



## Comparative Genomics Insight into Phytopathogenic *Xanthomonas arboricola* pathovar *corylina* Strains

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**Abstract:** *Xanthomonas arboricola* pathovar *corylina* (*Xac*) causes a bacterial blight disease (BBD) resulting in economic losses in young hazelnut trees worldwide. Although virulent *Xac* genomes were sequenced, there is no comparative genomics study on these genomes. In this study, all publicly available whole-genome sequences of *Xac* were compared by a comparative genomics approach. The results showed that *Xac* CFBP1159 and *Xac* CFBP2565 genomes are phylogenetically related to each other based on the orthology results. The genomic diversification of *Xac* strains was depended on mobile genetic elements. Interestingly, *Xac* NCCB100457 genome had additional motility genes than *Xac* CFBP1159 and *Xac* CFBP2565 genomes. All *Xac* genomes had shared virulence-related genes such as secretion systems and adherence factors. The comparative genomics approach of this study supports that *Xac* genomes have slight genetic variations, and the virulence-related proteins interacted with the host proteins. This comparative genomics approach will provide insights into the understanding of the *Xac* genomes.

**Keywords:** Comparative genomics, host-pathogen interaction, phytopathogenic, virulence, *Xanthomonas*.

## Fitopatojenik *Xanthomonas arboricola* patovar *corylina* Suşlarına Karşılaştırmalı Genomik Bakış

**Öz:** *Xanthomonas arboricola* patovar *corylina* (*Xac*), Dünya çapında genç fındık ağaçlarında ekonomik kayıplarla sonuçlanan fındık bakteriyel yanıklığı hastalığına neden olmaktadır. Virulent *Xac* suşlarının tüm genomları dizilenmiş olmasına rağmen, bu genomlar üzerinde bir karşılaştırmalı genomik çalışma yoktur. Bu çalışmada, *Xac* suşlarının halka açık tüm genom dizileri, karşılaştırmalı genomik yaklaşım kullanılarak karşılaştırılmıştır. Elde edilen sonuçlarda, *Xac* CFBP1159 ve *Xac* CFBP2565 genomlarının, ortoloji sonucuna göre filogenetik olarak birbirleriyle daha yakın ilişkili olduğu saptanmıştır. *Xac* suşlarının genomik çeşitliliğinin mobil genetik elementlerle ilişkisi olduğu anlaşılmıştır. İlginç bir şekilde, *Xac* NCCB100457 genomu, *Xac* CFBP1159 ve *Xac* CFBP2565 genomlarından daha fazla motilite genlerine sahiptir. Tüm *Xac* genomları, sekresyon sistemleri ve adhezyon faktörleri gibi virülans ile ilgili ortak genlere sahiptir. Bu çalışmanın karşılaştırmalı genomik yaklaşımı, *Xac* genomlarının bazı genetik varyasyonlara sahip olduğu ve virülans ile ilgili proteinlerin konakçı proteinlerle etkileşime girdiğini desteklemektedir. Bu çalışmanın karşılaştırmalı genomik yaklaşımı, *Xac* genomlarının anlaşılması için bir öngörü sağlamıştır.

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**Anahtar kelimeler:** Bitki patojeni, karşılaştırmalı genomik, konak patojen ilişkisi, virülans, *Xanthomonas*.

## INTRODUCTION

*Xanthomonas arboricola* is primarily known as a Gram-negative phytopathogenic bacterium that can infect economically important plant species such as prunus, walnut, and hazelnut (Vauterin et al., 1995). *X. arboricola* pathovar-assigned strains are mostly described as pathogenic bacteria although there are *X. arboricola* strains considered non-pathogenic bacteria. Moreover, there are nine pathovar strains (*arracaciae*, *celebensis*, *corylina*, *fragariae*, *guizotiae*, *juglandis*, *populi*, *pruni*, and *zantesdeschiae*) identified in *X. arboricola* (Fischer-Le Saux et al., 2015). As one of the most virulent pathovars, *Xanthomonas arboricola* pathovar *corylina* (*Xac*) is a causative agent of BBD of hazelnut trees (*Corylus* spp.). The BBD was first described on *C. maxima* in the USA (Barss, 1913). Later, the disease has been increasingly reported on *C. avellana* from hazelnut producer countries in the European Union (Eppo, 2004). *Xac* has been also prevalently isolated from the hazelnut orchards in the Black Sea Region of Turkey, which is a major hazelnut producer in Europe (Karahan et al., 2013). The symptoms of BBD can appear on hazelnut tree's leaves and twigs. It has been also reported that the disease can also kill hazelnut trees or delay the growth of trees (Lamichhane et al., 2014).

