



Traditional Medicinal Uses, Phytochemicals, and Pharmacological Activities of Genus *Rhamnus*: A review

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Abstract: The genus *Rhamnus* belongs to the Rhamnaceae family, which contains approximately 137 species, traditionally used as folk medicine in East Asia, North and South America, and subtropical regions of Africa. The genus is used traditionally to treat diseases such as cancer, wound, jaundice, hepatitis, gonorrhea, laxative, hypertension, malaria, stomach ache, snake bite and diarrhea. Anthraquinones and flavonoids are the most cited compounds from the genus of which polyphenols were abundant with tremendous antioxidant, wound healing and antiinflammatory activities. Pharmacological activity evaluation of the extracts and isolated compounds revealed anti-inflammatory, antioxidant, antimalarial, antibacterial, anti-mutagenic, anti-genotoxic, hepatoprotective, anticancer, and anti-proliferative activity. The genus afforded drug leads such as 6-methoxysorigenin (12) and prinoidin (23) with anti-tyrosinase and cytotoxicity, respectively, as well as antioxidant drug leads such as Kaempferol-3-O-β-rhamninoside (31) rhamnetin-3-O-β-isorhamninoside (37) and isotorachrysone (55). The present review endeavors to provide a comprehensive and up to date compilation of documented traditional medicinal uses, phytochemicals and pharmacological activities of the genus and provided valuable information in support of its uses as an alternative medicine for future healthcare practice.

Keywords: Rhamnus, anthraquinones, flavonoids, pharmacological activities.

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INTRODUCTION

Traditional medicine has been in existence even before the advent of modern medicine. It continues to remain as an alternative care available for the majority of the developing countries due to its intrinsic qualities, unique, and holistic approaches as well as its accessibility and affordability (1, 2). The genus *Rhamnus* consists of 137 species (Figure 1) and 19 synonyms (3). The word *Rhamnus* means 'a kind of prickly plant' and 'buckthorn or Christ's thorn' in Greek and Latin languages (4). The genus is distributed in East Asia, North and South America and various parts of

subtropical Africa with a wide spectrum of traditional medicinal uses (5-13).

The chemistry of *Rhamanus* species does not exhibit great diversity. The main groups of secondary metabolites reported from the genus are anthrones, anthraquinones, and flavonoids of which polyphenols were abundant with tremendous antioxidant, wound healing, and antiinflammatory activities. The present review endeavors to provide a comprehensive and up-to-date compilation of documented biological activities and phytochemistry of the genus and provided valuable information in support of its uses as an alternative medicine for future healthcare practice.

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Figure 1: Some medicinally important species of the genus Rhamnus L. (14, 15).

Taxonomy of the genus Rhamnus:

Kingdom: *Plantae*, **Order**: *Rosales*, **Family**: *Rhamnaceae*, **Genus**: *Rhamnus*. The names redberry, red berry buckthorn, California redberry, evergreen buckthorn, spiny buckthorn, and holly leaf buckthorn have been used for multiple taxa of *Rhamnus* (16).

Botanical Description and Traditional Uses

The genus *Rhamnus* comprises 137 species of shrubs and small trees in temperate, sub-tropical and tropical countries (17). It is an evergreen or deciduous plant and resistant to frost. The leaves are either alternate or sub-opposite. The hermaphrodite small flowers are weakly scented (18). Botanical description and traditional uses of various *Rhamnus* species is summarized in Table 1 below.

Table 1: Botanical distribution and traditional medicinal uses of the genus Rhamnus.

Scientific name	Part	Distribution	Traditional use	Refs
R. alaternus L.	Leaf, Aerial	Algeria	as a digestive, diuretic, laxative, and for the therapeutics of	(19)
	Part,	_	hepatic and dermatological disorders	
	Leaf	Algeria	treatment of gastrointestinal system diseases (hepatitis)	(20)
	Bark	Algeria	Used to treat jaundice	(21)
	Aerial part	Algeria	Hepatic jaundice and chlorosis	(22)
	Root, Aerial Part	Spain, Iberian	Used to treat depurative (blood purification)	(23, 24)
	·	Peninsula		
	Aerial Part	Spain	therapeutics of hypercholesterolemia	(23)
	Aerial Part	Spain	therapeutics of antihypertensive (lowers blood pressure)	(23)
	Bark, Branch	Italy	to treat hemostatic, wounds, laxative	(25)
	Branch, leaf	Iberian Peninsula	to treat high blood pressure	(24, 26)
	Branch, Leaf	Israel, Algeria	to treat jaundice	(5, 22)
R. alnifolia L'Hér	Root, Bark	USA	to treat gonorrhea and cathartic	(7)
R. alpina L.	Branch	Italy	to treat cardiac disease, wounds	(27)
R. cathartica L.	Bark	Bosnia and	treatments of common buckthorn, diarrhea, diuretic	(28, 29)
		Herzegovina, Turkey	, ,	(, ,
	Fruit	Southeast Europe	antiseptic for wounds	(30)
	Fruit	Serbia	to treat laxative	(31)
R. fallax L.	Bark	Bosnia and	to treat and manage dermal diseases	
		Herzegovina	_	
	Bark	Montenegro/ Serbia	to treat constipation	(34)
R. heterophylla Oliver	Root, Leaf	China	to cease bleeding	(11)
R. ilicifolia Kellogg	Root	USA	laxative, diuretic and to treat gonorrhea	(16)
	Whole Part	USA	analgesic or antirheumatic	(35)
R. lycioides L.	Leaves, Shoot	Turkey	to treat pulmonary cancer	(36)
R. nepalensis Wall MA	Root	India	to treat the treatment of pneumonia	(37)
Lawson			·	. ,
<i>R. nitudus</i> Davis	Bark	Turkey	Used as emetic	(38)
<i>R. persica</i> Boiss.	Leaf	Iran	to treat allergy and itching in children, wound	(39)
R. prinoides L'Hér	Bark, Fruit,	Kenya	to treat sexually transmitted disease (gonorrhea),	(40)
•	Multiple Part	•		. ,
	Fruit, Stem,	Kenya	to treat gonorrhea, prostate, malaria, brucellosis	(41)
	Root, and Leaf		· • · · · · · · · · · · · · · · · · · ·	
	Root	Kenya	to treat muscular skeleton disorder (Arthritis, backaches,	(42)
			rheumatic)	
	Leaf	Ethiopia	to treat Snakebite	(43)
	Roots, Leaf	South Africa	Used for blood purifiers, pneumonia, emetics, purgative, colic,	(44)

			stimulants	
	Branch	South Africa	Herpes, diabetes, HIV related infections	(45)
	Root, Leaf, and Steam	Kenya	to treat ear, nose, and throat (ENT) diseases	(46)
	Leaf	Ethiopia, Uganda	to treat tonsillitis, wound, eczema, skin infection, fever in children, tuberculosis, dandruff, water-borne disease	(47-56)
	Seed	Ethiopia	to treat ringworm	(57)
	Root	Kenya	to treat sexually transmitted infection	(58)
	Root	Kenya	amoebiasis, bacillary dysentery, tonic, pneumonia	(59)
	Root	Ethiopia	to treat hepatic problems	(60)
R. purpureus Edgew.	Bark, Steam, Fruit, Leaf	Himalayas	to treat digestive disorders	(61, 62)
R. purshiana DC.	Bark	Algeria	to treat respiratory tract diseases (pharyngitis)	(20)
	shell	Mexico	to treat skin rash and stomachache	(63)
R. staddo A.Rich	Tree	Kenya	Used for strength/nutrient supplement, sexually transmitted diseases, flu/cold	(64)
	Multiple Part	Kenya	to treat diarrhea	(40)
	Root, Steam,	Kenya	to treat gonorrhea, diabetes, endometritis	(41)
<i>R. triqueter</i> Wall M. A. Lawson	Leaf, Fruit, Branch	Pakistan	to treat hemorrhagic septicemia	(65)
	Bark, Fruit	Himalaya	used for blood purifier, boils, scabies, skin diseases, tonic	(62)
<i>R. virgatus</i> Roxb.	Bark	Himalaya, India	to treat eczema and ringworms	(61, 66)
	Steam, Fruit, Bark	Himalaya	to treat emetic, purgative, eczema, ringworm, affection of spleen	(62)
	Fruit and Bark	Nepal/Iran	to treat diarrhea and dysentry	(67)
	Fruit, Bark	India	to treat emetic, spleen infection, and purgative, curing white dots of eyes	(61, 68)

Phytochemicals

Anthraquinones, flavonoids, naphthalene derivatives, terpenoids, alkaloids, steroids, organic acids are secondary metabolites reported from various *Rhamnus* species of which anthraquinones and flavonoids are the most cited ones (Figures 2-4, Table 2).

Anthraquinones

Several anthraquinones (1-30, Table 2) have been isolated from Rhamnus species. Of these, compounds, a cytotoxic compound prinoidin (23) was reported from the fruits of R. nepalensis against KB (human epidermoid carcinoma of the mouth) with IC₅₀ value of 0.045 μ M, which was four times more potent than the standard doxorubicin having IC_{50} value of 0.2 μM (69). A prominent anti-tyrosinase effect was displayed by 6-methoxysorigenin (12) reported from R. nakaharai with IC₅₀ vaue of 42.2 μM, which was twofold more potent than kojic acid with IC₅₀ value of 82.1 µM (70). In a related study, antioxidant alaternin (10) was reported from R. nakaharai with IC₅₀ value of 117.7 μM compared to ascorbic acid (IC₅₀ value of 63.7 μM) using DPPH assay method (70).

Flavonoids

Flavonoids, which are important secondary metabolites, are widespread in the plant kingdom, either in a free form or in the form of glycosides with wide spectrum of pharmacological application (71). Various flavonoids (31-52) and their derivatives have been reported from the genus Rhamnus. Of these, flavonoids 31, 33, 36, 37 and exhibited cytotoxic, antioxidant, antihyperlipidemia, anti-proliferative, antigenotoxic activity (72-75).

Naphthalene derivatives

To date, 7 naphthalene derivatives (**53-59**) have been reported from different parts of *Rhamnus* species such as *R. prinoides*, *R. cathartica*, *R.*

wightii, R. procumbens, R. makaharai, R. pallasri and R. serrutu. Of these, musizen (**54**) obtained from whole part of R. wightii exhibited antibacterial activities against S. aureus and K. pneumonia using agar disc diffusion assay with MIC value of 9 μ g/mL, which was more potent than streptomycin having MIC value of 120 μ g/mL (76).

Terpenoids

Up to date, four terpenoids (**60-63**) have been reported from leaves and bark extracts of *R. califormica* (77).

Alkaloids

Alkaloids are a large group of naturally occurring compounds with diverse pharmacological activity (78). To date, four alkaloids (**64-67**) have been reported from leaves and bark extracts of *R. califormica* (77).

Steroids

Previous studies reported limited number of steriods and steriod glycosides such as β -sitosterol (**68**) from roots of *R. formosana*, root bark and leaves of *R. alaternus* and leaves of *R. serrutu* (79-81) whereas stigmasterol- β -D-glycoside (**69**) and β -sitosterol-3-O-glycoside (**70**) were reported roots of *R. formosana* (79) and root bark and leaves of *R. alaternus* (80), respectively.

Organic acids

Organic acids are another important component of the genus *Rhamnus*. Previously studied revealed compounds **71-72** and **73-75** from leaves of *R. alaternus* (82) and leaves of *R. heterophylla* (83), respectively.

Miscellaneous Compounds

Compounds **76** and **77** were reported from root and bark of *R. serrutu* and *R. davurica*, respectively (81, 84). The later was also reported from heart wood parts of *R. nakaharai* (70).

Table 2: Compounds reported from the genus *Rhamnus*.

