**ORIGINAL ARTICLE / ÖZGÜN MAKALE** 



# ANTIOXIDANT, ANTIMICROBIAL AND ANTI-PROLIFERATIVE ACTIVITY OF *SUILLUS LUTEUS* (L.) ROUSSEL EXTRACTS

## SUILLUS LUTEUS (L).ROUSSEL EKSTRESİ'NİN ANTİOKSİDAN, ANTİMİKROBİYAL VE ANTİ-PROLİFERATİF AKTİVİTESİ

## Erdi Can AYTAR<sup>1,\*</sup>, Ilgaz AKATA<sup>2</sup>, Leyla AÇIK<sup>3</sup>

<sup>1</sup> Ondokuz Mayıs University, Faculty of Art and Science, Department of Biology, 55200, Samsun,

Turkey

<sup>2</sup>Ankara University, Faculty of Science, Department of Biology, 06560, Ankara, Turkey

<sup>3</sup>Gazi University, Faculty of Science, Department of Biology, 06500, Ankara, Turkey

## ABSTRACT

**Objective:** Many drug discovery have used nature as an inspiration for the design of naturel products like compound classes. From ancient times edible mushrooms have been used both as food and medicine. People living in Turkey widely consume Suillus luteus (L.) Roussel wild edible mushrooms In this study, we were investigated antioxidant, antimicrobial and cytotoxic activities of various extracts of S.luteus.

**Material and Method:** Antioxidant activity of S.luteus was detected method by DPHH free radical scavenging and ferrous ion chelating ability. In addition, the concent of the components with antioxidant properties, such as total phenols, $\beta$ -caratone and lycopene were determined by spectrophotometric methods. The antimicrobial potential was demonstrated with a agar well diffusion method on 14 microorganisms. Finally, the cytotoxic effect of methanolic extract of S. luteus on MCF-7 cancer cell lines were evaluated by using MTT method.

**Result and Discussion:** The results indicated that S.luteus methanolic and ethanolic extracts have more abundant phenols (153, 49.33 mg GAE/g extract, respectively).In addition  $\beta$ -caratone and lycopene content detected. (from 0.120 to 0.606µg/mL).S.luteus extracts had more potent free radical scavenging activity than standard antioxidants BHT. (Methanol extract (IC<sub>50</sub>: 63.72µg/mL) > Ethanol extract (IC<sub>50</sub>: 80.72 µg/mL) > BHT (IC<sub>50</sub>: 96.47µg/mL). In addition, methanol extracts possessed higher ferrous ion chelating ability than ethanol extracts(2.72, 3.45 µg/mL, respectively) .Generally, the tested mushroom extracts had relatively low antimicrobial activity against the tested microorganisms (9 and 10 mm zone diameter). Also, S.luteus methanolic extract was found to kill all cancer cells at a concentration of 1mg/mL. These results showed that S.luteus, especially methanol extracts, have potential medical.

Keywords: antimicrobial activity, antioxidant activity, anti-proliferative activity, Suillus luteus

Corresponding Author / Sorumlu Yazar: Erdi Can Aytar e-mail / e-posta: erdicanaytar@gmail.com, Phone / Tel.: +905326459086

Submitted / Gönderilme: 20.03.2020

Accepted / Kabul: 12.06.2020

## ÖΖ

**Amaç:** Birçok ilaç keşfinde doğa, doğal ürünler benzeri bileşik sınıflarının tasarımına ilham kaynağı olarak kullanmıştır. Eski zamanlardan beri yenilebilir mantarlar hem gıda hem de ilaç olarak kullanılmıştır. Türkiye'de yaşayan insanlar Suillus luteus (L.) Roussel yabani yenilebilir mantarlarını yaygın olarak tüketmektedir. Bu çalışmada, çeşitli S.luteus ekstraktlarının antioksidan, antimikrobiyal ve anti-proliferatif aktiviteleri araştırıldı.

**Gereç ve Yöntem:** S.luteus'un antioksidan aktivitesi, DPHH serbest radikal süpürme yöntemi ve demir iyonu şelatlama kabiliyeti saptandı. Ek olarak, toplam fenoller, β-karoten ve likopen gibi antioksidan özelliklere sahip bileşenlerin konsantrasyonu spektrofotometrik yöntemlerle belirlenmiştir. Antimikrobiyal potansiyel, 14 mikroorganizma üzerinde agar difüzyon yöntemi ile gösterilmiştir. Son olarak, S. luteus metanol ekstresinin MCF-7 kanser hücre hatları üzerindeki sitotoksik etkisi MTT yöntemi kullanılarak değerlendirildi.

**Sonuç ve Tartışma:** Sonuçlar S.luteus metanolik ve etanolik özütlerin daha bol fenollere sahip olduğunu gösterdi. (sırasıyla 153, 49.33 mg GAE/g ekstre,) Ayrıca  $\beta$ -karoten ve likopen içeriği saptandı (0.120 ile 0.606  $\mu$ g/mL arası) S. luteus'un metanol ve etanol özütlerinin, DPPH radikaline karşı antioksidan aktiviteleri aynı konsantrasyondaki standart antioksidanlar olan BHT'den daha yüksek aktivite göstermiştir (Metanol özütü (IC<sub>50</sub>: 63.72 $\mu$ g/mL) > Etanol özütü (IC<sub>50</sub>: 80.72 $\mu$ g/mL) > BHT (IC<sub>50</sub>: 96.47 $\mu$ g/mL). Ek olarak, metanol ektresinin etanol ekstresine göre daha yüksek demir iyonu şelatlama kabiliyetine sahiptir (sırasıyla 2.72, 3.45 $\mu$ g/mL). Genel olarak, test edilen mantar ekstreleri test edilen mikroorganizmalara karşı nispeten düşük antimikrobiyal aktiviteye sahiptir (9 ve 10mm zonçapı) Ayrıca, S. luteus'un metanol ekstresinin Img/mL konsantrasyonda kanser hücrelerinin tamamını öldürdüğü tespit edildi. Bu çalışmanın sonuçları incelendiğinde S.luteus'un özellikle metanol ekstresinin potansiyel medikal özelliklere sahip olduğu gösterilmiştir.