The genus of *Xanthomonas* include a variety of plant pathogenic bacterial species, and these bacteria can utilize diverse virulence mechanisms such as secretion systems, effector proteins of diverse secretion systems,

flagella, and small RNAs (Timilsina et al., 2020). Recently, a comparative genomics study on virulent and non-virulent *Xanthomonas arboricola* pv. *pruni* (*Xap*) strains showed genomic differences between virulent and non-virulent *Xap* strains (Garita et al., 2017). On the other hand, the *Xac* strain was isolated from the leaf spots of *C. colurna* L. in Colorado, and the first DNA sequence of *Xac* was revealed in 2013 (Caballero et al., 2013). There are also sequences of two *Xac* genomes (*Xac* CFBP1159 and *Xac* CFBP2565) publicly stored in the National Center for Biotechnology Information (NCBI). However, these genomes were not used for any comparative genomics analysis.

Although several *Xac* genomes have been sequenced and their genomes were publicly available, there is no comparative genomics study on *Xac*'s genomes. Thus, the goal of this study was to compare *Xac* genomes that are publicly available in the NCBI database. This study provides a comparative insight into the comparison of *Xac* genomes, genomic variations, orthologous clusters at the protein level, annotation and gene predictions, prediction of virulence factors, and interactions of host-pathogen proteins.

## MATERIAL AND METHOD

**Bacterial genome data:** The *Xac* genomes data were obtained from NCBI (as of 01/11/2020). The *Xac* genomes were listed with features in Table 1.

**Table 1.** Genome features of *Xac* strains.

Species	Strain	Host	Location	Size (Mbp)	G+C	Level	Accession #	Reference
<i>Xac</i>	CFBP2565	<i>C. avellana</i>	France	5.05	65.6	Contig	NZ_MDSJ01000001.1	INRA
<i>Xac</i>	CFBP1159	<i>C. maxima</i>	USA	5.11	65.5	Scaffold	NZ_MDEA01000001.1	INRA
<i>Xac</i>	NCCB100457	<i>C. colurna</i>	USA	5.23	65.5	Contig	NZ_APMC02000173.1	Caballero et al., 2013

Location: country of origin, Level: genome assembly status, Mbp: million base pairs, *Xac*: *Xanthomonas arboricola* pathovar *corylina*, *C*: *Corylus*.

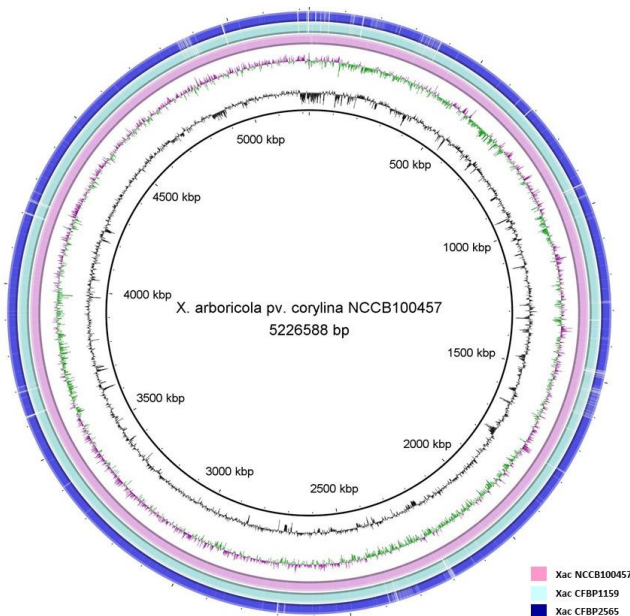
**Comparative genome analyses:** Blast Ring Image Generator (BRIG) was used to visualize the comparative genome analysis of *Xac* strains (Alikhan et al., 2011). The orthologous clusters of *Xac* strains at the protein level were calculated by using OrthoVenn2 (Wang et al., 2015). Prokaryotic Genomes Automatic Annotation Pipeline (NCBI PGAAP) and Rapid Annotation using Subsystem Technology (RAST) annotation pipelines were used for the annotation and gene prediction of bacterial genomes (Angiuoli et al., 2008; Overbeek, 2014). The potential protein-protein interactions between 3 of *Xac* whole proteins and the complete proteins of the eukaryotic host *C. avellana* (European hazelnut) genome (GenBank accession #: CAAJGP010000000) were determined using the Host-Pathogen Interaction Database (HPIDB) by the default upload options (Ammari et al., 2016; Kumar and Nanduri, 2010). The predicted putative virulence factors were determined by downloading the full dataset from the

Virulence Factors Database (VFDB) and by uploading them to BioEdit software (Lihong et al., 2016). The local BLAST feature of BioEdit was used with a cutoff E-value of  $10^{-50}$  (Hall, 1999).