Compound	Plant species	Part used	Reference
Anthraquinones	·		
Chryisophanol (1)	R.formosana and R. serrata	Root	(79, 81)
	R.prinoides	Leaves, Stem	(85)
	R. alaternus	Stem, bark	(86)
	R.frangula	Stem bark and branch	(87)
	R.sphaerosperma	Stem	(88)
	R.alpinus and R.saxatilis	Bark	(89-91)
	R.nepalensis	Fruit	(69)
	R.californica	Leaf and bark	(77)
Emodin (2)	R.formosanaand R. serrata	Root	(79, 81)
	R.pumila	Stem,bark	(86)
	R.prinoides	Fruit, Leaf	(92, 93)
	R.cathartica, R.pubescens, R. alaternus and R.heterophylla	Leaf	(83, 94-96)
	R.frangula	Stem bark and branch	(87)
	R.sphaerosperma	Stem	(88)
	R.procumbens	Whole part	(97)
	R.alpinus and R.saxatilis	Bark	(89, 90)
	R.nakaharai	heartwood	(70)
	R.nepalensis	Fruit	(69)
	R.californica	Leaf and bark	(77)
Physcion (3)	R.formosana	Root	(79)
	R.fallax	Stem, bark	(86)
	R.intermedia	Stem	(86)
	R.prinoides	Leaf and Stem	(85, 92)
	R.frangula	Stem bark and branch	(87)

	R.serrate, R.alaternus and R. alaternus	Root	(81, 98)
	R.sphaerosperma	Stem	(88)
	R.davurica, R.alpinus and R.saxatilis	Bark	(84, 89, 91)
	R.procumbens	Whole part	(97)
	R.nepalensis	Fruit	(69)
	R.californica	Leaf and bark	(77)
Emodinanthrone (4)	R.prinoides	Leaves, Stem	(85, 93)
Emodinbianthrone (5)	R.prinoides	Fruits	(93)
	R.nepalensis	Fruit	(69)
Chrysophanol-emodinbianthrones (6)	R.nepalensis	Fruit	(69)
Chrysophanolbianthrone (7)	R.nepalensis	Fruit	(69)
1,2,6,8-tetrahydroxy-3- methylanthraquinone-8-O-β- glucopyranoside (8)	R.nakaharai	heartwood	(70)
emodin-8-O-β-glucopyranoside (9)	R.nakaharai	heartwood	(70)
Alaternin (10)	R.nakaharai	heartwood	(70)
6-methoxysorigenin-8-O-β-glucopyranoside (11)	R.nakaharai	heartwood	(70)
6-methoxysorigenin (12)	R.nakaharai	heartwood	(70)
Aloe-emodin (13)	R.alaternus	Root	(98)
	R.alpinus and R.saxatilis	Bark	(89-91)
Rhein (14)	R.alaternus	Root	(98)
	R.alpinus and R.saxatilis	Bark	(89-91)
Madagascin (15)	R.saxatilis and R. alpinus	Bark	(90)
	R.cathartica and R. intermedia	Fruit	(99)
3-geranyloxyemodin (16)	R.saxatilis and R. alpinus	Bark	(90)
emodin-6-O-arabinopyranoside- 3',4'-diacetate (17)	R.alaternus	Fruit	(100)
emodin-6-O-arabinopyranoside- 2',3',4'-triacetate (18)	R.alaternus	Fruit	(100)
Emodin 6-O- β -L-rhamnose (19)	R.libanoticus	Bark	(101)
Emodin 8-O-β-D-glucoside (20)	R.libanoticus	Bark	(101)

Physcion 8-O-β-rutinoside (21)	R.libanoticus	Bark	(101)
	R.pallasri	Bark	(102)
Emodinanthrone-6-0- rhamnopyranoside-2',3',4'- triacetate (22)	R.prinoides	Fruit	(93)
Prinoidin (23)	R.prinoides	Fruit	(85, 93)
Prinoidin-emodinbianthrones (24)	R. nepalensis	Fruit	(69)
Rhamnepalins (25)	R. nepalensis	Fruit	(69)
Glucofrangulin (26)	R.prinoides	Fruit	(103)
	R.cathartica	Leaf	(94)
1,6,8-trihydroxy-3- methylanthraquinone 1 -O- rhamnosyl (1→2) glucoside (27)	R.formosana	Root	(104)
1,8-dihydroxy-6-methoxy-3- methyl anthraquinones 8-0- rhamnosyl- (1→2)-glucoside (28)	R.formosana	Root	(79)
1,2,6,8 tetrahydroxy-3 methyl anthraquinone 8-O-ß-D- glucopyranoside (29)	R.alaternus	Root bark and Leaf	(80)
1,4,6,8 tetrahydroxy-3 methyl anthraquinones 1-O-β-D- glucopyranosyl-4,6-di-O-α-L- rhamnopyranoside (30)	R.alaternus	Root bark and Leaf	(80)
Flavonoids			
Kaempferol-3-O-β-rhamninoside	R.petiolaris	Fruit	(105)
(31)	R. nakaharai	Heartwood	(70)
	R. alaternus	Leaf	(72, 73)
Luteolin (32)	R.alaternus	Leaf	(82)
	R.davurica	Bark	(84)
Kaempferol (33)	R.alaternus	Leaf, Fruit	(96, 106)
	R.lycioides	Aerial parts	(107)
	R.davurica	Bark	(84)
	R.saxatilis, R.catharticus and R.disperma	Fruit	(106)
	R.californica	Leaf and bark	(77)
	R.pallasii	Bark	(108)

	R.heterophylla	Leaf	(83)
Quercetin (34)	R.lycioides	Aerial part	(107)
	R.pallasii and R.davurica	Bark	(84)
	R.saxatilis, R.catharticus, R.alaternus and R.disperma	Fruit	(106)
	R.californica	Leaf and bark	(77)
	R.heterophylla	Leaf	(83)
Rhamnazin-3- isorhamninoside (35)	R.formosana	Root	(104)
Rhamnocitrin 3-O-β-isorhamninoside	R.formosana	Root	(104)
(36)	R.nakaharai	heartwood	(70)
	R. alaternus	Leaf	(72, 73)
Rhamnetin 3-O-isorhamninoside (37)	R. alaternus	Leaf	(72, 73)
Rhamnetin 3-O-(3""-O-β- coumaroyl)-β – rhamninoside (38)	R.petiolaris	Fruit	(105)
Quercitrin 39)	R.petiolaris	Fruit	(105)
	R.pallnsii	Bark	(108)
Apigenin (40)	R.davurica	Bark	(84)
Rutin (41)	R.alaternus	Leaf	(82)
	R.cathartica	Leaf	(94)
Rhamnazin (42)	R.prinoides	Fruits, Leaf	(93)
	R.lycioides	Arial part	(107)
	R.disperma	Arial part	(109)
	R.heterophylla	Leaf	(83)
Rhamnetin (43)	R.lycioides	Arial Part	(107)
	R.disperma	Fruit	(70)
Aromadendrin (44)	R.lycioides	Arial Part	(110)
	R.pallasii	Bark	(108)
Eriodictyol (45)	R.lycioides	Arial Part	(110)
	R.pallasii	Bark	(108)
Rhamnocitrin (46)	R.prinoides	Leaf and Stem	(93)
	R.lycioides	Arial Part	(107)
	R.davurica	Bark	(84)
	R.saxatilis	Fruit	(106)

	R.catharticus	Fruit	(106)
	R.alaternus	Fruit	(106)
	R.heterophylla	Leaf	(83)
Taxifolin (47)	R.lycioides	Arial Part	(107)
	R.pallnsii	Bark	(108)
	R.davurica	Bark	(84)
	R.pallasii	Bark	(108)
3- methoxy flavone (48)	R.lycioides	Aerial part	(107)
3-O-Methylquercetin (49)	R.prinoides	Leaf and Stem	(93)
Pallasiin (50)	R.pallasii	Bark	(108)
Isorhamnetin (51)	R.pallasii	Bark	(108)
Mearnsetin (52)	R.pallasii	Bark	(108)
Naphthalene Derivatives			
Geshoidin (P-sorigenin-8-O-β-D-	R.prinoides	Leaf and stem	(85)
glucoside) (53)	R. cathartica	Leaf	
Musizin (54)	R.prinoides	Leaf and stem	(85)
	R.wightii and R. procumbens	Whole part	(76, 97)
Isotorachrysone (55)	R.nakaharai	Root bark	(111)
β-sorigenin (56)	R.prinoides	Leaf and stem	(85)
	R. cathartica	Leaf	(94)
a-sorinin (57)	R.pallasri	Bark	(102)
Eugenine (58)	R.serrutu	Root	(81)
3-hydroxyeugenine (59)	R.serrutu	Root	(81)
Terpenoids			
Umbellulone (60)	R.californica	Leaf and bark	(77)
1,8-cineole (61)	R.californica	Leaf and bark	(77)
a-terpineol (62)	R.californica	Leaf and bark	(77)
Thymol (63)	R.californica	Leaf and bark	(77)
Alkaloid			
Domesticine (64)	R.californica	Leaf and bark	(77)

Isoboldine (66)	R.californica	Leaf and bark	(77)
Bufotenine (67)	R.californica	Leaf and bark	(77)
Steroids			
Stigmasterol-β-D-glycoside (68)	R.formosana	Root	(79)
β-sitosterol (69)	R.formosana	Root	(79)
	R.alaternus	Root bark and Leaf	(80)
	R.serrutu	Leaf	(81)
β-sitosterol-3-O-glycoside (70)	R.alaternus	Root bark and Leaf	(80)
Organic Acid			-
P-coumaric acid (71)	R.alaternus	Leaf	(82)
Ferulic acid (72)	R.alaternus	Leaf	(82)
Gallic acid (73)	R.alaternus	Leaf	(82)
	R.heterophylla	Leaf	(83)
Malic acid (74)	R.heterophylla	Leaf	(83)
Salicylic acid (75)	R.heterophylla	Leaf	(83)
Miscellaneous Compounds			
5-hydroxy-7-methoxyphtali (76)	R.serrutu	Root	(81)
	R.davurica	Bark	(84)
p-hydroxybenzaldehyde (77)	R. nakaharai	Heart wood	(70)

Figure 2: Anthraquinones reported from the genus *Rhamnus*.

Figure 3: Flavonoids reported from the genus Rhamnus.

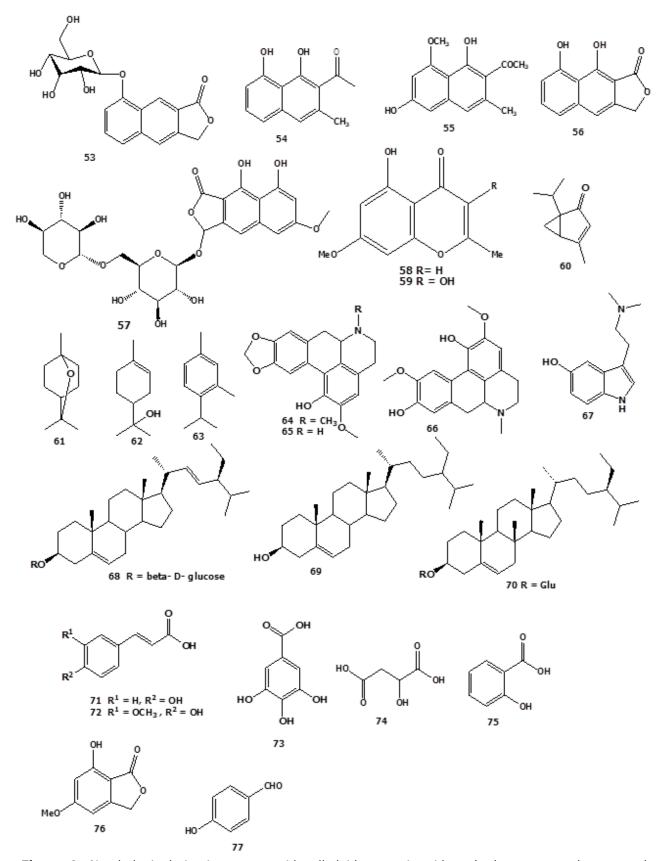


Figure 4 : Napthalenic derivatives, terpenoids, alkaloids, organic acids and other compounds reported from the *genus Rhamnus*.

Essential oils

The essential oils from plants are known with various pharmacological activities (112). Campbell et al., (2019) reported essential oils from the leaves of R. prinoides of which 4-hydroxy-4-methyl-2-pentanone and ethyl 4-ethoxybenzoate score more than 85% and exhibited significant anti-biofilm activity (113). In a related study, Chouitah et al., (2012) reported essential oils from the leaves of R.alaternus (114) of which camphene (17.63%), linalool (16.13%), pulegone (15.01%), naphthalene (14.66%), mequinol (2.77%) and borneol (2.13%) are among the major components.

Pharmacological activities Hepatoprotective activity

Berroukche et (2015)al. hepatoprotective activity of the macerated R. alaternus extract in Wistar rats treated with the toxic carbon tetrachloride (CCl₄) that causes hepatic damage through evaluation of both the biochemical and histopathological changes in rats. The extracts with bodily weight (250 mg/kg) reduced the elevated levels alkaline of phosphatase (ALP), Glutamic oxaloacetic transaminase (GOT),Glutamic pyruvic transaminase (GPT) and total bilirubin and significantly attenuated the deleterious histopathologic changes in the liver after carbon tetrachloride (CCl₄)-intoxication (14).

Anti-inflammatory activity

Thakru and Prasad (2019) evaluated *in vivo* anti-inflammatory activity of ethanolic extract of R.purpureus stem bark using the carrageenan-induced rat paw edema assay in adult Swiss albino mice, where 200 mg/kg bodily weight of the extract was administered orally to different groups of mice with indomethacin (10mg/kg) as the positive control. The crude ethanolic extract showed considerable (P < 0.05) anti-inflammatory activity with inhibition of 54.50% and 54.77% after 3 h and 4 h of treatment as compared to the standard drug indomethacin (10 mg/kg) showed the inhibition of 50.46%, and 51.78% after 3 h and 4 h of treatment, respectively (115).

Chen et al., (2018) evaluated the antiinflammatory activity of apigenin (40) and Kaempferol (33) isolated from 80% methanol bark extract of *R.davurica* Pall using the cyclooxygenase (COX-2) inhibition assay, with aspirin as the positive control. Apigenin (40) and Kaempferol (33) exhibited anti-inflammatory activity with IC₅₀ values of 10.14 and 9.27 μ g/mL, respectively (74). Chen et al., (2020) evaluated anti-inflammatory activity of 60% ethanol stem and stem bark semi-R.prinoides purified extracts of using cyclooxygenase (COX-2) inhibition assay, with

aspirin as the positive control. The semi-purified extract exhibited activity with IC_{50} value of 20.6 μ g/mL, which was weak activity compared to IC_{50} value of 6.33 μ g/mL exhibited by ascorbic acid (116).

Antibacterial activity

Molla et al., (2016) evaluated antibacterial activities of methanol and chloroform solvent fractions of R.prinoides crude leaves extract against S. aureus, S. pyogen, S. pneumoniae, and S. typhi using agar well diffusion methods with ampicillin and ciprofloxacin as positive controls. Methanol and chloroform extracts antibacterial activities at different concentrations (78 mg/well, 39 mg/well, and 19.5 mg/well). The average minimum inhibitory concentration of the methanol and chloroform extracts ranged from mg/mL and 8.13-16.25 8.13-32.5 respectively (117).