Anahtar Kelimeler: antimikrobial aktivite, antioksidan aktivite, antiproliferatif aktivite, Suillus luteus

### **INTRODUCTION**

Oxidation is an essential process for the production of energy to many organisms. Under physiological conditions, however, the concentrations of reactive oxygen species (ROS) are usually over physiological limits leading to oxidative stress [1,2]. Overproduction of free radicals can cause oxidative damage to biomolecules, (lipids, proteins, DNA), eventually leading to many chronic diseases such as cancer, cardio-vascular diseases and inflammation in humans. Oxidative stress in cells can result from either an increase in the levels of reactive oxygen species, or a reduction of the natural cell antioxidant capacities [3]. Antioxidants can be defined as molecules that can delay or prevent oxidation of the substrate when they encounter a low amount of oxidizable substrate [4]. In some cases, the amount of antioxidant in cell may be insufficient for intracellular protection. In such cases, external antioxidant supplementation will contribute to the renovation of this balance again [5]. Currently, synthetic antioxidants had unwanted side effects mostly [6]. Therefore, it is essential to develop and utilize effective natural antioxidants to replace the synthetic antioxidants [7].

Natural products (such as secondary metabolites) and their analogs are the source of inspiration in the production of new drugs. Active ingredients of many drugs such as antibiotics (penicillin, tetracycline and erythromycin), anti-parasites (such as avermectin), antimalarials (such as uinine, arminthymine), lipid control agents (lovastatin and analogues), immunosuppressants for organ transplantation (cyclosporine, rapamycin), anti-cancer drugs (Toxol, doxorubicin) are derived from natural resources [8].

Mushrooms are used by people since ancient times as they have significant nutritional values and medical property, especially in Asian countries [9]. Edible mushrooms are rich in high minerals (potassium, phosphorus, iron), essential amino acids, vitamins (B12 and D) and source of some fiber [10-12]. Mushrooms use their own metabolic pathways throughout their life cycles. Mushroom also produces a variety of secondary metabolites, such as numerous alkaloids, terpenes, steroids and phenolic compounds that can be used for therapeutic purposes [13]. The mushroom's compounds possess antimicrobial activity [14], antigenotoxic [15], antioxidant [16], antiproliferative [17], anticancer [18], antihyperlipidemic [19], anti-hypertensive, anti-nociceptive and immunostimulanting [20], hypocholesterolemic, anti-atherogenic [21], anti-allergic [22], neuroprotective and antidepressan effect [23,24].Bioactive compounds isolated from mushroom include small molecule compounds, polysaccharides, proteins, polysaccharide-protein compounds. Amongst bioactive compounds, polysaccharides have been studied in the broadest field [25-27]. These polysaccharides are actively involved in the life cycle of organisms and have biological activities such as anti-cancer, anti-fungal, antioxidant [28,29]. Among the polysaccharides found in mushroom,  $\beta$ -glucan is used as a chemotherapeutic drug in cancer treatments and various diseases [30,31]. In recent years, therapeutic agents which affected apoptosis, angiogenesis, metastasis, cell cycle and signal transduction control has been used in oncology [32]. The use of polysaccharide and polysaccharide-protein complexes isolated from edible mushrooms has proven to be a source of therapeutic agents due to their immunomodulatory and anti-tumor effects [33].

Over 2600 macrofungi species have so been reported from Turkey and approximately 300 of them are edible [34-36]. Today, a significant amount of cork exports are made in Turkey and 171 million US dollars was recovered from exports from 2007 until 2017 [37, 38]. *S. luteus*, a member of *Boletales* in *Agaricomycetes* is an ectomycorrhizal fungus that solely associates with *Pinaceae* plants in the Northern Hemisphere, such as *Pinus densiflora*, *Pinus thunbergii*, *Pinus sylvestris*, *Pinus strobus*, and *Picea glehnii*. The mushrooms are widely consumed in central Europe [39, 40]. In our country, it is necessary to determine the nutritional and medicinal properties of fungi because of this variety and economic value of their potential.

The main objectives of the current study were to evaluate the phenolic,  $\beta$ -carotene and lycopene content and antioxidant, antimicrobial and anti-proliferative activity of *S. luteus* in Turkey.

#### **MATERIAL AND METHOD**

#### **Mushroom material**

*S. luteus* samples were collected from Ankara and Tokat province in 2013. The samples used in this study were identified by Dr. Ilgaz Akata from Ankara University. The identified specimens were deposited at the herbarium of Ankara University.

#### Chemicals

Chloroform, Folin-Ciocalteu's phenol reagent, ethanol, methanol were purchased from Merck (Darmstadt, Germany). 3-[4,5-Dimethylthiazole-2-yl]-2,5-diphenyltetrazoliumbromide (MTT), Tween 40, dimethylsulphoxide (DMSO),  $\beta$ -carotene, 2,2-diphenyl-1-picrylhydrazyl (DPPH), 3-(2-pyridyl)-5,6-bis(4-phenyl-sulphonic acid)-1,2,4-triazine (ferrozine), gallic acid, 2,6-di-tert-butyl-4-methylphenol (BHT) and linoleic acid were purchased from Sigma-Aldrich GmbH (Steinheim, Germany). Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum were purchased from Gibco BRL (Gaithersburg, MD, USA). All other chemicals were analytical grade and obtained from either Sigma or Merck.