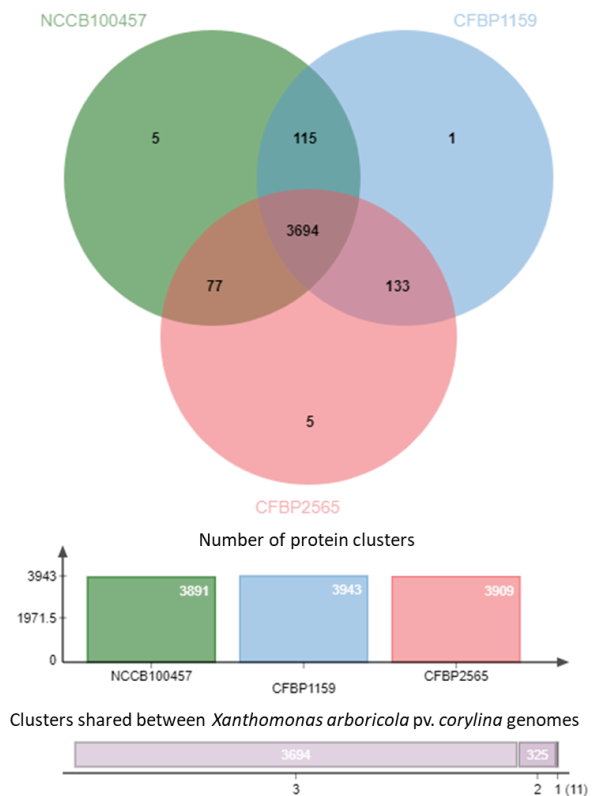
## RESULTS

**Genome features of *Xac* genomes:** The genomic information about *Xac* genomes (plant host, location, and genome-level) was summarized in Table 1. The three genomes of *Xac* was represented and stored in NCBI. The average genome size of *Xac* genomes is 5.13 million base-pairs (Mbp). The average G+C content of *Xac* genomes is 65.53 mol%. The NCBI genome data indicated that *Xac* genomes were isolated from different hazelnut species (Table 1). There are no plasmids sequences reported for *Xac* strains.

**Comparative analyses of *Xac* genomes:** The visualization of the comparison of genes by BRIG showed that most of the genes among all *Xac* genomes were conserved (Fig 1). In orthology analysis, the comparison of proteins encoded by *Xac* genomes was determined by OrthoVenn2 (Fig 2). All strains shared 3694 clusters of orthologous proteins. The unique protein clusters identified such as 1 cluster (Non-ribosomal peptide synthetase) in *Xac* CFBP1159, 5 clusters (Phage portal protein, P-type conjugative transfer protein TrbJ, DDE-type integrase/transposase) in *Xac* CFBP2565, and 5 clusters (SDR family oxidoreductase, UvrD helicase, virulence RhuM family protein, transcriptional regulator) in *Xac* NCCB100457 was unique to genomes. The annotation and categorization of genes based upon the RAST showed that there is a close relationship between *Xac* CFBP1159 and *Xac* CFBP2565 genomes in the subsystems categorization of genes (Fig 3). However, *Xac* NCCB100457 had slightly more genes in some subcategories compared to other *Xac* genomes. Interestingly, *Xac* NCCB100457 had motility and chemotaxis genes, which other *Xac* genomes do not encode any of these genes (Fig 3). In host-pathogen interactions (HPIs), the interactions were predicted between a total of 26 proteins of *Xac* and 34 proteins of *C. avellana* (Fig 4). The virulence factors detected in *Xac* genomes were categorized based on the function such as adherence, adhesion, and secretion systems (Table 2). The pathogen proteins interacted with the host proteins are listed in Table 3.



