

Original article (Orijinal araştırma)

Current occurrence and prevalence of root-knot nematodes species, *Meloidogyne* spp. Goeldi, 1892 (Tylenchida: Meloidogynidae) in ware potato fields of Turkey

Yemeklik patates alanlarında kök-ur nematodu türlerinin, *Meloidogyne* spp. Goeldi, 1892 (Tylenchida: Meloidogynidae) mevcut durumu ve yaygınlığı

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Abstract

The study was conducted to determine the occurrence, frequency and density of species of *Meloidogyne* Goeldi, 1892 (Tylenchida: Meloidogynidae) in ware potato cultivation areas of Turkey in Plant Protection Central Research Institute. Soil samples were taken from Afyonkarahisar (149), Aksaray (69), Bolu (94), Kayseri (94), Konya (127), Nevşehir (91), Niğde (226) and Sivas (77) Provinces in 2018-2019. *Meloidogyne* juveniles were extracted by modified Baermann funnel and counted under inverted microscope. *Meloidogyne* populations were identified by species-specific primers. *Meloidogyne* spp. were detected in 84 of 927 soil samples. *Meloidogyne* spp. was detected in Aksaray, Kayseri, Nevşehir, and Niğde while was not found Afyonkarahisar, Bolu, Konya, and Sivas Provinces. In survey areas, the occurrence of *Meloidogyne chitwoodi* Golden, O'Bannon, Santo & Finley, 1980 and *Meloidogyne hapla* (Chitwood, 1949) (Tylenchida: Meloidogynidae) was 8.7 and 1.5%, respectively. Also, *M. chitwoodi* and *M. hapla* mixed populations were found in 1.2% of samples. Mean density of *Meloidogyne* spp. J2s was determined as 182, 175, 162 and 90 J2s/100 ml of soil in Niğde, Nevşehir, Aksaray and Kayseri Provinces, respectively. In conclusion, important ware potato cultivation areas of Turkey were found to be infested with *Meloidogyne* spp., but the important seed potato-growing areas were found to be free of *Meloidogyne* spp.

Keywords: *Meloidogyne chitwoodi*, *Meloidogyne hapla*, seed potato, survey, ware potato

Öz

Bu çalışma Türkiye'deki yemeklik patates ekiliş alanlarında *Meloidogyne* Goeldi, 1892 (Tylenchida: Meloidogynidae) türlerinin bulunuş, yaygınlık ve yoğunluklarının belirlenmesi amacıyla Ziraî Mücadele Merkez Araştırma Enstitüsü'nde yapılmıştır. Toprak örnekleri 2018-2019 yıllarında Afyonkarahisar (149), Aksaray (69), Bolu (94), Kayseri (94), Konya (127), Nevşehir (91), Niğde (226) ve Sivas (77) illerinden alınmıştır. *Meloidogyne* ikinci dönem larvaları modifiye Baermann huni tekniği ile elde edilmiş ve inverted mikroskopta sayılmıştır. *Meloidogyne* spp. popülasyonları türe spesifik primerler kullanılarak teşhis edilmiştir. Çalışma sonucunda 927 toprak örneğinin 84'ünün *Meloidogyne* spp. ile bulaşık olduğu belirlenmiştir. Aksaray, Kayseri, Nevşehir ve Niğde illerinde *Meloidogyne* spp. tespit edilirken Afyonkarahisar, Bolu, Konya ve Sivas illerinde bulunmamıştır. Sürvey alanında *Meloidogyne chitwoodi* Golden, O'Bannon, Santo & Finley, 1980 ve *Meloidogyne hapla* (Chitwood, 1949) (Tylenchida: Meloidogynidae) yaygınlığı sırasıyla %8.7 ve %1.5 olarak belirlenmiştir. Ayrıca *M. chitwoodi* ve *M. hapla* karışık popülasyonlarının yaygınlığı ise %1.2 olarak belirlenmiştir. *Meloidogyne* spp. J2 ortalama yoğunluğu Niğde, Nevşehir, Aksaray ve Kayseri illerinde sırasıyla 182, 175, 162 ve 90 J2/100 ml toprak/J2 olarak belirlenmiştir. Çalışma sonucunda, önemli yemeklik patates ekiliş alanlarının yoğun olarak *Meloidogyne* spp. ile bulaşık olduğu ancak Türkiye'deki önemli tohumluk patates ekiliş alanlarının ise *Meloidogyne* spp. açısından temiz olduğu sonucuna varılmıştır.

