RESEARCH ARTICLE

GABAergic Effects of Some Food Extracts Via Inhibition of GABA-Transaminase

Sumeyye Sahin ^{1(ID)}, Sahra Haas ^{2(ID)}

¹Department of Food Engineering, Ordu University, Ordu, Turkey ²Food Chemistry Unit, Department of Chemistry and Pharmacy, Emil Fischer Center, Friedrich-Alexander Universität Erlangen-Nürnberg (FAU), Schuhstr, Germany

Copyright@Author(s) - Available online at https://dergipark.org.tr/en/pub/mbsjohs Content of this journal is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License,



Received: 03 August 2021, Accepted: 13 December 2021, Published online: 31 December 2021 © Ordu University Institute of Health Sciences, Turkey, 2021

Abstract

Objective: GABAergic system is a target for various groups of medications including sedatives, anxiolytics, muscle relaxants, antidepressants and antiepileptics. Several foods or food ingredients are able to affect the GABAergic system by the inhibition of the γ -aminobutyric acid (GABA) degrading enzymes including the GABA-transaminase and succinate semialdehyde dehydrogenase. The purpose of this study to investigate the inhibitory effects of tea (*Camellia sinensis*), coffee (*Coffea arabica L.*), peppermint (*Mentha piperita L.*), thyme (*Thymus vulgaris L.*), and cinnamon (*Cinnamonum zeylanicum*) on GABA degrading enzymes.

Methods: The inhibition of the GABA-T by aqueous extracts of tea (*Camellia sinensis*), coffee (*Coffea arabica L.*), peppermint (*Mentha piperita L.*), cinnamon (*Cinnamomum zeylanicum*), and thyme (*Thymus vulgaris L.*) was investigated using a fluorometric microplate enzyme assay. Dose-dependent inhibition of the GABA-degrading enzymes was attained by all the food extracts tested. For determination of the IC50 values of the extracts (\pm 95 % CI), a linear regression was performed using Origin® (Origin® 2015G von Origin Lab Corporation, Northampton, MA 01060 USA).

Results: The aqueous extract of black tea presented the strongest inhibitory activity with an IC50-value (half maximal inhibitory concentration) of 13.0 (11.0-15.3) μ g/mL. The tested food extracts were successful in inhibiting the GABA-degrading enzymes even at low concentrations.

Conclusion: In conclusion, the selected food extracts could serve as natural inhibitors for GABA-degrading enzymes thus, they could increase the GABA concentration in the brain.

Key words: γ -Amino butyric acid, GABA degrading enzymes, enzyme inhibition, black tea, thyme

Suggested Citation: Sahin S, Haas S. GABAergic effects of some food extracts via inhibition of GABA-transaminase. Mid Blac Sea Journal of Health Sci, 2021; 6(3):423-428.

Address for correspondence/reprints:

Sumeyye Sahin

Telephone number: +90 (452) 226 52 00-6305

E-mail: sumeyyesahin@odu.edu.tr

Introduction

GABA is formed from glutamate-by-glutamate decarboxylase, which is a pyridoxal phosphatedependent enzyme, and has two isoforms (65-kDa GAD and 67-kDa) in the human organism (1-5). The degradation of GABA (Fig. 1) occurs via a transamination reaction catalyzed by the GABA transaminase (GABA-T). GABA-T catabolizes GABA to the succinic semialdehyde that is either oxidized to the succinate by a reductase enzyme, the succinic semialdehyde dehydrogenase (SSA-DH; EC 1.2.1.24)) or reduced to the γ -hydroxybutyrate by the succinic semialdehyde reductase (SSA-R; EC 1.1.1.61)) (6-8).

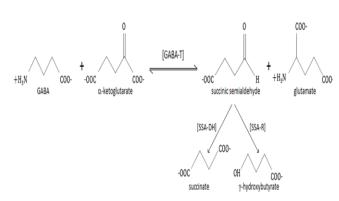


Figure 1. Schematic overview of degradation of neurotransmitter GABA; GABA-T: GABA transaminase; SSA-DH: Succinic semialdehyde dehydrogenase; SSA-R: Succinic semialdehyde reductase (6).