**Figure 1.** Comparative circular visualization of *Xac* genomes. The reference strain is *Xac* NCCB100457. The two inner rings represent the G+C content (black) and GC-skew (green/purple). The three outside rings represent a genomics comparison between *Xac* CFBP1159 and *Xac* CFBP2565 strains and the reference *Xac* NCCB100457 strain.



**Figure 2.** Venn diagram of protein clusters encoded by *Xac* genomes based on orthology. The Venn diagrams and bar charts show the numbers of unique and shared orthologous genes of each genome.

**Table 2.** Virulence factors of *Xac* strains.

Class	Virulence Factor	<i>Xanthomonas arboricola</i> pv. <i>corylina</i>		
		CFBP1159	CFBP2565	NCCB100457
Adherence	Type IV pili	+	+	+
Adhesion	Outer membrane protein	+	+	+
Adhesion	Autotransporter-like protein	+	+	+
Secretion system	Type II secretion system	+	+	+
Secretion system	Type III secretion system	+	+	+

(+): Presence of virulence factor in the genome.

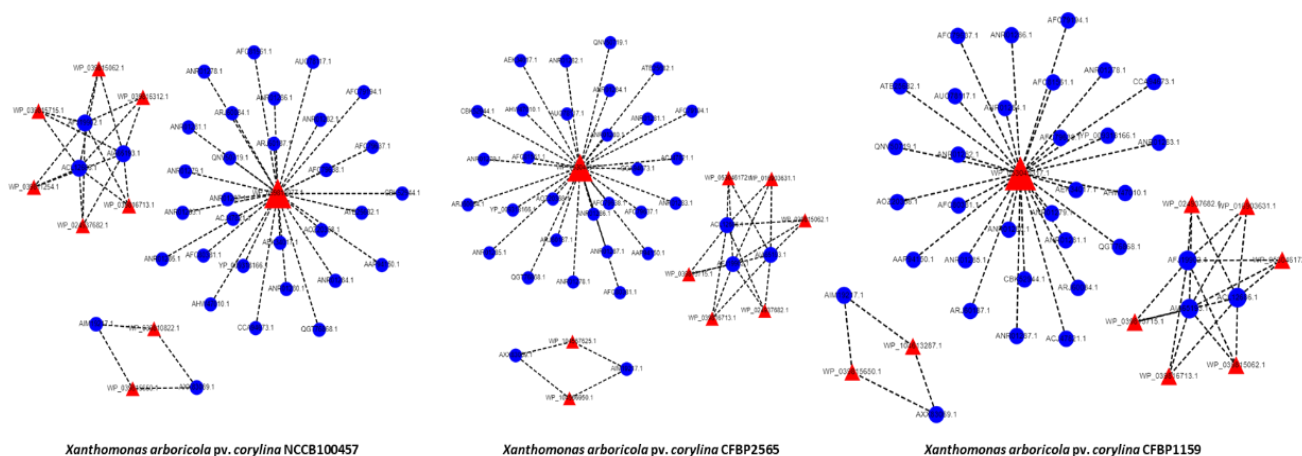
**Table 3** Predicted pathogen proteins in host-pathogen interactions.

Species	Protein ID	Pathogen Protein
<i>Xac</i> CFBP1159	WP_016903631.1	Diaminopimelate epimerase
	WP_024937682.1	ParA family protein
	WP_039815062.1	tRNA preQ1(34) S-adenosylmethionine ribosyltransferase-isomerase
	WP_039815650.1	Response regulator
	WP_039815715.1	Histidinol dehydrogenase
	WP_039816713.1	ParA family protein
	WP_053045317.1	Thioredoxin TrxC
	WP_053046172.1	Mannitol dehydrogenase family protein
	WP_104613287.1	CHASE3 domain-containing protein
	WP_016903631.1	Diaminopimelate epimerase
<i>Xac</i> CFBP2565	WP_024937682.1	ParA family protein
	WP_039815715.1	Histidinol dehydrogenase
	WP_039816713.1	ParA family protein
	WP_053045317.1	Thioredoxin TrxC
	WP_053046172.1	Mannitol dehydrogenase family protein
	WP_104566950.1	CHASE3 domain-containing protein
	WP_104567625.1	Response regulator
	WP_024937682.1	ParA family protein
	WP_039810077.1	Thioredoxin TrxC
	WP_039810822.1	CHASE3 domain-containing protein
<i>Xac</i> NCCB100457	WP_039811254.1	Diaminopimelate epimerase
	WP_039815062.1	tRNA preQ1(34) S-adenosylmethionine ribosyltransferase-isomerase
	WP_039815650.1	Response regulator
	WP_039815715.1	Histidinol dehydrogenase
	WP_039816312.1	Mannitol dehydrogenase, partial
	WP_039816713.1	ParA family protein

*Xac*: *Xanthomonas arboricola* pathovar *corylina*.