Anahtar sözcükler: *Meloidogyne chitwoodi*, *Meloidogyne hapla*, tohumluk patates, sürvey, yemeklik patates

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Received (Alınış): 16.03.2021

Accepted (Kabul edilmiş): 04.06.2021

Published Online (Çevrimiçi Yayın Tarihi): 15.06.2021

Introduction

Potato, *Solanum tuberosum* L. (Solanaceae) is the most important non-grain crop in the world and the fourth major food crop after maize, rice and wheat with an annual production of 370 Mt (FAOSTAT, 2019). Considered one of the most promising products in the fight against hunger and poverty, potatoes are highly recommended by FAO as a food security crop due to the growing population and food demand (Thomas & Sansonetti, 2009). Potato crops are damaged by numerous pests and pathogens, including nematodes (Niere & Karuri, 2018). Plant-parasitic nematodes are one of the crucial factors restricting potato yield and quality in many potato-growing areas (Lima et al., 2018). Potato cyst nematodes, *Globodera rostochiensis* (Wollenweber, 1923) Skarbilovich, 1959 and *Globodera pallida* (Stone, 1973) Behrens, 1975 (Tylenchida: Heteroderidae) and root-knot nematodes, *Meloidogyne* spp. Goeldi, 1892 (Tylenchida: Meloidogynidae) (RKN) are the major parasites of potato (Lima et al., 2018). Estimated annual yield losses in potato due to plant-parasitic nematodes is over 13% worth about US\$6 billion worldwide (Ravichandra, 2014). While cyst nematodes have been more troublesome in the past, there have been drastic rises in root-knot nematodes due to the ban of some effective nematicides and soil fumigants such as 1,3-D, cadusafos and ethoprophos and also lacking resistant cultivars (Wesemael et al., 2011).

Root-knot nematodes include the most economically damaging plant-parasitic nematodes to crops worldwide (Karssen & Moens, 2006). Although the genus *Meloidogyne* includes about 100 species, *Meloidogyne incognita* (Kofoid & White, 1919) Chitwood, 1949, *Meloidogyne javanica* (Treub, 1885) Chitwood, 1949, *Meloidogyne arenaria* (Neal, 1889) Chitwood, 1949, *Meloidogyne chitwoodi* Golden, O'Bannon, Santo & Finley, 1980, *Meloidogyne fallax* Karssen, 1996 and *Meloidogyne hapla* Chitwood, 1949 (Tylenchida: Meloidogynidae) are the most common species. These species account for over 95% of the *Meloidogyne* occurrences in the world (Adam et al., 2007; Jones et al., 2013). The impact of these species is increasing due to their wide host ranges and it is estimated that they can affect more than 5,500 plant species (Trudgill & Blok, 2001) with crop losses of about 5% worldwide (Agrios, 2005).

Potato is one of the main hosts of *Meloidogyne* spp., and the prevalence and damage of these species have been increasing in recent decades. The damage of RKN to potatoes has been determined in all continents except Antarctica, including the major potato producing countries including China (Mao et al., 2019), India (Singh & Kumar, 2015), the USA (Golden et al., 1980), Germany (Müller et al., 1996), France (Djian-Caporalino, 2012), Holland (Keidel et al., 2007), South Africa (Fourie et al., 2001), and Australia (Nobbs et al., 2001). In potato production, *M. arenaria*, *Meloidogyne enterolobii* Yang & Eisenback, 1983, *M. incognita* and *M. javanica* and are present in warmer climates, while *M. chitwoodi*, *M. fallax*, *M. hapla* and *Meloidogyne minor* Karssen et al., 2004 are encountered in relatively colder climates (Nyczepir et al., 1982; Nobbs et al., 2001; Thoden et al., 2012; Onkendi & Moleleki, 2013; Medina et al., 2017). Of these, *M. arenaria*, *M. chitwoodi*, *M. fallax*, *M. hapla*, *M. incognita* and *M. javanica* are significant for potato production (Lima et al., 2018). In Turkey, *M. chitwoodi*, *M. hapla*, *M. incognita*, and *M. javanica* have been determined in potato-growing areas, however *M. chitwoodi* is the most common one among them (Kepenekci et al., 2006; Özarslandan & Elekcioğlu, 2010; Erdoğuş et al., 2011; Evlice & Bayram, 2016; Demirbaş Pehlivan et al., 2020).

There are no reports of specific aboveground symptoms in potato associated with root-knot nematode infestation, however varying degrees of stunting, yellowing, and wilting under moisture stress can be seen in some of the infested plants. Root-knot nematodes reproduce on potato roots and tubers, but the size and shape of galls or knots could vary depending on the species and density of nematodes (Niere & Karuri, 2018). The economic threshold on potato cultivation in the USA and the Netherlands are 1 and 10 J2s/100 ml soil for *M. chitwoodi*, respectively (Brodie et al., 1993; Norshie et al., 2011) and 50 eggs/250 ml soil for *M. hapla* (Brodie et al., 1993).

Galls of temperate species, *M. chitwoodi*, *M. fallax*, *M. hapla* and *M. minor*, are usually smaller than those caused by other species and cause extensive lateral root formation. However, these species cause numerous small pimple-like swelling on the tuber surface (Elling, 2013). Even 5% necrotic spotting in the flesh of tubers make them inappropriate for the fresh market, and if 5-15% of the tubers have defects when harvested, the whole field crop can be discarded (King & Taberna, 2013).