#### **Preparation of the extracts**

The dried mushroom samples were extracted by maceration in 1:4 (w/v) biomass /solvent ratio with methanol and ethanol for 2 weeks at room temperature in a dark environment. The obtained methanolic and ethalolic extracts were filtered through filter paper. After filtration the solvent was evaporated in a rotary evaporator (Heidolph, Germany) at 50°C under reduced pressure and the solid extracts were stored at  $+4^{\circ}$ C until further use.

## **Determination of Total Phenolics Content**

Total phenolic of each mushroom extract was quantified according to the method of Folin-Ciocalteu [41] using gallic acid as standard. Briefly, 0.1 mL of extracts (1 mg/mL) were mixed with 0.2 mL of diluted Folin-Ciocalteu reagent (1:1 with water). After incubation at room temperature for 3 min,1 mL 2% sodium carbonate was added to the reaction mixture. The absorbance was read at 760 nm by spectrophotometer after 1 h of incubation at room temperature in the dark. The total phenolic concent values are expressed as gallic equivalent (GAE) in milligrams per gram of dried extract (mg GAE/g). All measurements were performed in triplicate.

#### Determination of $\beta$ -Carotene and lycopene content

 $\beta$ -Carotene and lycopene content of the extracts were determined according to the method described by (42) with slight modification. Briefly, dried samples (100 mg) were mixed with acetone/hexane (4:6, v/v). After incubation for 1 min. The absorbance of the supernatants was read at 453, 505, 645 and 663 nm by spectrophotometer. Contents of  $\beta$ -carotene and lycopene were calculated according to the following equation:

Lycopene (mg/100 ml) =  $-0.0458 A_{663} + 0.372 A_{505} - 0.0806 A_{453}$ ;  $\beta$ -carotene (mg/100 ml) =  $0.216 A_{663} - 0.304 A_{505} + 0.452 A_{453}$ .

#### Antioxidant activity

#### 1-Diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity

The free radical scavenging capacity of the extracts were analyzed according to the method described by (43) with slight modifications. Briefly,0.5 mL extracts with different concentrations were mixed with a methanolic solution of DPPH radical (0.1mM) freshly prepared. After incubation for 30 min at room temperature in the dark, absorbance was read at was added to extracts solutions at 517 nm by spectrophotometer (Shimadzu UV-1800,Japan) against a blank (extract only). Same procedure with a solution without the extract was applied as a control group. Butylated hydroxytoluene (BHT) was used as a reference standard. The percentage of DPPH radical scavenging effect was calculated according to the following equation:

DPHH scavenging activity (%inhibation) = [(Acontrol-Asample)/Aconrtol]×100, where A control is the absorbance of the control and Asample is the absorbance of the reaction mixture or standard. A curve of extract concentration versus % inhibation was created to determine the concentration of the extract needed to cause a %50 decrease of the beginning DPHH concentration. This value calculated by linear regression analysis is known as a  $IC_{50}$ .The lower  $IC_{50}$  value indicates better antioxidant activity.

#### Ferrous ion chelating ability

Ferrous ion chelating ability of the extracts were determined according to the method described by (44) with slight modifications. 0.5 mL of the extracts at different concentrations were mixed with 1.35 mL of methanol and ethanol. 50  $\mu$ l of 2 mM FeCl<sub>2</sub> were added to extract solution and stand for 5 min. Thereafter, 100  $\mu$ l of 5 mM ferrozine solution were added to this mixture and incubated for 10 min. After incubation, absorbance was read at 562 nm by spectrophotometer (Shimadzu UV-1800, Japan) against a blank (extract and FeCl<sub>2</sub> only). In the control group, extract was substituted with methanol and ethanol. EDTA (Ethylene diamine tetraacetate) was used as a positive control. Percentage of the ability of the sample to helate ferrous ion was calculated according to the following equation:

Ferrous ion chelating ability (%)= $[(A_{control}-A_{sample})/A_{control}]x100$ , where  $A_{control}$  is the absorbance of the control and  $A_{sample}$  is the absorbance of the reaction mixture. The IC<sub>50</sub> value, which is the concentration of the extracts that chelate 50% of the ferrous ion, was calculated by linear regression curve.

#### Antimicrobial activity

The antimicrobial activities of mushroom extracts were determined by agar well method and evaluated against bacterial strains on *Staphylococcus aureus* ATCC 25923, *Bacillus cereus* NRRL B-

3711, Bacillus subtilis ATCC 6633, Escherichia coli ATCC 35218, Escherichia coli ATCC 25922, Enterococcus faecalis ATCC 29212, Enterococcus hirae ATCC 9790, Klebsiella pneumaniae ATCC 13883, Pseudomonas aeruginosa ATCC 27853, Proteus vulgaris RSKK 96029, Salmonella typhimurium ATCC 14028 and fungal strains Candida tropicalis Y-12968, Candida albicans ATCC 10231, Candida krusei ATCC 6258. For comparison ampicillin and chloramphenicol were used a standard antibiotics.

In the agar well method, bacterial strains were allowed to incubate at 37°C for 24 hours in Nutrient Agar medium and yeast strains were incubated for 48 hours at 30°C in Malt Extract Agar medium. The post-incubation microorganisms were adjusted to 0.5 McFarland blur. Muller – Hinton Agar (for bacterial strains) and Malt Extract Agar (for yeast strains) were spread on a petri with a 1% suspension of microorganism suspension. With the punch, 6 mm in diameter wells are opened at specific points of the medium. The opened wells were placed in a volume of 50  $\mu$ L from mushroom extracts at a concentration of 150 mg/mL and left to incubate. The diameter of the inhibition zones formed after incubation is measured in mm. Chloramphenicol, ampicillin were used for antimicrobial activity.