Strains	Subsystems																										
	Amino Acids and Derivatives	Carbohydrates	Membrane Transport	Protein Metabolism	Cofactors, Vitamins, Prosthetic Groups, Pigments	Respiration	DNA Metabolism	Stress Response	Fatty Acids, Lipids, and Isoprenoids	Nucleosides and Nucleotides	RNA Metabolism	Cell Wall and Capsule	Metabolism of Aromatic Compounds	Miscellaneous	Virulence, Disease and Defense	Regulation and Cell Signaling	Phosphorus Metabolism	Sulfur Metabolism	Potassium Metabolism	Iron Acquisition and Metabolism	Secondary Metabolism	Nitrogen Metabolism	Phages, Prophages, Transposable Elements, Plasmids	Dormancy and Sporulation	Motility and Chemotaxis	Cell Division and Cell Cycle	Photosynthesis
Xanthomonas arboricola pv. corylina CFBP1159	288	207	185	184	131	93	88	79	66	55	41	31	29	27	25	25	23	17	11	11	8	6	2	1	0	0	0
Xanthomonas arboricola pv. corylina CFBP2565	288	208	180	182	129	93	87	79	64	55	39	28	29	27	23	26	23	17	11	6	8	6	3	1	0	0	0
Xanthomonas arboricola pv. corylina NCCB100457	295	217	197	192	129	93	103	80	70	57	41	29	29	28	28	26	26	19	11	11	8	6	0	1	27	0	0
	Number of Genes																										
	295	288	208	180	129	103	80	70	55	39	29	17	8	0													

**Figure 3.** SEED subsystem categorization and RAST annotation of *Xac* genomes. The colors indicate the number of functional categorization of genes in *Xac* genomes.



**Figure 4.** Visualization of predicted interactions network of host *C. avellana* proteins and bacterial *Xac* proteins. The shapes and colors represent host proteins (blues circles) and bacterial proteins (red triangles).

## DISCUSSION AND CONCLUSION

In this research, all publicly available genomes of *Xac* were analyzed by comparative genomics methods. This is the first comparative genomics analysis of *Xac* genomes isolated from three different hazelnut species.

Orthology analysis provides more accurate information about the phylogenetic relationship between closely related bacterial strains (Ullah et al., 2015). Based on the orthology analysis of *Xac* genomes in this study, *Xac* CFBP1159 and *Xac* CFBP2565 genomes are more closely related than *Xac* NCCB100457 genome. On the other hand, *Xac* strains share 3694 orthologous clusters. The genetic differences among genomes depend on mobile elements. This finding suggests that genetic differentiation of *Xac* genomes is linked to the acquisition of different mobile elements. Interestingly, *Xac* NCCB100457 genome possesses additional motility and chemotaxis gene sets.

This variation may be related to location and host adaptation.

The prediction of potential interactions between host and pathogen proteins may provide valuable information about an infection process (Durmus et al., 2015). The HPIs analysis in this study showed that a total of 26 proteins of *Xac* proteins interacted with host *C. avellana* proteins. These results confirm that multiple potential interactions occur between *Xac* and *C. avellana* in the host-pathogen interactions. Thus, the HPIs result demonstrated evidence of *Xac* virulence during infection. Determination of virulence factors of bacterial pathogens is important to understand the pathogenesis of bacteria during the infection process (Wu et al., 2008). The whole-genome sequencing of *Xac* strains enables identifying virulence-related factors. Identification of virulence factors in this study revealed that *Xac* genomes have important virulence-related factors such as type II secretion system (T2SS), type III secretion system (T3SS), and adhesion

factors (Table 2). The bacterial adhesion of *Xanthomonas* strains is crucially important for the invasion of plant tissue during the diseases process (Mhedbi-Hajri et al., 2011). While *Xac* strains invade plant tissues, degradative enzymes and secretion system-dependent effector proteins play a major role in establishing a successful bacterial infection. For example, T3SS and effector proteins are identified as important virulence factors in plant immunity suppression by *Xac* strains (Hajri et al., 2011; Jacques et al., 2016). In addition to T3SS, T2SS also plays an important role in secreting a variety of degradation enzymes in the genus of *Xanthomonas* (Szczeny et al., 2010). Overall, these virulence factors may contribute to the bacterial pathogenicity of *Xac* strains in the host.