Ammar *et al.*, (2007) evaluated the antibacterial activities of petroleum ether, chloroform, ethyl acetate, methanol, and total Oligomers flavonoids (TOF) enriched leaves extracts of *R.alaternus* against *S. aureus*, *E. faecalis*, *E. coli*, *S. enteritidis* and *S. typhimurium* using micro dilution and agar dilution methods. The TOF extracts showed activities against *S. aureus*, *E. faecalis*, *E. coli*, *S. enteritidis* and *S. typhimurium* with MIC values of 120 μ g/mL,175 μ g/mL, 1.75 mg/mL, 125 μ g/mL and 62.5 μ g/mL, respectively, while the ethyl acetate extract exhibited with MIC values of 70 μ g/mL, 150 μ g/mL, 3.75 mg/mL, 100 μ g/mL and 175 μ g/mL, respectively (118).

Chouitah et al., (2012) evaluated antibacterial activities of essential oils of R.alaternus leaves against P. aeroginosa, E. coli and S. typhimurium using the paper disc diffusion method. The essential oils exhibited activities with zones of inhibition 8, 17 and 15 mm, respectively (114). Carranza et al., (2015) evaluated antibacterial activities of methanol extracts of leaves and bark of R. californica against B. cereus, S. pyogenes, M. smegmatis, S. aureus, methicillin-resistant S. aureus (MRSA), E. coli, and P. aeruginosa using disc diffusion and minimal inhibitory concentration (MIC) assays. Both extracts inhibited MRSA growth and other Gram-positive bacteria with MICs of 3.3-6.0 mg/mL (77). Raja et al., (2018) evaluated antibacterial activities of ethyl acetate extract of R.wightii whole part against S. aureus, B. cereus, E. faecalis, K. pneumonia, P. aeruginosa and E. coli using agar disc diffusion method with streptomycin and gentamycin as positive controls. The ethyl acetate extract of the whole part of R. wightii revealed inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and 12, respectively, which is highly comparable with the positive control, streptomycin (25 µg/disc) and gentamycin (50

 $\mu g/disc)$. The isolated compound musizen (**54**) and standard drugs have additionally inhibited *S. aureus* and *K. pneumonia* growth at a concentration (MIC value) of 9 $\mu g/mL$ and 120 $\mu g/mL$, respectively (76).

Kosalec et al., (2013) evaluated antibacterial activities of methanol bark extracts of R. alaternus, R. fallax, R. intermedia and R. pumila against S. aureus, P. aeruginosa and E. coli using microdilution broth assay. All plant extracts exhibited activities with MIC values of ranging from 1.25 to 2.5 µg/mL (86). Carranza et al., (2015) evaluated antibacterial activities of methanol leaf extracts of R.californica against S. aureus, Methicillin-resistant S. aureus, B. cereus, P. aeruginosa, S. pyogenes and E. coli using Kirby-Bauer disc diffusion assay with streptomycin as positive control. The extract exhibited activities with zone of inhibition ranging from 9 mm to 14.3 mm, which was moderate activities compared to the standard with zone of inhibition ranging from 17 mm to 23.8 mm (77).

Antifungal Activity

Kosalec *et al.*, (2013) evaluated antibacterial activities of methanol bark extracts of *R. alaternus*, *R. fallax*, *R. intermedia* and *R. pumila* against *C. albicans*, *A. niger* and *M. gypseum* using microdilution broth assay. All the plant extracts exhibited activities with MIC values of 0.625 mg/mL and 2.5 mg/mL against *Candida albicans* and *Aspergillus niger*, respectively, whereas extracts of *R. fallax*, *R. intermedia* and *R. pumila* exhibited with MIC value of 0.313 mg/mL against dermatophyte species (*Microsporum gypseum*) (86).

Antimalarial activity

Koch et al. (2009) evaluated antimalarial activities of chloroform root bark extracts of R. prinoides chloroquine-sensitive Plasmodium falciparum strain using ELISA assav chloroquine as standard drug. The extract exhibit with IC_{50} value of $3.53\mu g/mL$, which was weak activities compared to IC_{50} value of 0.004 $\mu g/mL$ exhibited by chloroguine the standard drug (119). Another study evaluated the anti-plasmodia activities of n-hexane, dichloromethane, and methanol root extracts of R.prinoides using the radioisotope method. All extracts were found to have in vitro antimalarial activity. The highest activity was displayed by n-hexane and dichloromethane extracts with IC_{50} values of 19.9 $\mu g/mL$ and 30.3 $\mu g/mL$, respectively (120). The naphthalene derivative geshoidin (53) from R.prinoides showed an IC50 value of 4.0 pM and 0.4 pM against the chloroquine-sensitive (D6) and chloroquine-resistant (W2) strains of *Plasmodium* falciparum (121). In a related study, in vivo antimalarial activity of aqueous extracts from leaves

and root barks of *R.staddo*, *R. prinoides* and their chloroquine (CQ) potential effects against a blood-induced CQ-resistant rodent parasite in mice showed high chemo suppression in the range 51% -75% (122). Results of those studies suggest that the extracts of *R.prinoides* have a promising antiplasmodial activity which supports the folkloric use of the plant for treating malaria.

Antioxidant activity

Bhouri et al., (2011) evaluated Kaempferol 3-O-βisorhamninoside (31) and rhamnocitrin $3-O-\beta$ isorhamninoside (36) isolated from soxhlet methanolic leaves extract of R.alaternus using superoxide radical scavenging activity riboflavin as reference signal. The compounds produced an 80.4% and 85.6% decrease in NBT/riboflavin photo reduction, respectively, at a dose of 150 µg/assay. However K3O-ir was more potent superoxide scavenger with an IC₅₀ value of 18.75 μ g/mL than R3O-ir (IC₅₀ = 22.5 μ g/mL)(73). Rocchetti et al., (2019) determined the antioxidant activities of methanol and aqueous unmature fruit extracts of *R.petiolaris* using radical scavenging activities (DPPH and ABTS assay) with reported as trolox equivalents (mgTE/g extract) as reference. The methanolic and aqueous unmature fruit extracts were the most effective 2,2-diphenyl-1-(DPPH) picrylhydrazyl and 2,2'-azino-bis(3ethylbenzothiazoline-6-sulfonic (ABTS) acid) scavenger (470.96 mg trolox equivalent (TE)/g and 394.96 mg TE/g) respectively (123).

Zeouk *et al.*, (2020) evaluated *in vivo* antioxidant activities of ethanolic extracts of *R.alaternus* leaves using scavenging activities (DPPH assay) with butylated hydroxytoluene (BHT) as a standard. The crude extract showed activities with IC_{50} value of $58~\mu g/mL$, which has good antioxidant activities when compared to IC_{50} value of $31~\mu g/mL$ exhibited by butylated hydroxytoluene (BHT), the positive control. Similarly, ethanolic extracts of *R.alaternus* leaves fraction have exhibited highest antioxidant activity with IC_{50} values of 32.76%, 27.01% and 38.87%, respectively(96).

Ammar et al., (2008) evaluated the antioxidant activity of aqueous extract and ethyl acetate fraction leaves of R.alaternus using Xanthine Oxidase (XOD) assay with allopurinol as positive control. The aqueous extract and ethyl acetate fraction exhibited high xanthine oxidase inhibiting with respective IC₅₀ values of 208 and 137 μg/mL and super oxide anion scavenging effects with IC₅₀ values of 132 and 117 µg/mL (124). Ammar et al., (2009) evaluated the antioxidant activities of methanolic and total oligomer flavonoid enriched extracts from R. alaternus leaves using DPPH radical scavenging and xanthine oxidase (XOD) assay with vitamin E and allopurinol as positive control, respectively. Rhamnetin-3-0isorhamninoside (37) showed DPPH activities with IC $_{50}$ value of 1.5 µg/mL which is more antioxidant activity as compared to IC $_{50}$ value of 3 µg/mL exhibited by vitamin E the standard drug. Similarly, the isolated compound exhibited xanthine oxidase (XOD) inhibiting with respective IC $_{50}$ values of 18, 81 and 40 µg/mL and superoxide anion scavenging effects with IC $_{50}$ values of 42,79 and 35 µg/mL as compared with the positive control allopurinol having IC $_{50}$ value of 37 and 4 µg/mL, respectively (72).

Ben Ammar et al., (2008) evaluated antioxidant activities of methanol extracts from R.alaternus leaves and root bark using DPPH radical scavenging and xanthine oxidase (XOD) assay with a-tocopherol and allopurinol as positive control, respectively. The root bark extract of *R.alaternus* revealed more effective than the leaves extract with IC_{50} values of 7.21 and 18.84 $\mu g/mL$, respectively, compared to IC₅₀ value of 3 μg/mL exhibited by a-tocopherol. Similarly, the leaves and root bark extract exhibited xanthine oxidase (XOD) inhibiting with respective IC₅₀ values of 103.96 and 83.33 µg/mL and superoxide anion scavenging effects with IC₅₀ values of 171 and 92 μg/mL compared to allopurinol having IC₅₀ value of 37.3 and 6 µg/mL, respectively(125).

Bhouri et al., (2012) evaluated antioxidant activities of Kaempferol-3-O-β-isorhamninoside and rhamnocitrin 3-O- β -isorhamninoside (37) isolated from leaves of R. alaternus using cupric reducing antioxidant capacity (CUPRAC), reducing power assay, and ferric reducing antioxidant power (FRAP) with Trolox (10-1000 μ g/mL) as a positive control. The compound K30-ir and exhibited a significant ability to reduce the Cu2+ neocuproine complex to Cu⁺ neocuproine in a dose dependent manner. The highest values obtained with 1 mg/mL of each compound, were 374 µg/mL and 310 µg/mL equivalent to Trolox, respectively. The reducing power assay evaluates antioxidant capacity of compounds based on their ability to reduce ferric (Fe³⁺) to ferrous (Fe²⁺) ion through the donation of an electron, with the resulting (Fe^{2+}) formation monitored ferrous ion spectrophotometrically at 700 nm. The tested compounds exhibited good reducing potential a concentration of 1 mg/mL. R3O-ir exhibited higher reducing power of iron (368 µg/mL equivalent of Trolox) than K3O-ir (330 µg/mL equivalent of Trolox) (126).

Chaouche et al., (2020) evaluated the antioxidant activities of methanol-acetone leaves and stem bark extracts of R. alaternus using DPPH radical scavenging and ferric reducing antioxidant potential (FRAP) assay with butylated hydroxyanisole (BHA) as a positive control. The leaves and stem bark extracts exhibited DPPH

activities with IC50 values of 10.5 and 51.2 µg/mL, respectively, which was weak activity compared to IC50 value of 5.6 µg/mL exhibited by BHA the positive control. Similarly, the leaves and stem bark extracts exhibited FRAP activities with EC50 values of 0.4 and 1.8 µg/mL, respectively, which was weak activity compared to EC50 value of 0.1 µg/mL exhibited by BHA (127).

Hsiao *et al.*, (1996) evaluated antioxidant activities of compound isotorachrysone (**55**) isolated from root bark extracts of *R.nakaharai* using ironinduced lipid peroxidation technique in rat brain homogenates with butylated hydroxytoluene (BHT), alpha tocopherol and desferrioxamine as a positive controls. The study revealed that isotorachrysone (**55**) exhibited IC₅₀ value of 1.64 μ M, which was comparable to IC₅₀ value of 1.08 μ M exhibited by BHT and was more potent than alpha tocopherol and desferrioxamine with IC₅₀ values of 3.71 and 97.10 μ M as standard drug (111).

Kosalec et al., (2013) evaluated antioxidant activities of bark extracts of R. alaternus, R. fallax, R. intermedia and R. pumila using β-carotenelinoleic acid, DPPH radical scavenging, reducing power assay, and chelating activity with BHA, ascorbic acid, quercetin, and EDTA as positive controls. All the plant extracts, R. alaternus, R. fallax, R. intermedia and R. pumila exhibited activities using β-carotene-linoleic acid assay with EC₅₀ values of 250, 289, 38 and 29.5 μ g/mL respectively, which was greater activity compared to EC₅₀ value of 852 µg/mL exhibited by ascorbic acid. Similarly, all the plant extracts, R. alaternus, R. fallax, R. intermedia and R. pumila exhibited activities using reducing power assay with EC₅₀ values of 0.91, 1.99, 0.81 and 0.99 $\mu g/mL$ respectively, which was comparable and greater activities compared to EC₅₀ values of 7.53, 1.8 and 7.59 µg/mL, respectively, exhibited by BHA, quercetin and ascorbic acid as standard drugs (86).

Lu et al., (2016) evaluated antioxidant activates of alaternin (10) and emodin-8-O-glucoside (20) isolated from methanol extracts of R.nakaharai heart wood using ABTS, DPPH and Superoxide dismutase (SOD-like) assay with ascorbic acid, 3-tbutyl-4-hydroxynisode (BHA) as positive control. The compound alaternin (10), showed DPPH activity with IC50 value of 117.7 μM , which was moderate activities compared to IC₅₀ value of 63.7 μM exhibited by ascorbic acid. Also, alaternin (10) and emodin-8-O-glucoside (20) exhibited SOD-like activities with IC50 values of 247 and 232 μM , respectively, which were better activities compared to IC₅₀ value of 292 µM exhibited by BHA (70). Chen et al., (2020) evaluated antioxidant activities of 60% ethanol stem and stem bark crude and semi purified extracts of R.prinoides using DPPH

and ABTS assay with butylated hydroxytoluene (BHT) as positive control. The semi-purified extract exhibit DPPH activities with IC_{50} value of 0.2 mg/mL, which was more potent than the standard BHT having IC_{50} value of 0.286 mg/mL. Similarly, the crude extracts exhibit ABTS activities with IC_{50} value of 0.0596 mg/mL, which was comparable to IC_{50} value of BHT (116).

