Egg White Cryogel for Removal of Methylene Blue from Aqueous Solution

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Abstract

Macroporous egg white (EW) cryogel synthesized by cryogelation at sub-zero temperature (-18°C) with crosslinker glutaraldehyde (GA) was tested in the adsorption of methylene blue (MB) as a model pollutant from aqueous solution. The characterization of obtained cryogel was carried out using Fourier-transform infrared spectroscopy and scanning electron microscopy. The effects of initial dye concentration, pH and contact time parameters on the adsorption were systematically evaluated. The pseudo-first order and pseudo-second order models were used to predict the adsorption kinetics. The adsorption followed the pseudo-second order model. Two isotherm models called Langmuir and Freundlich were applied on the experimental data to predict the maximum adsorption capacity and explain the process of adsorption. Langmuir isotherm model provided the best fit with maximum adsorption capacity of 56.18 mg MB/g cryogel. The dye adsorbed EW cryogel was easily regenerated and used several times with no noticeable reduction in its adsorption properties. The electrostatic attraction was the main adsorption mechanism of MB on the cryogel, especially at slightly basic pH. Its easy preparation, low-cost and good adsorption property make the EW cryogel a promising adsorbent for environmental applications.

Keywords: Cryogel, Biosorbent, Egg White, Methylene Blue, Adsorption

Introduction

Environmental pollution has increased seriously in direct proportion to the increase in industrial and human activities all over the world. Especially water is contaminated with toxic substances such as dyes and heavy metals, etc. Water pollution with dyes used in various areas such as textiles, printing, plastics and cosmetics is increasing more than ever, so that water treatment has become a very important issue for environmental protection today. These dyes can be dangerous to aquatic living organisms. Methylene blue (MB) is a cationic dye which can be more toxic than anionic dyes due to its interaction with anionic surface of cell membrane and penetration into the living cells (Bayramoglu et al., 2009; He et al., 2013). Moreover, MB as well as many dyes can cause many serious health problems in humans like increased heartbeat, vomiting, shock, cyanosis, jaundice, tissue necrosis and even cancer.

Biological treatment, coagulation/flocculation, photocatalysis, membrane filtration, adsorption and chemical oxidation are the most commonly applied techniques for removing pollutants from wastewaters (Bolto and Gregory, 2007; Hasanpour and Hatami, 2020; Kanaujiya et al., 2019; Karabayir et al., 2019; Kumar and Sivanesan, 2006). Among them, adsorption seems more effective and feasible thanks to its advantages such as ease of application, low-cost and availability of various adsorbents. During the last few decades, researchers have been introduced different types of adsorbents including natural materials, biosorbents and functional polymeric materials with high adsorption capacities, reusable and easy preparation (Crini, 2006; Dalaran et al., 2011; Erdem et al., 2017). Bio-based polymeric materials have attracted great attention due to their ability to be nontoxic and biodegradable. For example, alginate, chitin, chitosan, starch, carboxymethylcellulose based hydrogels have been tested as adsorbents (Crini and Badot, 2008; Dragan and Loghin, 2013; Huang et al., 2011; Rocher et al., 2010). Egg white (EW), mainly consists of ~ 90% water and ~10% protein, is a good alternative biopolymer to fabricate adsorbent owing to its cheap and biodegradable without leading to secondary pollution. For example, egg white/polyethyleneimine hydrogel crosslinked with epichlorohydrin was prepared for removing heavy metal ions (Godiya et al., 2020). In another study, EW biosorbent was prepared by crosslinking with N,N′-methylenebis(acrylamide) for removing anionic indigo carmine (Oymak and Bağda, 2018). Unlike these two studies, it was aimed here to prepare macroporous EW cryogel as a biosorbent.

Cryogelation or cryotropic gelation is one of the several methods to fabricate porous materials (Dinu et al., 2007; Lozinsky et al., 2001). The hydrogel obtained by this method is called cryogel. During cryogelation, solvent is frozen at temperatures below freezing point of the reaction solution and crystals are formed as a porogen. It is generally performed at sub-zero temperature because most of the time water is
used as a solvent. After thawing, the structure with interconnected large pores in the cryogels is obtained, which provide not only sufficient surface area for adsorption but also fast mass transfer of substances. Interconnected macropores even could facilitate the flow of liquids such as wastewater at low back pressure.

