



The Effect of Lichen (*Xanthoparmelia somloensis*) Added in Broiler Rations on Performance Parameters and Organ Weights

Muhammet Ali TUNÇ^{1,*}, Mehmet Akif YÖRÜK², Ali ASLAN³

¹ Department of Animal Science, Narman Vocational College, Atatürk University, Erzurum, Turkey

² Department of Animal Nutrition and Nutritional Disorders, Veterinary Faculty, Atatürk University, Erzurum, Turkey

³ Department of Pharmacology, Faculty of Pharmacy, Van Yüzüncü Yıl University, Van, Turkey

*Corresponding author E-mail: matunc@atauni.edu.tr

HIGHLIGHTS

- > The effects of lichens on performance parameters in broiler nutrition were determined.
- > Lichens can be used as a feed additive in broiler nutrition.

ARTICLE INFO

Received : 10.14.2020

Accepted : 12.02.2020

Published : 12.15.2020

Keywords:

Broiler,
Lichen,
Performance

ABSTRACT

In this study, it was aimed to investigate the effect of feeding broilers with lichen supplemented feed on performance parameters and organ weight. 120 male Ross 308 animals (chicks) were used in the study. Chicks were randomly divided into 3 groups with 4 replications. There were 10 chicks in each subgroup. The control group (Group C) was formed with the group fed with additive-free feed. Lichen 1 group (Group L1) included the chicks fed with lichen added at the rate of 0.1% to their feed, while lichen 2 group (Group L2) included the ones fed with lichen added at the rate of 0.05% to their feed. In the 7-14 day period of the study, the highest body weight gain (BWG) and feed intake (FI) were in group L2 while the lowest feed conversion rate (FCR) was in the control group. When the whole study was evaluated, the highest FI and BW were in group L2 while the lowest FCR was determined in the control group. Group L2 had the lowest mortality rate in the study. The hottest and coldest carcass and neck, thigh, chest and heart weights in the study were observed in Group L2. Consequently, it was found that lichens can be used as antimicrobial feed additive to broiler rations at certain rates.

Contents

1. Introduction	18
2. Materials and Methods	18
2.1. Animals and Management	18
2.2. Experimental Design.....	18
2.3. Performance Measurements.....	18
2.4. Statistical Analysis.....	18
3. Results	19
4. Discussion	20
5. Conclusion.....	21
Acknowledgments	21
References	21

Cite this article Tunç MA, Yörük MA, Aslan A. The Effect of Lichen (*Xanthoparmelia somloensis*) Added in Broiler Rations on Performance Parameters and Organ Weights. *International Journal of Innovative Research and Reviews (INJIRR)* (2020) 4(2) 17-22

Link to this article: <http://www.injirr.com/article/view/62>



Copyright © 2020 Authors.

This is an open access article distributed under the [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/), which permits unrestricted use, and sharing of this material in any medium, provided the original work is not modified or used for commercial purposes.

1. Introduction

One of the main goals in animal feeding is to meet the nutritional needs of animals with minimum amount of feed raw materials without damaging the health of animals, and to obtain maximum yield and product [1]. Accordingly, the usage of antimicrobial feed additives gained importance at the end of the 20th century. Especially in the poultry industry, antimicrobials are highly preferred due to their disease preventive, therapeutic and growth promoting properties [2–4]. However, as a result of the excessive usage of antimicrobials, they generated residues in many animal products and this caused the development of cross-resistance against antibiotics and adverse effects on human health [5–7].

For this reason, the usage of antibiotics as feed additives was prohibited [8, 9] Thus, many aromatic plants and mushrooms, which have been used in alternative medicine since ancient times, started to be used as organic feed additives [10]. In this process, the possibility of using lichens, which are naturally found in nature and have many biological activities, as a feed additive emerged. Lichens are organisms that can survive by combining with a symbiotic relationship in environments where fungi and algae or cyanobacteria cannot live alone [11, 12].