Mazhar et al., (2013) evaluated antioxidant activities of methanol extract and their fractions (ethyl acetate, n-butanol, chloroform and nhexane) of R.triguetra aerial parts using DPPH assay with butylated hydroxytoluene (BHT) as a positive control. The crude extract and their fractions exhibited activities with IC50 values of 70.26, 7.59, 37.98, 60.09 and 182.99 $\mu g/mL$ respectively, of which the ethyl acetate fraction showed better activity among the extracts, compared to IC₅₀ value of 12.1 µg/mL exhibited by BHT (128). Boussahel et al., (2013) evaluated antioxidant activities of methanol and aqueous extract of R. alaternus leaves using DPPH and βcarotene-linoleic acid assay with butylated hydroxytoluene (BHT) as a positive control. The methanolic and aqueous extracts exhibited DPPH activities with IC_{50} values of 0.082 and 0.398 mg/mL, respectively, of which methanol extract is more active, compared to IC₅₀ value of 0.032 mg/mL exhibited by BHT. Similarly, the methanol extract exhibited activities using β-carotenelinoleic acid assay with 89% inhibition, which was comparable to 99.2% inhibition displayed by BHT Boussahel *et al.*, (2015) evaluated antioxidant activities of methanol bark extract of R. alaternus using oxygen radical absorbance capacity assay (ORAC) with trolox equivalent antioxidant capacity as a standard. The extract exhibited with 6.55 mmol TE/g extract, which was more active as compared to the standard TEAC with 0.75 mmol TE/g extract (130).

Antiproliferative Activity

Ben Ammar et al., (2008) evaluated the antiproliferative effect of root bark and leaves extracts obtained from R. alaternus against K562 human cell line and L1210 mouse lymphoma cells, at various concentrations comprised between 100 and 800 µg/mL using tetrazolium salt (3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (MTT) assay. The leaves and roots extracts from R. alaternus showed interesting antiproliferative in a dose-dependent manner. The root extract was more effective than the leaves, on both types of leukemia cells. Indeed, concerning the K562 human cell, the IC₅₀ values of roots and leaves extracts were determined at 165 and 260.69 μg/mL, respectively. Concerning the L1210 cells, the IC50 values of roots and leaves extracts were determined at 210.73 and 343.10 µg/mL,

respectively, in the presence of a-tocopherol as positive control (125).

Chen et al., (2016) evaluated the antiproliferative effect of 80% methanol extracts obtained from R.davurica using protein-staining sulforhodamine B (SRB) microculture colorimetric assay against human cancer cell lines of HT-29 (intestinal carcinoma) and SGC-7901 (gastric carcinoma). The extract exhibited significant dose-dependent antiproliferative activities against HT-29 and SGC-7901 cells with IC₅₀ values of 24.96 and 89.53 respectively. Meanwhile, activities against both HT-29 and SGC-7901 cells significantly increased by the treatment with R. davurica bark extract in a time-dependent manner from 24-96 h at a dose of 150 $\mu g/mL$, although there was a decrease on SGC-7901 cells at the time from 72 h-96 h (84).

Chen et al., (2018) evaluated the antiproliferative effect of compounds apigenin (40) and kaempferol (33) obtained from 80% methanol extracts of R.davurica bark using MTT colorimetric assay against three human cancer cell lines of Hep G2 (hepatic cancer), SGC-7901 (gastric carcinoma), and HT-29 (intestinal carcinoma). Kaempferol (33) exhibited antiproliferative activities against HT-29, SGC-7901 and Hep G2 cells with IC50 values of 25.7, 13.43 and 20 μ g/mL respectively, while the compound apigenin (40) exhibited with IC50 values 19.79, 17.76 and 10.20 μ g/mL, respectively (74).

Wound healing Activity

Tessema *et al.*, (2021) evaluated wound healing activities of methanol leaf extracts of *R.prinoides* using excision and incision models in adult Swiss albino mice, with nitrofurazone ointment as a standard. Treatment with 5 % and 10 % (w/w) methanol extract ointment exhibited significant wound recovery activities in both excision and incision models, which has higher activity when compared the standard nitrofurazone ointment (131).

Cytotoxicity and Toxicity Activity

Ahmadi et al., (2016) evaluated the cytotoxic activities of hydroalcoholic extracts of R.frangula against breast cancer cellline (MCF-7) using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay. The extract exhibited activities with half maximal cytotoxic concentration (CC₅₀) value of 10 mg/mL (132). Ben Ammar et al., (2008) evaluated the cytotoxic activities of petroleum ether, chloroform, ethyl methanol and total oligomers flavonoids (TOF) enriched leaves extracts of R.alaternus against human chronic myelogenous K562 and murine Leukaemia L1210 cells using 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The TOF extract exhibited

with IC₅₀ values of 75 μ g/mL and 198 μ g/mL against K562 and L1210 cells, respectively. Similarly, the ethyl acetate extract showed activities with IC₅₀ values of 232 μ g/mL and 176 μ g/mL respectively (125).

Bhouri et al., (2011) evaluated the cytotoxic activities of kaempferol 3-O-\(\beta\)-isorhamninoside and rhamnocitrin 3-O-β-isorhamninoside isolated from methanol leaves extracts of *R.alaternus* using Alamar blue assay against human lymphoblastoid TK6 cells, with cells treated by 0.5% DMSO as a control. The compound neither K3O-ir nor R3O-ir reached 50% inhibition of TK6 cell proliferation (75). Chen et al., (2016) evaluated in vitro toxicity activity of 80% ethanol bark extracts of R. davurica against normal human hepatic cells (L-O2) using protein-staining sulforhodamine B (SRB) microculture colorimetric assay. The extract exhibited activities with IC50 value of 229.19 μg/mL on L-O2, which suggested that R. davurica bark extract showed very low or no toxicity on hepatic cell viability (84).

Mai *et al.*, (2001) evaluated cytotoxicity activity of prinoidin (**23**) isolated from methanol extracts of *R.nepalensis* fruit against KB (human epidermoid carcinoma of the mouth) cell using MTT assay with doxorubicin as a positive control. Prinoidin (**23**) exhibited IC_{50} value of 0.045 μ M, which was four times more potent than the standard, doxorubicin, having IC_{50} value of 0.2 μ M (69). Boussahel *et al.*, (2015) evaluated cytotoxicity of methanol extract of *R.alaternus* bark against human monocytic leukemia cells (U937) using trypan blue assay with taxol as standard drug. The extract exhibited activities with IC_{50} values of 6.39 μ g/mL, which was comparable to IC_{50} value of 2.47 μ g/mL exhibited by taxol the standard drug (130).

Anti-tyrosinase Activity

Lu *et al.*, (2016) evaluated the anti-tyrosinase activity of 6-methoxysorigenin (**12**) isolated from methanol extracts of *R.nakaharai* using mushroom tyrosine inhibitory assay with kojic acid as positive control. The study revealed that 6-methoxysorigenin (**12**) exhibited activities with IC₅₀ value of 42.2 μ M, which was twofold inhibitory effect than the positive control kojil acid having IC₅₀ value of 82.1 μ M (70).

Antihyperlipidemic Activity

Tacherfiout et al., (2018) evaluated antihyperlipidemic activities of methanol extracts of *R.alaternus* leaf on circulating lipids in rats with Triton WR-1339-induced hyperlipidemia, intracellular lipid accumulation and expression of genes of fatty acid metabolism in human hepatoma HepG2 cells, and adipogenesis in the 3T3-L1 murine adipocyte cell model. The Oral crude extract administration decreased blood levels of

cholesterol and triacylglycerol in hyperlipidemic rats (by 60% and 70%, respectively, at 200 mg extract/kg). In HepG2 cells, the extract exposure dose-dependently decreased intracellular lipids and up-regulated gene expression of palmitoyl transferase 1 involved in fatty acid oxidation, while in the 3T3-L1 model the extract favored preadipocyte proliferation and adipogenesis, pointing to positive effects on adipose tissue expandability(133).

Ammar et al., (2009) evaluated the anti lipid peroxidation activity of kaempferol 3-O-Bisorhamninoside, rhamnocitrin-3-0isorhamninoside and rhamnetin-3-O-βisorhamninoside (37) from methanol leaf extracts of R. alaternuswas estimated by calculating the values of malondialdehyde (MDA) in cultured K562 human chronic myelogenous leukemia cells. In this study, the compounds displayed IC50 values of 180,320 and 106 μg/mL, respectively, compared to IC₅₀ value of 15µg/mL exhibited by vitamin C as a reference (72).

Antimutagenic activity

Ammar et al., (2008) evaluated the antimutagenic activity of leaves extracts by the Ames assay, using the mutagen Aflatoxin B1 (AFB1) at a concentration of 10 μ g/mL. The experiment was carried out with two strains of Salmonella Typhimurium (i.e., TA98 and TA100) in the presence of various extracts, and spontaneous revertant was used as control. Petroleum ether, chloroform, methanol, water, and total oligomers flavonoids (TOF) extracts obtained by R. alaternus were investigated at various doses (10, 50, and 250 µg/mL) and remarkably reduced the AFB1induced mutagenicity. The study revealed that ethyl acetate extract to be the most effective at a dose of 250 µg/mL. At such dose, the inhibition percentage of mutagenicity was determined by the Ames assay up to 78% for the TA98 strain (124).

Antigenotoxic activity

Bhouriet al., (2011) evaluated the antigenotoxic activity of Kaempferol 3-O- β -isorhamninoside and rhamnocitrin3-O- β -isorhamninoside isolated from methanol extract of leaves of R. alaternus on E. coli PQ37 using SOS chromo test with two positive control snifuroxazide and aflatoxin B1 used at 10 $\mu g/assay$ and 5 $\mu g/assay$, respectively. The assay carried out in absence of both aflatoxin B1 and extracts constituted the negative control. For the three flavonoid concentrations studied (1, 5, and 10 $\mu g/assay$), the antigenotoxic activity of rhamnocitrin 3-O- β -isorhamninoside was higher than the one determined for Kaempferol 3-O- β -isorhamninoside (73).

Bhouri et al., (2012) evaluated antigenotoxic properties of Kaempferol $3-O-\beta$ -isorhamninoside

(**31**) and rhamnocitrin 3-O-β-isorhamninoside (**36**) isolated from leaves of *R. alaternus* (i.e.,) using comet assay on human lymphoblastoid cells TK6 and NH32. Quantification of the comet data was reported as Total DNA damage (TDD). The compound exhibited no significant difference was detected between the TDD induced by K3O-ir (TDD=212, 151 and 67 at concentrations of respectively of 800, 400 and 200 μg/mL) and that induced by R3O-ir (TDD=238, 139 and 110) at the same tested concentrations in TK6 cells and the negative control (non-treated cells; TDD=163) on the other hand. In the opposite, a significant increase of the total DNA damage (TDD=348) was

observed in TK6 cells exposed to 75 μ M of H₂O₂, compared to the untreated cells. Likewise, K3O-ir and R3O-ir revealed a non genotoxic effect at the doses of (200 and 400 μ g/ml) whereas the highest tested concentration (800 μ g/mL) exhibited a genotoxic effect when tested with NH32 cells. The TDD values were 240 and 226 with respectively K3O-ir and R3O-ir, suggesting inducing of DNA breakage in p53 deficient lymphoblastoid human cells (126).

Summary of pharmacological activity of *Rhamanus* species is presented in Table 3 below.

Table 3: Pharmacological activities of extracts and isolated compounds from *Rhamnus* species