#### Cultiring of cell lines

Human breast adenocarcinoma cell line MCF-7 was purchased from American Type Tissue Culture Collection (USA) and cultured in RPMI 1640 (Sigma Chemicals) media containing 10% FBS and 1% of sodium pyruvate, amphotericin B, penicillin and streptomycin. Cells were maintained at 37 °C and 5% CO<sub>2</sub> under humidified condition.

#### Antiproliferative activity

Cells were grown in culture flask at a range of 10,000-100,000 cells per ml. Mushroom extracts were applied at increasing concentrations (25, 50,100, 250,500 and 1000 mg/mL) for 24, 48 and 72 hours. Viable cells in the control and application groups were determined by MTT [3- (4,5-dimethyl thiazol-2-yl) -2,5-diphenyl tetrazolium bromide] staining method [45]. The solution was measured by spectrophotometer (Thermo/LabSystems 352 Multiskan MS Microplate Reader) at 590 nm. All experiments were performed with 3 replications.

 $[(C_{72h+extract} - C_{24h+extract}) / (C_{72h-control} - C_{24h-control})] \times 100 = \% \text{ dividing cell viability}$ 

C<sub>72h+ extract</sub>: Live cell measurement 72 hours after manipulation

C<sub>24h+ extract</sub>: Live cell measurement 24 hours after manipulation

C72h- control: 72 hours after live cell measurement without extract manipulation

C<sub>24h- control</sub>: 24 hours after live cell measurement without extract manipulation

#### Statistical analysis

SPSS 11 were used for statistical analyses. Experimental results were expressed as mean  $\pm$  S.D of three parallel measurements. *P*-values < 0.05 were regarded as significant.

## **RESULT AND DISCUSSION**

The extracts of *S.luteus* was studied regarding antioxidant capacity potential. The standard curve equation is, y (absorbance) =0.008 5x (µg gallic acid)-0.0209, R<sup>2</sup>=0.999 9. The data of the sample regarding the content of total phenolics,  $\beta$ -carotene and lycopene are presented in Table 1. As a shown in table, the mushrooms of *S.luteus* methanolic and ethanolic extracts presented phenolic contents with 153 ± 3.54 and 49.33 ± 0.14 mg GAE/g extract, respectively. The results suggest that most of the phenolic compounds in methanolic extract. In addition, ethanolic extract had more  $\beta$ -carotene and lycopene content (0.606 ± 0.05, 0.357 ± 0.02 µg/mL, respectively) than methanolic extract (0.220 ± 0.01, 0.120 ± 0.05 µg/mL, respectively).

**Table 1.** Total phenolic,  $\beta$ -Carotene and lycopene content in the extracts of *S.luteus* and  $\pm$ SD\*(n=3).

| Sample              | Total phenolic content<br>(mg GAE/g extract) | β -Carotene<br>(μg/mL) | Lycopene<br>(µg/mL) |
|---------------------|--|------------------------|---------------------|
| Methonolic extracts | $153\pm3.54$                                 | $0.220 \pm 0.01$       | $0.120 \pm 0.05$    |
| Ethanolic extracts  | 49.33 ±0.14                                  | $0.606\pm0.05$         | $0.357 \ \pm 0.02$  |

\*Standart devidation

The antioxidant activity of mushrooms increased with the increased in the concentration of samples, higher the antioxidant properties lower the IC<sub>50</sub> values. A lower IC<sub>50</sub> values means better radical scavenging activity [46]. The scavenging DPPH radicals of the studied methanolic and ethanolic extracts are indicated in Table 2. As a shown in table, the free radical scavenging activity of the mushroom extracts was evaluated by DPPH assay comparing the IC<sub>50</sub> value of synthetic chemical BHT, which was 96,47  $\pm$  0.57 µg/mL. Antioxidant activity was detected method by DPHH free radical scavenging activity than BHT (Methanol extract IC<sub>50</sub>: 63.72  $\pm$  0.89 µg/mL > Ethanol extract IC<sub>50</sub>: 80.72  $\pm$  0.58 µg/mL > BHT: IC<sub>50</sub>: 96.47  $\pm$  0.57 µg/mL). Besides, ferrous ion chelating activities of the extracts expressed as IC<sub>50</sub> values are shown in Table 3. As a shown in table, *S.luteus* methanolic extract (2.72  $\pm$  0.06, 3.45  $\pm$  0.05 mg/mL, respectively). EDTA showed very powerful activity.

**Table 2.** DPPH radical scavenging activities of the *S. luteus* extracts. Scavenging activity is expressed as  $IC_{50}$  (µg/mL) ± SD (n=3).

| Sample             | IC50 (μg/mL)     |  |  |
|--------------------|------------------|--|--|
| Methanolic extract | $63.72 \pm 0.89$ |  |  |
| Ethanolic extract  | $80.72\pm0.58$   |  |  |
| BHT                | $96.47 \pm 0.57$ |  |  |

\*Standart devidation

**Table 3.** Ferrous ion chelating activities of the *S. luteus* extracts. Chelating activity is expressed as  $IC_{50}$  (mg/mL)  $\pm$  SD (n=3).

| Sample             | IC <sub>50</sub> (mg/mL) |  |  |
|--------------------|--------------------------|--|--|
| Methanolic extract | $2.72\pm0.06$            |  |  |
| Ethanolic extract  | $3.45 \pm 0.05$          |  |  |
| EDTA               | $0.018 \pm 0.001$        |  |  |

\*Standart devidation

Antimicrobial activities of the mushrooms extract against the test microorganisms is shown in Table 4. *S. luteus* methanolic extract formed against to *E. faecalis* ATCC 29212, *B.subtilis* ATCC 6633, *K.pneumaniae* ATCC 13883 9 mm inhibition zone diameter. Ethanolic extract formed against to *E. faecalis* ATCC 29212, *S. aureus* ATCC 25923, *S.typhimurium* ATCC14028 9 mm and *P. aeruginosa* ATCC 27853 10 mm inhibition zone diameter.