As a result, the comparative genomics data presented in this study showed that *Xac* strains are closely related to each other. The virulence-related factors of *Xac* strains are also important in host-pathogen interactions.

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#### REFERENCES

- Alikhan, N.F., Petty, N.K., Ben Zakour, N.L. & Beatson, S.A. (2011). Blast ring image generator (BRIG): simple prokaryote genome comparisons. *BMC Genomics*, *12*, 402.
- Ammari, M.G., Gresham, C.R., McCarthy, F.M. & Nanduri, B. (2016). HPIDB 2.0: a curated database for host-pathogen interactions. *Database*, (Oxford), *2016*, baw103.
- Angiuoli, S.V., Gussman, A., Klimke, W., Cochrane, G., Field, D., Garrity, G., Kodira, C.D., Kyrpides, N., Madupu, R., Markowitz, V., Tatusova, T., Thomson, N. & White, O. (2008). Toward an online repository of Standard Operating Procedures (SOPs) for (meta) genomic annotation. *OMICS*, *12*, 137-141.
- Barss, H.P. (1913). A new filbert disease in Oregon. *Oregon Agricultural Experiment Station Biennial Crop Pest and Horticulture Report*, *14*, 213-23.
- Caballero, J.I., Zerillo, M.M., Snelling, J., Boucher, C. & Tisserat, N. (2013). Genome sequence of *Xanthomonas arboricola* pv. *corylina*, isolated from Turkish Filbert in Colorado. *Genome Announcement*, *1*(3), e00246-13. DOI: [10.1128/genomeA.00246-13](https://doi.org/10.1128/genomeA.00246-13)
- Durmus, S., Cakir, T., Ozgur, A. & Guthke, R. (2015). A review on computational systems biology of pathogen-host interactions. *Frontiers in Microbiology*, *6*, 235. DOI: [10.3389/fmicb.2015.00235](https://doi.org/10.3389/fmicb.2015.00235)
- EPPO. (2004). (2004). Diagnosis protocols for regulated pests *Xanthomonas arboricola* pv. *corylina*. *EPPO Bulletin*, *34*, 155-7.
- Fischer-Le Saux, M., Bonneau, S., Essakhi, S., Manceau, C. & Jacques, M.A.A. (2015). Aggressive emerging pathovars of *Xanthomonas arboricola* represent widespread epidemic clones distinct from poorly pathogenic strains, as revealed by multilocus sequence typing. *Applied Environmental Microbiology*, *81*, 4651-4668. <https://doi.org/10.1128/AEM.00050-15>
- Garita, C.J., Palacio-Bielsa, A., López, M.M. & Cubero, J. (2017). Pan-Genomic analysis permits differentiation of virulent and non-virulent Strains of *Xanthomonas arboricola* that Cohabit *Prunus* spp. and elucidate bacterial virulence factors. *Frontiers in Microbiology*, *8*, 573. DOI: [10.3389/fmicb.2017.00573](https://doi.org/10.3389/fmicb.2017.00573)
- Hajri, A., Pothier, J.F., Saux, M.F.L. Bonneau, S., Poussier, S., Boureau, T., Duffy, B. & Manceau, C. (2011). Type three effector gene distribution and sequence analysis provide new insights into the pathogenicity of plant-pathogenic *Xanthomonas arboricola*. *Applied and Environmental Microbiology*, *78*(2), 371-384. DOI: [10.1128/AEM.06119-11](https://doi.org/10.1128/AEM.06119-11)
- Hall, T.A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, *41*, 95-98.
- Jacques, M.A., Arlat, M., Boulanger, A., Boureau, T., Carrère, S., Cesbron, S., Chen, N.W., Cociancich, S., Darrasse, A., Denancé, N., Fischer-Le Saux, M., Gagnevin, L., Koebnik, R., Lauber, E., Noël, L.D., Pieretti, I., Portier, P., Pruvost, O., Rieux, A., Robène, L., Royer, M., Szurek, B., Verdier, V. & Vernière, C. (2016). Using Ecology, Physiology, and Genomics to Understand Host Specificity in *Xanthomonas*. *Annual Reviews Phytopathology*, *4*(54), 163-87. DOI: [10.1146/annurev-phyto-080615-100147](https://doi.org/10.1146/annurev-phyto-080615-100147)
- Karahan, A., Altundağ, Ş., Duran, H. & Kılınc, A.O. (2013). Karadeniz Bölgesinde fındık bakteriyel yanıklığı [*Xanthomonas arboricola* pv. *corylina* (Miller et al.) Vauterin et al.] hastalığının yaygınlığı üzerine araştırmalar. *Bitki Koruma Bülteni*, *53*(3), 159-174.
- Kumar, R. & Nanduri, B. (2010). HPIDB - a unified resource for host-pathogen interactions. *BMC Bioinformatics*, *11*(6), S16. DOI: [10.1186/1471-2105-11-S6-S16](https://doi.org/10.1186/1471-2105-11-S6-S16)
- Lamichhane, J.R. & Varvaro, L. (2014). *Xanthomonas arboricola* disease of hazelnut: current status and future perspectives for its management. *Plant Pathology*, *63*(2), 243-254. DOI: [10.1111/ppa.12152](https://doi.org/10.1111/ppa.12152)
- Lihong, C., Dandan, Z., Bo, L., Jian, Y. & Qi, J. (2016). VFDB 2016: hierarchical and refined dataset for big data analysis-10 years on. *Nucleic Acids*

