



## IMMUNOMODULATORY POTENTIAL OF *CLAUSENA EXCAVATA* LEAVES FRACTIONS VIA DECREASING THE PRODUCTION OF REACTIVE OXYGEN SPECIES FROM IMMUNE CELLS

*CLAUSENA EXCAVATA* YAPRAK FRAKSİYONLARININ BAĞIŞIKLIK HÜCRELERİNDE REAKTİF OKSİJEN TÜRLERİNİN ÜRETİMİNİ AZALTARAK GÖSTERDİĞİ IMMÜNOMODÜLATÖR ETKİNLİK

Shaymaa Fadhel Abbas ALBAAYIT<sup>1\*</sup> , Rukesh MAHARJAN<sup>2</sup> 

<sup>1</sup>Department of Biology, College of Science, University of Baghdad, Baghdad, Iraq

<sup>2</sup>H.E.J Research Institute of Chemistry, International Center for Chemical and Biological Sciences, University of Karachi-75270, Karachi, Pakistan

### ABSTRACT

**Objective:** *Clausena excavata* is known to possess anti-oxidant property. However, this property through which mechanism it affects the immune cells and suppresses the production of reactive oxygen species (ROS) has not been explored.

**Material and Method:** This study evaluated the immunomodulatory activities of ethyl acetate, petroleum ether, chloroform, and methanol *C. excavata* leaf extracts by decreasing the production of ROS from whole blood, polymorphonuclears (PMNs) cells and macrophages.

**Result and Discussion:** Among the fractions tested, ethyl acetate *C. excavata* extract (EACE) showed potent anti-oxidant property and significantly ( $p < 0.001$ ) suppressed intracellular and extracellular phagocytic oxidative ROS burst produced by the zymosan and PMA-activated whole blood, PMNs, and macrophages cells with 50% inhibitory concentration ( $IC_{50}$ ) values of  $5.7 \pm 0.01$ ,  $1.3 \pm 0.01$ , and  $0.7 \pm 0.03$   $\mu\text{g/mL}$  respectively. This study provides information regarding the mechanism behind its anti-oxidant property and its herbal use in treating various higher oxidative stress associated diseases.

**Keywords:** *Clausena excavata*, inflammation, macrophage, ROS

### ÖZ

\* **Corresponding Author / Sorumlu Yazar:** Shaymaa Fadhel Abbas Albaayit  
**e-mail / e-posta:** shaymaa\_albaayit@yahoo.com, **Phone / Tel.:** +9647808430086

**Amaç:** *Clausena excavata*'ın anti-enflamatuvar etkinlik gösterdiği bilinmektedir. Bununla birlikte, bu özelliği, bağışıklık hücrelerine hangi mekanizma ile etki ettiği ve reaktif oksijen türlerinin (ROS) üretimini baskıladığı araştırılmamıştır.

**Gereç ve Yöntem:** Bu çalışmada, *C. excavata* yapraklarının etil asetat, petrol eteri, kloroform ve metanol ekstraktlarının tam kan, polimorfonkleer (PMN) hücreler ve makrofajlardan ROS üretimini azaltarak gösterdikleri immunomodülatör etki değerlendirilmiştir.

**Sonuç ve Tartışma:** Test edilen fraksiyonlar arasından, *C. excavata* etil asetat ekstresi en güçlü anti-enflamatuvar etkiyi göstermiş ve zimosan ve PMA tarafından aktive edilmiş tam kan, PMN ve makrofaj hücrelerinde, intraselüler ve ekstraselüler fagositik oksidatif ROS üretimini anlamlı oranda ( $p < 0.001$ ) baskılamış ve %50 inhibisyon konsantrasyonları ( $IC_{50}$ ) sırasıyla  $5.7 \pm 0.01$ ,  $1.3 \pm 0.01$ , and  $0.7 \pm 0.03$   $\mu\text{g/mL}$  olarak tespit edilmiştir. Bu çalışma, bitkinin anti-enflamatuvar aktivitesinin mekanizması ve çeşitli enflamatuvar rahatsızlıkların tedavisinde kullanımına dair bilgi sağlamaktadır.