For management, in most countries, at least a 2-year rotation is used in potato production. There are no commercially available potato cultivars resistant to root-knot nematodes. Under such conditions, *Meloidogyne* spp. populations can increase rapidly. For this reason, the most important and widely used management strategy is to apply nematicides.

This study has been conducted to determine (1) occurrence of the root-knot nematodes and species, (2) frequency and density of root-knot nematode species in ware potato-growing areas in Turkey.

Materials and Methods

Sampling strategy and sample processing

Surveys were conducted in 8 potato-growing provinces (Afyonkarahisar, Aksaray, Bolu, Kayseri, Konya, Nevşehir, Niğde and Sivas) between 2018 and 2019. A total of 927 soil samples were taken from fields immediately after potato harvest, 0.1% of the annual ware potato production area in each province (Figure 1). Soil sample of 2-3 kg consisting of 50 subsamples according to a grid pattern from 0-30 cm depth was randomly obtained from each field using an auger. The soil samples and symptomatic potato tubers (Figure 2) were placed in a plastic container. For each sampled area, GPS coordinates of sampling sites were recorded.

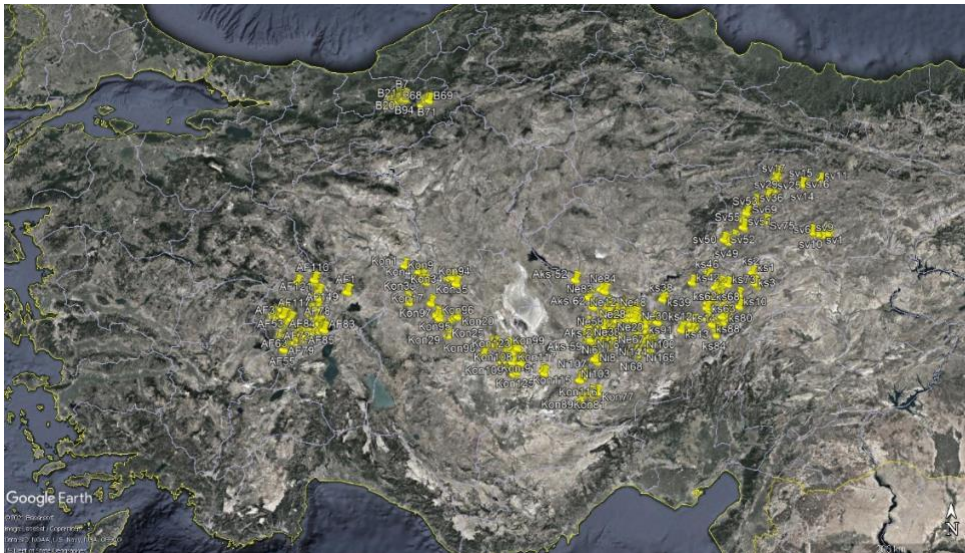


Figure 1. Soil samples taken from potato-growing areas in this study (mapped using www.earth.google.com).

Soil samples were mixed thoroughly and 200 ml subsamples were processed for nematode extraction for each field. Root-knot nematodes were extracted by the modified Baermann funnel technique (Hooper, 1986). After 24 h, nematode suspension was transferred to 100 ml graduated cylinder and collected in a 50 ml sample bottle for examination. The whole suspension was checked for *Meloidogyne* spp. and counted under an invert microscope (Leica DMI 400B).

DNA extraction

DNA were extracted from second-stage juveniles (J2s) for each isolate according to Waeyenberge et al. (2000). For this purpose, 10 *Meloidogyne* spp. J2s was picked from extracted soil samples using a small needle and cut into two pieces with a sterile scalpel under a stereomicroscope (Leica M165C) in 20 μ l of molecular grade water on a glass microscope slide, then 10 μ l with the nematode pieces was transferred by pipette into a 0.2 ml centrifuge tube containing 8 μ l of worm lysis buffer (WLB; 500 mM KCl, 100 mM Tris-Cl pH 8.3, 15 mM MgCl₂, 10 mM DTT, 4.5% Tween 20, 0.1% gelatin). After adding 2 μ l of proteinase K (600 g/ml; Thermo Fisher Scientific, Waltham, MS, USA) the suspension was centrifuged at 13,500 rpm for 2 min and frozen to -80°C for 10 min. After freezing, the contents were thawed rapidly. The lysis mix was incubated at 65°C for 1 h followed by 95°C for 10 min for a proteinase K inactivation. After incubation, the tubes were centrifuged for 1 min at 14,000 rpm and stored at -20°C until used.



Figure 2. Potato tuber infested with *Meloidogyne chitwoodi*, pimple-like galls and necrotic spots on their surface.