In the present study, inexpensive and abundant EW as a biopolymer was crosslinked with glutaraldehyde (GA) at -18°C to prepare a cryogel for the removal of MB. The synthesis procedure was simple and consists of only mixing with GA and subsequently cooling. The functional groups and morphology of prepared cryogel were analyzed. The effects of pH (3-9), initial dye concentration (10-180 mg/L) and time (0.5-48h) on the adsorption were systematically investigated. The adsorption mechanism of MB on the EW cryogel was predicted by applying various isotherm and kinetic models on the experimental data.

**Materials and Methods**

Chicken eggs (Local market), methylene Blue (MB, Merck), glutaraldehyde solution 25% (GA, Merck), hydrochloric acid (HCl, Merck), sodium hydroxide (NaOH, Merck) and n-hexane (Lab-scan) were used without further purification.

EW cryogel was fabricated using glutaraldehyde as a crosslinker (Balaji et al., 2019). Firstly, the EW was carefully separated from the yolk in Figure 1. Collected EW was stirred for 10 min to make homogeneous. Then, 10 mL of EW was mixed with 200 µL glutaraldehyde solution (final concentration 0.5%) and 40 µL concentrated HCl. The reaction solution was immediately poured into 1 mL plastic syringes and placed at -18°C for 24 h. After cryogelation, the cryogel samples were immersed in large amount of distilled water at room temperature to extract dissolved and unreacted reagents by changing water over several days. The samples were then dried.

The dried EW cryogels were swollen in water up to the equilibrium to determine the swelling ratio (SR) which is calculated by equation given below,

$$SR = \frac{m_s}{m_d}$$  \hspace{1cm} (Eq. 1)

where $m_d$ and $m_s$ are the weights of dried and corresponding swollen gel sample at equilibrium, respectively. The pore volume ($V_p$) was determined by immersing the dried sample in $n$-hexane as a poor solvent for 1h and measuring the weight of the sample at swollen state (Dinu et al., 2011). $V_p$ was calculated by equation given below,

$$V_p = \frac{m_s - m_d}{d_n m_d}$$ \hspace{1cm} (Eq. 2)

where $d_n$ is the density of $n$-hexane. The porosity ($P\%$) was estimated by squeezing the cryogel swollen in water and removing free water from macropores (Henderson et al., 2013). $P\%$ was calculated by equation given below,

$$P\% = \frac{m_s - m_{sd}}{m_s} \times 100$$ \hspace{1cm} (Eq. 3)

where $m_{sd}$ is the weight of squeezed cryogel sample.

The functional groups in the cryogel were analyzed using a Fourier transform infrared spectroscopy (FTIR, Perkin-Elmer) in the range 4000-450 cm$^{-1}$. The microstructure of the cryogel was visualized using a scanning electron microscope (SEM, FEI/Quanta FEG 250) at various magnifications between 30 and 1000 times. The sample was sprayed with platinum and observed under an operating voltage of 20 kV. The pore sizes were measured from SEM images using Image J software.

The dye adsorption in aqueous solution was systematically evaluated by a batch procedure. UV spectrophotometry (Shimadzu UV 2600) was used to monitor the spectra and measure the absorbance of dye in solution. Before adsorption experiment, the calibration curve of MB at 664 nm was obtained by preparing solutions with different concentrations. ~10 mg cryogel samples were added into 10 mL solution of a certain concentration of MB at different initial pH values. The pH was adjusted using 1M NaOH and 1M HCl solutions. The experiments were performed in an isothermal shaker at 25°C and 120 rpm. The absorbance of residual dye in solution was measured and then the concentration was determined from calibration curve. The equilibrium adsorption capacity ($q_e$ (mg/g) (dye/dried cryogel) was calculated by equation given below,

$$q_e = \frac{(c_o - c_e)V}{m}$$ \hspace{1cm} (Eq. 4)

where $C_o$ and $C_e$ (mg/L) are the initial and residual concentrations of dye in aqueous solutions, respectively. $V$ (mL) is the volume of dye solution and $m$ (mg) is the weight of dried EW cryogel.