Lichens may contain hundreds of bioactive components in their structures. These components may have a ratio between 0.1–30% of the dry weight of lichens [13]. Additionally, lichens contain polysaccharides and macrophages [14]. In a study on broilers, it was reported that lichens have antifungal [15], analgesic [16] and anti-inflammatory [17] effects, as well as anticoccidial effects [18] thanks to these components they have. It was also shown that a polysaccharide derived from lichen has antiviral activity against yellow fever virus and RNA viruses in poultry [19]. In another study, it was reported that usnic acid is effective in treating trichomoniasis in pigeons [20]. Moreover, it was reported that usnic and vulpinic acid, which are lichen compounds, have an antimicrobial effect against some Gram-positive (*Enterococcus faecalis*, *S. aureus*, *E. faecium*) bacteria [13, 21]. Considering all these features of lichens, it was thought that they can be used in poultry both for health and as a feed additive.

In this study, the effects of powdered lichens, which were added to broiler rations, on performance parameters and organ weights were determined.

2. Materials and Methods

2.1. Animals and Management

In the study was used of 120 male Ross 308 chicks. Chicks were purchased from a commercial enterprise with vaccinations at the age of 1 day. The trial lasted 42 days. The chicks were bred for adaptation in the first 7 days of the experiment, without grouping. Then the chicks were randomly divided into 3 groups with 4 replications. There are 10 chicks in each subgroup. The chicks were housed in floor cages measuring 1.5 m x 2.5 m, littered with 8cm deep wood shavings on the floor, a hanging drinker and a feeder. The ambient temperature was set to 33°C on the first day, then it

was stabilized by gradually lowering it to 23°C. During the study, water and feed were given to chicken *ad libitum*.

In the study was used *Xanthoparmelia somloensis* type lichen. Lichens were collected in Turkey's Erzurum province. Species of lichen were determined using various flora books and papers [22, 23]. The lichen dried at room temperature was powdered.

2.2. Experimental Design

The rates of lichen included in diets in the study are below the toxic effect limits [18, 24]. In the study, broilers were divided into 3 groups after 7 days of adaptation. The control group (Group C) was formed with the group fed without any feed additives in their feed (Table 1 and Table 2). Lichen 1 group (Group L1) was created by adding 0.1% lichen to feeds. Lichen 2 group (Group L2) was created by adding 0.05% lichen to feeds. Lichen was added to the feed at the rates specified in the commercial feed factory where the feed was prepared.

2.3. Performance Measurements

Chicks body weights and feed consumption were taken at the same day and time every week. Performance parameters (body weight gain and feed conversion rate) were calculated at 14, 21, 28, 35, 42 days and all of the work. Mortality was calculated between the 7th and 42nd days of the study.

2.4. Statistical Analysis

Statistical analysis was performed with SPSS package program version 20.00 with a General Linear Model. Ago group effects were tested, then post-hoc Tukey tests were used to compare group differences. Data are expressed as mean (M) ± standard deviation (SD).

Table 1 Composition of the experimental diet

Ingredients (%)	1-42 days
Maize	36.00
Full fat soybean meal	19.70
Dry soybean	17.20
Wheat	12.90
Vegetable fat	5.10
Poultry meal	3.00
Maize gluten	1.50
Meat and bone meal	2.45
DCP (Dicalcium phosphate)	0.73
Methionine	0.23
Vitamin-mineral premix	0.50
Sodium chloride	0.18
Sodium bicarbonate	0.15
Antitoxin	0.10
Choline chloride	0.09
Threonin	0.07
Lysine	0.10
TOTAL	100
C alculated Values	
Dry matter	89.20
Crude protein	22.05
Metabolisable energy (MJ/kg)	13.8
Crude fat	7.40
Crude fibre	3.78
Ca	0.88
P	0.44

Table 2 Supplied per kg of diet

Ingredient	Amount
Vitamin A	10000 IU
Vitamin E	12 mg
Vitamin D	2000 IU
Niacin	36 mg
D-pantothenic acid	10 mg
Riboflavin	3.61 mg
Pyridoxine	3.52 mg
Thiamine	2.41 mg
Folic acid	1.39 mg
Biotin	0.16 mg
Vitamin B	0.03 mg
Manganese	59 mg
Zinc	41 mg
Iron	1281 mg
Copper	7.9 mg
Iodine	0.31 mg
Selenium	0.22 mg