DNA Amplification protocols

The primers used in PCR amplification for the identification of root-knot nematodes are given in Table 1. All multiplex PCR amplifications were conducted in a total volume of 25 μ l consisting of 10x PCR Buffer, 200 μ M dNTPs 0.2 μ M of each primer, 1 unit Taq DNA polymerase (Thermo Fisher Scientific) and 2 μ l template DNA. All DNA amplifications were conducted with a thermocycler (Techne, TC-5000).

Table 1. Primers used in the multiplex PCR to identify the species of *Meloidogyne* spp.

Species	Fragments	Primer	Primer sequences (5'-3')	Reference
<i>Meloidogyne chitwoodi</i>	540 bp	JMV1	GGATGGCGTGCTTTCAAC	
<i>Meloidogyne fallax</i>	670 bp	JMV2	TTTCCCCTTATGATGTTTACCC	Wishart et al., 2002
<i>Meloidogyne hapla</i>	440 bp	JMVhapla	AAAAATCCCCTCGAAAAATCCACC	
<i>Meloidogyne incognita</i>	150 bp	MincF1 MincR1	GCACCTCTTTCATAGCCACG GGTGCGCGATTGAACTGAGC	Devran et al., 2018
<i>Meloidogyne javanica</i>	670 bp	Fjav Rjav	GGTGCGCGATTGAACTGAGC CAGGCCCTTCAGTGGAACTATAC	Zijlstra et al., 2000
<i>Meloidogyne arenaria</i>	420 bp	Far Rar	TCGGCGATAGAGGTTAAATGAC TCGGCGATAGACACTACAACT	Zijlstra et al., 2000

The cycling parameters were as follows for *M. chitwoodi*, *M. fallax*, and *M. hapla*: an initial denaturation step at 94°C for 3 min followed by 35 cycles at 94°C for 30 s, 55°C for 30 s, then 72°C for 90 s and final extension step at 72°C for 10 min; and for *M. incognita*, *M. javanica*, *M. arenaria*: initial denaturing step for 3 min at 95°C followed by 35 cycles at 95°C for 30 s, 56°C for 30 s, then 72°C for 60 s and a final extension step at 72°C for 7 min. PCR products were separated by electrophoresis in Tris-EDTA buffer with 1.7% agarose gel stained with Pronosafe (Condalab, Spain) at 80 V for 1 h and then visualized with an image acquisition system (Vilber QUANTUM ST4 1000, Collégien, France).

Meloidogyne spp. community analyses

The incidence, occurrence, and frequency of root-knot nematodes determined in the study at the genus and species level in potato-growing areas were calculated according to the formulas below (Norton, 1978).

$$\text{Occurrence of genus or species} = \frac{\text{Number of sample with root - knot nematodes infection}}{\text{Total number of sample surveyed}} \times 100$$

$$\text{Absolute frequency} = \frac{\text{Number of sample containing species}}{\text{Number of sample collected}} \times 100$$

$$\text{Relative frequency} = \frac{\text{Frequency of occurrence of species}}{\text{Sum of frequency of all Meloidogyne spp.}} \times 100$$

Results

Species identification

Multiplex PCR analysis by using JMV1/JMV2/JMVhapla primers produced a DNA fragment size of 540 bp specific to *M. chitwoodi* for 70 populations, 440 bp specific to *M. hapla* for three populations, and also both 540 bp and 440 bp for 11 populations. However, there was no 670-bp amplification product which was characteristic for the *M. fallax*. In addition, primer sets Far/Rar, MincF1/MincR1 and Fjav/Rjav, specific for *M. arenaria*, *M. incognita* and *M. javanica*, respectively, did not yield any amplification products. These results showed that 70 populations were *M. chitwoodi*, three populations were *M. hapla*, and 11 populations were mixed *M. chitwoodi* and *M. hapla*. There was no evidence of the existence of *M. arenaria*, *M. fallax*, *M. incognita* or *M. javanica* as previously reported in potato areas in Central Anatolia (Table 2).

Meloidogyne spp. community analyses

J2s of *Meloidogyne* spp. were detected in 84 of 927 soil samples in all surveyed areas. *Meloidogyne* spp. was detected in Aksaray, Kayseri, Nevşehir and Niğde Provinces, but not in Afyonkarahisar, Bolu, Konya and Sivas Provinces (Tables 2 & 3). The occurrence, frequency and population density of *Meloidogyne* genus and species differed according to the location sampled. The occurrence of RKNs in all surveyed areas was 9.1% with a mean density of 176 J2s/100 ml of soil. *Meloidogyne chitwoodi* was the most abundant species in all provinces infested with *Meloidogyne* spp. The occurrence of *M. chitwoodi* and *M. hapla* in the field surveys were 8.7 and 1.5%, respectively, and *M. chitwoodi* and *M. hapla* mixed populations were 1.2%. *Meloidogyne chitwoodi* was the most prevalent species in all surveyed areas, with 96.4 and 85.3% absolute and relative frequency, respectively. This was followed by *M. hapla* with 16.7 and 14.7% absolute and relative frequency, respectively.