The antimicrobial activity was compared with the standard antibiotics, ampicillin and chloramphenicol. The results showed that standard antibiotics had stronger activity than tested samples as shown in Table 4. In a negative control, DMSO had no inhibitory effect on the tested organisms.

| Test microorganisms      | Methanolic<br>extract | Ethanolic<br>extract | Ampicillin | Chloramphenicol |
|--------------------------|-----------------------|----------------------|------------|-----------------|
| E. faecalis ATCC 29212   | 9±1                   | 9±0                  | 27±0       | 20±0            |
| K. pneumaniae ATCC 13883 | 9 ±0                  | -                    | -          | 31±1            |
| B. subtilis ATCC 6633    | 9±1                   | -                    | 23±1       | 21±0            |
| S. aureus ATCC 25923     | -                     | 9±0                  | 44±1       | 24±0            |
| P. aeruginosa ATCC 27853 | -                     | 10±1                 | 60±0       | 34±0            |
| S.typhimurium ATCC 14028 | -                     | 9±0                  | 19±1       | 38±1            |

**Table 4**. Antimicrobial activity results (zone diameter / mm) and  $\pm$  SD.

\*Standart devidation

Antiproliferative activity was studied in methanolic extract of *S.luteus*, since methanol extract is more effective than antioxidant activity and antioxidant containing ingredients than ethanol extract. The experimental data of antiproliferative activity of *S. luteus* methanolic extract on MCF-7 cell line by MTT method are shown in Figure1. It was found that the cells exposed to 25, 50, 100, 250, 500 µg/mL and 1 mg/mL concentrations of methanolic extract resulted in %,87.83 ,%78.82, %86.48, %34.68, %15.76 and % 0 cell viability reduction compared to the negative control group, respectively and these reductions were statistically significant in comparison to negative control group (P < 0.05). The IC<sub>50</sub> value of methanol extract was calculated to be approximately 173µ/mL. Observed to cause damage to the breast cancer cells.



Concentration

Figure 1. Percentage of viability of MCF-7 lysed breast cancer cells after exposure to various concentrations of methanol extract of *S. luteus at* 24 h

Many research studies have shown that *S.luteus* extract has antioxidant activity and total phenolic contents. Previous reports have demonstrated that the DPPH radical scavenging effect *S.luteus* of ethanol extract was found IC<sub>50</sub>: 0.66 mg/mL and total phenolic content was found 27.7  $\pm$  4.0 mg GAE/g [47]. Another study indicated that DPPH radical scavenging effect methanolic extract of *S.luteus* IC<sub>50</sub>: 1.92  $\pm$  0.08 mg/mL. *S.luteus* showed the high concentration of phenolic acids (0.72 mg/100 g), due to the contribution of protocatechuic (0.47 mg/100 g) and cinnamic acid (0.41 mg/100 g) [48]. According to Keleş et al. reported that antioxidant activity was measured by the FRAP method, methanol extract of *S.luteus* shound EC<sub>50</sub>: 4.76 mg/mL [49]. It was also concluded that totel total phenolic content of the methanol extract was found 1.72  $\pm$  0.02 mg GAE/g. [50] and Jowarska et al. reported that DPPH activity in methanol extract was found IC<sub>50</sub>: 3.48  $\pm$  0.20 mmol TE and using the FRAP method it was 9.15 mmol Fe<sub>2</sub><sup>+</sup>[51]. In the study of Heleno et al. the antioxidant DPPH activity of *Suillus collinitus* and *Suillus mediterraneenis* methanol extracts were examined IC<sub>50</sub>: 14.05  $\pm$  1.24 11 mg/mL and 2.90  $\pm$  0.11

mg/mL, respectively. Methanol and ethanol extracts of *S.luteus* fungus used in our study have higher activity than *S.collinitus* and *S.mediterraneenis* [52]. Our results indicate that *S.luteus* have more abundant phenolic components, higher antioxidant activity and ferrous ion chelating ability. According to the results of our study, it is clearly indicated that the ethanolic and methanolic extracts of *S.luteus* have significant phenolic content and antioxidant activity. Furthermore, a good correlation was also observed between the total phenolic content and antioxidant activity. These differences can be attributed to differences in soil conditions in different geographical locations and the sub-species ability to synthesize phenolic compounds.

 $\beta$ -carotene is a light yellow or orange pigment that is the precursor of vitamin A. Antioxidant  $\beta$ carotene prevents oxidation of unsaturated fats and the formation of free radicals [34]. Lycopene, an important derivative of carotenoids, is the most powerful antioxidant in vitro and has more radical scavenging activity [54,55]. The extracts of *S.luteus* of  $\beta$ -carotene and lycopene content are presented in Table 1.Jowarska et al. investigated *S. luteus* total polyphenol and flavonoids and B group vitamin contents. *Suillus* species are richer in polyphenols compared to other fungi in the literature [51].

Antimicrobial activities of the *S.luteus* extracts against the test microorganisms are shown in Table 4. Antimicrobial activities of the extracts were determined on five Gram-positive, six Gramnegative bacteria and three yeasts. In reported studies methanolic and ethanolic extracts from *S.luteus* showed similar antimicrobial activity against microorganisms [56,57]. In this study, the antibacterial properties of *S.luteus* were not as effective as the commercial drugs.

