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Review article

Medicinal values of a Saiva ritual plant-Bauhinia tomentosa L.

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Abstract

Bauhinia tomentosa L. is a small tree that belongs to the Fabaceae family and it is distributed in Asia, Africa, North America, and Oceania. B. tomentosa is used to treat some diseases including liver inflammation, abscess, tumors, wounds, and hyperlipidemia in ethnomedicines in Asia and Africa. Compounds like phytone, β -cubebene, β -caryophyllene, 3-O-methyl-d-glucose, and phthalic acid have been isolated from leaves of this plant species. This review article aims to analyze, document, and summarize the reported bioactivities of this plant species. A literature review was conducted using electronic databases like the Web of Science, Scopus, PubMed, and Science Direct to identify the relevant published studies from the year 1900 to November 2020. Various parts of B. tomentosa exhibited bioactivities such as analgesic, anti-anxiety, antibacterial, anticatatonic, anticonvulsant, antidepressant, antidiabetic, antifungal, anthelminthic, antihyperlipidemic, antinociceptive, antioxidant, antipyretic, anti-ulcerative colitis, motor coordination, nephroprotective, nootropic, and wound healing activities in various assays and animal models. However, no bioactive compound has been isolated from this plant species. It was observed that a daily dose of 3000 mg/kg was safe in animal models. Hence, further phytochemical and bioactivities should be conducted to explore more about this plant species. This work analyzed, documented, and summarized the reported bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities and bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities and bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities of B. tomentosa that will be very useful for further phytochemical and bioactivities of B. tomentos

Keywords: Bauhinia tomentosa; bioactivity; cancer; Fabaceae; microbiota; Sri Lanka

1. Introduction

Bauhinia tomentosa L. is a small tree that grows from 1 to 8 m in height belongs to the *Fabaceae* family (Fig. 1). It is called Thiruvaaththi in Tamil. This plant species is native to Asia (Sri Lanka, Yemen, and India) and Africa (Angola, Burundi, Ethiopia, Kenya, Malawi, Mozambique, Somalia, Sudan, Swaziland, South Africa, Tanzania, Zambia, Zaïre, and Zimbabwe). Anyway, it has been introduced into Asia (Andaman Islands, China, Malaysia, Taiwan, Thailand, and Vietnam), Africa (Cameroon, Gambia, Ghana, Guinea, Nigeria, and Sierra Leone), North America (Cuba, Dominican Republic, Haiti, Puerto Rico, and Trinidad and Tobago), and Oceania (Australia) (Royal Botanic Gardens, Kew, 2020) (Fig. 2). *B. tomentosa* is utilized to treat liver inflammation, abscess, tumors, wounds, hyperlipidemia, bleeding, diabetes, diarrhea, animal bites, helminthiasis, infections, fever, and abdominal, skin, and urinary tract illnesses in ethnomedicines in Asia and Africa (Chopra et al., 1992; Sastri, 1995; Kirtikar and Basu, 2005; Orwa et al., 2009). This plant species is also grown as an ornamental plant in gardens and also used as a hedge. Fiber obtained from the trees are employed to make baskets and timber is utilized as beams for sheds. Further,

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leaves are used to prepare a yellow dye and flowers are used in Saiva rituals in Sri Lanka (Orwa et al., 2009). Compounds like Phytone, β -cubebene, β -caryophyllene, 3-O-methyl-d-glucose, phthalic acid, ethyl pentyl ester, 2-butanone, 3-methoxy-3-methyl, 2,2-dimethylpropionic acid, cyclopentyl ester, 2-hexen-1-ol, 2-ethyl, 5-hydroxy-2,2-dimethylhexan-3-one, pentanoic acid, 2-methyl, butane, and 1-bromo-2-methyl have been iso-lated from leaves of *B. tomentosa* (Vasudevan et al., 2014; Panda et al., 2019).