Low levels of GABA in the brain are associated with some neurophysiological diseases such as epilepsy, depression, panic disorders, anxiety disorders or sleep disorders (9-15). In the human study, Wood et al. (13), reported that epileptic patients have significantly low levels of GABA in cerebrospinal fluid. In the brain of depressed patients, a significantly decreased GABA concentration was also detected (14). Since the permeability of the blood-brain barrier for GABA is very low (16, 17), GABA supplementation cannot lead to an increase in GABA levels in the brain. For example, no effect on the GABA concentration in the brain of rats could be demonstrated by intraperitoneal injection of GABA However, the increasing of GABA (18).concentration is possible by the inhibition of GABA degradation (19-23).

GABA-T is a target for antiepileptic drugs, which can increase GABA levels in the brain via inhibition of GABA-T, thereby eliciting an antiepileptic or anticonvulsant effect (19, 21). Some anticonvulsants such as vigabatrin, a vinyl derivative of GABA, act as a competitive inhibitor of GABA-T (7). In addition, the inhibitory effects of several anxiolytic plants on GABA-T have been reported (24-26). Since the inhibition of GABA-T causes an increase in GABA level in the brain, there is a growing interest in the inactivation of GABA-T by foods.

Although the anxiolytic, sedative, hypnotic, and calming effects of some food plants and their ingredients have been studied in numerous studies, their physiological targets have not been completely elucidated. For example, animal model studies demonstrated that epigallocatechin gallate, a major catechin found in green tea (*Camellia sinensis*), acts as sedatives and as hypnotics in the brain (27). Coffee

(*Coffea arabica L*) is consumed because coffee reduces stress, drowsiness, and neuralgia (28). Peppermint (*Mentha piperita L*.) has a relaxation effect on the muscular actions and secretory processes of the gastrointestinal tract, analgesic, and anesthetic effects in the central and peripheral nervous system (29). Some Thymus spp. are used to produce antiseptic, antispasmodic, antioxidative, and sedative effects (30). Cinnamon (*Cinnamonum zeylanicum*) exhibited anti-depressant and anti-anxiety like effect in mice (31). The present study evaluated the effects of tea, coffee, peppermint, thyme, and cinnamon on GABA-T to explain their physiological action mechanism in the central nervous system.

Methods

Chemicals

GABA, GABase from *Pseudomonas fluorescens*, and α -ketoglutarate were purchased from Sigma-Aldrich (Taufkirchen, Germany). β -nicotinamide adenine dinucleotide phosphate (NADP) disodium salt was obtained from AppliChem (Darmstadt, Germany) and vigabatrin from British Pharmacopoeia Commission Laboratory (London, United Kingdom).

Plant Material and Preparation of Extracts

Peppermint leaves (Mentha piperita L.), thyme leaves (Thymus vulgaris L.), black tea leaves (Camellia sinensis) and cinnamon powder (Cinnamomum zeylanicum) were obtained in crushed form from a local pharmacy. Roasted coffee beans (Coffea arabica L.) were finely ground before extraction. 100 mL of boiling water were added to 2.5 g of the test material. The mixtures were stirred (60 min at 90 °C) and filtered through filter paper. The solvents in the filtrates were removed by freezedrying (Lyophilizer Savant Novalyphe NL150). After lyophilization, the samples were kept at -20 °C and dissolved in water (1 mg/mL) before use.

Fluorometric Microplate Enzyme Assay

The activity of GABAse composed of GABA-T and SSADH was determined spectrophotometrically according to literature (32) with some modifications. The reaction mixture contained potassium pyrophosphate buffer (150 mM, pH= 8.0), α ketoglutarate (228,5 mM), GABA (873 mM), and GABAse (1,5-1,7 units/mg). The extracts were added to the reaction mixture and preincubated (30 min at 37 °C). After preincubation, NADP+ (26.1 mM) was added to the mixture and incubated (30 and 60 min at 37 °C). GABA-T activity was monitored by measuring the absorbance changes at 340 nm due to the reduction of NADP+. The absorbance was measured using a microplate spectrophotometer (μ Quant BioTek). Vigabatrin, an inhibitor of GABA-T, was used as the positive control. Water was used instead of the inhibitor for the negative control.

Statistical analysis

For determination of the IC50 values of the extracts (\pm 95 % CI), a linear regression was performed using Origin® (Origin® 2015G von OriginLab Corporation, Northampton, MA 01060 USA).