3. Results

In the 7-14 day period of the study, the highest body weight gain (BWG) and feed intake (FI) were in group L2, while the lowest feed conversion rate (FCR) was in the control group ($P<0.05$). In the 15-21 day period, the best body weight (BW) and BWG were observed in group L2, and the highest FI and FCR were observed in group L1 ($P<0.05$). In the 22-28 days period, the highest values of BW and BWG and FI were obtained in group L2, and FCR was obtained in group L1 ($P<0.05$). In the 29-35 day phase of the trial, the lowest BW and FCR, the highest BWG and FI were in group L1 ($P<0.05$). In the 36-42 days period, L2 had the heaviest BW while the least FI was in the control group ($P<0.05$). During this period, there was no significant difference in BWG and FCR ($P<0.05$). When the whole study (1-42 days) was evaluated, it was seen that the highest BW and FI were in group L2, while the lowest FCR was in the control group. Furthermore, the lowest mortality rate in the study was group L2 (Table 3)

Table 3 Performance parameters

	Control group	Group L1	Group L2	P
Starting weight	48.2	46.8	47.6	
7-14 days				
BW (g)	355.75±13.24	344.75±15.04	356.0±15.14	0.068
BWG (g/d)	189.38±4.17 ^a	180.38±3.58 ^b	192.50±4.84 ^a	0.041
FI (g/d)	260.87±5.21 ^b	271.87±5.11 ^a	278.66±6.47 ^a	0.027
FCR	1.38±0.03 ^b	1.51±0.04 ^a	1.45±0.03 ^{ab}	0.034
15-21 days				
BW (g)	700.25±21.18 ^b	693.0±20.54 ^b	723.02±19.82 ^a	0.037
BWG (g/d)	344.50±15.03 ^b	348.0±15.26 ^b	367.00±15.43 ^a	0.029
FI (g/d)	571.25±18.62 ^b	588.25±19.32 ^a	580.33±19.25 ^a	0.033
FCR	1.66±0.06 ^a	1.69±0.02 ^a	1.58±0.04 ^b	0.042
22-28 days				
BW (g)	1157.75±25.14 ^b	1090.25±26.07 ^c	1204.33±26.59 ^a	0.028
BWG (g/d)	457.50±16.81 ^b	397.25±15.23 ^c	481.33±17.08 ^a	0.035
FI (g/d)	755.75±19.47 ^b	737.0±18.51 ^c	789.67±18.79 ^a	0.021
FCR	1.65±0.04 ^b	1.85±0.03 ^a	1.64±0.03 ^b	0.032
29-35 days				
BW (g)	1616.75±28.05 ^b	1589.0±27.17 ^b	1676.0±28.84 ^a	0.019
BWG (g/d)	459±16.27 ^b	498.75±17.86 ^a	471.67±17.21 ^b	0.023
FI (g/d)	849.5±20.16 ^b	879.75±20.65 ^a	878.0±20.41 ^a	0.037
FCR	1.85±0.06 ^a	1.76±0.04 ^b	1.86±0.04 ^a	0.029
36-42 days				
BW (g)	2068.50±32.83 ^b	2052.75±32.57 ^b	2136.67±33.42 ^a	0.043
BWG (g/d)	451.75±15.73	463.75±15.33	460.67±15.81	0.087
FI (g/d)	899.50±21.49 ^c	939.75±22.58 ^a	918.0±22.88 ^b	0.027
FCR	1.99±0.08	2.03±0.12	1.99±0.11	0.062
Finishing data				
BW (g)	2068.50±32.83 ^b	2052.75±32.57 ^b	2136.67±33.42 ^a	0.043
FI (g/d)	3544.37±59.47 ^b	3691.06±59.15 ^a	3732.39±61.41 ^a	0.032
FCR	1.71±0.05 ^b	1.80±0.06 ^a	1.75±0.05 ^b	0.037
Mortality (%)	6	8	2	