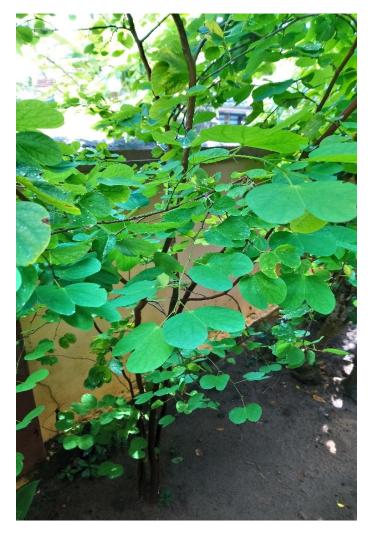


Fig. 1. B. tomentosa in a home garden in Batticaloa, Sri Lanka.

Until now, there is no comprehensive systematic review of bioactivities of *B. tomentosa*. Therefore, this review article aims to analyze, document, and summarize the reported bioactivities of this plant species. This work would be useful for further phytochemical and bioactivities related researches.

For this aim, a literature review was conducted using electronic databases namely the Web of Science, Scopus, PubMed, and Science Direct to identify the relevant published studies from 1900 to November 2020. The scientific name (*Bauhinia tomentosa*) was applied as a search term.

2. Bioactivities of B. tomentosa

Details of the level of scientific evidence, bioactivity, part used, extract/compound, assay/model, dose/concentration, duration, and reference are presented in Table. So far, only *in vitro* and *in vivo* reported studies are available and the majority of the

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studies have been conducted in *in vivo* models. Antioxidant activities have both *in vitro* and *in vivo* scientific evidence.

A greater number of researches were carried out to study the antioxidant activities of this plant species (Kannan et al., 2010; Krishnaswamy et al., 2013; Tiwari and Singh, 2013; Banerjee and De, 2014). Leaves unveiled several bioactivities including antibacterial (Mythreyi et al., 2005; Dugasani et al., 2010), anticonvulsant (Risa et al., 2004), anti-anxiety, anticatatonic, antidepressant (Sathya et al., 2011), antidiabetic (Mannangatti et al., 2010a; Devaki et al., 2011; Kaur et al., 2011; Tiwari and Singh, 2013), anti-ulcerative colitis (Kannan and Guruvayoorappan, 2013), motor coordination (Sathya et al., 2011), nephroprotective (Kannan et al., 2016), and nootropic (Sathya et al., 2011) activities in both *in vitro* and *in vivo* levels.

Further, ethanol extract was used in the majority of the reviewed studies. Anyway, no bioactive compound has been isolated from this plant species. As stated earlier, *B. tomentosa* has a range of uses in ethnomedicines. On the other hand, only ethnomedicinal treatments for inflammation, infections, diabetes, helminthiasis, hyperlipidemia, and would healing activities have scientific evidence currently. Only higher scientific level (*in vivo*) studies according to the lower dose and duration of treatment are deliberated below.

3. Reported in vivo studies

3.1. Analgesic activity

In a study carried out by Tiwari and Singh (2013), aqueous and methanol extracts of the root (200 mg/kg) were orally administered to mice. After 300 minutes, in eddy's hot plate method significant analgesic activity was observed.

3.2. Anti-anxiety activity

The anti-anxiety property was noticed in the elevated plusmaze model, hole board test, and light-dark models after administering 200 mg/kg of leaf ethanol extract (Sathya et al., 2011).

3.3. Anticatatonic activity

Leaf ethanol extract of dose 200 mg/kg administered to haloperidol-induced catalepsy animal models showed anticatatonic activity (Sathya et al., 2011).

3.4. Antidepressant activity

In an investigation conducted by Sathya et al. (2011), an extract prepared using leaves and ethanol (200 mg/kg) was administered revealed antidepressant activity and improved the depressant conditions in forced swim tests and diazepam-induced sleeping time models.

3.5. Antidiabetic activity

A dose of 100 mg/kg of flower ethanol extract was orally administered to Streptozotocin-induced diabetic animals reduced elevated blood glucose concentrations (Mannangatti et al., 2010a).

