



## ARAŞTIRMA / RESEARCH

# Effectiveness of melatonin in preventing vancomycin-induced nephrotoxicity: an experimental study

Vankomisin kaynaklı nefrotoksisiteyi önlemede melatoninin etkinliği: deneysel bir çalışma

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### Abstract

**Purpose:** The aim of the study explores probable toxic effects of vancomycin on kidney and analysis of the probable protective effects of melatonin.

**Materials and Methods:** In this study, rats were randomly divided into 4 groups: the control group; the melatonin (10 mg/kg/day) group; the vancomycin-treated (200 mg/kg) group; and the vancomycin (200 mg/kg) + melatonin (10 mg/kg/day) group. Rats in the treatment group were given two doses of vancomycin a day with an interval of seven consecutive days and melatonin (10 mg/kg/day) once daily for seven consecutive days. The experiment was continued for 15 days. In each group, seven rats were grouped together. 15 days after the experiment, the rats were sacrificed under anesthesia and among all groups. Kidney tissues were collected and processed for further TNF- expression analysis, as well as histological analyses such as hematoxylin and eosin (H&E), Masson's tricom, and Periodic acid schiff (PAS) staining to assess pathological severity. In addition, a terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay was performed to evaluate apoptosis.

**Results:** While vancomycin upregulated TNF- $\alpha$  expression, melatonin reduced levels of TNF- $\alpha$  immunoreactivity intensity and clearly improved pathological severity in rat kidneys. Further, melatonin significantly inhibited vancomycin-induced TUNEL-positive cell numbers.

**Conclusion:** Melatonin has protective activity against vancomycin-induced pro-inflammatory and proapoptotic

### Öz

**Amaç:** Bu çalışmanın amacı, vankomisinin böbrek üzerindeki olası toksik etkilerinin belirlenmesi ve melatoninin olası koruyucu etkilerini araştırmaktır.

**Gereç ve Yöntem:** Bu çalışmada, sıçanlar rastgele 4 gruba ayrıldı: kontrol grubu, melatonin (10 mg/kg/gün) grubu, vankomisin uygulanan (200 mg/kg) grup ve vankomisin (200 mg/kg) +melatonin (10 mg/kg/gün) grubu. Tedavi grubundaki sıçanlara art arda yedi gün boyunca günde iki kez vankomisin ve ardından yedi gün boyunca günde bir kez melatonin (10 mg/kg/gün) verildi. Her grupta yedi sıçan yer aldı ve deneye 15 gün devam edildi. Deneden 15 gün sonra, sıçanlar anestezi altında sakrifiye edildi. Sıçanlara ait böbrek dokuları alındı ve patolojik şiddeti değerlendirmek için TNF- $\alpha$  ekspresyon analizi, hematoksilin ve eozin (H&E), Masson's tricrom ve Periodic acid schiff (PAS) gibi histolojik analizler yapıldı. Ek olarak, apoptozu değerlendirmek için Terminal deoksiniükleotidil transferaz dUTP nick-end etiketleme (TUNEL) yöntemi uygulandı.

**Bulgular:** Vankomisin, TNF- $\alpha$  ekspresyonunu yükseltirken; melatonin, TNF- $\alpha$  immünoaktivite yoğunluğunu azalttı ve sıçanların böbreklerinde açıkça patolojik şiddeti iyileştirdi. Ayrıca melatonin, vankomisinin neden olduğu TUNEL pozitif hücre sayılarını önemli ölçüde inhibe etti.

**Sonuç:** Bulgularımız, melatoninin organ koruma süresi boyunca böbreklerde vankomisinin neden olduğu proinflatuar ve proapoptotik etkilere karşı koruyucu ve böbrek fonksiyonlarını iyileştirici etkiye sahip olduğunu gösterdi.

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effects in kidneys during organ preservation time and improves kidney function.

**Keywords:** Kidney damage, apoptosis, pro-inflammatory cytokines, melatonin, vancomycin

**Anahtar kelimeler:** Böbrek hasarı, apoptoz, pro-inflamatuar sitokinler, melatonin, vankomisin

## INTRODUCTION

Vancomycin is the most effective antibiotic for treating gram-positive bacteria like *Staphylococcus epidermidis* and methicillin-resistant *Staphylococcus aureus* (MRSA) infections<sup>1,2</sup>. Also, these bacteria can evolve resistance to this antibiotic. Although it has potential side effects such as nephrotoxicity, a high dose of vancomycin (every 8–12 h, 15–20 mg/kg body weight) is recommended in the literature<sup>3-5</sup>. The molecular mechanism of nephrotoxicity induced by vancomycin is not fully understood, but it is thought to play a role in the pathogenesis of kidney injury by triggering apoptosis, oxidative stress, and inflammation<sup>6,7</sup>. As a result, research into the mechanism of vancomycin-induced nephrotoxicity is required in order to determine how to effectively overcome this antibiotic adverse effect. Protecting the kidney from vancomycin-induced damage is very important to benefit from the positive effects. Therefore, recent studies research the oxidative effects of vancomycin induced kidney damage<sup>8</sup>.