induced rat paw edema assay Induced rat paw edmand frum induced restricties with ICso value of 20.6 µg/mL Induced rat paw edmand from 8.13 µg/mL to 10.50 value of 6.33 µg/mL to 10.50 value of 10.14 and 9.27 µg/mL respectively Induced rat paw edmand from 8.13 pay edman	Activity	Plant species	Extract	Plant Part	Method	Effect	Refs
Anti-inflammatory R. prinoides ethanol service bark bark (COX-2) assay bark (COX-2) assay (COX-2) as	Hepatoprotective	R. alaternus	aqueous	leaves	histopathological changes in Wistar	Glutamic oxaloacetic transaminase (GOT), Glutamic pyruvic transaminase (GPT) and total bilirubin and significantly attenuated deleterious histopathological	(14)
Anti-inflammatory R. davurica methanol bark Cyclooxygenase (COX-2) assay methanol bark Cyclooxygenase (COX-2) assay methanol bark Cyclooxygenase (COX-2) assay (COX-2) ass	Anti-inflammatory	R. alaternus	ethanol	Stem bark	induced rat paw	of 54.50% and 54.77% after 3 hr and 4 hr of treatment as compared the standard drug indomethacin (10 mg/kg) showed the inhibition of 50.46%, and 51.78% after 3hr	(115)
Antibacterial R. prinoides Methanol and chlorofor m Methanol ch	Anti-inflammatory	,		bark	, , , ,	which was weak activities compared to IC_{50} value of 6.33 $\mu g/mL$ exhibited by ascorbic acid the standard	(116)
Antibacterial R. alaterus Ethyl leaves acetate and Total Oligomers flavonoids (TOF) Antibacterial R. wightii Ethyl R. wightii Ethyl acetate Antibacterial R. wightii Ethyl acetate Antibacterial R. wightii Ethyl acetate Antibacterial R. wightii Ethyl R. wightii Ethyl R. wightii acetate Antibacterial R. wightii Ethyl R. wightii Ethyl R. wightii acetate Agar disc diffusion S. pneumoniae, and S. typhi with MIC value of 3.13 mg/mL to 16.25 mg/mL, respectively. Antibacterial R. wightii Ethyl R. wightii Ethyl R. wightii acetate Agar disc diffusion Ethyl R. wightii Ethyl R. wightii acetate Agar disc diffusion Ethyl R. wightii Ethyl R. whole part acetate exhibited activities against S. aureus, E. acetate exhibited activities and S. typhimurium with MIC values of 120 µg/mL, 1.75 mg/mL, 1.75 µg/mL, 150µg/mL, 3.75 mg/mL, 100 µg/mL and 175 µg/mL respectively Antibacterial R. wightii Ethyl R. whole part acetate Ethyl R. wightii Ethyl R. whole part acetate Ethyl R. wightii Inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and 12 respectively	, 		methanol	bark	(COX-2) assay	(33) exhibited activities with IC_{50} values of 10.14 and 9.27 $\mu g/mL$ respectively	(74)
Antibacterial R. alaterus Ethyl leaves acetate and Total Oligomers flavonoids (TOF) Antibacterial R. wightii Ethyl acetate Antibacterial R. wightii Ethyl acetate Antibacterial R. wightii Ethyl leaves Agar disc diffusion Extract exhibited activities against S. aureus, E. aureus, E. flavonoids (TOF) Agar disc diffusion Extract exhibited activities against S. aureus, E. aureus, E. coli, S. enteritidis and S. typhimurium with MIC values of 120 μg/mL,175 μg/mL, 1.75 mg/mL, 1.25 μg/mL and 62.5 μg/mL respectively, while the ethyl acetate extract exhibited with MIC values of 70μg/mL, 150μg/mL, 3.75 mg/mL, 100 μg/mL and 175 μg/mL respectively Agar disc diffusion Extract exhibited activities against S. aureus, B. cereus, E. faecalis, K.pneumonia, P. aeruginosa and E. coliwith inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and 12 respectively	Antibacterial	R. prinoides	and chlorofor	leaves	Agar well diffusion	S. pneumoniae, and S. typhi with MIC value of methanol and chloroform fractions ranged from 8.13 mg/mL to 32.5 mg/mL and from 8.13 mg/mL to 16.25 mg/mL,	(117)
acetate agar dilution faecalis, E. coli, S. enteritidis and S. typhimurium with and Total Oligomers playmL and 62.5 μg/mL respectively, while the ethyl acetate extract exhibited with MIC values of 70μg/mL, 150μg/mL, 3.75 mg/mL, 100 μg/mL and 175 μg/mL respectively Antibacterial R. wightii Ethyl Whole part acetate Ethyl Whole part acetate Agar disc diffusion acetate extract exhibited activities against S. aureus, B. cereus, E. faecalis, K.pneumonia, P. aeruginosa and E. coliwith inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and 12 respectively	Antibacterial	R. californica	methanol	Leaf and bark	Disc diffusion	pyogenes, M. smegmatis, S. aureus, methicillin-resistant	(77)
Antibacterial R. wightii Ethyl Whole part Agar disc diffusion Extract exhibited activities against S. aureus, B. cereus, (76) acetate E. faecalis, K.pneumonia, P. aeruginosa and E. coliwith inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and 12 respectively	Antibacterial	R. alaterus	acetate and Total Oligomers flavonoids	leaves		faecalis, E. coli, S. enteritidis and S. typhimurium with MIC values of 120 μ g/mL,175 μ g/mL, 1.75 mg/mL, 125 μ g/mL and 62.5 μ g/mL respectively, while the ethyl acetate extract exhibited with MIC values of 70 μ g/mL, 150 μ g/mL, 3.75 mg/mL, 100 μ g/mL and 175 μ g/mL	(118)
	Antibacterial	R. wightii	•	Whole part	Agar disc diffusion	Extract exhibited activities against <i>S. aureus</i> , <i>B. cereus</i> , <i>E. faecalis</i> , <i>K.pneumonia</i> , <i>P. aeruginosa</i> and <i>E. coli</i> with inhibition zones (in mm) of 15, 16.66, 15, 19, 10.66 and	(76)
	Antibacterial	R.	methanol	Bark	Micro-dilution	· · · · · · · · · · · · · · · · · · ·	(86)

	alaternus,R. fallax,R. intermedia and R. pumila			broth assay	aeruginosa and E. coli with MIC value of ranging from 1.25 to 2.5 μg/mL	
Antibacterial	R. califormica	methanol	Leaves	Kirby-Bauer disc diffusion	Extract exhibited activities against Staphylococcus aureus, Methicillin-resistant Staphylococcusaureus, Bacillus cereus, Pseudomonas aeruginosa, Streptococcus pyogenes and Escherichia coliwith zone of inhibition ranging from 9 mm to 14.3 mm	(77)
Antibacterial	R. wightii	Ethyl acetate	Whole part	Agar disc diffusion	Musizen (54) exhibited activities against <i>S. aureus</i> and <i>K. pneumonia</i> with MIC value of 9 μ g/mL, which was more potent than the standard drug Streptomycin having MIC value of 120 μ g/mL	(76)
Antifungal	R. alaternus, R. fallax,R. intermedia and R. pumila	methanol	Bark	micro-dilution broth assay	All extract exhibited activities against <i>C. albicans</i> , <i>A. niger</i> and <i>M.gypseum</i> with MIC values of 0.625 mg/mL and 2.5 mg/mL against <i>C. albicans</i> and <i>A.niger</i> respectively, while the plant extracts <i>R. fallax</i> , <i>R. intermedia</i> and <i>R. pumila</i> exhibited with MIC value of 0.313 mg/ml against dermatophyte species (<i>M. gypseum</i>)	(86)
Antimalarial	R. prinoides	Chlorofor m	Root bark	ELISA assay	extract exhibited activities against chloroquine-sensitive <i>Plasmodium falciparum</i> strain with IC_{50} value of 3.53 μ g/mL, which was weak activities compared to IC_{50} value of 0.004 μ g/mL exhibited by chloroquine the standard drug	(119)
Antimalarial	R. prinoides	Hexane& dichlorom ethane	root	Radioisotope	extracts of hexane and dichloromethane exhibited activities anti-plasmodia with IC_{50} values of 19.9 μ g/mL and 30.3 μ g/mL, respectively	(120)
Antimalarial	R. prinoides and R. staddo	aqueous	Leaves and root bark	blood-induced CQ- resistant rodent parasite in mice	The plant extract and standard drug,chloroquine (CQ) potential effects against a blood-induced CQ-resistant rodent parasite in mice showed high chemo suppression in the range 51% -75%	(122)
Antioxidant	R. alaterus	methanol	Root bark and leaves	DPPH , Xanthine Oxidase (XOD) and Superoxide anion scavenging effects	The root bark and Leaves extract exhibited DPPH activities with IC_{50} values of 7.21 and 18.84 µg/mL, respectively, when compared to IC_{50} value of 3 µg/mL exhibited by a-tocopherol the positive control. Similarly, the leaves and root bark extract exhibited xanthine oxidase (XOD) inhibiting with respective IC_{50} values of 103.96 and 83.33 µg/mL and superoxide anion scavenging effects with IC_{50} values of 171 and 92 µg/mL as compared with the positive control allopurinol having	(125)

					IC50 value of 37.3 and 6 μg/mL respectively	
Antioxidant	R. petiolaris	Methanol and aqueous	fruit	DPPH and ABTS assay	Extracts exhibited activities with 470.96 mg trolox equivalent (TE)/g and 394.96 mg TE/g) respectively	(123)
Antioxidant	R. alaternus	methanol	leaves	DPPH, Xanthine Oxidase and Superoxide anion scavenging effects	Rhamnetin-3-O- β -isorhamninoside (38) exhibited DPPH activities with IC ₅₀ value of 1.5 µg/mL which is more antioxidant activity as compared to IC ₅₀ value of 3 µg/mL exhibited by vitamin E the standard drug. Similarly, kaempferol 3-O- β -isorhamninoside (31), rhamnocitrin-3-O- β -isorhamninoside (36) and rhamnetin-3-O-isorhamninoside (37) exhibited xanthine oxidase (XOD) inhibiting with respective IC ₅₀ values of 18, 81 and 40 µg/mL and superoxide anion scavenging effects with IC ₅₀ values of 42, 79 and 35µg/mL as compared with the positive control allopurinol having IC ₅₀ value of 37 and 4µg/mL respectively	(72)
Antioxidant	R, alaternus	methanol	leaf	Super oxide radical scavenging	Kaempferol 3-O- β -isorhamninoside (31) and rhamnocitrin 3-O- β -isorhamninoside exhibited activities with IC ₅₀ values 18.75 and 22.5 μg/mL respectively	(73)
Antioxidant	R. nakaharai	methanol	Heart wood	DPPH assay	The isolated compound, alaternin ($\bf{10}$) exhibited activities with IC ₅₀ value of 117.7 μ M, which was moderate activities compared to IC ₅₀ value of 63.7 μ M exhibited by ascorbic acid the standard drug	(70)
Antioxidant	R. nakaharai		Root bark	Iron induced lipid peroxidation	The isolated compound, isotorachrysone ($\bf 55$) exhibited activities with IC ₅₀ value of 1.64 µM, which was comparable to IC ₅₀ value of 1.08 µM exhibited by the standard butylated hydroxyl toluene (BHT) and was more potent than a - tocophenol and desferrioxamine with IC ₅₀ values of 3.71 and 97.10 µM respectively	(111)
Antioxidant	R. alaternus	ethanol	leaves	DPPH assay	extract exhibited activities with IC_{50} value of 58 µg/mL, which has good antioxidant activities when compared to IC_{50} value of 31 µg/mL exhibited by butylatedhydroxytoluene (BHT), the positive control	(96)
Antioxidant	R. alaterus	Aqueous and ethyl acetate fraction	leaves	Xanthine Oxidase (XOD) and Super oxide anion scavenging	aqueous extract and ethyl acetate fraction exhibited high xanthine oxidase inhibiting with respective IC ₅₀ values of 208 and 137μg/mL, and super oxide anion scavenging effects with IC ₅₀ values of 132 and 117μg/mL	(124)
Antioxidant	R. alaterus	Methanol- acetone	Leaf and stem bark	DPPH assay and ferric reducing antioxidant potential	leaves and stem bark extracts exhibited DPPH activities with IC_{50} values of 10.5 and 51.2 μ g/mL respectively, which was weak activity compared to IC_{50} value of 5.6 μ g/mL exhibited by BHA the positive control and FRAP	(127)

Antioxidant	R. alaternus		leaves	Cupric reducing antioxidant (CUPRAC), reducing power assay and ferric reducing	activities with EC ₅₀ values of 0.4 and 1.8 μ g/mL respectively, which was weak activity compared to EC ₅₀ value of 0.1 μ g/mL exhibited by BHA the positive control Kaempferol 3-O- β -isorhamninoside (31) and rhamnocitrin 3-O- β -isorhamninoside (36) exhibited CUPRAC and reduced power assay activities with IC ₅₀ value of 1 mg/mL, while FRAP activities exhibited at the same concentration 1000 μ g/mL reduce a maximum of iron ion by 300 μ g/mL and 320 μ g/mL equivalent of Trolox	(126)
Authoritan	0 -/-/		D- d	antioxidant power (FRAP)	respectively	
Antioxidant	R. alaternus, R. Fallax, R. intermedia and R. pumila		Bark	beta-Carotene- linoleic acid, DPPH radical scavenging, reducing power assay and Chelating activity	respectively, which was greater activity compared to EC_{50} value of 852 µg/mL exhibited by ascorbic acid the positive control. Similarly, all the plant extracts exhibited activities using reducing power assay with EC_{50} values of 0.91, 1.99, 0.81 and 0.99 µg/mL respectively, which was comparable and greater activities compared to EC_{50} values of 7.53, 1.8 and 7.59 µg/mL respectively exhibited by BHA, quercetin and ascorbic acid as standard drugs	(86)
Antioxidant	R. prinoides	ethanol	Stem and stem bark	DPPH and ABTS assay	The semi purified extract exhibited DPPH activities with IC_{50} value of 0.2 mg/mL, which was more potent than the standard BHT having IC_{50} value of 0.286 mg/mL. Similarly, the crude extracts exhibit ABTS activities with IC_{50} value of 0.0596 mg/mL, which was comparable to IC_{50} value of BHT, the positive control	(116)
Antioxidant	R. triquetra	methanol	Aerial part	DPPH assay	crude extract and their fractions (ethyl acetate , n-butanol, chloroform and n-hexane) exhibited activities with IC $_{50}$ values of 70.26, 7.59, 37.98, 60.09 and 182.99 μ g/mL respectively, which was the ethyl acetate fraction, is more active compared to IC $_{50}$ value of 12.1 μ g/mL exhibited by the standard (BHT)	(128)
Antioxidant	R. alaternus	Methanol and aqueous	leaves	DPPH and β-carotene-linoleic acid assay	The methanol and aqueous extracts exhibited DPPH activities with IC_{50} values of 0.082 and 0.398 mg/mL respectively, which was the methanol extract, is more active compared to IC_{50} value of 0.032 mg/mL exhibited by the standard (BHT). Similarly, the methanol extract exhibit activities using β -carotene-linoleic acid assay with 89% inhibition, which was comparable to 99.2% inhibition by BHT the standard	(129)