Values are expressed as means±SEM. ^{a-c}Mean values within rows with different letters are significantly different

The hottest and coldest carcass and neck, leg, chest and heart weights in the study were in group L2, while gizzard was in the control group and liver was in group L1 ($P<0.05$). There

was no significant difference in gizzard and heart weights between the groups (Table 4)

Table 4 Organ weights

	Control group	Group L1	Group L2	P
Hot carcass (g)	1585±18.20 ^c	1620±18.57 ^b	1722±18.61 ^a	0.022
Cold carcass (g)	1556±18.03 ^c	1595±17.84 ^b	1696±18.27 ^a	0.036
Neck weight (g)	112.75±11.44 ^b	120.86±10.93 ^a	125.58±11.87 ^a	0.035
Leg weight (g)	586.50±10.11 ^b	618.26±11.67 ^a	625.00±11.34 ^a	0.031
Chest weight (g)	702.11±13.71 ^b	752.00±14.53 ^a	759.00±14.19 ^a	0.027
Liver weight (g)	42.90±1.73 ^b	51.24±1.67 ^a	44.82±1.82 ^a	0.043
Gizzard weight (g)	48.24±1.27	47.05±1.64	46.37±1.51	0.124
Heart weight (g)	11.04±0.33	12.65±0.41	12.71±0.37	0.092

Values are expressed as means±SEM. ^{a-c} Mean values within rows with different letters are significantly different.

4. Discussion

Recently, with the prohibition of the usage of antimicrobials as feed additives in terms of food safety and public health, local organic plants that do not require high costs started to be used as feed additives in animal diets. Lichens have been used as medicinal plants since ancient times and have been the subject of scientific studies [25]. Studies showed that lichens are mainly composed of pigments such as chitin (in hyphal walls), pectins, polyalcohols, lichen, isolycenin, hemicellulose, disaccharides, enzymes, amino acids, algal chromophores and chlorophyll, β carotenes, and xanthophylls [26]. As secondary metabolites, many polyphenolic compounds (gallic acid, usnic acid, epigallocatechin gallate, curcumin, quercetin, eugenol), lichesterinic acid and polysaccharide was found in its structure [27]. These phenolic compounds give lichens a powerful antioxidant property, thus they provide intracellular antioxidant-oxidant balance. If this balance is disrupted by the increase of reactive oxygen species (ROS) formed during cellular metabolism and the inadequacy of antioxidants that detoxify them, oxidative stress occurs [28], which negatively affects the integrity and functions of cell components, causing health deterioration [29]. Furthermore, phytochemicals such as phenolics and flavonoids, which are naturally found in the structure of lichens, was reported to increase the number of beneficial bacteria (probiotic effect) that prevent the proliferation of harmful bacteria in the intestines [30–32]. Thus, it is thought that it can provide better usage of nutrients by creating a positive effect on both cellular balance and intestinal health, and consequently, it increases body weight, body weight increase and feed consumption. Guven et al. (2016) reported that as a result of the relaxation of the intestinal mucosa in broilers, lichens improved the increase in body weight and feed intake and the feed conversion rate [18].

In addition to lichens, studies have been carried out in which many medicinal and aromatic plants with antimicrobial properties are used as feed additives. These studies have also reported that tarragon, which is included in the ration, does not affect the carcass weight of the broiler [33, 34], and there are studies reporting that it increases the carcass weight [35]. In another study, while sumac berries (*Rhus coriaria L.*) and thyme (rate of 2%) (*Thymus vulgaris*) added to broiler feed reduced feed intake, it had no effect on feed conversion rate and body weight, in the same study, it was reported that the addition of sumac also reduced the carcass weight [36]. Zhu et al. (2014) showed that thyme oil has no effect on feed consumption in chickens, but increases the growth rate [37].