**Anahtar Kelimeler:** *Clausena excavata*, enflamasyon, makrofaj, ROS

## INTRODUCTION

Prolonged inflammation plays detrimental role in the progression of various chronic diseases such as gastritis, atherosclerosis, cancer, diabetes, and other various diseases. Phagocytes, known for its role in first line of immune defense system provide protection in the site of inflammation by releasing sudden burst of reactive oxygen species so that it could counteract with various types of invading agents [1,2]. During higher oxidative stress, the antioxidant system of host cells and tissues could not counteract over produced ROS due to which excess ROS react with DNA, protein consequently leading to the progression of chronic inflammatory related diseases. In natural products, there are many compounds which possess antioxidant property therefore these can prevent oxidative stress and prevent damage to cells caused by foreign xenobiotics and thereby balancing oxidative stress level and prevent progression of inflammatory diseases [3-6].

*Clausena excavata* Burm .f. can be found in tropical and subtropical Asian regions [7]. In many countries, still the leaves of *C. excavata* are being practiced in traditional medicine to treat wound, abdominal pain, headache, diarrhea, and snake-bite [8]. Some of the major bioactive compounds like alkaloids, flavonoid, carbazole, glycosides, coumarins etc are reported to be present in the leaves of *C. excavata* [9]. Even though, many biological activities of *C. excavata* like anticancer, antiinflammatory, antioxidant, and antiulcer properties has been reported but till now [10], the immunomodulatory ability of this plant to decrease oxidative stress has not been reported yet. In pursuance to this unreported activity, we investigated the ROS inhibiting ability of *C. excavata* by using chemiluminescence assay and further gave more evidence regarding its potent antioxidant property.

## MATERIAL AND METHOD

### Extraction of *C. excavata*

*C. excavata* plant was collected and submitted to Biodiversity Unit, Institute of BioScience, Universiti Putra Malaysia. It was thoroughly evaluated by Dr. Shamsul Khamis, botanist and after authentication, a specimen voucher no: TI-013201-CE was issued. At room temperature, fresh leaves were dried, powdered, and extracted according to the procedure described previously [11]. Briefly, the extraction was done using 5:1 petroleum ether: dried plant (weight to volume) suspension for 4 days. Thus, the filtrate collected was subjected to further extraction with chloroform, ethyl acetate, and methanol. Filtrates of all extracts were dried by evaporating in a rotary evaporator under reduced pressure to obtain the crude extract.

### **Immunomodulatory studies**

#### **Estimation of intracellular ROS production**

Fresh human blood approximately 2-3 mL was withdrawn from a healthy person with consent which was approved from independent ethics committee with protocol no (IEC-047-HB-2019/PROTOCOL/1.0), University of Karachi. Ficoll-hypaque density gradient centrifugation method was used to isolate polymorphonuclear cells (PMNs). Equal volume (2 mL) of blood, lymphocytes separation medium, and Hank's Balance Salt solution (HBSS-- ) were added and mixed. RBCs were lysed using hypotonic solution for 1 min incubation so that lymphocytes purified will be freed from contaminating RBCs. Lysis process was stopped by adding HBSS solution. Thus, obtained PMNs were resuspended in (HBSS++) solution and adjusted to  $0.5-1 \times 10^6$  cells/ mL [12].

#### **Estimation of extracellular ROS production**

One Balb/c mice (25 g) was taken from animal house facility of ICCBS, with approved animal study protocol from animal house facility of ICCBS, University of Karachi. It was important to immunize mice therefore 1 mL of fetal bovine serum (FBS) was injected intraperitoneally. After 3 days, animal was sacrificed by cervical dislocation. Whole animal was dipped in 70% ethanol to sterilize whole body. After sterilization process, 5 mL of 10% complete RPMI medium was injected into the peritoneal cavity, massaged for 2 min and peritoneal cavity was exposed by cutting abdominal skin from lower side. Previously injected RPMI containing peritoneum exudate cells having macrophage was collected and centrifuged at 500 rpm for 5 min at 4°C. Cell pellet was formed at bottom, discarded supernatant and again cells were resuspended in incomplete RPMI medium containing HBSS++ solution. Equal volume (10  $\mu$ L) of macrophages and trypan blue was mixed and counted cells using hemocytometer, and cells concentrations adjusted to  $2 \times 10^6$  cells/mL [13].