The highest occurrence of RKNs was found in Nevşehir with 25.3%, followed by Niğde, Aksaray, and Kayseri with 23.5, 8.7, and 2.1%, respectively. The occurrence and absolute frequencies were 25.3, 22.1%, 8.7 and 2.1%, and 100, 94.3, 100 and 100% in Nevşehir, Niğde, Aksaray, and Kayseri Provinces, respectively. *Meloidogyne hapla* was only found as the sole species in Niğde, but it was found in mixed populations with *M. chitwoodi* in Aksaray and Nevşehir Provinces. Additionally, *M. hapla* was not detected in any samples from Kayseri Province. Its occurrence and absolute frequencies were found to be 4.4, 3.5

and 2.9%, and 17.4, 15.1 and 33.3% in Nevşehir, Niğde and Aksaray Provinces, respectively. Mean density of *Meloidogyne* spp. J2s were similar in Nevşehir (175 J2s/100 ml of soil), Niğde (182 J2s/100 ml of soil) and Aksaray (162 J2s/100 ml of soil) Provinces, but lower in Kayseri (90 J2s/100 ml of soil).

Table 2. Occurrence of *Meloidogyne* spp. on potatoes collected from ware potato fields in Turkey

Province	District	Village	Altitude (m)	Coordinate	Species	J2s/100 ml soil	
Aksaray	Gülağaç	-	1192	38°21'8.15"N 34°20'49.16"E	<i>M. chitwoodi</i>	44	
		Demirci	1190	38°20'42.49"N 34°18'40.94"E	<i>M. chitwoodi</i>	108	
	Güzelyurt	Alanyurt	1204	38°20'1.76"N 34°19'7.08"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	237	
			1212	38°20'40.34"N 34°20'16.93"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	194	
		Bozcayurt	1200	38°21'30.02"N 34°22'23.10"E	<i>M. chitwoodi</i>	83	
Merkez	Yenipınar	1412	38°12'37.95"N 34°16'50.61"E	<i>M. chitwoodi</i>	305		
Kayseri	İncesu	Örenşehir	1046	38°41'42.26"N 35°14'46.92"E	<i>M. chitwoodi</i>	83	
	Melikgazi	Yeşilyurt	1086	38°48'43.61"N 35°36'49.44"E	<i>M. chitwoodi</i>	97	
Nevşehir	Acıgöl	Kurugöl	1288	38°22'32.75"N 34°32'27.47"E	<i>M. chitwoodi</i>	124	
		Çakıllı	1342	38°24'28.98"N 34°39'13.37"E	<i>M. chitwoodi</i>	149	
		Doğala	1387	38°25'37.96"N 34°36'26.69"E	<i>M. chitwoodi</i>	53	
		Kuyulutatlar	1305	38°22'2.92"N 34°32'4.62"E	<i>M. chitwoodi</i>	196	
			1292	38°23'20.81"N 34°31'48.68"E	<i>M. chitwoodi</i>	79	
		Til	1500	38°25'29.67"N 34°47'23.46"E	<i>M. chitwoodi</i>	148	
		Yazıhüyük	1287	38°21'22.85"N 34°36'42.45"E	<i>M. chitwoodi</i>	207	
	Derinkuyu	-	1389	38°20'50.37"N 34°46'51.10"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	417	
		-	1320	38°20'37.51"N 34°44'35.28"E	<i>M. chitwoodi</i>	202	
		-	1336	38°21'23.18"N 34°44'23.79"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	581	
-		1363	38°23'6.14"N 34°44'52.66"E	<i>M. chitwoodi</i>	299		
-		1323	38°20'34.63"N 34°44'36.57"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	45		
-		1336	38°21'23.28"N 34°44'23.78"E	<i>M. chitwoodi</i>	217		
-		1361	38°22'51.59"N 34°43'22.27"E	<i>M. chitwoodi</i>	102		
-	1449	38°27'26.94"N 34°42'11.31"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	52			
-	1392	38°21'26.63"N 34°45'39.54"D	<i>M. chitwoodi</i>	74			
Merkez	Çardak	1373	38°32'43.71"N 34°46'8.02"E	<i>M. chitwoodi</i>	48		
		1409	38°32'0.91"N 34°45'33.11"E	<i>M. chitwoodi</i>	294		
	İcik	1384	38°29'56.68"N 34°35'42.48"E	<i>M. chitwoodi</i>	61		
		1363	38°30'51.49"N 34°35'55.92"E	<i>M. chitwoodi</i>	121		
		Kaymaklı	1423	38°28'56.74"N 34°45'10.41"E	<i>M. chitwoodi</i>	56	
1406	38°27'13.72"N 34°44'38.68"E	<i>M. chitwoodi</i>	81				
Ürgüp	Bahçeli	1474	38°31'0.78"N 34°47'23.01"E	<i>M. chitwoodi</i>	427		
Niğde	Altınhisar	-	1212	38°0'30.00"N 34°19'39.00"E	<i>M. chitwoodi</i>	95	
			1572	38°12'39.63"N 34°31'2.85"E	<i>M. hapla</i>	61	
		Azatlı	1582	38°13'0.93"N 34°31'22.52"E	<i>M. chitwoodi</i>	349	
			1541	38°12'9.00"N 34°28'35.00"E	<i>M. chitwoodi</i>	57	
			1580	38°11'51.02"N 34°29'9.67"D	<i>M. chitwoodi</i>	134	
			1575	38°13'10.00"N 34°29'10.00"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	167	
		Çiftlik	Bozköy	1591	38°13'43.00"N 34°29'10.00"E	<i>M. chitwoodi</i>	69
				1569	38°13'4.00"N 34°29'18.00"E	<i>M. chitwoodi</i>	60
				1546	38°12'15.00"N 34°28'51.00"E	<i>M. chitwoodi</i>	45
		Divarlı	1542	38°12'49.93"N 34°27'55.76"E	<i>M. hapla</i>	301	
			1643	38°13'45.00"N 34°28'16.00"E	<i>M. chitwoodi</i>	59	
			Kitreli	1422	38°11'48.28"N 34°21'31.36"E	<i>M. chitwoodi</i>	74