3.6. Antihyperlipidemic activity

Mannangatti et al. (2010a) observed significant antihyper-

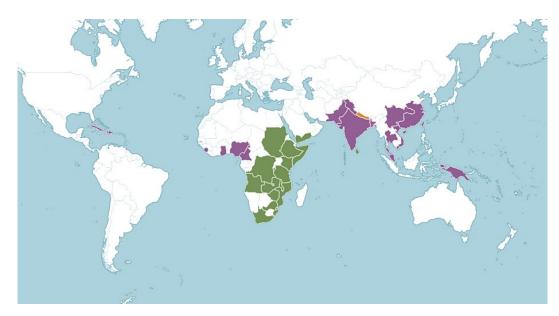


Fig. 2. Distribution map of *B. tomentosa* (Source: Plants of the World Online: Royal Botanic Gardens, Kew. Available at http://plantsofthe-worldonline.org/taxon/urn:lsid:ipni.org;names:30193-2). Keys: Orange: Doubtful; Green: Native and Purple: Introduced

lipidemic properties in Streptozotocin-induced diabetic animals after orally administrating 100 mg/kg of flower ethanol extract for 7 days.

3.7. Antinociceptive activity

An extract prepared using roots and ethanol (200mg/kg) was orally administered to mice unveiled antinociceptive potentials (Tiwari and Singh, 2013).

3.8. Antioxidant activity

Streptozotocin-induced diabetic animal models were orally administered 100 mg/kg of flower ethanol extract for 15 days showed antioxidant activity (Mannangatti et al., 2010b).

3.9. Antipyretic activity

In research performed by Tiwari and Singh (2015), root and stem 70% methanol extracts (200 mg/kg) were separately orally directed to yeast-induced hyperthermia models. After 300 min.s it was noticed that the hypothermia condition was reduced.

3.10. Anti-ulcerative colitis activity

An extract was made using leaves and 70% methanol and it was orally administered to colonic inflammation animals at a dose of 20 mg/kg. After 5 days, it was spotted significant antiulcerative colitis activity (Kannan and Guruvayoorappan, 2013).

3.11. Motor coordination activity

Ethanol extract of leaves (200 mg/kg) administered to animals showed motor coordination activity after 30 minutes in the Rotarod test (Sathya et al., 2011).

3.12. Nephroprotective activity

Kannan et al. (2016) administered 250 mg/kg leaf methanol extract to cisplatin-induced renal damaged models. After 5 days, it was observed that an elevation in antioxidant enzymes such as glutathione, catalase, and superoxide dismutase. Also, the bodyweight rose and reduced creatinine, serum urea, and lipid peroxidation. This study explains that this plant species has nephroprotective effects.

3.13. Nootropic activity

In a study carried out by Sathya et al. (2011), leaf methanol extract at a dose of 400 mg/kg was orally administered to the elevated plus-maze models exhibited nootropic properties.

3.14. Wound healing activity

The methanol extract was applied to both incision and excision wound models healed the wounds after 14 days (Panda et al., 2018).

4. Toxicity study

A study was carried out to observe the toxic and identify the safest dose (ED) of stem and root 70% methanol extracts separately. A dose of 3000 mg/kg orally administered to mice for 1 week showed no toxic effects and it is considered to be safe (Tiwari and Singh, 2015).

5. Conclusion

This review work revealed that *B. tomentosa* has a range of ethnomedicinal uses and scientific evidence is available for some of the ethnomedicinal utilizations. Hence, further bioactivities and phytochemical studies should be conducted to produce more scientific evidence to confirm the ethnomedicinal uses for standardization, safety, and efficacy purposes.

Also, the bioactive compounds should be discovered from this plant species, and they might be a candidate as a lead compound in future researches to combat diseases like cancer. Then these useful bioactive compounds could be synthesized in the laboratory to produce a large scale.

So far, an enormous number of bioactive phytochemicals have been isolated from several plant species. Anyway, not all the compounds or extracts have *in vivo* and clinical trial eviden-

ce and mechanisms of action for their bioactivities. Therefore, there is an urgent requirement to conduct these studies to find more effective drugs with few or no side effects than currently available drugs.

This work analyzed, documented, and summarized the reported bioactivities of *B. tomentosa*. Further, this work will be

Table

Reported bioactivities of *B. tomentosa*.

very useful for the researchers who are interested to study further bioactivity and phytochemical studies using this plant species.