Antioxidant	R. alaternus	methanol	leaves	Oxygen radical absorbance capacity assay (ORAC)	extract exhibited with 6.55 mmol TE/g extract, which was more active as compared to the standard TEAC with 0.75 mmol TE/g extract	(130)
Anti-hyperlipidemia	R. alaternus	methanol	leaves	Calculating Malondialdehyde in cultured K562 cells	kaempferol 3-O- β -isorhamninoside (31), rhamnocitrin-3-O- β -isorhamninoside (36) and rhamnetin-3-O-isorhamninoside (37) from methanol leaf extracts exhibited by calculating the values of malondialdehyde (MDA) in cultured K562 human chronic myelogenous leukemia cells with IC ₅₀ values of isolated compound 180,320 and 106 μg/mL, respectively	(72)
Anti-hyperlipidemia	R. alaternus	methanol	leaves	Using Hyperlipidemia rats	The Oral crude extract administration decreased blood levels of cholesterol and triacylglycerols in human hepatoma HePG2 and 3T3-L1 murine dipocyte cell hyperlipidemic rats model (by 60% and 70%, respectively, at 200 mg extract/kg)	(133)
Anti-proliferative	R. alaternus	methanol	Root bark and leaf	MTT assay	The root barks and leaf extract exhibited activities against K562 cells with IC_{50} value of 165 and 260.69 μ g/mL. Similarly the extracts exhibited activities against L1210 cells with IC_{50} value of 210.73 and 343.10 μ g/mL, respectively	(125)
Anti-proliferative	R. davurica	methanol	bark	sulforhodamine B (SRB)micro culture colorimetric assay	The extract exhibited activities against human cancer cell lines HT-29 and SGC-7901 with IC50 values of 24.96 and 89.53 μ g/mL. respectively	(84)
Anti-proliferative	R. davurica	methanol	bark	MTT colorimetric assay	Kaempferol (33) exhibited activities against human cancer cell lines HT-29, SGC-7901 and HePG2 with IC50 values of 25.7, 13.43 and 20 μg/mL respectively, while the compound apigenin (40) exhibited with IC50 values 19.79, 17.76 and 10.20 μg/mL respectively	(74)
Wound healing	R. prinoides	methanol	leaves	Excision and incision models in adult Swiss albino mice	Treatment with 5 % and 10 % (w/w) methanol extract ointment exhibited significant wound recovery activities in both excision and incision models	(131)
Cytotoxicity	R. frangula	hydroalco holic		MTT assay	Extract exhibited activities against breast cancer cell lines (MCF-7) with half maximal cytotoxic concentration (CC_{50}) value of 10 mg/mL.	(132)
Cytotoxicity	R. alaternus	Ethyl acetate and Total Oligomers	leaf	MTT assay	TOF extract exhibited activities against human chronic myelogenous K562 and murine leukemia L1210 with IC $_{50}$ values of 75 μ g/mL and 198 μ g/mL against K562 and L1210 cells respectively. Similarly, the ethyl acetate	(125)

		flavonoids			extract showed activities with IC_{50} values of 232 $\mu g/mL$ and 176 $\mu g/mL$ respectively	
Toxicity	R. davurica	ethanol	bark	Sulforhodamine B (SRB) microculture colorimetric assay	Extract exhibited activities against normal human hepatic cells (L-02) with IC $_{50}$ value of 229.19 $\mu g/mL$.	(84)
Cytotoxicity	R. alaternus	methanol	leaf	Alamar blue assay	kaempferol 3-O-β-isorhamninoside (31) and rhamnocitrin 3-O-β-isorhamninoside (36) exhibited activities against human lymphoblastoid TK6 cells, the compound neither K3O-ir nor R3O-ir reached 50% inhibition of TK6 cell proliferation	(75)
Cytotoxicity	R. nepalensis	methanol	fruit	MTT assay	The isolated compound, prinoidin (23) exhibited activities against KB (human epidermoid carcinoma of the mouth) with IC50 value of 0.045 μ M, which was four times more potent than the standard, doxorubicin having IC50 value of 0.2 μ M	(69)
Cytotoxicity	R. alaternus	methanol	bark	Trypan blue assay	extract exhibited activities against human monocytic leukemia cells (U937) with IC $_{50}$ values of 6.39 $\mu g/mL$, which was comparable to IC $_{50}$ value of 2.47 $\mu g/mL$ exhibited by taxol the standard drug	(130)
Anti-tyrosinase	R. nakaharai	methanol	Heart wood	Mushroom tyrosine inhibitory assay	6-methoxysorigenin (12) exhibited activities with IC_{50} value of 42.2 μ M, which was twofold inhibitory effect than the positive control kojil acid having IC_{50} value of 82.1 μ M	(70)
Anti-mutagenicity	R. alaternus	aqueous	leaf	Ames assay	The ethyl acetate fraction exhibited against <i>Salmonella Typhimurium</i> (TA98) strains with a dose of 250 µg/mL and 78% inhibition mutagenicity	(124)
Antigenotoxic	R. alaternus	methanol	leaf	SOS chromo test	kaempferol 3-O-β-isorhamninoside (31) and rhamnocitrin 3-O-β-isorhamninoside (36) on <i>E. coli</i> PQ37 at different concentration (1,5 and 10 μ g/mL) showed antigenotoxicity activities	(73)
Antigenotoxic	R. alaternus	methanol	leaf	Comet assay	Kaempferol 3-O-β-isorhamninoside (31) and rhamnocitrin 3-O-β-isorhamninoside (36) exhibited activities against human lymphoblastoid cells TK6 at the same tested concentration, the total DNA damage induced by K3O-ir and R3O-ir showed no significant difference was detected	(126)

CONCLUSION

Traditional medicine continues as an alternative care available for the majority of the developing countries due to its intrinsic qualities, unique and holistic approaches as well as its accessibility and affordability. The present review endeavors to up to date provide a comprehensive and compilation of documented traditional medicinal pharmacological phytochemicals and uses, activities of the genus and provided valuable information in support of its uses as an alternative future healthcare for practice. **Phytochemicals** including anthraquinones flavonoids are the most dominant compounds reported from the genus of which polyphenols were abundant with tremendous antioxidant, wound healing and antiinflammatory activities. The genus afforded exemplary drug leads such as 6methoxysorigenin (12) and prinoidin (23) with anti-tyrosinase and cytotoxicity as well as antioxidant drug leads such as Rhamnetin-3-O-βisorhamninoside (37) and isotorachrysone (55). Nevertheless, more attention should be paid to the considering its wide spectrum pharmacological properties. Further investigation should be conducted to evaluate promising cruds extracts as well as compounds in search for new drug candidates.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

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REFERENCES

- 1. Nigussie G. A review on traditionally used medicinal plants for scabies therapy in Ethiopia. ADV TRADIT MED (ADTM). 2021 Jun;21(2):199–208. LODI.
- 2. Nigussie G, Ibrahim F, Neway S. A Phytopharmacological Review on a Medicinal Plant: Cordia africana Lam. J Trop Pharm Chem. 2021 Jan 18;5(3):254–63. CODI>.
- 3. Anonymous. Plants of the World online [Internet]. 2021. <URL>.

- 5. Dafni A, Yaniv Z, Palevitch D. Ethnobotanical survey of medicinal plants in northern Israel. Journal of Ethnopharmacology. 1984 May;10(3):295–310. <DOI>.
- 6. Watson RR, Preedy VR, editors. Arab herbal medicine. In: Botanical medicine in clinical practice. Wallingford, UK; Cambridge, MA: CABI; 2008. p. 31–9. ISBN: 978-1-84593-413-2.
- 7. Moerman DE. Native American ethnobotany. Portland, Or: Timber Press; 1998. 927 p. ISBN: 978-0-88192-453-4.
- 8. Zevin IV, Altman N, Zevin LV. A Russian herbal: traditional remedies for health and healing [Internet]. Rochester, Vt.: Healing Arts Press; 1997 [cited 2021 Jul 29]. ISBN: 978-0-89281-549-4
- 9. Watson RR, Preedy VR, editors. Botanical medicine in clinical practice. Wallingford, UK; Cambridge, MA: CABI; 2008. 915 p. ISBN: 978-1-84593-413-2.
- 10. Page L. Detoxification: all you need to know to recharge, renew and rejuvenate your body, mind and spirit! Carmel Valley, CA: Traditional Wisdom, Inc.; 2002. ISBN: 978-1-884334-54-2.
- 11. Xutian S. Handbook of traditional chinese medicine. Singapore: World Scientific; 2014. ISBN: 978-981-4571-34-0.
- 12. Tucakov J. [Ethnophytotherapy of diabetes. Critical view on the use of medicinal plant extracts in our national medicine in the treatment of diabetes mellitus]. Srp Arh Celok Lek. 1978 Feb;106(2):159–73.
- 13. Chhabra S, Uiso F. Antibacterial activity of some Tanzanian plants used in traditional medicine. Fitoterapia. 1991;62(6):499–503.
- 14. Abdelkrim B, Khaled K, Miloud S, Imane D, Kheira A. Hepatoprotective effects of the decoction and macerated leaves of Rhamnus alaternus L. on rats exposed to carbon tetrachloride. J Pharmacognosy Phytother. 2015 Oct 31;7(10):253–62. DOI
- 15. Amabye TG. Evaluation of Phytochemical, Chemical Composition, Antioxidant and Antimicrobial Screening Parameters of Rhamnus prinoides (Gesho) Available in the Market of Mekelle, Tigray, Ethiopia. Nat Prod Chem Res [Internet]. 2016;04(01):1000198, 1-6. CDDI.
- 16. Montalvo A, Riordan E, Beyers J. Plant profile for Rhamnus crocea and Rhamnus ilicifolia. Native

- Plant Recommendations for Southern California Ecoregions. 2020;
- 17. Evans WC, Evans D. Trease and Evans pharmacognosy. Edinburgh; London; New York: Saunders Elsevier; 2009. ISBN: 978-0-7020-2933-2.
- 18. Kubitzki K. Flowering plants Dicotyledons: Celastrales, oxalidales, rosales, cornales, ericales [Internet]. 2004 [cited 2021 Jul 29]. ISBN: 978-3-662-07257-8.
- 19. Boukef K. Rhamnus alaternus. Essaydali. 2001;81:34–5.
- 20. Benarba B. Medicinal plants used by traditional healers from South-west Algeria: an ethnobotanical study. J Intercult Ethnopharmacol. 2016;5(4):320. <a href="Moleoner-Policy Republic Policy Republic Pol
- 21. Nazim B, Houari T, Ismail B. Ethnobotanical Survey of Some Plants Used in Tessala Region, Algeria. Current Perspectives on Medicinal and Aromatic Plants (CUPMAP). 2020 Apr 1;3(1):18–24. <DOI>.
- 22. Miara MD, Bendif H, Rebbas K, Rabah B, Hammou MA, Maggi F. Medicinal plants and their traditional uses in the highland region of Bordj Bou Arreridj (Northeast Algeria). Journal of Herbal Medicine. 2019 Jun;16:100262. < Northeast Algeria).
- 23. Calvo MI, Cavero RY. Medicinal plants used for cardiovascular diseases in Navarra and their validation from Official sources. Journal of Ethnopharmacology. 2014 Nov;157:268–73. <a href="mailto:color:blue:c
- 24. Menendez-Baceta G, Aceituno-Mata L, Molina M, Reyes-García V, Tardío J, Pardo-de-Santayana M. Medicinal plants traditionally used in the northwest of the Basque Country (Biscay and Alava), Iberian Peninsula. Journal of Ethnopharmacology. 2014 Feb;152(1):113–34. DOI>.
- 25. Leto C, Tuttolomondo T, La Bella S, Licata M. Ethnobotanical study in the Madonie Regional Park (Central Sicily, Italy)—Medicinal use of wild shrub and herbaceous plant species. Journal of Ethnopharmacology. 2013 Mar;146(1):90–112. <DOI>.
- 26. Akerreta S, Cavero RY, López V, Calvo MI. Analyzing factors that influence the folk use and phytonomy of 18 medicinal plants in Navarra. J Ethnobiology Ethnomedicine. 2007 Dec;3(1):16. DOI>..

- 27. Fortini P, Di Marzio P, Guarrera PM, Iorizzi M. Ethnobotanical study on the medicinal plants in the Mainarde Mountains (central-southern Apennine, Italy). Journal of Ethnopharmacology. 2016 May;184:208–18. DOI>.
- 29. Altundag E, Ozturk M. Ethnomedicinal studies on the plant resources of east Anatolia, Turkey. Procedia Social and Behavioral Sciences. 2011;19:756–77. CDOI.
- 31. Jarić S, Popović Z, Mačukanović-Jocić M, Djurdjević L, Mijatović M, Karadžić B, et al. An ethnobotanical study on the usage of wild medicinal herbs from Kopaonik Mountain (Central Serbia). Journal of Ethnopharmacology. 2007 Apr;111(1):160–75. <u>DOI></u>.
- 32. Šamec D, Kremer D, Grúz J, Jurišić Grubešić R, Piljac-Žegarac J. Rhamnus intermedia Steud. et Hochst.—a new source of bioactive phytochemicals. Croatica Chemica Acta. 2012;85(2):125–9. <URL>.
- 33. Šarić-Kundalić B, Dobeš C, Klatte-Asselmeyer V, Saukel J. Ethnobotanical survey of traditionally used plants in human therapy of east, north and north-east Bosnia and Herzegovina. Journal of Ethnopharmacology. 2011 Feb;133(3):1051–76. <a href="mailto:color:black:col
- 34. Menković N, Šavikin K, Tasić S, Zdunić G, Stešević D, Milosavljević S, et al. Ethnobotanical study on traditional uses of wild medicinal plants in Prokletije Mountains (Montenegro). Journal of Ethnopharmacology. 2011 Jan;133(1):97–107. <DOI>.
- 35. Mohagheghzadeh A, Faridi P, Shams-Ardakani M, Ghasemi Y. Medicinal smokes. Journal of Ethnopharmacology. 2006 Nov;108(2):161–84. DOI>..
- 36. Bozyel ME, Merdamert Bozyel E, Canli K, Altuner EM. Türk Geleneksel Tıbbında Tıbbi Bitkilerin Antikanser Kullanımları. Kahramanmaraş Sütçü İmam Üniversitesi Tarım ve Doğa Dergisi. 2019 Dec 31;22(Suppl2):465–84.