The regeneration and reusability of the cryogel were performed in repetitive adsorption/desorption cycles. The dye adsorbed cryogel after the adsorption step (15 mg/L of initial dye concentration) was immersed into 1M HCl solution (20 mL) and shaken for 2h. After this process was repeated twice, the cryogel was removed from the solution and rinsed with water for neutralization. In this way, the cryogel was regenerated for re-adsorption. The reusability was investigated by applying four adsorption/desorption cycles. From the absorbed dye concentration, the regeneration capacity ($R\%$ for each cycle was calculated by equation given below (Godiya et al., 2020),

$$R\% = \frac{c_o - c_r}{c_o - c_i} \times 100$$ \hspace{1cm} (Eq. 5)
where $C_i$ and $C_r$ are the residual concentrations of dye in aqueous solution (mg/L) after initial cycle and regeneration, respectively.

For kinetic study, the adsorption experiments as a function of contact time (15 mg/L of aqueous dye solution at pH9) were carried out. The adsorption capacity in any time $q_t$ (mg/g) was calculated by equation given below,

$$q_t = \frac{(C_0 - C_t)V}{m}$$

(Eq. 6)

where $C_t$ is the dye concentration in aqueous solution (mg/L) at any time $t$.

**Results and Discussion**

In our approach, the EW cryogel was obtained by crosslinking with GA via cryogelation method. The characterization of network structure was performed by FTIR. The spectra of the EW crosslinked with and without GA are given in Figure 2. The peaks at 1538 and 1627 cm$^{-1}$ correspond to Amide II (N–H bending vibration) and Amide I (C=O stretching vibration), respectively. These two characteristic peaks related to the functional groups are commonly observed in proteins (Garidel & Schott, 2006). The strong peak appeared at 3281 cm$^{-1}$ corresponds to the Amide A band (N–H stretching vibration). Also, the peaks at 1397 and 2874 cm$^{-1}$ belong to the amide bands. According to these spectra, it is seen that the EW retained its functional groups after crosslinking (Balaji et al., 2019).

![Figure 2. FTIR spectra of the EW cryogel and native EW without crosslinking.](image)

**Table 1. The parameters of the EW cryogel.**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>$SR$ (g/g)</td>
<td>9.7±0.7</td>
</tr>
<tr>
<td>$V_p$ (mL/g)</td>
<td>2.7±0.2</td>
</tr>
<tr>
<td>$P%$</td>
<td>71±2</td>
</tr>
</tbody>
</table>

![Figure 3. SEM images of dry EW cryogel. Scaling bars = 2 mm (left) and 500 μm (right).](image)

**Adsorption of dye**

The effects of solution pH, initial dye concentration and contact time were studied in detail. The pH is one of the main factors affecting mechanism of adsorption or desorption of dyes (Hu et al., 2018). The experiments were initially performed at various pH values in the range of 3 and 9 to determine the optimum solution medium. In Figure 4, the adsorption capacity of MB on EW cryogel was pH dependent and reached high values above pH6. The amount of MB adsorption significantly increased from ~1 up to ~11 mg/g as the pH of solution was increased from 3 to 9. This behavior was attributed to the electrostatic attraction between EW and MB. The proteins found in EW comprise of many amino acids which have amino
and carbonyl functional groups. These active sites of the bioadsorbent can be strongly cationic or anionic depending on the pH of the medium. EW mainly consists of ovalbumin, ovotransferrin, lysozyme and ovomucin proteins. Ovalbumin (isoelectric point $\sim$4.5) content is about 54% of total protein of EW, so that it dominates the most of the functional groups in the biosorbent (Croguennec et al., 2000; Pereira et al., 2016). On the other hand, MB is heterocyclic aromatic chemical compound with a positive charge. Since the amino groups of EW are protonated at low pH of the solution, the repulsion occurred between the cryogel and cationic dye MB. The cryogel offered almost no adsorption towards to MB at pH = 3 and 4, indicating that the electrostatic attraction was the main adsorption mechanism. When the pH of solution was above isoelectric point, the positive charged MB was interacted with negative charged EW due to the dissociation of carbonyl groups and as a result, the adsorption capacity enhanced.

The adsorption performance of the cryogel was studied at dye concentration range from 10 to 180 mg/L at pH9 for 24h contact time (Salazar-Rabago et al., 2017). The adsorption amount rose sharply until initial dye concentration increased to 100 mg/L and then reached a plateau value of $\sim$52±3 mg/g (Figure 5). It is explained by the fact that the adsorption capacity remained stable at high initial dye concentrations due to almost occupation of all active sites of the adsorbents, while the adsorption capacity was low due to the excess of adsorbent dose at low initial dye concentrations (Uddin et al., 2009).