In this study, the antiproliferative activity of the methanol extract of *S. luteus* on MCF-7 cell lines in 24 hours was studied by MTT method. Methanol extract was found to kill all cancer cells at a concentration of 1mg/mL. The IC<sub>50</sub> value of methanol extract was calculated to be approximately IC<sub>50</sub>: 173µg/mL. Previous studies had investigated the effect of *S.luteus* methanolic exttract on colon cancer cell line by MTT method. The most sensitive amount was found to be IC<sub>50</sub>= 17.75 ± 1.6 µg/mL on HCT-15 cell line which is the colon cancer cell line [58]. Vaz et al., also concluded antiproliferative activity of *S. collinitus* on ASG gastric cancer cell line. They found that the cell line had IC<sub>50</sub>: 79.2 ± 15.5 µg/mL [59]. The results of the study show that *S. luteus* is a potential anticancer agent. No data are available against the antiproliferative activity of *S. luteus* MCF-7 on the cancer cell line. It is predicted that these new findings added to the literature will be effective in further studies.

As a result, *S. luteus* have high antioxidant activity at low concentrations of methanolic and ethanolic extracts, but have low antimicrobial activity. Especially methanol extract has antiproliferative activity on MCF-7 breast cancer cell line. Nowadays, the emergence of some side effects of the drugs used in the treatment of diseases causes increasing interest in the treatment with natural resources. Multidrug resistance is still a major problem in cancer chemotherapy [60]. Researchers should focus on solving this problem. Our studies have tried to determine some basic concepts about the applicability of

this kind of research in practice. *S. luteus*, especially methanol extract is recommended to be included in further studies.

## REFERENCES

- 1. Liu, Q., Jiang, J. (2012). Antioxidative activities of medicinal plants from TCM. *Mini-Reviews in Medicinal Chemistry*, *12*(11), 1154-1172.
- 2. Liu, Z., Jiao, Y.C., Lu, H., Shu, X., Chen, Q. (2020). Chemical characterization, antioxidant properties and anticancer activity of exopolysaccharides from *Floccularia luteovirens*. *Carbohydrate Polymers*, 229, 115432.
- 3. Racchi, M., Daglia, M., Lanni, C., Papetti, A., Govoni, S., Gazzani, G. (2002). Antiradical activity of water soluble components in common diet vegetables. *J. Agric. Food. Chem*, 50, 1272-1277.
- 4. Becker, E.M., Nissen, L.R. and Skibsted, L.H. (2004). Antioxidant evaluation protocols: Food quality or health effects. *European Food Research and Technology*, *19*, 561-571.
- 5. Valko, M., Leibfritz, D., Moncol, J., Cronin, M.T., Mazur, M., Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry & Cell Biology*, *39*, 44-84.
- Qi, H., Zhang, Q., Zhao, T., Chen, R., Zhang, H., Niu, X., Li, Z. (2005). Antioxidant activity of different sulfate content derivatives of polysaccharide extracted from Ulva pertusa (Chlorophyta) in vitro. *International Journal of Biological Macromolecules*, 37(4), 195-199.
- 7. Singh, N., Rajini, P.S. (2004). Free radical scavenging activity of an aqueous extract of potato peel. *Food Chemistry*, 85(4), 611-616.
- 8. Li, J.W., Vederas, J.C. (2009). Drug discovery and natural products: End of an era or an endless frontier. *Science*, *32*, 161-165.
- 9. Hoshi, H., Yagi, Y., Iijima, H., Matsunaga, K., Ishihara, Y., & Yasuhara, T. (2005). Isolation and characterization of a novel immunomodulatory r-glucan-protein complex from the mycelium of *Tricholoma matsutake* in Basidiomycetes. *Journal of Agricultural and Food Chemistry*, 53, 8948–8956.
- Feeney, M.J., Dwyer, J., Hasler-Lewis, C.M., Milner, J.A., Noakes, M., Rowe, S., ... Wu, D. (2014). Mushrooms and Health Summit Proceedings. *Journal of Food and Nutrition Research*, 144(7), 1128-1136.
- 11. Manninen, H., Rotola-Pukkila, M., Aisala, H., Hopia, A., Laaksonen, T. (2018). Free Amino Acids and 5'-Nucleotides in Finnish Forest Mushrooms. *Food Chem*, 247, 23-28.
- 12. Roupas, P., Keogh, J., Noakes, M., Margetts, C., Taylor, P. (2012). The role of edible mushrooms in health: Evaluation of the evidence. *J. Funct. Foods*, *4*, 687-709.