Results

Each food extract was tested for its inhibitory activity at various concentrations (4-180 µg/mL). The activity of GABA-T decreased with increasing concentrations of black tea extract (Fig. 2a). Similarly, all other plant extracts exhibited inhibitory effects on GABA-T activity in a dose-dependent manner (Fig. 2 b, c, d, e). At the concentration of 18 µg/mL, cinnamon powder, coffee beans, peppermint leaves and thyme leaves extracts led to very weak inhibition (5.1 %, 8 %, 12.8 % and 21.5 % inhibition of GABA-T, respectively), whereas the extract of black tea induced the strongest inhibitory effect (56.6 % inhibition). When black tea extract was tested at the lower concentrations of 18 µg/mL, it also showed high inhibition indicating that it is the most potent inhibitor. Black tea showed 74 % inhibition at the highest concentration tested (27) $\mu g/mL$). Interestingly, the extracts of thyme and coffee showed similar GABA-T inhibition effects (65 % inhibition) at the concentration of 108 µg/mL. Compared to thyme and coffee in the same concentration (108 μ g/mL), the extracts of peppermint and cinnamon induced lower activity (55 % and 34 % inhibition, respectively). The cinnamon extract exhibited only 55% inhibition at the highest concentration used (180 μ g/mL).

The inhibitory effect of each food extract was evaluated by determining its respective IC50 values (the half-maximal inhibitory concentration). The IC50 values of extracts ranged between 13 and 174.2 µg/mL (Table 1). The strongest inhibitory effect was seen with the aqueous extract of black tea (IC50=13.0 µg/mL). The extracts of coffee, peppermint and thvme showed fifty percent inhibition at concentrations lower than 100 µg/mL. On the other hands, cinnamon extract was used at a much higher concentration (174.2 µg/mL) to inhibit 50 % of the enzyme.

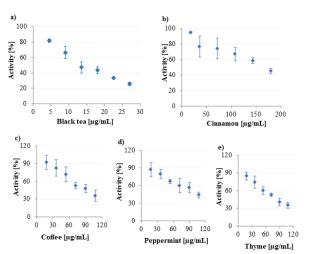


Figure 2. Dose response curve for the effect of the aqueous extract of black tea (a), cinnamon (b), coffee (c), peppermint (d), and thyme (e) on GABA-T activity expressed as percent of control. Values are the mean \pm SD from triplicate analysis

Table 1. IC50 of aqueous extracts on GABA-'	Т
---------------------------------------------	---

Botanical	IC ₅₀ (with 95 % confidence
	interval) [µg/mL]
Black tea	13.0 (11.0-15.3)
Thyme	72.6 (60.9-86.5)
Coffee	83.9 (71.1-98.9)
Peppermint	97.5 (81.3-116.9)
Cinnamon	174.2 (135.4-224.2)

Discussion

Vigabatrin, which was designed to increase the amount of GABA in the central nervous system (7), binds as an inhibitor selectively and irreversibly at the active site of GABA-T (33). As the positive control, vigabatrin was used at the concentration of 2.7 mM (348.7 μ g/mL) in the enzyme assay to confirm that detectable in vitro stimulation of total GABA-T inhibition could be measured under our experimental conditions. The food extracts showed their inhibitory effects in the concentrations of 4-180 μ g/mL despite using lower concentrations than vigabatrin.

Awad et al. (24) demonstrated that the inhibitory effects of the aqueous and ethanolic extracts of traditionally used anxiolytic botanicals (*Centella asiatica, Eschscholtzia californica, Humulus lupulus, Hypericum perforatum, Matricaria recutita, Melissa officinalis, Passiflora incarnata, Piper methysticum, Scutellaria lateriflora,* and *Valeriana officinalis*) on GABA-T. They reported that IC50 values of the anxiolytic botanicals ranged from 350 to > 4000 μ g/mL and the aqueous extract of M. officinalis showed the greatest inhibition of GABA-T with an IC50 of 350 μ g/mL (24). Similarly, the ethanolic extracts of 34 traditional Q'eqchi' Maya plants, which used to treat some mental diseases such as epilepsy and anxiety, were tested by Awad et al. (2009) for activity in the GABA-T. The IC50 value of the most active plant extract was 420 μ g/mL (34). Compared to these anxiolytic botanicals, the aqueous food extracts tested in this study exhibited a higher inhibitory activity against GABA-T with very low IC50-values (13.0-174.2 μ g/mL). Therefore, it can be concluded the aqueous food extracts (tea, thyme, coffee, peppermint, and cinnamon) may be more useful in the treatment of epilepsy and anxiety than the anxiolytic botanicals reported by Awad et al. 2007 and 2009.