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| Level of scien- tific evidence | Bioactivity | Part used | Extract/Compound | Assay/Model | Dose/Con- centration | Du- ra- tion | Reference |
|-----------------------------------|----------------|--------------|---------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------|--------------------|-----------------------------|
| In vitro | Antibacterial | Leaf | Chloroform, Metha- nol, Ethanol, Petro- leum ether, Ethyl a- cetate, Aqueous | Bacillus cereus, Bacillus subtilis, Escherichia coli, Pseudomonas ae- ruginosa, Staphylococcus aureus | 100 μg/ml (MIC) | NA | Mythreyi et al. (2005) |
| | | Leaf | Chloroform, Metha- nol, Ethanol, Petro- leum ether, Ethyl a- cetate, Aqueous | Candida albicans, Aspergillus niger | 100 μg/ml (MIC) | NA | |
| In vitro | Antibacterial | Root | Ethyl acetate | Proteus vulgaris (ATCC 12454) | 7 μg/ml (MIC) | NA | |
| | | Root | Ethyl acetate | Pseudomonas aeruginosa (ATCC 27853), Enterococcus faecalis (ATCC 2912), Bacillus subtilis (ATCC 10774), Bacillus pumilus (ATCC 14884), Escherichia coli (ATCC 25922) | 15 μg/ml (MIC) | NA | Dugasani et al. (2010) |
| | | Root | Ethyl acetate | Staphylococcus aureus (ATCC 25923) | 31 μg/ml (MIC) | NA | |
| | | Root | Hexane | Escherichia coli (ATCC 25922), Pseudomonas aeruginosa (ATCC 27853), Proteus vulgaris (ATCC 12454), Bacillus subtilis (ATCC 10774), Bacillus pumilus (ATCC 14884), Enterococcus faecalis (ATCC 2912), Staphylococcus au- reus (ATCC 25923) | 250 μg/ml (MIC) | NA | |
| | | Root | Methanol | Escherichia coli (ATCC 25922), Pseudomonas aeruginosa (ATCC 27853), Proteus vulgaris (ATCC 12454), Bacillus subtilis (ATCC 10774), Bacillus pumilus (ATCC 14884), Enterococcus faecalis (ATCC 2912) Staphylococcus aureus (ATCC | 31 μg/ml (MIC) 62 μg/ml | NA | |
| | | Root | Methanol | 25923) | (MIC) | NA | Disc et al |
| In vitro | Anticonvulsant | Leaf | Aqueous, Ethanol | GABA A-benzodiazepine receptor binding assay | 1 mg/ml | NA | Risa et al. (2004) |
| In vitro | Antifungal | Root | Aqueous, Ethanol Ethyl acetate | Candida krusei (ATCC 6258), Can- dida albicans (ATCC 10231) | 15 μg/ml (MIC) | NA | |
| | | Root | Hexane | Candida krusei (ATCC 6258), Can- dida albicans (ATCC 10231) | 250 μg/ml (MIC) | NA | Dugasani et a (2010) |
| | | Root | Methanol | Candida krusei (ATCC 6258), Can- dida albicans (ATCC 10231) | 31 μg/ml (MIC) | NA | |
| In vitro | Antihelminthic | Root | Ethanol, aqueous | Pheritema postuma, Ascaris lumbri- coides | 10% solu- tion | NA | Aditya et al (2013) |
| In vitro | Antioxidant | NS | NS | Mouse liver microsomes | 90 µg/ml (IC ₅₀) | NA | Kannan et a |
| | | NS | NS | NO radical scavenging | 65 μg/ml (IC ₅₀) | NA | (2010) |
| In vitro | Antioxidant | Pod, seed | NS | DPPH radical scavenging, NO radi- cal scavenging, OH radical scaveng- ing, ABTS radical scavenging, metal iron chelating, β-carotene-linoleate model system | NS | NA | Krishnaswan et al. (2013 |