In another study conducted on poultry, it was found that the application of plant extract as a feed additive did not affect the increase in body weight and feed intake, and it improved the feed conversion rate [38]. In another study, it was reported that St. John's Wort (*Hypericum perforatum*) powder added to broiler feed did not have a positive effect on growth performance, but it reduced live weight [39]. In addition, it was found that there was no significant difference in performance parameters of broiler chicks fed with anise extract (*Pimpinella anisum L.*) [40] and yarrow (*Achillea millefolium*) [41] supplemented feed.

Additionally, studies in rats [42] and cattle [43] reported a positive effect on body weight gain. The data obtained in this study are consistent with the literature. Moreover, hot-cold carcass, neck, thigh and breast weights analyzed in the study increased in parallel with the increasing body weight.

It was reported that lichens have an antibacterial effect against various pathogenic bacteria. It was determined that especially usnic acid, vulpinic acid, pulvinic acid, depsidones and lichesterinic acids, which are among the lichen components, have antibiotic properties. But more importantly, some lichen compounds were also found to act against bacteria resistant to some multidrug [44, 45]. *E. Faecalis*, one of the Gram-positive bacteria, causes many pathogenic conditions in poultry. In particular, it causes an increase in chick mortality rate [46], as well as pulmonary hypertension syndrome and amyloid atropia [47, 48]. However, usnic acid was reported to have a strong antibacterial effect, especially against *E. Faecalis* [13]. In a study, it was stated that a synergistic effect was tried to be generated by combining lichens with antibiotics. One of the most important features emerging from studies showed that it inhibits bacterial growth at much lower concentrations than other sources of antibiotic therapy [49, 50]. Thus, chickens, whose diets includes lichen, may have resistance to many bacterial infections, which may cause a decrease in the mortality rate in chickens. The low mortality rate in the study is considered as a reflection of this feature.

Different medicinal plant derivatives (powder, oil extract, hydraulic or organic extracts and infusion) were used on the organ weights of poultry. It has been reported that tarragon added to broiler feed has no effect on gizzard, heart and liver weights [33]. In another study using the tarragon additive diet, it was found that it did not affect intestinal length and weight, pancreatic and gizzard weights [34]. On the other hand, sumac berries (*Rhus coriaria L.*) and thyme (*Thymus vulgaris*) added to broiler feeds do not affect heart, neck and testicular weights, while sumac supplementation has been reported to be affected by the weight of the head, lungs and

kidneys [36]. In another study, it was reported that liver weight did not change in broilers fed with phytogetic extract supplemented feed [51]. Lichens were found to increase liver weight both in this study and in the study performed on rats [42]. It is thought that this result may be caused by the hepatotoxic effect of usnic acid, one of the lichen compounds, causing necrosis and causing dysfunction by affecting mitochondrial functions [52], as well as by the mobilization of lipids by the thermogenic effect of usnic acid and its accumulation on the ground tissue of the liver [42].

5. Conclusion

In conclusion, it is thought that lichens can be used in a moderate amount as feed additive to broiler rations. However, since the properties of lichens depend on the compounds they contain, it should be kept in mind that these compounds can change under the influence of many factors, including the geographical origin of the lichen, sampling time, drying and extraction method. Much more scientific studies are needed so that lichens can be used as the feed additives in poultry.

Acknowledgments

This study was conducted in accordance with ethical norms approved by Atatürk University Experimental Animal Education and Research Center Ethics Committee (No: 2011/181).