Table 2 Continued

Province	District	Village	Altitude (m)	Coordinate	Species	J2s/100 ml soil
Niğde	Merkez	Ağcaşar	1320	38°18'35.03"K 34°45'15.70"D	<i>M. chitwoodi</i> , <i>M. hapla</i>	1159
		Alay	1302	38°17'5.84"N 34°41'32.33"E	<i>M. chitwoodi</i>	34
			1311	38°16'14.72"N 34°40'44.60"E	<i>M. chitwoodi</i>	56
		Bağlama	1308	38°16'2.60"N 34°40'35.93"E	<i>M. chitwoodi</i>	67
			1330	38°16'29.16"N 34°39'25.84"E	<i>M. chitwoodi</i>	295
			1325	38°16'16.63"N 34°39'32.55"E	<i>M. chitwoodi</i>	86
			1318	38°15'23.97"N 34°39'40.18"E	<i>M. chitwoodi</i>	45
			1315	38°16'8.33"N 34°40'25.80"E	<i>M. chitwoodi</i>	413
		Edikli	1368	38°14'7.20"N 34°55'10.86"E	<i>M. chitwoodi</i>	102
			1363	38°14'2.05"N 34°55'10.63"E	<i>M. chitwoodi</i>	147
			1428	38°17'0.66"N 34°53'35.87"E	<i>M. chitwoodi</i>	309
			1429	38°17'43.11"N 34°53'18.59"E	<i>M. chitwoodi</i>	416
		Kiledere	1306	38°16'36.40"N 34°40'45.45"E	<i>M. chitwoodi</i>	74
			1297	38°17'11.49"N 34°41'13.01"E	<i>M. chitwoodi</i>	56
			1300	38°17'34.37"N 34°41'11.16"E	<i>M. chitwoodi</i>	85
		Konaklı	1343	38°7'31.52"N 34°49'52.42"E	<i>M. chitwoodi</i>	167
			1339	38°8'40.37"N 34°49'41.42"E	<i>M. chitwoodi</i>	191
			1335	38°10'48.69"N 34°50'5.69"E	<i>M. chitwoodi</i>	368
			1343	38°11'37.52"N 34°49'23.86"E	<i>M. chitwoodi</i>	152
			1366	38°13'33.99"N 34°51'5.63"E	<i>M. chitwoodi</i>	202
			1426	38°16'39.74"N 34°52'28.59"E	<i>M. chitwoodi</i>	64
			1452	38°19'29.20"N 34°50'21.20"E	<i>M. chitwoodi</i>	445
		Orhanlı	1426	38°17'16.40"N 34°52'58.64"E	<i>M. chitwoodi</i>	83
			1425	38°16'16.69"N 34°52'27.42"E	<i>M. chitwoodi</i>	367
			1452	38°18'27.21"N 34°52'48.99"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	281
			1459	38°18'56.21"N 34°51'54.59"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	384
			1474	38°19'38.19"N 34°56'26.21"E	<i>M. chitwoodi</i>	194
			1465	38°19'1.57"N 34°55'27.45"E	<i>M. chitwoodi</i>	226
			1529	38°20'43.58"K 34°48'48.41"D	<i>M. chitwoodi</i>	416
		Ovacık	1332	38°4'35.66"N 34°48'1.81"E	<i>M. chitwoodi</i>	281
			1383	38°8'33.71"K 34°46'39.37"D	<i>M. chitwoodi</i>	67
		Tırhan	1304	38°14'35.99"N 34°42'25.47"E	<i>M. chitwoodi</i>	81
			1296	38°15'0.00"N 34°43'50.54"E	<i>M. hapla</i>	194
			1293	38°15'9.37"N 34°43'28.00"E	<i>M. chitwoodi</i>	84
			1296	38°14'41.51"N 34°42'54.20"E	<i>M. chitwoodi</i> , <i>M. hapla</i>	106
			1297	38°14'17.73"N 34°43'4.81"E	<i>M. chitwoodi</i>	73
		Yeşilgölcük	1307	38°16'27.08"N 34°45'37.98"E	<i>M. chitwoodi</i>	124
			1308	38°15'56.28"N 34°45'41.84"E	<i>M. chitwoodi</i>	64
			1308	38°15'17.14"N 34°46'0.56"E	<i>M. chitwoodi</i>	81
			1304	38°14'57.90"N 34°45'51.97"E	<i>M. chitwoodi</i>	50
			1305	38°13'59.33"N 34°46'13.14"E	<i>M. chitwoodi</i>	63