- 13. Jayakumar, T., Thomas, P.A., Sheu, J.R. and Geraldine, P. (2011). In-vitro and in-vivo antioxidant effects of the oyster mushroom Pleurotus ostreatus. *Food Research International*, 44, 851-861.
- Osaki, K., Suyama, S., Sakuno, E., Ushijima, S., Nagasawa, E., Maekawa, N., Ishihara, A. (2019). Antifungal activity of the volatile compound isovelleral produced by ectomycorrhizal Russula fungi against plant-pathogenic fungi. J. Gen. Plant Pathol, 85, 428-435.
- Zivkovic, L., Bajic, V., Bruic, M., Borozan, S., Popic, K., Topalovic, D., Santibanez, J., Potparevic, B. (2019). Antigenotoxic and antioxidant potential of medicinal mushrooms (Immune Assist) against DNA damage induced by free radicals-an in vitro study. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 845, 403078.
- Song, X., Ren, Z., Wang, X., Jia, L., Zhang, C. (2020). Antioxidant, anti-inflammatory and renoprotective effects of acidic-hydrolytic polysaccharides by spent mushroom compost (*Lentinula edodes*) on LPS-induced kidney injury. *International Journal of Biological Macromolecules*, 151, 1267-1276.
- 17. Aytar, E., Özmen, A. (2020). Cytotoxic and apoptotic activities of *Rhizopogon roseolus* (Corda) Th. Fr. extracts. *International Journal of Secondary Metabolite*, 7(1), 54-62.
- Kim, S.H., Jakhar, R., Kang, S.C. (2015). Apoptotic properties of polysaccharide isolated from fruiting bodies of medicinal mushroom *Fomes fomentarius* in human lung carcinoma cell line. *Saudi J. Biol. Sci.*, 22, 484-490.
- Opletal, L., Jahodar, L., Chobot, V., Zdansky, P., Lukes, J., Bratova, M., Solichova, D., Blunden, G., Dacke, C.G. and Patel, A.V. (1997) Evidence for the antihyperlipidemic activity of edible fungus *Pleurotus ostreatu. Brit J Biomed Sci.*, 54,240-243.
- 20. Vaz, J.A., Barros, L., Martins, A., Santos-Buelga, C., Vasconcelos, M., Ferreira, I.C. (2011). Chemical composition of wild edible mushrooms and antioxidant properties of their water soluble polysaccharidic and ethanolic fractions. *Food Chemistry*, *126*, 610-616.
- 21. Han, E.H., Hwang, Y.P., Kim, H.G., Choi, J.H., Im, J.H., Yang, J.H., Lee, H.U., Chun, S.S., Chung, Y.C., Jeong, H.G. (2011). Inhibitory effect of *Pleurotus eryngii* extracts on the activities of allergic mediators in antigen-stimulated mast cells. *Food Chem. Toxicol*, 27, 1199-1128.
- 22. Nguyen, T.M., Le, H.G., Le, B.V., Kim, Y.H.K., Hwang, I. (2020). Anti-allergic effect of inotodiol, a lanostane triterpenoid from Chaga mushroom, via selective inhibition of mast cell function. *International Immunopharmacology*, *81*, 106244.
- 23. Choi, Y.J., Park, I.S., Kim, M.H., Choi, J.H.I Im, J.H., Yang, J.H., Lee, H.K., Chun, S.S., Chung, Y.C., Jeong, H.G. (2018). The medicinal mushroom *Auricularia auricula-judae* (Bull.) extract has antioxidant activity and promotes procollagen biosynthesis in HaCaT cells. *Nat Prod Res*, 4,1-4.
- 24. Donatini, B. (2011). *Hericium erinaceus*: properties mostly related to the secretion of neuronal growth factor. *Phytothérapie*, *9*, 48-52.
- 25. Kozarski, M., Klaus, A., Niksic, M., Jakovljevic, D., Helsper, J.P.F.G. and Van Griensven, L.J.L.D. (2011). Antioxidative and immunomodulating activities of polysaccharide extracts of

the medicinal mushrooms Agaricus bisporus, Agaricus brasiliensis, Ganoderma lucidum and Phellinus linteus, Food Chemistry, 129, 1667-1675.

- 26. Pan, Y., Dong, S., Hao, Y., Zhou, Y., Ren, X., Wang, J., Wang, W., Cheu, T. (2010). Ultrasonicassisted extraction process of crude polysaccharides from Yunzhi mushroom and its effect on hydroxyproline and glycosaminoglycan levels. *Carbohydate Polymers*, *81*, 93-96.
- Zhang, B.Z., Yan, P.S., Chen, H. and He, J. (2012). Optimization of production conditions for mushroom polysaccharides with high yield and antitumor activity. *Carbohydrate Polymers*, 87, 2569-2575.
- 28. Lu, R., Conrad, P., Yacine, H. (2012). Antitumor activity of mushroom polysaccharides. *Food Function*, *3*, 1118–1130.
- Trajkovic, L.M.H., Mijatovic, S.A., Maksimov Iclvanic, D.D., Stojanovic, I.D., Momcilovic, M.B. (2009). Anticancer properties of *Ganoderma lucidum* methanol extracts in vitro and in vivo. *Nutrition and Cancer*, 61, 696–707.
- 30. Chan, G.C., Chan, W.K. and Sze, D.M. (2009). The effects of β-glucan on human immune and cancer cells. *Journal of Hematology Oncology*, 2, 25.
- 31. Kidd, P.M. (2000). The use of mushroom glucans and proteoglycans in cancer treatment. *Alternative Medicine Review*, 5, 4–27.
- 32. Ouyang, F., Wang, G., Guo, W., Zhang, Y., Xiang, Y. and Zhao, M. (2013). AKT signalling and mitochondrial pathways are involved in mushroom polysaccharide-induced apoptosis and G1 or S phase arrest in human hepatoma cells. *Food Chemistry*, *138*, 2130-2139.
- 33. Ooi, V.E., Liu, F. (2000). Immunomodulation and anti-cancer activity of polysaccharidprotein complexes. *Current Medicinal Chemistry*, 7, 715–729.
- 34. Akata, I., Altuntaş, D., Kabaktepe, Ş. (2019). Fungi determined ın Ankara University Tandoğan campus area (Ankara-Turkey). *Trakya University Journal of Natural Sciences*, 20 (1): 47-55.
- 35. Akata, I., Doğan, H.H. (2015). Orbiliaceae for Turkish Ascomycota: Three new records. *Bangladesh Journal of Botany*, 44(1), 91-95.
- Sesli, E., Denchev, C.M. (2008). Checklists of the myxomycetes, larger ascomycetes, and larger basidiomycetes in Turkey. *Mycotaxon*, 106, 65–67.
- Bulam, S., Ustun, N.Ş., Pekşen, A. (2018). Mushroom Foreign trade of Turkey in the last decade. Conference: International Congress on Engineering and Life Science (ICELIS) 26-29 April 2018 at Kastamonu Turkey. Proceeding Book 779-784.
- 38. Bahadori, M.B., Sarikurkcu, C., Yalcin, O.U., Cengiz, M. and Gungor, H. (2019). Metal concentration, phenolics profiling, and antioxidant activity of two wild edible Melanoleuca mushrooms (*M. cognata* and *M. stridula*). *Microchemical Journey*, *150*, 140172.
- Murata, H., Yamada, A., Yokota, S., Maruyama, T., Shimokawa, T., Neda, H. (2015). Innate traits of Pinaceae-specific ectomycorrhizal symbiont *Suillus luteus* that differentially associates with arbuscular mycorrhizal broad-leaved trees *in vitro*. *Mycoscience*, 56(6), 606-611.