- 37. Purkayastha J, Nath SC, Islam M. Ethnobotany of medicinal plants from Dibru-Saikhowa Biosphere Reserve of Northeast India. Fitoterapia. 2005 Jan;76(1):121-7. < DOI>.
- 38. Fakir H, Korkmaz M, Güller B. Medicinal Plant Diversity of Western Mediterrenean Region in Turkey. Journal of Applied Biological Sciences. 2019;3(2):33–43. <a href="https://www.uks.ncbe.new.org/linearin
- 39. Maleki T, Akhani H. Ethnobotanical and ethnomedicinal studies in Baluchi tribes: A case study in Mt. Taftan, southeastern Iran. Journal of Ethnopharmacology. 2018 May;217:163–77. <a href="mailto:color:blue
- 40. Nankaya J, Nampushi J, Petenya S, Balslev H. Ethnomedicinal plants of the Loita Maasai of Kenya. Environ Dev Sustain. 2020 Mar;22(3):2569–89. <u>OOI></u>.
- 41. Muthee JK, Gakuya DW, Mbaria JM, Kareru PG, Mulei CM, Njonge FK. Ethnobotanical study of anthelmintic and other medicinal plants traditionally used in Loitoktok district of Kenya. Journal of Ethnopharmacology. 2011 Apr;135(1):15–21. <DOI>...
- 42. Nankaya J, Gichuki N, Lukhoba C, Balslev H. Medicinal Plants of the Maasai of Kenya: A Review. Plants. 2019 Dec 27;9(1):44. <a h
- 43. Mesfin F, Seta T, Assefa A. An ethnobotanical study of medicinal plants in Amaro Woreda, Ethiopia. Ethnobotany Research and Applications. 2014;12:341–54.
- 44. Elgorashi E. Screening of medicinal plants used in South African traditional medicine for genotoxic effects. Toxicology Letters. 2003 Jul 20;143(2):195–207. Logoration.
- 45. Seleteng Kose L, Moteetee A, Van Vuuren S. Ethnobotanical survey of medicinal plants used in the Maseru district of Lesotho. Journal of Ethnopharmacology. 2015 Jul;170:184–200. DOI>.
- 46. Njoroge GN, Bussmann RW. Traditional management of ear, nose and throat (ENT) diseases in Central Kenya. J Ethnobiology Ethnomedicine. 2006 Dec;2(1):54. <DOI>...
- 47. Kidane B, van Andel T, van der Maesen L, Asfaw Z. Use and management of traditional medicinal plants by Maale and Ari ethnic communities in southern Ethiopia. J Ethnobiology Ethnomedicine. 2014;10(1):46. \leq DOI>.
- 48. Teka A, Asfaw Z, Demissew S, Van Damme P. Traditional uses of medicinal plants practiced by

- the indigenous communities in Gurage Zone, south central Ethiopia. Ethnobotany Research and Applications. 2020;19:1–31.
- 49. Tamene S. Ethnobotanical study of indigenous knowledge on medicinal plant uses and threatening factors around the Malga District, Southern Ethiopia. International Journal of Biodiversity and Conservation. 2020;12(3):215–26.
- 50. Bitew H, Gebregergs H, Tuem KB, Yeshak MY. Ethiopian medicinal plants traditionally used for wound treatment: a systematic review. Ethiopian Journal of Health Development. 2019;33(2).
- 51. Araya S, Abera B, Giday M. Study of plants traditionally used in public and animal health management in Seharti Samre District, Southern Tigray, Ethiopia. J Ethnobiology Ethnomedicine. 2015 Dec;11(1):22. DOIS.
- 52. Teklehaymanot T, Giday M. Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. J Ethnobiology Ethnomedicine. 2007 Dec;3(1):12. <DOI>...
- 53. Asmare T, Yilkal B, Mekuannint T, Yibeltal A. Traditional medicinal plants used to treat maternal and child health illnesses in Ethiopia: an ethnobotanical approach. J Tradit Med Clin Natur. 2018;7(277):2.
- 54. Tabuti JRS, Kukunda CB, Waako PJ. Medicinal plants used by traditional medicine practitioners in the treatment of tuberculosis and related ailments in Uganda. Journal of Ethnopharmacology. 2010 Jan;127(1):130-6. LDOI.
- 55. Mesfin K, Tekle G, Tesfay T. Ethnobotanical study of traditional medicinal plants used by indigenous people of Gemad District, Northern Ethiopia. Journal of Medicinal Plants Studies. 2013;1(4):32–7. <URL>...
- 56. Siyum D, Woyessa D. Assessment of bacteriological quality and traditional treatment methods of water-borne diseases among well water users in Jimma Town, South West Ethiopia. ARPN J Ag & Bio Sci. 2013;8:477–86.
- 57. Abdeta D, Amante M, Tamiru Y. Survey on Ethno Botany and Medicinal Animals at Sayo and Hawa Gelan Districts of Kelem Wollega Zone, Western Ethiopia. preventive medicine. 2020;28(2):21408–20.
- 58. Kamanja I, Mbaria J, Gathumbi P, Mbaabu M, Lanyasunya A, Gakuya D, et al. Medicinal plants used in the management of sexually transmitted infections by the Samburu Community Kenya. Int J Pharm Res. 2015;7:44–52.

- 59. Amuka O, Mbugua PK, Okemo PO. Ethnobotanical survey of selected medicinal plants used by the Ogiek communities in Kenya against microbial infections. Ethnobotany Research and Applications. 2014;12:627–41.
- 60. Garedew B, Bizuayehu B. A Review on Ethnobotanical Study of Traditional Medicinal Plants Used for Treatment of Liver Problems in Ethiopia. EJMP. 2018 Dec 1;26(1):1–18. \leq DOI>.
- 61. Malik ZA, Bhat JA, Ballabha R, Bussmann RW, Bhatt AB. Ethnomedicinal plants traditionally used in health care practices by inhabitants of Western Himalaya. Journal of Ethnopharmacology. 2015 Aug;172:133–44. CDOI.
- 62. Sharma P, Samant S. Diversity, distribution and indigenous uses of medicinal plants in Parbati Valley of Kullu district in Himachal Pradesh, Northwestern Himalaya. Asian J Adv Basic Sci. 2014;2:77–98.
- 63. Josabad Alonso-Castro A, Jose Maldonado-Miranda J, Zarate-Martinez A, Jacobo-Salcedo M del R, Fernández-Galicia C, Alejandro Figueroa-Zuñiga L, et al. Medicinal plants used in the Huasteca Potosina, México. Journal of Ethnopharmacology. 2012 Aug;143(1):292–8. DOI>.
- 64. Kiringe JW. A survey of traditional health remedies used by the Maasai of Southern Kaijiado District, Kenya. Ethnobotany Research and Applications. 2006;4:061–74. <u>kllptchar.</u>
- 65. Ajaib M, Khan Z, Khan N, Wahab M. Ethnobotanical studies on useful shrubs of district Kotli, Azad Jammu & Kashmir, Pakistan. Pak J Bot. 2010;42(3):1407–15. CURL>.
- 66. Phondani P, Maikhuri R, Bisht N. Medicinal plants used in the health care system practiced by traditional Vaidyas in Alaknanda catchment of Uttarakhand, India. Ethnobotanical Leaflets. 2009;2009(12):4. <u > Leaflets</u>.
- 67. Rokaya MB, Münzbergová Z, Timsina B. Ethnobotanical study of medicinal plants from the Humla district of western Nepal. Journal of Ethnopharmacology. 2010 Aug;130(3):485–504. <DOI>...
- 68. Rana TS, Datt B. E THNOBOTANICAL O BSERVATION AMONG J AUNSARIS OF J AUNSAR -B AWAR, D EHRA D UN (U.P.), I NDIA. International Journal of Pharmacognosy. 1997 Jan;35(5):371-4. <a href="mailto:color:blue)color: blue by the color: blue by th

- 69. Mai LP, Guéritte F, Dumontet V, Tri MV, Hill B, Thoison O, et al. Cytotoxicity of Rhamnosylanthraquinones and Rhamnosylanthrones from Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnosylanthrones from Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. J Nat Prod. 2001 Sep;64(9):1162–8. Cytotoxicity of Rhamnus nepalensis. Sep;64(9):116
- 70. Lu T-M, Ko H-H. A new anthraquinone glycoside from Rhamnus nakaharai and antityrosinase effect of 6-methoxysorigenin. Natural Product Research. 2016 Dec 1;30(23):2655–61. <DOI>...
- 71. Liao J-C, Deng J-S, Chiu C-S, Huang S-S, Hou W-C, Lin W-C, et al. Chemical Compositions, Anti-Inflammatory, Antiproliferative and Radical-Scavenging Activities of Actinidia callosa var. ephippioides. Am J Chin Med. 2012 Jan;40(05):1047–62. DOI>.
- 72. Ammar RB, Bhouri W, Sghaier MB, Boubaker J, Skandrani I, Neffati A, et al. Antioxidant and free radical-scavenging properties of three flavonoids isolated from the leaves of Rhamnus alaternus L. (Rhamnaceae): A structure-activity relationship study. Food Chemistry. 2009 Sep;116(1):258–64. <DOI>...
- 73. Bhouri W, Sghaier MB, Kilani S, Bouhlel I, Dijoux-Franca M-G, Ghedira K, et al. Evaluation of antioxidant and antigenotoxic activity of two flavonoids from Rhamnus alaternus L. (Rhamnaceae): Kaempferol $3\text{-O-}\beta\text{-isorhamninoside}$ and rhamnocitrin $3\text{-O-}\beta\text{-isorhamninoside}$. Food and Chemical Toxicology. 2011 May;49(5):1167–73. <DOI>...
- 74. Chen G, Wu J, Li N, Guo M. Screening for antiproliferative and anti-inflammatory components from Rhamnus davurica Pall. using bio-affinity ultrafiltration with multiple drug targets. Anal Bioanal Chem. 2018 Jun;410(15):3587–95. DOI>.
- 75. Bhouri W, Bouhlel I, Boubaker J, Kilani S, Ghedira K, Ghedira LC. Induction of apoptosis in human lymphoblastoid cells by kaempferol 3-O- β -isorhamninoside and rhamnocitrin 3-O- β -isorhamninoside from Rhamnus alaternus L. (Rhamnaceae): Induction of apoptosis in human lymphoblastoid cells. Cell Proliferation. 2011 Jun;44(3):283-90. DOI.
- 76. Raja WRT, Antony S, Entomology Research Institute, Loyola College, Chennai 600034, India, Pachaiyappan SK, Entomology Research Institute, Loyola College, Chennai 600034, India, Amalraj J, et al. Antibacterial Activity study of Musizin isolated from Rhamnus wightii Wight & Arn. Bioinformation. 2018 Dec 31;14(9):511–20. <DOI>...

- 77. Carranza MG, Sevigny MB, Banerjee D, Fox-Cubley L. Antibacterial activity of native California medicinal plant extracts isolated from Rhamnus californica and Umbellularia californica. Ann Clin Microbiol Antimicrob. 2015 Dec;14(1):29. DOI>..
- 78. Bribi N. Pharmacological activity of alkaloids: a review. Asian Journal of Botany. 2018;1:1–6.
- 79. Lin C-N, Chung M-I, Lu C-M. Anthraquinones from Rhamnus formosana. Phytochemistry. 1990;29(12):3903–5. CDOI>.
- 80. Ben Ammar R, Miyamoto T, Chekir-Ghedira L, Ghedira K, Lacaille-Dubois M-A. Isolation and identification of new anthraquinones from Rhamnus alaternus L and evaluation of their free radical scavenging activity. Natural Product Research. 2019 Jan 17;33(2):280–6. CDOI.
- 81. Hernández-Carlos B, Fernández R, Delgado F, Tamariz J, Zepeda LG, Joseph-nathan P. The Chemical Constituents of Rhamnus serrata var. serrata. Natural Product Letters. 1996 Feb;8(1):39–42. <DOI>...
- 82. Moussi K, Nayak B, Perkins LB, Dahmoune F, Madani K, Chibane M. HPLC-DAD profile of phenolic compounds and antioxidant activity of leaves extract of Rhamnus alaternus L. Industrial Crops and Products. 2015 Nov;74:858–66. <DOI>.
- 83. Wang L, Fan S, Wang X, Wang X, Yan X, Shan D, et al. Physicochemical Aspects and Sensory Profiles as Various Potential Factors for Comprehensive Quality Assessment of Nü-Er-Cha Produced from Rhamnus heterophylla Oliv. Molecules. 2019 Sep 4;24(18):3211. DOI>.
- 84. Chen G, Li X, Saleri F, Guo M. Analysis of Flavonoids in Rhamnus davurica and Its Antiproliferative Activities. Molecules. 2016 Sep 23;21(10):1275. CDOI.
- 85. Abegaz BM, Kebede T. Geshoidin: A bitter principle of Rhamnus prinoides and other constituents of the leaves. Bulletin of the Chemical Society of Ethiopia. 1995;9(2):107–14. URL>..
- 86. Kosalec I, Kremer D, Locatelli M, Epifano F, Genovese S, Carlucci G, et al. Anthraquinone profile, antioxidant and antimicrobial activity of bark extracts of Rhamnus alaternus, R. fallax, R. intermedia and R. pumila. Food Chemistry. 2013 Jan;136(2):335–41. <a href="mailto:cooling:c
- 87. Gonçalves RS, Silva EL, Hioka N, Nakamura CV, Bruschi ML, Caetano W. An optimized protocol for anthraquinones isolation from Rhamnus frangula L.