Table 3. Community analyses of *Meloidogyne* spp. on potato in the survey areas

	Survey area	Afyonkarahisar	Aksaray	Bolu	Kayseri	Konya	Nevşehir	Niğde	Sivas
Number of samples	928	149	69	94	94	127	91	226	77
<i>Meloidogyne</i> spp. infestation	84	0	6	0	2	0	23	53	0
<i>M. chitwoodi</i> infestation	81	0	6	0	2	0	23	50	0
<i>M. hapla</i> infestation	14	0	2	0	0	0	4	8	0
<i>M. chitwoodi</i> + <i>M. hapla</i> infestation	11	0	2	0	0	0	4	5	0
Occurrence of <i>Meloidogyne</i> spp.	9.1%	-	8.7%	-	2.1%	-	25.3%	23.5%	-
Occurrence of <i>M. chitwoodi</i>	8.7%	-	8.7%	-	2.1%	-	25.3%	22.1%	-
Occurrence of <i>M. hapla</i>	1.5%	-	2.9%	-	-	-	4.4%	3.5%	-
Occurrence of <i>M. chitwoodi</i> + <i>M. hapla</i>	1.2%	-	-	-	-	-	4.4%	2.2%	-
Absolute frequency of <i>M. chitwoodi</i>	96.4%	-	100%	-	100%	-	100%	94.3%	-
Absolute frequency of <i>M. hapla</i>	16.7%	-	33.3%	-	-	-	17.4%	15.1%	-
Relative frequency of <i>M. chitwoodi</i>	85.3%	-	75%	-	100%	-	85.2%	86.2%	-
Relative frequency of <i>M. hapla</i>	14.7%	-	25%	-	-	-	14.8%	14.8%	-

Discussion

The survey area covered about 58% of the potato production and harvested area of Turkey (TUIK, 2019). The findings showed that a total of 927 soil samples were obtained from potato-growing areas in eight provinces of Turkey, of which 9.1% were infested with *Meloidogyne* spp. *Meloidogyne chitwoodi* and *M. hapla* were detected at 8.7 and 1.5%, respectively, and *M. chitwoodi* and *M. hapla* were found in mixed populations in 1.2% of the samples. Nine root-knot species, *M. arenaria*, *Meloidogyne artiellia* Franklin, 1961, *M. chitwoodi*, *Meloidogyne ethiopica* Whitehead, 1968, *Meloidogyne exigua* Goeldi, 1887, *M. hapla*, *M. incognita*, *M. javanica* and *Meloidogyne thamesi* Chitwood, 1952 have been identified in different regions and different crops in Turkey (Yüksel, 1966; 1967; Elekcioğlu, 1992; Özarslandan et al., 2009; Aydınli et al., 2013; Kepenekci et al., 2014; İmren et al., 2014). However, four root-knot species, *M. chitwoodi*, *M. hapla*, *M. incognita* and *M. javanica*, have been found in association with potato in Turkey (Kepenekci et al, 2006; Özarslandan et al., 2009; Erdoğan et al., 2011; Demirbaş Pehlivan et al., 2020). The first report of root-knot nematode in potatoes appears to have been made by Kepenekci et al (2006) as *M. hapla* from potato tubers collected from Aksaray, Nevşehir and Niğde Provinces. Following that, *M. chitwoodi* was found in Niğde potato-growing areas by Özarslandan et al (2009). This finding has been supported by other studies in which only *M. chitwoodi* was detected and *M. chitwoodi* has since be found in Aksaray, Balıkesir, Bitlis, Isparta, İzmir, Kayseri, Konya, Kütahya, Manisa and Nevşehir Provinces (Özarslandan & Elekcioğlu, 2010; Ulutaş et al., 2012; Özarslandan et al., 2013; Evlice & Bayram, 2016). The present findings are similar to previous studies, and the relative frequency of *M. hapla* (14.7%) was quite low compared to *M. chitwoodi* (85.3%). *Meloidogyne incognita* was first identified in the potato-growing areas of Edirne, then in İzmir (Erdoğan et al., 2011; Demirbaş Pehlivan et al., 2020). Additionally, *M. javanica*, *M. hapla* and *M. chitwoodi* was determined in potato fields in İzmir (Yıldız et al., 2009; Demirbaş Pehlivan et al., 2020). The occurrence of *Meloidogyne* spp. in potato cultivation areas was higher in İzmir (18.4%) than found overall in the present survey (9.1%) but lower than Nevşehir (25.3%) and Niğde (23.5%). Differing from the present findings, the dominant species in the potato cultivation areas of İzmir Province was determined as *M. incognita* and the relative frequency of *M. incognita*, *M. javanica*, *M. hapla* and *M. chitwoodi* was found 61, 24.4, 12.2 and 2.4%, respectively (Demirbaş Pehlivan et al., 2020). This was an expected result due to the widespread cultivation of good hosts of root-knot nematodes in the Aegean Region and the prevalence of tropical root-