#### **Chemiluminescence assay**

The protocol of Mesaik et al., 2012 was performed for luminol enhanced chemiluminescence assay. 1 mL of whole blood was diluted 20 times in sterile HBSS++. Briefly, 25  $\mu$ L of this diluted blood

suspension, 25  $\mu\text{L}$  of PMNs ( $1 \times 10^6$  cells/mL), 25  $\mu\text{L}$  of mice macrophage ( $2 \times 10^6$  cells/mL) in different white 96 well plate, were mixed with 25  $\mu\text{L}$  of different concentration of plant extract (1, 10, and 100  $\mu\text{g}/\text{mL}$ ) in triplicate and incubated at  $37^\circ\text{C}$ . Ibuprofen was used as a drug control. After 15 min incubation, 25  $\mu\text{L}$  of 0.3% serum opsonized zymosan and 25  $\mu\text{L}$  of luminol dye ( $7 \times 10^{-5}$  M) were added to those wells containing PMNs and whole human blood. Whereas 25  $\mu\text{L}$  of PMA dye and 25  $\mu\text{L}$  lucigenin dye were added to those wells containing macrophages. The final volume in each well became 100  $\mu\text{L}$ . Plate was inserted inside the luminometer and chemiluminescence was monitored as relative light units (RLU). The level of the ROS was recorded and inhibition of ROS production (%) was calculated using following formula and  $\text{IC}_{50}$  values were determined. Experiment was carried out in six wells per concentration and was done in triplicate in three different days.

$$\% \text{ inhibition of ROS production} = 100 - \frac{\text{Average reading of test plant extracts}}{\text{Average reading of positive control}} \times 100.$$

### Statistical analysis

One way (ANOVA) with Tukey's post-hoc test ( $p < 0.05$ ) was applied to compare the data (mean  $\pm$  SD) of treated samples with untreated control. All calculations were calculated by using GraphPad prism 6.0 statistical software.

## RESULT AND DISCUSSION

Since many centuries, different medicinal plants present in different geographical locations had been used locally as medicinal agent to treat various diseases, therefore, medical practitioners, chemical and biological scientists have been focusing in the area of ethnopharmacology to further explore undiscovered benefits of those plants, especially natural drug with potent immunomodulation activities by inhibiting the production of ROS from immune cells in inflammatory processes [14-24]. Previous studies reported that *C. excavata* can modulate the immune cell through different mechanism [7, 10]. In pursuance of those studies, this current study intends to evaluate the immunomodulation effects of *C. excavata* that could be potential treatment for chronic inflammatory diseases. Activator agents such as PMA/zymosan are always used for preliminary screening of the immunomodulatory efficiency of a product. Luminol and lucigenin chemiluminescence dye are used in measuring extracellular and intracellular ROS, respectively [12-14].

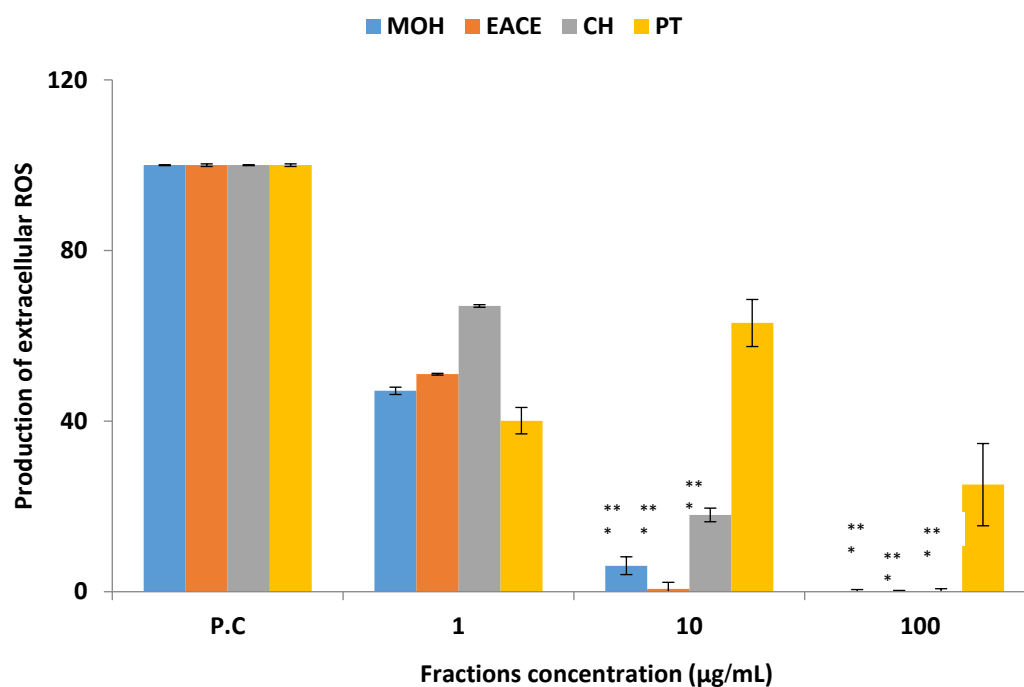
Methanol, chloroform, and ethyl acetate extracts exhibited a significant oxidative burst inhibition in whole blood, PMNs and macrophages. Table 1 shows the potent inhibition of whole blood and PMNs cells generated ROS. Among the tested extracts on whole blood for inhibiting ROS production, the ethyl acetate extract showed potent activity ( $p < 0.01, **$ ) with the lowest  $\text{IC}_{50}$  of  $< 10 \mu\text{g}/\text{mL}$ , followed by