Tea is consumed all over the world to relieve stress, drowsiness, and neuralgia (35). It is rich in flavonoids that have antioxidant, antitoxic. anticarcinogenic, antispastic, and antiviral activities. A study in the chick brain showed that epigallocatechin gallate, a major flavonoid of tea, acts as sedatives and as hypnotics in the brain, thereby moderating the acute stress response (27). Additionally, the anxiolytic effect of epigallocatechin gallate on mice has been reported (36). Furthermore, a human study described the anxiolytic effect of Ltheanine, which is an important bioactive compound of tea (37). Like tea, coffee is consumed because coffee reduces stress (28). Peppermint, one of the most popular single ingredient herbal teas, has a relaxation effect on the muscular actions and secretory processes of the gastrointestinal tract, analgesic, and anesthetic effects in the central and peripheral nervous system, immunomodulating actions and chemo preventive potential (29). The primary constituent of peppermint oil is menthol (38) which has analgesic (39) and anesthetic activities (40). Thymol, a structural analogue of menthol, is found in thyme that is used to produce the antiseptic, antispasmodic, antioxidative, and sedative effects (30). Cinnamon showed an anti-depressant-like and anti-anxiety like effects in animal studies (31).

Conclusion

In conclusion, since low levels of GABA in the brain are associated with some neurophysiological diseases such as epilepsy, depression, panic disorders, anxiety disorders, or sleep disorders (9-15), it can be deduced that these food plants (tea, thyme, coffee, peppermint, and cinnamon) may produce their reported sedative, anti-depressant, anti-anxiety or anxiolytic effects by increasing the level of GABA due to inhibition of GABA-T.

Acknowledgements

We thank Prof. Dr. Monika Pischetsrieder (Chair of Food Chemistry, Department of Chemistry and Pharmacy, Friedrich-Alexander University) for providing access to her lab for preparation of extracts and enzyme assay.

Ethics Committee Approval: Ethics committee approval is not required for this study.

Peer-review: Externally peer-reviewed.

Author Contributions:

Concept: S.S., S.H., Design: S.S., S.H; Literature search: S.S., S.H., Data Collection and Processing: S.S., S.H., Analysis or Interpretation: S.S., S.H., Writing: S.S., S.H.,

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: The authors declared that this study hasn't received no financial support.

References

- 1. Awapara J, Landua AJ, Fuerst R, Seale B. Free γaminobutyric acid in brain. J Biol Chem. 1950;187:35-39.
- Baxter CF, Roberts E. The γ-Aminobutyric Acidα-Ketoglutaric Acid Transaminase of Beef Brain. J Biol Chem. 1958;233(5):1135-1139.
- 3. Roberts E, Frankel S. γ-Aminobutyric acid in brain: Its formation from glutamic acid. J Biol Chem. 1950;187:55-63.
- 4. Wingo WJ, Awapara J. Decarboxylation of Lglutamic acid by brain. J Biol Chem. 1950;187:267-71.
- 5. Bu DF, Erlander MG, Hitz BC, Tillakaratne NJK, Kaufman DL, Wagner-McPherson CB, Evans G A,Tobin AJ. Two human glutamate decarboxylases, 65-kDa GAD and 67-kDa GAD, are each encoded by a single gene. Proc Natl Acad Sci USA. 1992; 89:2115-2119.
- Tsukatani T, Higuchi T, Matsumoto K. Enzymebased microtiter plate assay for γ-aminobutyric acid: Application to the screening of γaminobutyric acid-producing lactic acid bacteria. Analytica Chimica Acta. 2005;540(2):293-297.
- 7. Yogeeswari P, Sriram D, Vaigundaragavendran J. The GABA shunt: an attractive and potential therapeutic target in the treatment of epileptic disorders. Curr Drug Metab. 2005;6(2):127-139.