### Adsorption Kinetics

The MB adsorption on the cryogel as a function of contact time was also studied to evaluate the kinetics and gain insight into the underlying mechanisms. Figure 6 shows the adsorption performance depending on the time with a dye concentration of 15mg/L at pH9. Initially, the adsorption rate was high due to rich active sites for dye binding and decreased as time progressed. The adsorption almost reached to the equilibrium after $\sim$24h.

The adsorption rate, the model of process and interaction between MB and EW cryogel were analyzed by fitting the experimental data to pseudo-first order and pseudo-second order kinetic models using equation 7 and 8, respectively. (Gerente et al., 2007).
Pseudo-first order model (Lagergren, 1898):

\[
\log(q_e - q_t) = \log q_e - \frac{k_1}{2.303} t
\]  

(Eq. 7)

Pseudo-second order model (Ho, 2006):

\[
\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{t}{q_e}
\]  

(Eq. 8)

where \(k_1\) and \(k_2\) are the rate constants of models. Adsorption kinetic models provide insight about whether the adsorption process is physical or chemical. The slope lines of kinetic models for MB adsorption on the EW cryogel were drawn in Figure 7 and the corresponding kinetic parameters were collected in Table 2. The linear regression correlation coefficient (\(R^2=0.99\)) value from the pseudo-second order model was close to unity and higher than that (\(R^2=0.89\)) from the pseudo-first order model. Moreover, the experimental adsorption capacity \((q_{e,exp}=10.40 \text{ mg/g})\) was similar to the calculated equilibrium adsorption capacity \((q_{e,cal}=10.88 \text{ mg/g})\) of the pseudo-second order in contrast to the pseudo-first order \((q_{e,cal}=5.93 \text{ mg/g})\), indicating the adsorption process matched the pseudo-second order model and chemical adsorption dominated the process (Kuang et al., 2020). This behavior was consistent with many studies of MB adsorption on cryogel sorbents (Dragan & Loghin, 2013).

The intra-particle diffusion model was also applied on kinetic data by using Weber–Morris equation given below (Weber & Morris, 1963). According to this model, the adsorption process is controlled by only intra-particle diffusion if the plot of \(q_t\) against \(t^{0.5}\) gives a single straight line through the origin. Otherwise it is controlled by more than one step.

\[
q_t = k_{id} t^{0.5} + C
\]  

(Eq. 9)

where \(k_{id}\) (mg/(g min^{0.5})) is rate constant and \(C\) (mg/g) is the constant related to boundary layer thickness. As can be seen in Figure 8, nonzero values (2.37 and 8.78) of constant \(C\) and the multilinearity with two straight lines indicate that the adsorption mechanism consisted of multistep process (Doğan et al., 2009). This can be explained by that the diffusion of MB to the surface of EW cryogel and into the interior part through macropores is the first step, and reaching saturation with MB in active sites is the second step.

Table 2. Calculated parameters from kinetic models for MB adsorption onto EW cryogel.

<table>
<thead>
<tr>
<th>Pseudo-first order model</th>
<th>Pseudo-second order model</th>
</tr>
</thead>
<tbody>
<tr>
<td>(C_o) (mg/L)</td>
<td>(q_e) (mg/g)</td>
</tr>
<tr>
<td>15</td>
<td>5.93</td>
</tr>
</tbody>
</table>

Figure 7. (Top) Kinetic and (bottom) isotherm models of the MB adsorption on the EW cryogel.
Figure 8. Intra-particle diffusion model of the MB adsorption on the EW cryogel.

**Adsorption Isotherms**

The experimental data obtained from the adsorption at various initial dye concentrations in the range from 10 to 180 mg/L (pH 9, ~10 mg sorbent and 24h contact time) were analyzed using most commonly known Langmuir and Freundlich isotherm models (Foo & Hameed, 2010; Freundlich, 1907; Langmuir, 1918). The equations of these isotherm models are given below in equations 10 and 11.

Langmuir isotherm:

\[
\frac{C_e}{q_e} = \frac{1}{K_L q_m} + \frac{C_e}{q_m} \quad \text{(Eq.10)}
\]

Freundlich isotherm:

\[
\ln q_e = \log K_F + \frac{1}{n} \ln C_e \quad \text{(Eq.11)}
\]

where \(q_m\) (mg/g) is the maximum adsorption capacity, \(K_L\) (L/mg), \(K_F\) (mg/g)/(mg/L)\(^{1/n}\) and \(n\) are constants.