methanol ( $20.2 \pm 0.3 \mu\text{g/mL}$ ) ( $p < 0.05, *$ ) and chloroform ( $27.9 \pm 0.4 \mu\text{g/mL}$ ) ( $p < 0.05, *$ ) fractions. In another experiment, ethyl acetate and chloroform showed significant ( $p < 0.001, ***$ ) inhibition of oxidative burst generated from zymosan-activated PMNs at lower concentrations ( $\text{IC}_{50}$  of  $1.3 \pm 0.3$  and  $2.1 \pm 0.1 \mu\text{g/mL}$  respectively).

**Table 1.** Intracellular ROS production after treatment with *C. excavata* fractions.

Fraction/Drug	WB ROS/ $\text{IC}_{50} \pm \text{SD}$ ( $\mu\text{g/mL}$ )	PMNs ROS/ $\text{IC}_{50} \pm \text{SD}$ ( $\mu\text{g/mL}$ )
Methanol	$20.2 \pm 0.3$ (*)	$7.1 \pm 0.05$ (*)
Ethyl acetate	$<10$ (**)	$1.3 \pm 0.3$ (***)
Chloroform	$27.9 \pm 0.4$ (*)	$2.1 \pm 0.1$ (***)
Petroleum ether	$93.1 \pm 0.5$	$33.1 \pm 2.1$
Standard (Ibuprofen)	$10.1 \pm 1.7$	$3.0 \pm 0.5$

WB: Whole Blood, ROS: Reactive Oxygen Species, PMNs: Polymorphonuclear cells (\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ )



**Figure 1.** Extracellular ROS production after treatment with *C. excavata* fractions

**Abbreviations:** P.C, positive control; MOH, methanol; EA, ethyl acetate; CH, chloroform; PT, petroleum ether.  
(\* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ )

The immunomodulatory activity of different fractions of *C. excavata* leaves to suppress extracellular ROS production from macrophages is presented in Fig. 1. Chloroform, methanol and ethyl acetate of *C. excavata* showed significant ( $p < 0.001$ ) decreasing of oxidative burst generated from PMA-activated macrophages ( $\text{IC}_{50}$   $2.3 \pm 0.01$ ,  $1.7 \pm 0.01$ , and  $0.7 \pm 0.03 \mu\text{g/mL}$  respectively).