- 8. Beleboni RO, Carolino ROG, Pizzo AB, Castellan-Baldan L, Coutinho-Netto J, dos Santos WF. Coimbra NC. Pharmacological and **Biochemical** GABAergic Aspects of Pathological Neurotransmission: and Neuropsychobiological Relationships. Cellular and Molecular Neurobiology. 2004;24(6):707-728.
- Nitz D, Siegel JM. GABA release in the locus coeruleus as a function of sleep/wake state. Neuroscience. 1977;78(3):795–801.
- 10.Gottesmann C. Brain inhibitory mechanisms involved in basic and higher integrated sleep processes. Brain research Brain research reviews. 2004;45(3):230-249.
- 11.Streeter CC, Whitfield TH, Owen L, Rein T, Karri SK, Yakhkind A, Perlmutter R, Prescot A, Renshaw PF, Ciraulo DA, Jensen JE. Effects of yoga versus walking on mood, anxiety, and brain GABA levels: a randomized controlled MRS study. The Journal of Alternative and Complementary Medicine. 2010;16(11):1145-1152.
- 12. Chang L, Cloak CC, Ernst T. Magnetic resonance spectroscopy studies of GABA in neuropsychiatric disorders. The Journal of Clinical Psychiatry 2003;64 (3):7-14.
- 13.Wood JH, Hare TA, Glaeser BS, Ballenger JC, Post RM. Low cerebrospinal fluid γ-aminobutyric acid content in seizure patients. Neurology. 1979;29(9, Part 1):1203-1208.
- 14.Sanacora G, Mason GF, Rothman DL, Behar KL, Hyder F, Petroff OA, Berman RM, Charney DS, Krystal JH. Reduced cortical γ-aminobutyric acid levels in depressed patients determined by proton magnetic resonance spectroscopy. Arch Gen Psychiatry. 1999 56(11):1043-1047.
- 15.Rimón R, Lepola U, Jolkkonen J, Halonen T, Riekkinen P. Cerebrospinal fluid gammaaminobutyric acid in patients with panic disorder. Biol Psychiatry 1995;38(11):737-741.
- 16.Knudsen GM, Schmidt J, Almdal T, Paulson OB, Vilstrup H. Passage of Amino Acids and Glucose Across the Blood-brain Barrier in Patients with Hepatic Encephalopathy. Hepatology. 1993;17(6):987-992.
- 17.Meldrum BS. GABAergic mechanisms in the pathogenesis and treatment of epilepsy. Brit J Clin Pharmaco. 1989;27:3S-11S.
- Van Gelder NM, Elliott KAC. Disposition of γ-Aminobutyric Acid Administered to Mammals. Journal of Neurochemisrry. 1958;3:139-143.
- 19. Storici P, De Biase D, Bossa F, Bruno S, Mozzarelli A, Peneff C, Silverman RB, Schirmer

T. Structures of g-aminobutyric acid (GABA) aminotransferase, a pyridoxal 5'-phosphate, and [2Fe-2S] cluster-containing enzyme, complexed with g-ethynyl-GABA and with the antiepilepsy drug vigabatrin. J Biol Chem. 2004;279(1):363-373.

- 20.Schechter PJ, Tranier Y, Jung MJ, Böhlen P. Audiogenic seizure protection by elevated brain GABA concentration in mice: effects of γacetylenic GABA and γ-vinyl GABA, two irreversible GABA-T inhibitors. European Journal of Pharmacology. 1977;45 319-328.
- 21.Ben-Menachem E. Mechanism of action of vigabatrin: correcting misperceptions. Acta neurologica Scandinavica Supplementum. 2011;124(Suppl. 192):5-15.
- 22. Waterhouse EJ, Mims KN, Gowda SN. Treatment of refractory complex partial seizures: role of vigabatrin. Neuropsych Dis Treat. 2009;5 505– 515.
- 23.Beleboni RO, Carolino ROG, Pizzo AB, Castellan-Baldan L, Coutinho-Netto J, Dos Santos WF. Coimbra NC. Pharmacological and **Biochemical** Aspects of GABAergic Neurotransmission Pathological and Neuropsychobiological Relationships. Cellular and Molecular Neurobiology. 2004;24(6):707-728.
- 24. Awad R, Levac D, Cybulska P, Merali Z, Trudeau VL, Arnason J. T. Effects of traditionally used anxiolytic botanicals on enzymes of the g-aminobutyric acid (GABA) system. Can J Physiol Pharmacol. 2007;85(9):933-942.
- 25.Awad R, Muhammad A, Durst T, Trudeau VL, Arnason JT. Bioassay-guided Fractionation of Lemon Balm (Melissa officinalis L.) using an In Vitro Measure of GABA Transaminase Activity. Phytotherapy Research. 2009;23(8):1075–1081.
- 26.Awad R, Ahmed F, Bourbonnais-Spear N, Mullally M, Ta CA, Tang A, Merali Z, Maquin P, Caal F, Cal V, Poveda L, Vindas PS, Trudeau VL, Arnason JT. Ethnopharmacology of Q'eqchi' Maya antiepileptic and anxiolytic plants: Effects on the GABAergic system. Journal of Ethnopharmacology. 2009;125(2):257-264.
- 27. Adachi N, Tomonag S, Tachibana T, Denbow DM, Furuse M. (-)-Epigallocatechin gallate attenuates acute stress responses through GABAergic system in the brain. Eur J Pharmacol. 2006;531(1-3):171-175.
- 28.Hossain SJ, Aoshima H, Koda H, Kiso Y. Effects of coffee components on the response of GABAA receptors expressed in Xenopus oocytes. J Agr Food Chem. 2003;51:7568-7575.