Table 3. Calculated parameters from isotherm models for MB adsorption on the EW cryogel.

<table>
<thead>
<tr>
<th>Cycle (C_o) (mg/L)</th>
<th>(q_m) (mg/g)</th>
<th>(K_L) (L/mg)</th>
<th>(R^2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>15</td>
<td>56.18</td>
<td>0.064</td>
<td>0.99</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Cycle (C_o) (mg/L)</th>
<th>(n)</th>
<th>(K_F) (mg/g)/(mg/L)(^{1/n})</th>
<th>(R^2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>15</td>
<td>2.41</td>
<td>7.06</td>
<td>0.89</td>
</tr>
</tbody>
</table>

The linear regression lines of the models for MB adsorption onto the EW cryogel were drawn in Figure 7 and the corresponding parameters were collected in Table 3. The results indicated that the Langmuir model with correlation coefficient of \(R^2\sim0.99\) gave a better description of adsorption than the Freundlich model whose correlation coefficient value (\(R^2\sim0.89\)) was not close to unit. The maximum adsorption capacity \(q_m\) of the cryogel found from Langmuir model was 56.18 mg/g, which was close to equilibrium adsorption capacity at plateau value of \(\sim52\pm3\) mg/g in Figure 5. It can be assumed that the adsorption occurred on a homogeneous surface and in a monolayer rather than a reversible heterogeneous surface due to \(R^2_{\text{Langmuir}} > R^2_{\text{Freundlich}}\) (Eq.12).

The separation factor \(R_L\) is also used to depict the feasibility of the adsorption (Hall et al., 1966). It is calculated from \(K_L\) using below equation,

\[
R_L = \frac{1}{1+K_L C_0} \quad \text{(Eq. 13)}
\]

The type of the isotherm can be irreversible \((R_L = 0)\), favorable \((0 < R_L < 1)\), linear \((R_L = 1)\), or unfavorable \((R_L > 1)\) according to the value of \(R_L\). The calculated values of \(R_L\) between 0.05 and 0.61 are plotted against the initial dye concentration in Figure 9, indicating the adsorption of MB on the EW cryogel was favorable for all studied dye concentrations.

**Desorption and reusability**

Figure 9. The separation factor \(R_L\) values of MB adsorption on the EW cryogel.

![Figure 10](image)

Figure 10. (Top) Digital photos of the EW cryogel before and after adsorption, and desorption of MB, respectively. (Bottom) the reusability of the EW cryogel \((C_o=15\text{ mg/L}, \text{pH}=9\text{ and } t=24\text{ h})\).
The reusability of the EW cryogel was conducted with four adsorption/desorption cycles. In the regeneration process, the dye adsorbed cryogel sample was firstly treated with 20 mL of 1M HCl solution for 2h (Figure 10). Then, the cryogel was separated and rinsed several times with water until neutral. The desorbed cryogel was used again for next adsorption cycle. The adsorption capacity of the cryogel remained almost the same and $R_\%$ decreased only by ~5 without losing its mechanical stability after repeating cycles (Figure 10). These results show that almost complete desorption and reusability make the EW cryogel an economical and practical adsorbent in the wastewater treatment.

**Conclusion**

The study presented that the easily prepared EW cryogel can be efficiently used as sorbent for the removal MB from water. The adsorption amount of dye changed depending on the solution pH, initial dye concentration and contact time. The equilibrium time was about 24h and the removal efficiency was high at slightly basic pH. The pseudo-second order kinetic model fitted ($R^2 > 0.99$) with adsorption dynamics. Intraparticle diffusion model showed that there was more than one diffusion mode in the adsorption. According to the correlation coefficient ($R^2$) values, the Langmuir isotherm model was more applicable with the adsorption data than that of the Freundlich one, confirming the monolayer sorption. The maximum MB adsorption capacity of the EW cryogel was 56.18 mg/g. Furthermore, the prepared EW cryogel may be used as a potential biosorbent for removal of other contaminants from aqueous solution.

**Acknowledgements**

The author is grateful to Sevgi GULYUZ from TUBITAK MAM for FTIR and SEM measurements.

**References**