References

- [1] Yavuz HM. *Çiftlik hayvanlarının beslenmesinde temel prensipler ve karma yem üretiminde bazı bilimsel yaklaşımlar [Basic principles in feeding farm animals and some scientific approaches in compound feed production]*. İstanbul: Farmavet (2001).
- [2] Landoni MF, Albarellos G. The use of antimicrobial agents in broiler chickens. *The Veterinary Journal* (2015) **205**(1):21–27.
- [3] Agunos A, Léger D, Carson C. Review of antimicrobial therapy of selected bacterial diseases in broiler chickens in Canada. *The Canadian Veterinary Journal* (2012) **53**(12):1289.
- [4] Page SW, Gautier P, others. Use of antimicrobial agents in livestock. *Revue Scientifique et Technique-OIE* (2012) **31**(1):145.
- [5] Prestinaci F, Pezzotti P, Pantosti A. Antimicrobial resistance: a global multifaceted phenomenon. *Pathogens and global health* (2015) **109**(7):309–318.
- [6] Goetting V, Lee KA, Tell LA. Pharmacokinetics of veterinary drugs in laying hens and residues in eggs: a review of the literature. *Journal of veterinary pharmacology and therapeutics* (2011) **34**(6):521–556.
- [7] Reig M, Toldrá F. Veterinary drug residues in meat: Concerns and rapid methods for detection. *Meat science* (2008) **78**(1-2):60–67.
- [8] Newman KE. Antibiotic resistance is a reality: novel techniques for overcoming antibiotic resistance when using new growth promoters. Nutritional biotechnology in the feed and food industries. *Proceedings of Alltech's 18th Annual Symposium, Nottingham University Press* (2002):98–106.
- [9] European Commission. *Ban on antibiotics as growth promoters in animal feed enters into effect* (2005). Available from: https://ec.europa.eu/commission/presscorner/detail/en/IP_05_1687.
- [10] Pereira C, Barros L, Ferreira IC. A comparison of the nutritional contribution of thirty-nine aromatic plants used as condiments and/or herbal infusions. *Plant Foods for Human Nutrition* (2015) **70**(2):176–183.
- [11] Huneck S. The significance of lichens and their metabolites. *Naturwissenschaften* (1999) **86**(12):559–570.
- [12] Müller K. Pharmaceutically relevant metabolites from lichens. *Applied Microbiology and Biotechnology* (2001) **56**(1-2):9–16.
- [13] Basiouni S, Fayed MAA, Tarabees R, El-Sayed M, Elkhatam A, Töllner K-R, et al. Characterization of Sunflower Oil Extracts from the Lichen *Usnea barbata*. *Metabolites* (2020) **10**(9):353.
- [14] Ingoldsdottir K, Jurcic K, Fischer B, Wagner H. Immunologically active polysaccharide from *Cetraria islandica*. *Planta medica* (1994) **60**(06):527–531.
- [15] Schmieda-Hirschmann G, Tapia A, Lima B, Pertino M, Sortino M, Zacchino S, et al. A new antifungal and antiprotozoal depside from the Andean lichen *Protousnea poeppigii*. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives* (2008) **22**(3):349–355.
- [16] Okuyama E, Umeyama K, Yamazaki M, Kinoshita Y, Yamamoto Y. Usnic acid and diffractaic acid as analgesic and antipyretic components of *Usnea diffracta*. *Planta medica* (1995) **61**(02):113–115.
- [17] Vijayakumar CS, Viswanathan S, Reddy MK, Parvathavarthini S, Kundu AB, Sukumar E. Anti-inflammatory activity of (+)-usnic acid. *Fitoterapia* (2000) **71**(5):564–566.
- [18] Guven E, Avcioglu H, Aslan A, Hayirli A. Anticoccidial efficacy of usnic acid in broilers. *Kafkas Univ Vet Fak Derg* (2016) **22**(4):551–556.
