


