The inhibited level of different extracts might be related to high phenolic contents that could be main contributor to antioxidant and immunomodulatory capacity of the extract, as previously reported publication [11]. The phenolic compounds can inhibit protein kinase C (PKC) and complement system; therefore, it is possible that these compounds present in this plant decrease ROS production by inhibiting NADPH oxidase enzyme [21-28]. Phenolic compounds also block the mitochondrial respiratory chain and ATPase [29]. Different polyphenolic compounds like curcumin [30], resveratrol [31], quercetin [32], ellagic acid [33], chlorogenic acid [34], exert their antioxidant properties by regulating the antioxidant enzyme genes through PKC signaling. The phytochemical analysis of essential oil obtained from the leaves of *C. excavata* through GC-MS showed anethole, and estragole as the major constituents and these are reported for their strong antioxidant properties [35]. Current findings of this study are in agreement with a previous outcome [11,36,37], in which LCMS/MS analysis of methanolic and ethyl acetate leaf extracts of *C. excavata* contained higher total phenolic contents, total flavonoid content, quercetin, furocoumarin, 8-geranyloxy psoralen, myricetin glucoside conjugate, kaempferol conjugate, caffeic acid and showed anti-inflammatory and antioxidant activities. Based on the outcomes in this study and those reported earlier, the leaves extract of *C. excavata* especially ethyl acetate extract has strong therapeutic to be developed into a controlling inflammation agent.

In conclusion, the ethyl acetate, chloroform, and methanol extract possess significant immunomodulatory activity as evidenced from decreased ROS production from activated immune cells. Thus, these extracts have potential to retard the progression of acute and chronic inflammatory conditions, support, and encourage for its traditional use in the folklore herbal medicine to treat inflammatory-related conditions.

## ACKNOWLEDGEMENTS

The corresponding author is thankful to Prof. Dr. M. Iqbal Choudhary for providing NAM-ICCBS fellowship, 2018 in International Center for Chemical and Biological Sciences, University of Karachi, Karachi, Pakistan

## AUTHOR CONTRIBUTIONS

Conception: *S.F.A.A.*; Design: *S.F.A.A.*; Supervision: *S.F.A.A., R.M.*; Resources: *S.F.A.A., R.M.*; Data Collection and/or processing: *S.F.A.A.*; Analysis and/or interpretation: *S.F.A.A.*; Literature search: *S.F.A.A.*; Writing manuscript: *S.F.A.A., R.M.*; Critical review: *S.F.A.A., R.M.*; Other: *R.M.*

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest for this article.

## ETHICS COMMITTEE APPROVAL

ICCBS/IEC-028-HB-2017/PROTOCOL/1.0, University of Karachi

## REFERENCES

1. Bryan, N., Ahswin, H., Smart, N., Bayon, Y., Wohlert, S., Hunt, J.A. (2012). Reactive oxygen species (ROS) –a family of fate deciding molecules pivotal in constructive inflammation and wound healing. *European Cells and Materials*, 24(249), e65. [\[CrossRef\]](#)
2. Albaayit, S.F.A., Al-Khafaji, A.S.K., Alnaimy, H.S. (2019). *In vitro* macrophage nitric oxide and interleukin-1 beta suppression by *Moringa peregrina* seed. *Turkish Journal of Pharmaceutical sciences*. 16(3), 362–365. [\[CrossRef\]](#)
3. Poljsak, B. (2011). Strategies for reducing or preventing the generation of oxidative stress. *Oxidative medicine and cellular longevity*, 2011. [\[CrossRef\]](#)
4. Albaayit, S.F.A., Abdullah, R., Abdullah, N. (2020). Zerumbone-loaded nanostructured lipid carrier gel facilitates wound healing in rats. *Revista Brasileira de Farmacognosia*, 30(2), 272–278. [\[CrossRef\]](#)
5. Mohammed, E. S., Al-Taie, W. F., Numan, I. T., Hussain, S. A. (2012). Changes in Serum Levels of Tumor Necrosis Factor-Alpha and Antioxidant status in Different Stages of Malignant Prostate Cancer Patients in Iraq. *Iraqi Journal of Pharmaceutical Sciences*, 21(1), 56-60. [\[CrossRef\]](#)
6. Ali, S. H., Hamadi, S. A., Al-Jaff, A. N. (2007). Effect of ergotamine and its combination with vitamin E or melatonin on total antioxidant status in migraine patients. *Iraqi Journal of Pharmaceutical Sciences*, 16(2), 27-33. [\[CrossRef\]](#)
7. Manosroi, A., Saraphanchotiwitthaya, A., Manosroi, J. (2003). Immunomodulatory activities of *Clausena excavata* Burm. f. wood extracts. *Journal of Ethnopharmacology*, 89(1), 155–160. [\[CrossRef\]](#)
8. Rahman, M.T., Alimuzzaman, M., Shilpi, J.A., Hossain, M.F. (2002). Antinociceptive activity of *Clausena excavata* leaves. *Fitoterapia*, 73(7-8), 701–703. [\[CrossRef\]](#)
9. Huang, L., Zhe-Ling, F.E.N.G., Yi-Tao, W.A.N.G., Li-Gen, L.I.N. (2017). Anticancer carbazole alkaloids and coumarins from *Clausena* plants: A review. *Chinese Journal of Natural Medicines*, 15(12), 881–888. [\[CrossRef\]](#)
10. Albaayit, S.F.A., Rasedee, A., Abdullah, N., Abba, Y. (2020). Methanolic extract of *Clausena excavata* promotes wound healing via antiinflammatory and anti-apoptotic activities. *Asian Pacific Journal of Tropical Biomedicine*, 10(5), 232-238. [\[CrossRef\]](#)
11. Albaayit, S.F.A., Khan, M., Rasedee, A., Noor, M.H.M. (2021). Ethyl acetate extract of *Clausena excavata* induces growth inhibition of non-small-lung cancer, NCI-H460, cell line via apoptosis. *Journal of Applied Biomedicine*, 19(1), 40–47. [\[CrossRef\]](#)