- [19] Esimone CO, Ofokansi KC, Adikwu MU, Ibezim EC, Aboniyi DO, Odaibo GN, et al. In vitro evaluation of the antiviral activity of extracts from the lichen *parmelia perlata* (L) Ach. against three RNA viruses (2007).
- [20] Wu J, Zhang M, Ding D, Tan T, Yan B. Effect of *Cladonia alpestris* on *Trichomonas vaginalis* in vitro. *Zhongguo ji Sheng Chong xue yu ji Sheng Chong Bing za zhi= Chinese Journal of Parasitology & Parasitic Diseases* (1995) **13**(2):126–129.
- [21] Lauterwein M, Oethinger M, Belsner K, Peters T, Marre R. In vitro activities of the lichen secondary metabolites vulpinic acid,(+)-usnic acid, and (-)-usnic acid against aerobic and anaerobic microorganisms. *Antimicrobial agents and chemotherapy* (1995) **39**(11):2541–2543.
- [22] Aslan A, Öztürk A. Oltu (Erzurum) yöresine ait liken florası üzerine çalışmalar [Studies on lichen flora of Oltu (Erzurum) region]. *Turk J Bot* (1994) **18**:103–106.
- [23] Aslan A, Budak G, Karabulut A. The amounts Fe, Ba, Sr, K, Ca and Ti in some lichens growing in Erzurum province (Turkey). *Journal of Quantitative Spectroscopy and Radiative Transfer* (2004) **88**(4):423–431.
- [24] Abo-Khatwa AN, Al-Robai AA, Al-Jawhari DA. The Uncoupling of Oxidative Phosphorylation of Mouse-Liver Mitochondria in vivo by Usnic Acid. *Science* (2005) **17**(1).
- [25] Shukla V, Joshi GP, Rawat MS. Lichens as a potential natural source of bioactive compounds: a review. *Phytochemistry reviews* (2010) **9**(2):303–314.
- [26] Podterob AP. Chemical composition of lichens and their medical applications. *Pharmaceutical Chemistry Journal* (2008) **42**(10):582–588.
- [27] Halici M, Odabasoglu F, Suleyman H, Kahir A, Aslan A, Bayir Y. Effects of water extract of *Usnea longissima* on antioxidant enzyme activity and mucosal damage caused by indomethacin in rats. *Phytomedicine* (2005) **12**(9):656–662.
- [28] Özcan O, Erdal H, Çakırca G, Yönden Z. Oksidatif stres ve hücre içi lipid, protein ve DNA yapıları üzerine etkileri [Oxidative stress and its effects on intracellular lipid, protein and DNA structures] (2015).
- [29] McCord JM. The evolution of free radicals and oxidative stress. *The American journal of medicine* (2000) **108**(8):652–659.
- [30] Gupta VK, Sharma SK. Plants as natural antioxidants. *Natural Product Radiance* (2006) **5**(4):326–334.
- [31] Marin L, Miguélez EM, Villar CJ, Lombó F. Bioavailability of dietary polyphenols and gut microbiota metabolism: antimicrobial properties. *BioMed research international* (2015) **2015**.
- [32] Pacheco-Ordaz R, Wall-Medrano A, Goñi MG, Ramos-Clamont-Montfort G, Ayala-Zavala JF, González-Aguilar GA. Effect of phenolic compounds on the growth of selected probiotic and pathogenic bacteria. *Letters in applied microbiology* (2018) **66**(1):25–31.
- [33] Yıldırım F, Tunç MA. The effect of dietary tarragon (*Artemisia dracunculus*) powders in different levels on carcass characteristics and some internal organ's weight of broiler chickens. *Brazilian Journal of Poultry Science* (2018) **20**(1):179–182.
- [34] Hosseinzadeh Z, Moghaddam G. Effects of Tarragon Powders different Levels (*Artemisia Dracunculus*) on General Performance And Anatomic Properties of Digestive System of Male Broiler Chickens (2014).
- [35] Gharetappe FK, Hassanabadi A, Semnaninezhad H, Nassiry. The Effect of Dietary Tarragon (*Artemisia dracunculus*) and Peppermint