- 40. Kalogeropoulos, N., Mylona, A., Chiou, A., Ioannou, M.S., Andrikopoulos, N.K. (2007). Retention and distribution of natural antioxidants ( $\alpha$ -tocopherol, polyphenols and terpenic acids) after shallow frying of vegetables in virgin olive oil *LWT*, 40(6),1008-1017.
- 41. Siddhuraju, P., Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. *Journal of Agricultural and Food Chemistry*, 51(8), 2144–2155.
- 42. Nagata, M. and Yamashita, I. (1992). Simple method for simultaneous determination of chlorophyll and carotenoids in tomato fruit. *Nippon Shokuhin Kogyo Gakkaish*, *39*, 925–928.
- 43. Braca, A., De Tommasi, N., Di Bari, L., Pizza, C., Politi, M., Morelli, I. (2001). Antioxidant principles from bauhinia tarapotensis. *Journal of Natural Products*, *64*(7), 892-895.
- 44. Dinnis, T., Madeira, V., Almeida, L. (1994) Action of phenolic derivative (acetoaminophene salycilate and 5-amino solycilate as inhibitors of membrane lipid peroxidation and as peroxyl radical scavengers). *Archives Biochemistry and Biophyics*, *315*, 161–169.
- 45. Dabur, R.R., Sharma, G. L. (2002). Studies on antimycotic properties of *Datura metel J. Ethnopharmacol*, 80, 193-197.
- 46. Abdelaaty, S., Abeer, I., Mansour, A.S. (2015). Antioxidant capacity and polyphenolic content of seven Saudi Arabian medicinal herbs traditionally used in Saudi Arabia. *Indian J Tradit Knowledge*, 14(3), 28-35.
- 47. Macakovaa, K., Opletala, L., Polasekb, M., Samkovac, V., Jahodara, L. (2009). Free-radical scavenging activity of some European Boletales. *Natural Product Communications*, 2, 261-264.
- 48. Reis, F.S., Heleno, S.A., Barros, L., Sousa, M.J., Martins, A., Santos-Buelga, C., Ferreria, I.C.F.R. (2011). Toward the antioxidant and chemical characterization of mycorrhizal mushrooms from Northeast Portugal. *Journal of Food Science*, *76*, 6.
- 49. Keleş, A., Koca, İ., Gençcelep, H. (2011). Antioxidant properties of wild edible. *Journal Food Process Technology*, 2, 130.
- Zeng, X., Suwandi, J., Fuller, J., Doronila, A., Ng, K. (2011). Antioxidant capacity and mineral contents of ediblewild Australian mushrooms. *Food Science and Technology International*, 18(4), 367–379.
- 51. Jaworska, G., Pogon, K., Bernas, E., Gabor, A. (2014). Vitamins, phenolics and antioxidant activity of culinary prepared *Suillus luteus (L.)* Roussel mushroom. *Food Science and Technology*, 99, 701-706.
- 52. Heleno, S.A., Barros, L., Sousa, M.J., Martins, A., Ferreira, I.C.F.R. (2010). Tocopherols composition of Portuguese wild mushrooms with antioxidant capacity. *Food Chemistry*, *119*,1443-1450.
- 53. Miura, Y., Kondo, K., Saito, T., Shimada, H., Fraser, P. and Misawa, N. (1998). Production of the carotenoids lycopene, carotene, and astaxanhin in the food yeast candida utilis. *Applied and Environmental Microbiology*, *64*, 1226–1229.

- 54. Lindshield, B.L., Canene-Adams, K., Erdman Jr, J.W. (2007). Lycopenoids. *Journal of Nutrition*, 122(11), 2161-2166.
- 55. Yaping, Z., Suping, Q., Wenli, Y., Zheng, X., Hong, S., Side, Y., Da-pu, W. (2002). Antioxidant activity of lycopene extracted from tomato paste towards tri chloro methyl peroxyl radical CC13O2. *Food Chemistry*, 77, 209-212.
- 56. Barranco, P.G., Ocanas, L.G., Cabrera, L.V., Carmona, M.C.S., Ocanas, F.G., Gomez, X.S.R., Rangel,R.L. (2010). Evaluation of antioxidant, immunomodulating, cytotoxic and antimicrobial properties of different strains of Basidiomycetes from Northeastern Mexico. *Journal of Medicinal Plants Research*, 4(17), 1762-1769.
- Duman, R., Doğan, H.H., Ateş, A. (2003).*Morchella conica* (Pers.) Boudier and *Suillus luteus* (L.) S. F. Gray Makrofunguslarının Antimikrobiyal Aktiviteleri. *Selçuk Üniversitesi Fen Edebiyat Dergisi*, 22, 19-24.
- 58. Santos, T., Tavares, C., Sousa, D., Vaz, J.A., Calhelha, R.C., Martins, A., Ferreira, I.C.F.R., Vasconcelos, H.M. (2013). *Suillus luteus* methanolic extract inhibits cell growth and proliferation of a colon cancer cell line. *Food Research Intertational*, *53*, 476-481.
- 59. Vaz, J.A., Ferreira, I.C., Tavares, C., Almedia, G.M., Martins, A., Vesconcelos, M. (2012). *Suillus collinitus* methanolic extract increases P53 expression and causes cell cycle arrest and apoptosis in a breast cancer cell line. *Food Chem*, *135*, 596–602.
- 60. Lage, H. (2008). An overview of cancer multidrug resistance: a stil unsolved problem. *Cellular and Molecular Life Sciences*, 65, 3145-3167.