knot nematodes species in these areas (Kaşkavalcı & Öncüer, 1999; Yağcı & Kaşkavalcı, 2018). Similar results were obtained in other surveys conducted in warmer regions worldwide (Okendi & Moleleki, 2013; Medina et al., 2017). Temperate species such as *M. chitwoodi* and *M. hapla* are cold temperature nematodes whereas tropical species such as *M. incognita* and *M. javanica* prefer at higher temperatures and cannot tolerate cold temperatures (Evans & Perry, 2009). The base threshold temperature is 4°C and 8°C for *M. chitwoodi* and *M. hapla* while 10°C and 13°C for *M. incognita* and *M. javanica*, respectively (Insera et al., 1983; Lahtinen et al., 1988; Madulu & Trudgill, 1994; Ploeg & Maris, 1999).

The results presented here show that the mean density of *Meloidogyne* spp. ranged from 34 to 1159 J2s/100 ml of soil. *Meloidogyne chitwoodi* and *M. hapla* are the most important species within *Meloidogyne* for potatoes (Brodie et al., 1993). The economic threshold of *M. chitwoodi* in potatoes is reported to be 1 and 10 J2s/100 ml soil in the USA and Holland, respectively, whereas it is as 50 eggs/250 ml soil for *M. hapla* (Brodie et al., 1993; Norshie et al., 2011). Therefore, in the present study, the population density of RKN was above the economic damage threshold in all areas where it was determined. Crop loss from RKN infestation can be about 25%, depending on cultivars, environmental factors and population density, but may reach 100% in potato fields (Mai et al., 1981; Lima et al., 2018). The economic loss of cultivated potatoes caused by *M. chitwoodi* can reach US\$ 9 900/ha (Ingham et al., 2007). Potato roots and tubers can be invaded by root-knot nematodes. However, the first generation resides mostly on the root system, with subsequent generations entering the tubers (Pinkerton et al., 1991). Depending on the nematode density and species, infested roots and tubers may have galls of varying sizes and shapes. *Meloidogyne chitwoodi* and *M. hapla* produced galls that are normally smaller than those induced by other species (Niery & Karuri, 2018). Even 5% necrotic spotting in the flesh of tubers makes them commercially unacceptable for the fresh market. The entire crop can be rejected if 5 to 15% of the field tubers have visual defects when processed (King & Taberna, 2013). While potato genotypes resistant to *M. chitwoodi* have been identified, there is currently no commercially available resistant cultivars (Brown et al., 2006; Brown et al., 2009; Norshie et al., 2011). For this reason, the application of nematicides is the most commonly used control measure (Jones et al., 2017). The approved rates in the USA for some nematicides for control of *M. chitwoodi* are higher than for other *Meloidogyne* spp. (Lima et al., 2018). Crop rotation or succession with non- or poor hosts, shortening the growing period, planting certified seed and destruction of volunteer potato plants are some of the other control methods (Jones et al., 2017).

This study demonstrated that important ware potato cultivation areas, such as in Nevşehir and Niğde, where seed potato cultivation has been prohibited, are heavily infested with *M. chitwoodi*. However, the most important seed potato-growing areas in Turkey including Afyonkarahisar, Konya and Sivas were free from *Meloidogyne* spp. Therefore, quarantine measures should be applied meticulously to prevent infestations in these areas. Turkey's annual requirement is about 510 kt of seed potatoes and the use of seed potatoes is about 340 kt (BÜGEM, 2019; TÜİK, 2019). The difference of about 45% between production and use is due to the use of non-seed potatoes for planting. Although *Meloidogyne* spp. causes pimple-like swellings on the surface of potato tubers, these symptoms depend on the population density and the duration of vegetative growth. For this reason, asymptotically infested tubers are observed in most cases, and the use of these tubers as seeds must be avoided to prevent the spread of *Meloidogyne* spp. to uninfested areas.

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