- (*Mentha piperita*) Leaves on Growth Performance and Antibody Response of Broiler Chickens. *Iranian Journal of Applied Animal Science* (2015) **5**(2).
- [36] Ahmadian A, Seidavi A, Phillips CJC. Growth, Carcass Composition, Haematology and Immunity of Broilers Supplemented with Sumac Berries (*Rhus coriaria* L.) and Thyme (*Thymus vulgaris*). *Animals* (2020) **10**(3):513.
- [37] Zhu X, Liu W, Yuan S, Chen H. The Effect of Different Dietary Levels of Thyme Essential Oil on Serum Biochemical Indices in Mahua Broiler Chickens. *Italian Journal of Animal Science* (2014) **13**(3):3238. doi:10.4081/ijas.2014.3238.
- [38] Ciftci M, Şimşek ÜG, Azman MA, Cerci IH, Tonbak F. The effects of dietary rosemary (*Rosmarinus officinalis* L.) oil supplementation on performance, carcass traits and some blood parameters of Japanese quail under heat stressed condition. *Kafkas Univ Vet Fak Derg* (2013) **19**(4):595–599.
- [39] Landy N, Ghalamkari GH, Toghyani M. Evaluation of St Johns Wort (*Hypericum perforatum* L.) as an antibiotic growth promoter substitution on performance, carcass characteristics, some of the immune responses, and serum biochemical parameters of broiler chicks. *Journal of Medicinal Plants Research* (2012) **6**(3):510–515.
- [40] Ciftci M, Guler T, Dalkılıç B, Ertas ON. The effect of anise oil (*Pimpinella anisum* L.) on broiler performance. *International Journal of Poultry Science* (2005) **4**(11):851–855.
- [41] Cross DE, McDevitt RM, Hillman K, Acamovic T. The effect of herbs and their associated essential oils on performance, dietary digestibility and gut microflora in chickens from 7 to 28 days of age. *British poultry science* (2007) **48**(4):496–506.
- [42] Al-Ahmadi AA, Al-Robai AA, Abo-Khatwa AN, Ali SS. Assessment of Usnic Acid (*Lichen Usnea Articulata* Extract) Safety on Lipid Profile, Adipocytes Morphology and Liver Functions in Adult Male Rats. *Journal of King Abdulaziz University: Medical Sciences* (2013) **20**(2):45–66.
- [43] Kuzmina IY, Ginter EV, Kuzmin AM. The effect of feed additives from mountain pine and lichens on the young cattle productivity in Magadan Region. *IOP Conference Series: Earth and Environmental Science* (2020) **547**(1):12020. doi:10.1088/1755-1315/547/1/012020.
- [44] Martins MCB, Lima MJG de, Silva FP, Azevedo-Ximenes E, Da Silva NH, Pereira EC. *Cladia aggregata* (lichen) from Brazilian northeast: chemical characterization and antimicrobial activity. *Brazilian archives of biology and technology* (2010) **53**(1):115–122.
- [45] Kokubun T, Shiu WKP, Gibbons S. Inhibitory activities of lichen-derived compounds against methicillin- and multidrug-resistant *Staphylococcus aureus*. *Planta medica* (2007) **73**(02):176–179.
- [46] Olsen RH, Frantzen C, Christensen H, Bisgaard M. An investigation on first-week mortality in layers. *Avian diseases* (2012) **56**(1):51–57.
- [47] Blanco AE, Barz M, Icken W, Cavero D, Mazaheri A, Voss M, et al. Twenty years of amyloid arthropathy research in chickens. *World's Poultry Science Journal* (2016) **72**(3):495–508.
- [48] Tankson JD, Thaxton JP, Vizzier-Thaxton Y. Pulmonary hypertension syndrome in broilers caused by *Enterococcus faecalis*. *Infection and immunity* (2001) **69**(10):6318–6322.
- [49] Weckesser S, Engel K, Simon-Haarhaus B, Wittmer A, Pelz K, Schempp C. Screening of plant extracts for antimicrobial activity against bacteria and yeasts with dermatological relevance. *Phytotherapy* (2007) **14**(7-8):508–516.
- [50] Gordien AY, Gray AI, Ingleby K, Franzblau SG, Seidel V. Activity of Scottish plant, lichen and fungal endophyte extracts against *Mycobacterium aurum* and *Mycobacterium tuberculosis*. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives* (2010) **24**(5):692–698.
- [51] Jamroz D, Orda J, Kamel C, Wiliczkiwicz A, Wartecki T, Skorupinska J. The influence of phyto-genic extracts on performance, nutrient digestibility, carcass characteristics, and gut microbial status in broiler chickens. *Journal of Animal and Feed Sciences* (2003) **12**(3):583–596.
- [52] Araújo AA, Melo MG de, Rabelo TK, Nunes PS, Santos SL, Serafini, et al. Review of the biological properties and toxicity of usnic acid. *Natural product research* (2015) **29**(23):2167–2180.