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| In vitro | Antioxidant | Flower | Aqueous | DPPH radical scavenging | 74 μg/ml (IC ₅₀) | NA | Banerjee and |
|------------|-------------------------|---------------|-------------------------------|-------------------------------------------------------------|----------------------------------|------------|-------------------------------------------|
| | | Flower | Aqueous | Total antioxidant capacity | 265 μg/ml (IC ₅₀) | NA | De (2014) |
| | | NS | Aqueous | DPPH radical scavenging | 85 μg/ml | NA | |
| | | NS | Aqueous | NO radical scavenging | 310 µg/ml | NA | |
| In vitro | | NS | Ethanol | DPPH radical scavenging | 167 μg/ml (IC ₅₀) | NA | |
| | Antioxidant | NS | Ethanol | NO radical scavenging | 220 µg/ml | NA | Tiwari and |
| | | NS | Methanol | DPPH radical scavenging | 65 μg/ml (IC ₅₀) | NA | Singh (2013) |
| | | NS | Methanol | NO radical scavenging | 150 μg/ml | NA | |
| In vivo | | Root | Aqueous, methanol | Eddy's hot plate method in mouse | 200 mg/kg | 300 min | |
| | Analgesic | Root | Ethanol | Eddy's hot plate method in mouse | 400 mg/kg | 300 min | Tiwari and Singh (2013) |
| | | Root | Ethanol, Aqueous, Methanol | Acetic acid-induced writhing test in mouse | 400 mg/kg | 300 min | |
| | Anti-anxiety | Leaf | Ethanol | Elevated plus maze model, hole board test, light dark model | 200 mg/kg | NS | |
| In vivo | Anticatatonic | Leaf | Ethanol | Haloperidol-induced catalepsy | 200 mg/kg | NS | Sathya et al. (2011) |
| | Antidepressant | Leaf | Ethanol | Forced swim test, diazepam-induced sleeping time | 200 mg/kg | NS | (2011) |
| In vivo | Antidiabetic | Flower | Ethanol | Streptozotocin-induced diabetic | 100 mg/kg | 7 d | Mannangatti et al. (2010a) |
| In vivo | Antidiabetic | Leaf | Aqueous | Alloxan-induced diabetic | 300 mg/kg | 180 min | Devaki et al. (2011) |
| In vivo | Antidiabetic | Root | Petroleum ether | Alloxan-induced diabetic | 250 mg/kg | 14 d | Kaur et al. (2011) |
| In vivo | Antidiabetic | Stem | Aqueous, Ethanol | Streptozotocin-induced diabetic | 250 mg/kg | 21 d | Tiwari and Singh (2014) |
| In vivo | Antihyper- lipidemic | Flower | Ethanol | Streptozotocin-induced diabetic | 100 mg/kg | 7 d | Mannangatti et al. (2010a) |
| In vivo | Antinociceptive | Root | Ethanol | Mouse | 200 mg/kg | NS | Tiwari and Singh (2013) |
| | | Root, | M (1 1 (700/) | Mouse (Acetic acid-induced writh- | 250 // | 30 | |
| 7 . | . | Stem | Methanol (70%) | ing test) | 250 mg/kg | min | Tiwari and |
| In vivo | Antinociceptive | Root, Stem | Methanol (70%) | Mouse (Eddy's hot plate method) | 250 mg/kg | 120 min | Singh (2015) |
| In vivo | Antioxidant | Flower | Ethanol | Streptozotocin-induced diabetic | 100 mg/kg | 15 d | Mannangatti et al. (2010b) |
| In vivo | Antipyretic | Root, Stem | Methanol (70%) | Yeast-induced hyperthermia | 200 mg/kg | 300 min | Tiwari and Singh (2015) |
| In vivo | Anti-ulcerative colitis | Leaf | Methanol (70%) | Colonic inflammation | 20 mg/kg | 5 d | Kannan and Guruvayoorap- pan (2013) |
| In vivo | Motor coordina- tion | Leaf | Ethanol | Rota rod test | 200 mg/kg | 30 min | Sathya et al. (2011) |
| In vivo | Nephroprotective | Leaf | Methanol | Cisplatin-induced renal damage | 250 mg/kg | 5 d | Kannan et al. (2016) |
| In vivo | Nootropic | Leaf | Ethanol | Elevated plus maze model | 400 mg/kg | NS | Sathya et al. (2011) |
| In vivo | Wound healing | NS | Methanol | Incision wound, excision wound | NS | 14 d | Panda et al. |

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