12. Orhan, I. E., Mesaik, M. A., Jabeen, A., & Kan, Y. (2016). Immunomodulatory properties of various natural compounds and essential oils through modulation of human cellular immune response. *Industrial Crops and Products*, 81, 117-122. [CrossRef]
13. Albaayit, S.F.A., Maharjan, R. (2018). Immunomodulation of zerumbone via decreasing the production of reactive oxygen species from immune cells. *Pakistan Journal of Biological Sciences*, 21(9), 475–479. [CrossRef]
14. Majdalawieh, A.F., Fayyad, M.W. (2015). Immunomodulatory and anti-inflammatory action of *Nigella sativa* and thymoquinone: A comprehensive review. *International Immunopharmacology*, 28(1), 295–304. [CrossRef]
15. Albaayit, S.F.A. (2021). Evaluation of anti-methicillin resistant *Staphylococcus aureus* property of *Clausena excavata* leaves by using atomic force microscopy and flowcytometry techniques. *Pakistan Journal of Agricultural Sciences*, 58(1), 315–320. [CrossRef]
16. Albaayit, S.F.A., Maharjan, R., Khan, M. (2021). Evaluation of hemolysis activity of Zerumbone on RBCs and brine shrimp toxicity. *Baghdad Science Journal*, 18(1), 65–69. [CrossRef]
17. Al-Bahrani, R. M., Radif, H. M., Albaayit, S. F. A. (2020). Evaluation of potent silver nanoparticles production from *Agaricus bisporus* against *Helicobacter pylori*. *Pakistan Journal of Agricultural Sciences*, 57(4), 1197–1201. [CrossRef]
18. Albaayit, S. F. A., Mariam, K. H. A. N., & Abdullah, R. (2021). Zerumbone induces growth inhibition of Burkitt's lymphoma cell line via apoptosis. *Natural Volatiles and Essential Oils*, 8(3), 56-63. [CrossRef]
19. Al-Ani, L. K. T., Yonus, M. I., Mahdii, B. A., Omer, M. A., Taher, J. K., Albaayit, S. F. A., Al-Khoja, S. B. (2018). First record of use *Fusarium proliferatum* fungi in direct treatment to control the adult of wheat flour *Tribolium confusum*, as well as, use the entomopathogenic fungi *Beauveria bassiana*. *Ecology, Environment and Conservation*, 24(3), 29-34.
20. Albaayit, S. F. A. (2021). Enzyme inhibitory properties of zerumbone. *Pakistan Journal of Agricultural Sciences*, 58(3), 1207-1209. [CrossRef]
21. Albaayit, S.F.A. (2020). *In Vitro* Evaluation of Anticancer Activity of *Moringa Peregrina* Seeds on Breast Cancer Cells. *The Eurasia Proceedings of Science Technology Engineering and Mathematics*, 11,163-166. <http://www.epstem.net/en/pub/issue/58065/838264>.
22. Albaayit, S.F.A., Maharjan, R., Abdullah, R., Noor, M.H.M. (2022). Evaluation of anti-methicillin-resistant *Staphylococcus aureus* property of zerumbone. *Journal of Applied Biomedicine*, 20(1), 22–36.
23. Albaayit, S. F. A., Abba, Y., Abdullah, R., & Abdullah, N. (2016). Prophylactic effects of *Clausena excavata* Burum. f. leaf extract in ethanol-induced gastric ulcers. *Drug Design, Development and Therapy*.
24. Albaayit, S. F. A., Abba, Y., Abdullah, R., & Abdullah, N. (2015). Effect of *Clausena excavata* Burm. f.(Rutaceae) leaf extract on wound healing and antioxidant activity in rats. *Drug Design, Development and Therapy*.