- Research*, **44**, D694-D697. DOI: [10.1093/nar/gkv1239](https://doi.org/10.1093/nar/gkv1239)
- Mhedbi-Hajri, N., Darrasse, A., Pigne, S., Durand, K., Fouteau, S., Barbe, V., Manceau, C., Lemaire, C. & Jacques, M.A. (2011).** Sensing and adhesion are adaptive functions in the plant pathogenic xanthomonads. *BMC Evolutionary Biology*, **11**, 67.
- Overbeek, R., Olson, R., Pusch, G.D., Olsen, G.J., Davis, J.J., Disz, T., Edwards, R.A., Gerdes, S., Parrello, B., Shukla, M., Vonstein, V., Wattam, A.R., Xia, F. & Stevens, R. (2014).** The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Research*, **42**, D206-D214. DOI: [10.1093/nar/gkt1226](https://doi.org/10.1093/nar/gkt1226)
- Szczesny, R., Jordan, M., Schramm, C., Schulz, S., Cogež, V., Bonas, U. & Büttner, D. (2010).** Functional characterization of the Xcs and Xps type II secretion systems from the plant pathogenic bacterium *Xanthomonas campestris* pv *vesicatoria*. *New Phytologist*, **187**(4), 983-1002. DOI: [10.1111/j.1469-8137.2010.03312.x](https://doi.org/10.1111/j.1469-8137.2010.03312.x)
- Timilsina, S., Potnis, N., Newberry, E.A., Liyanapathiranage, P., Iruegas-Bocardo, F., White, F.F., Goss, M.M. & Jones, J.B. (2020).** *Xanthomonas* diversity, virulence and plant-pathogen interactions. *Nature Reviews Microbiology*, **18**, 415-427.
- Ullah, I., Sjöstrand, J., Andersson, P. & Sennblad, B. (2015).** Integrating sequence evolution into probabilistic orthology analysis. *Systematic Biology*, **64**(6), 969-982. DOI: [10.1093/sysbio/syv044](https://doi.org/10.1093/sysbio/syv044)
- Vauterin, L., Hoste, B., Kersters, K. & Swings, J. (1995).** Reclassification of *Xanthomonas*. *International Journal of Systematic and Evolutionary Microbiology*, **45**(3), 472-489. DOI: [10.1099/00207713-45-3-472](https://doi.org/10.1099/00207713-45-3-472)
- Wang, Y., Coleman-Derr, D., Chen, G. & Gu, Y.Q. (2015).** OrthoVenn: a web server for genome wide comparison and annotation of orthologous clusters across multiple species. *Nucleic Acids Research*, **43**, W78-W84. DOI: [10.1093/nar/gkv487](https://doi.org/10.1093/nar/gkv487)
- Wu, H.J., Wang, A.H.J. & Jennings, M.P. (2008).** Discovery of virulence factors of pathogenic bacteria. *Current Opinion in Chemical Biology*, **12**(1), 93-101. DOI: [10.1016/j.cbpa.2008.01.023](https://doi.org/10.1016/j.cbpa.2008.01.023)