- Mid Blac Sea J Health Sci 2021;7(3):423-428
- 29.McKay DL, Blumberg JB. A review of the bioactivity and potential health benefits of peppermint tea (Mentha piperita L.). Phytotherapy research: PTR. 2006;20(8):619-933.
- 30.Fachini-Queiroz FC, Kummer R, Estevao-Silva CF, Carvalho MD, Cunha JM, Grespan R, et al. Effects of Thymol and Carvacrol, Constituents of Thymus vulgaris L. Essential Oil, on the Inflammatory Response. Evid Based Complement Alternat Med. 2012;2012:657026.
- 31.Sohrabi R, Pazgoohan N, Seresht HR, Amin B. Repeated systemic administration of the cinnamon essential oil possesses anti-anxiety and antidepressant activities in mice. Iran J Basic Med Sci. 2017;20(6):708-714.
- 32.Jakoby WB. Enzymes of γ-Aminobutyrate Metabolism (Bacterial). Colowick SP, Kaplan, N.O., editor. New York: Academic Press; 1962. 765-778.
- 33.Lippert BMBW, Jung MJ, Casara P. 4-Aminohex-5-enoic Acid, a Selective Catalytic Inhibitor of 4-Aminobutyric-Acid Aminotransferase in Mammalian Brain. Eur J Biochem. 1977;74:441-445.
- 34.Awad R, Ahmed F, Bourbonnais-Spear N, Mullally M, Ta CA, Tang A, et al. Ethnopharmacology of Q'eqchi' Maya antiepileptic and anxiolytic plants: effects on the GABAergic system. J Ethnopharmacol. 2009;125(2):257-64.
- 35.Hossain SJ, Hamamoto K, Aoshima H, Hara Y. Effects of tea components on the response of GABAA receptors expressed in Xenopus Oocytes. J Agric Food Chem. 2002;50:3954–3960.
- 36. Vignes M, Maurice T, Lante F, Nedjar M, Thethi K, Guiramand J, Recasens M. Anxiolytic properties of green tea polyphenol (-)-epigallocatechin gallate (EGCG). Brain Res. 2006;1110(1):102-115.
- 37.Higashiyama A, Htay HH, Ozeki M, Juneja LR, Kapoor MP. Effects of L-theanine on attention and reaction time response. Journal of Functional Foods. 2011;3(3):171-178.
- 38.Ali B, Al-Wabel NA, Shams S, Ahamad A, Khan SA, Anwar F. Essential oils used in aromatherapy: A systemic review. Asian Pacific Journal of Tropical Biomedicine. 2015;5(8):601-611.
- 39.Haeseler G, Maue D, Grosskreutz J, Bufler J, Nentwig B, Piepenbrock S, Dengler R, Leuwer M. Voltage-dependent block of neuronal and skeletal muscle sodium channels by thymol and menthol. European Journal of Anaesthesiology 2002;19(8):571–579.

40.Galeotti N, Ghelardini C, Mannelli LDC, Mazzanti G, Baghiroli L, Bartolini A. Local Anaesthetic Activity of (+)- and (±)-Menthol. Planta medica. 2001;67 174-6.