25. Yahfoufi, N., Alsadi, N., Jambi, M., Matar, C. (2018). The immunomodulatory and anti-inflammatory role of polyphenols. *Nutrients*, 10(11), 1618. [\[CrossRef\]](#)
26. Das, J., Ramani, R., Suraju, M.O. (2016). Polyphenol compounds and PKC signaling. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1860(10), 2107–2121. [\[CrossRef\]](#)
27. Albaayit, S. F. A., Maharjan, R., Abdullah, R., Noor, M. H. M. (2021). Anti-Enterococcus Faecalis, Cytotoxicity, Phytotoxicity, and Anticancer Studies on *Clausena excavata* Burum. f.(Rutaceae) Leaves. *BioMed Research International*, 2021. [\[CrossRef\]](#)
28. Albaayit, S.F.A., Khan, M., Rasedee, A., Noor, M.H.M. (2022). Zerumbone-Loaded Nanostructured Lipid Carrier Gel Enhances Wound Healing in Diabetic Rats. *BioMed Research International*, 2022.
29. Ahmad, Z., Hassan, S.S., Azim, S. (2017). A therapeutic connection between dietary phytochemicals and ATP synthase. *Current Medicinal Chemistry*, 24(35), 3894–3906. [\[CrossRef\]](#)
30. Pérez-Lara, Á., Corbalán-García, S., Gómez-Fernández, J. C. (2011). Curcumin modulates PKC $\alpha$  activity by a membrane-dependent effect. *Archives of Biochemistry and Biophysics*, 513(1), 36-41. [\[CrossRef\]](#)
31. Menard, C., Bastianetto, S., Quirion, R. (2013). Neuroprotective effects of resveratrol and epigallocatechin gallate polyphenols are mediated by the activation of protein kinase C gamma. *Frontiers in Cellular Neuroscience*, 7, 281. [\[CrossRef\]](#)
32. Ferriola, P. C., Cody, V., Middleton Jr, E. (1989). Protein kinase C inhibition by plant flavonoids: kinetic mechanisms and structure-activity relationships. *Biochemical Pharmacology*, 38(10), 1617-1624. [\[CrossRef\]](#)
33. Mishra, S., Vinayak, M. (2015). Role of ellagic acid in regulation of apoptosis by modulating novel and atypical PKC in lymphoma bearing mice. *BMC Complementary and Alternative Medicine*, 15(1), 1-8. [\[CrossRef\]](#)
34. Szafer, H., Kaczmarek, J., Rybczyńska, M., Baer-Dubowska, W. (2007). The effect of plant phenols on the expression and activity of phorbol ester-induced PKC in mouse epidermis. *Toxicology*, 230(1), 1-10. [\[CrossRef\]](#)
35. Tanruean, K., Poolprasert, P., Suwannarach, N., Kumla, J., Lumyong, S. (2021). Phytochemical Analysis and Evaluation of Antioxidant and Biological Activities of Extracts from Three Clauseneae Plants in Northern Thailand. *Plants*, 10(1), 117. [\[CrossRef\]](#)
36. Albaayit, S. F. A., Abba, Y., Abdullah, R., Abdullah, N. (2014). Evaluation of antioxidant activity and acute toxicity of *Clausena excavata* leaves extract. *Evidence-Based Complementary and Alternative Medicine*, 2014, 1-10. [\[CrossRef\]](#)
37. Elumalai, K., Id, K. (2016). Antioxidant activity and phytochemical screening of different solvent extracts *Clausena excavata* burm F. Rutaceae) *MOJ Ecology & Environmental Sciences*, 1(1), 1. [\[CrossRef\]](#)