- Natural Product Research. 2018 Feb 1;32(3):366–9. \leq DOI>.
- 88. Moreira TF, Sorbo JM, Souza F de O, Fernandes BC, Ocampos FMM, de Oliveira DMS, et al. Emodin, Physcion, and Crude Extract of Rhamnus sphaerosperma var. pubescens Induce Mixed Cell Death, Increase in Oxidative Stress, DNA Damage, and Inhibition of AKT in Cervical and Oral Squamous Carcinoma Cell Lines. Oxidative Medicine and Cellular Longevity. 2018 Jul 3;2018:1–18. <DOI>...
- 89. Genovese S, Tammaro F, Menghini L, Carlucci G, Epifano F, Locatelli M. Comparison of three different extraction methods and **HPLC** determination of the anthraquinones aloeemodine, emodine, rheine, chrysophanol and physcione in the bark of Rhamnus alpinus L. (Rhamnaceae): Determination of Anthraquinones in Rhamnus Alpinus. Phytochem Anal. 2010 May;21(3):261-7. <DOI>...
- 90. Locatelli M, Genovese S, Carlucci G, Kremer D, Randic M, Epifano F. Development and application of high-performance liquid chromatography for the study of two new oxyprenylated anthraquinones produced by Rhamnus species. Journal of Chromatography A. 2012 Feb;1225:113–20. <DOI>...
- 91. Locatelli M, Tammaro F, Menghini L, Carlucci G, Epifano F, Genovese S. Anthraquinone profile and chemical fingerprint of Rhamnus saxatilis L. from Italy. Phytochemistry Letters. 2009 Nov;2(4):223–6. CDOI.
- 92. Abegaz B, Dagne E. Anthracene derivatives of Rhamnus prinoides. Bulletin of the Chemical Society of Ethiopia. 1988;2(1):15–20. <u > . <u >
- 93. Abegaz BM, Peter MG. Emodin and emodinanthrone rhamnoside acetates from fruits of Rhamnus prinoides. Phytochemistry. 1995 Aug;39(6):1411-4. <a href="https://doi.org/10.1001/journal.o
- 94. Hamed MM, Refahy LA, Abdel-Aziz MS. Evaluation of antimicrobial activity of some compounds isolated from Rhamnus cathartica L. Orient J Chem. 2015;31(2):1133–40.
- 95. Sharp H, Latif Z, Bartholomew B, Thomas D, Thomas B, Sarker SD, et al. Emodin and syringaldehyde from Rhamnus pubescens (Rhamnaceae). Biochemical systematics and ecology. 2001;29(1):113–5.
- 96. Zeouk I, Ouedrhiri W, Jiménez IA, Lorenzo-Morales J, Bazzocchi IL, Bekhti K. Intra-combined antioxidant activity and chemical characterization of three fractions from Rhamnus alaternus extract:

- Mixture design. Industrial Crops and Products. 2020 Feb;144:112054. <a href="Mailto:NOTON: NOTON: NOTO: NO
- 97. Dwivedi SPD, Pandey VB, Shah AH, Rao YB. Chemical constituents ofRhamnus procumbens and pharmacological actions of emodin. Phytother Res. 1988 Mar;2(1):51–3. CDOI.
- 98. Ben Ghezala H, Chaouali N, Gana I, Snouda S, Nouioui A, Belwaer I, et al. Toxic Effects of Rhamnus alaternus: A Rare Case Report. Case Reports in Emergency Medicine. 2015;2015:1–5. Localization-left:2015: Localization-left:2015: <a href="Localization-le
- 99. Epifano F, Genovese S, Kremer D, Randic M, Carlucci G, Locatelli M. Re-investigation of the Anthraquinone Pool of Rhamnus spp.: Madagascin from the Fruits of Rhamnus cathartica and R. intermedia. Natural Product Communications. 2012 Aug;7(8):1934578X1200700. DOI.
- 100. Cuoco G, Mathe C, Vieillescazes C. New emodin arabinoside acetates from fruits of Rhamnus alaternus. Current Topics in Phytochemistry, Volume 10. 2011;61–6.
- 101. Coskun M, Toshiko S, Hori K, Saiki Y, Tanker M. Anthraquinone glycosides from Rhamnus libanoticus☆. Phytochemistry. 1990;29(6):2018–20. <<u>DOI</u>>..
- 102. Coskun M, Tanker N, Sakushima A, Kitagawa S, Nishibe S. An anthraquinone glycoside from Rhamnus pallasii. Phytochemistry. 1984 Jan;23(7):1485–7. CDOI.
- 104. Lin C-N, Chung M-I, Gan K-H, Lu C-M. Flavonol and anthraquinone glycosides from Rhamnus formosana. Phytochemistry. 1991 Jan;30(9):3103–6. CDOI.
- 105. Özipek M, Çaliş İ, Ertan M, Rüedi P. Rhamnetin 3-p-coumaroylrhamninoside from Rhamnus petiolaris. Phytochemistry. 1994 Jan;37(1):249–53. CDOI.
- 106. Cuoco G, Mathe C, Vieillescazes C. Liquid chromatographic analysis of flavonol compounds in green fruits of three Rhamnus species used in Stil de grain. Microchemical Journal. 2014 Jul;115:130-7. < DOI>.
- 107. Payá M, Máñez S, Villar A. Flavonoid Constituents of Rhamnus lycioides L. Zeitschrift für Naturforschung C. 1986 Dec 1;41(11−12):976−8. < DOI>...

- 108. Sakushima A, Coşkun M, Hisada S, Nishibe S. Flavonoids from Rhamnus pallasii. Phytochemistry. 1983 Jan;22(7):1677–8. <a href="Do
- 109. Marzouk MS, El-Toumy SAA, Merfort I, Nawwar MAM. Polyphenolic metabolites of Rhamnus disperma. Phytochemistry. 1999 Nov;52(5):943–6. ODI.
- 110. Máñez S, Payá M, Terencio C, Villar A. Flavonoids of Rhamnus lycioides; Part 2. Planta Med. 1988 Apr;54(02):187–8. < CDOI>.
- 111. Hsiao G, Ko F-N, Lin C-N, Teng C-M. Antioxidant properties of isotorachrysone isolated from Rhamnus nakaharai. Biochimica et Biophysica Acta (BBA) Protein Structure and Molecular Enzymology. 1996 Nov;1298(1):119–30. DOI>.
- 112. Sut S, Dall'Acqua S, Ibrahime Sinan K, Bene K, Kumar G, Fawzi Mahomoodally M, et al. Cola caricifolia (G.Don) K. Schum and Crotalaria retusa L. from Ivory Coast as sources of bioactive constituents. Industrial Crops and Products. 2020 May;147:112246. DOI>...
- 113. Campbell M, Zhao W, Fathi R, Mihreteab M, Gilbert ES. Rhamnus prinoides (gesho): A source of diverse anti-biofilm activity. Journal of Ethnopharmacology. 2019 Sep;241:111955. <a href="Moleon Color: Note of the Color:
- 114. Chouitah O, Meddah B, Aoues A. Analysis of the Chemical Composition and Antimicrobial Activity of the Essential Oil from Rhamnus alaternus. Journal of Biologically Active Products from Nature. 2012 Jan;2(5):302–6. CDOI.
- 115. Thakur C, Prasad B. Antiinflammatory Activity of Stem Bark of Rhamnus purpureus. Int J Adv Microbiol Health Res. 2019;3(1):21–8.
- 116. Chen G-L, Munyao Mutie F, Xu Y-B, Saleri FD, Hu G-W, Guo M-Q. Antioxidant, Anti-inflammatory Activities and Polyphenol Profile of Rhamnus prinoides. Pharmaceuticals. 2020 Mar 26;13(4):55. <DOI>...
- 117. Molla Y, Nedi T, Tadesse G, Alemayehu H, Shibeshi W. Evaluation of the in vitro antibacterial activity of the solvent fractions of the leaves of Rhamnus prinoides L'Herit (Rhamnaceae) against pathogenic bacteria. BMC Complement Altern Med. 2016 Dec;16(1):287. COMPLE: COMPLE: COMPL: COMPLE: COMPLE: COMPLE: COMPLE: COMPLE: COMPLE: COMPLE: COMPL
- 118. Ben Ammar R, Kilani S, Bouhlel I, Skandrani I, Naffeti A, Boubaker J, et al. Antibacterial and cytotoxic activities of extracts from (Tunisian)Rhamnus alaternus (Rhamnaceae). Ann Microbiol. 2007 Sep;57(3):453–60. OOI>.

- 119. Koch A, Tamez P, Pezzuto J, Soejarto D. Evaluation of plants used for antimalarial treatment by the Maasai of Kenya. Journal of Ethnopharmacology. 2005 Oct; 101(1-3): 95-9. $\leq DOI > ...$
- 120. Bosire K. An investigation of a traditional herbal therapy used to treat malaria in Kisii [Internet] [Master of Science Thesis]. [Nairobi, Kenya]: University of Nairobi; 2003. .
- 121. Gebru M. Phytochemical and antiplasmodial investigation of rhamnus prinoides and kniphofia foliosa [Internet] [Master of Science Thesis]. [Nairobi, Kenya]: University of Nairobi; 2010. <ur>URL>...
- 122. Muregi FW, Ishih A, Suzuki T, Kino H, Amano T, Mkoji GM, et al. In Vivo antimalarial activity of aqueous extracts from Kenyan medicinal plants and their Chloroquine (CQ) potentiation effects against a blood-induced CQ-resistant rodent parasite in mice. Phytother Res. 2007 Apr;21(4):337–43. CDOI.
- 123. Rocchetti G, Miras-Moreno MB, Zengin G, Senkardes I, Sadeer NB, Mahomoodally MF, et al. UHPLC-QTOF-MS phytochemical profiling and in vitro biological properties of Rhamnus petiolaris (Rhamnaceae). Industrial Crops and Products. 2019 Dec;142:111856. <DOI>...
- 124. Ammar RB, Sghaier MB, Boubaker J, Bhouri W, Naffeti A, Skandrani I, et al. Antioxidant activity and inhibition of aflatoxin B1-, nifuroxazide-, and sodium azide-induced mutagenicity by extracts from Rhamnus alaternus L. Chemico-Biological Interactions. 2008 Jul;174(1):1–10. DOI.
- 125. Ben Ammar R, Kilani S, Bouhlel I, Ezzi L, Skandrani I, Boubaker J, et al. Antiproliferative, Antioxidant, and Antimutagenic Activities of Flavonoid-Enriched Extracts from (Tunisian) Rhamnus alaternus L.: Combination with the Phytochemical Composition. Drug and Chemical Toxicology. 2008 Jan;31(1):61–80. <a href="Moleonoon-english-bound-color: blue color: Logical Color: Logic
- 126. Bhouri W, Boubaker J, Kilani S, Ghedira K, Chekir-Ghedira L. Flavonoids from Rhamnus alaternus L. (Rhamnaceae): Kaempferol 3-O- β -isorhamninoside and rhamnocitrin 3-O- β -isorhamninoside protect against DNA damage in human lymphoblastoid cell and enhance antioxidant activity. South African Journal of Botany. 2012 May;80:57–62. DOI.
- 127. Chaouche TM, Haddouchi F, Boudjemai O, Ghellai I. Antioxidant and hemolytic activity of Ziziphus jujuba Mill and Rhamnus alaternus L (Rhamnaceae) extracts from Algeria. Activité antioxydante et hémolytique des extraits de

- Ziziphus jujuba Mill et Rhamnus alaternus (Rhamnaceae) d'Algérie. Bulletin de la Société Royale des Sciences de Liège [Internet]. 2020; <URL>...
- 128. Mazhar F, Jahangir M, Abbasi MA, Ilyas SA, Khalid F, Khanum R, et al. Rhamnus triquetra: A Valuable Source of Natural Antioxidants to Shield from Oxidative Stress. Asian Journal of Chemistry. 2013;25(15):8569–73.
- 129. Boussahel S, Dahamna S, Ruberto G, Siracusa L, Harzallah D. Phytochemical Study and Antioxidant Activities of Leaves Extracts from Rhamnus alaternus L. Pharmacognosy Communications. 2013;3(1):46–53.
- 130. Speciale A, Ferlazzo G, Harzallah D, Boussahel S, Dahamna S, Amar Y, et al. Flavonoid profile, antioxidant and cytotoxic activity of different extracts from Algerian Rhamnus alaternus L. bark. Phcog Mag. 2015;11(42):102. DOI>.
- 131. Tessema Z, Molla Y. Evaluation of the wound healing activity of the crude extract of root bark of Brucea antidysentrica, the leaves of Dodonaea angustifolia and Rhamnus prinoides in mice. Heliyon. 2021 Jan;7(1):e05901. <DOI>...
- 132. Ahmadi R, Hatami F, Molseghi M. The Effects of Co-administration of Frangula alnus and Rhamnus frangula Extract on Breast Cancer Cells in Cell Culture. In: BIOES-16 [Internet]. 2016. p. 78–80. <URL>...
- 133. Tacherfiout M, Petrov PD, Mattonai M, Ribechini E, Ribot J, Bonet ML, et al. Antihyperlipidemic effect of a Rhamnus alaternus leaf extract in Triton-induced hyperlipidemic rats and human HepG2 cells. Biomedicine & Pharmacotherapy. 2018 May;101:501–9. DOI>.

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