Melatonin (N-acetyl-5-methoxytryptamine) which is released from the pineal gland as an endocrine hormone, is also synthesized at numerous extra pineal sites<sup>9</sup>, as a potential useful agent in the treatment of several diseases and conditions. Melatonin has been considered as a potential endogenous free radical scavenger due to its protective effects against mitochondrial damage and tissue injury by scavenging scavenging RNS (reactive nitrogen species) or ROS (reactive oxygen species) in vitro and in vivo<sup>10</sup>. In addition to protecting cells and tissues from radical damage as a strong antioxidant<sup>11</sup>, melatonin reduces NF- $\kappa$ B by inhibiting proinflammatory cytokines such as interleukin (IL)-1 $\beta$ , IL-6, and tumour necrosis factor (TNF)- $\alpha$ , during hepatic fibrosis progression<sup>12</sup>.

Also, apoptotic cell death is known to be triggered by oxidative stress, which induces the caspase system<sup>13</sup>. Except for anti-inflammatory effects, melatonin is known to have anti-apoptotic effects by affecting pro-or anti-apoptotic proteins such as Bax and Bcl-2<sup>14</sup>. The use of mouse-derived mesenchymal stem cells in chronic kidney disease, has resulted in accelerated senescence as a treatment strategy,

melatonin administration in these stem cell-based treatments has been shown to protect cells against senescence<sup>15</sup>. In experimental models, it has also been shown that, administration of melatonin prevents structural and functional injuries in the kidney<sup>16,17,18</sup>. Moreover, melatonin is potentially helpful in the treatment of various diseases including diabetes, cerebrovascular diseases, cancer, and cognitive decline associated with aging<sup>19</sup>. We hypothesize that melatonin drives the observed increase in pro-inflammatory cytokines in vancomycin-induced kidney damage. Here, we show that melatonin treatment of vancomycin-induced kidney damage regulates apoptotic cell death and reduces pro-inflammatory cytokin levels. The molecular mechanism and downstream effector of melatonin's therapeutic effects have not been explored yet. The current study sought to investigate the degenerative effects of vancomycin on the kidney as well as the protective effects of melatonin against vancomycin administration for the first time in the literature. Importantly, melatonin could offer a novel therapy for patients with intractable kidney damage caused by vancomycin.

## MATERIALS AND METHODS

### Experiment procedure

The experimental protocol for experiments on animals was approved by the Erciyes University Experimental Animals Ethics Committee (Decision number: 21/221) and all experiments were performed according to the animal ethics protocols regulated by the committee. All experimental procedures were performed at the Experimental and Clinical Research Center of Erciyes University. Subjects were kept in well-ventilated polypropylene cages with tap water and food ad libitum under controlled laboratory conditions with a normal temperature ( $25 \pm 2$  °C) and normal light/dark cycle to acclimatize for two weeks. The 28 healthy female adult Wistar albino rats were divided into four groups, each containing seven rats, and the experimental procedure was continued for 15 days. A normal diet and water were given to the control group. The melatonin-treated group received

melatonin (10 mg/kg/day) intraperitoneally (i.p.) once daily for seven consecutive days. The Vancomycin treated group received vancomycin (200 mg/kg) i.p. two times a day with an interval of 12 hours for seven consecutive days. The vancomycin and melatonin treated group received vancomycin once a day with an interval of 12 hours at a concentration of 200 mg/kg for seven consecutive days<sup>20</sup> and then melatonin (10 mg/kg/day) was administered once daily for seven consecutive days started on day 8th day. Kidney tissue samples were collected under general anesthesia with xylazine (10 mg/kg) and ketamine (50 mg/kg) at the end of the application period.

### Histological analysis

For the light microscopy examinations, kidney tissue samples of rats from all groups were fixed in 10% formaldehyde for histological analysis. Following the washing and dehydration, the kidney tissues were embedded in paraffin. Then these tissue blocks were sectioned at a 5 µm thickness. Thereafter, the staining with hematoxylin and eosin (H&E), Masson's trichrom and Periodic acid schiff (PAS), histopathological changes in the kidney tissue sections of the control and experimental groups were monitored. Also, the apoptotic cells were analysed in the vancomycin treated groups to compare with the control and melatonin treated groups.

### Immunohistochemistry analysis

Tissue paraffin sections were stained for TNF- $\alpha$  immunoreactivity in the kidney using the avidin-biotin-peroxidase method (Thermo Scientific, Waltham, MA), as recommended by the manufacturer. After incubation for 5 minutes in distilled water at room temperature, sections were washed in PBS and blocked with 3% hydrogen peroxide for 10 min at room temperature. Then, sections were kept in the microwave at 95 oC for 20 minutes for antigen recovery in 0.01 M sodium citrate buffer (pH 6.0). Sections were washed in PBS after being incubated overnight at 4 oC with primary antibody TNF- $\alpha$  antibody diluted 1:50 (sc.25280; Santa Cruz Biotechnology, Santa Cruz, CA). Following the application of biotinylated secondary antibodies for 15 min at room temperature, slides were washed in PBS again, and stained with diaminobenzidine (DAB). In addition to that, counterstaining with hematoxylin was followed. TNF- $\alpha$  positive cells were observed to stain brown

after dehydration and covering. The TNF- $\alpha$  immunoreactivity intensities for each section were calculated as the mean immunoreactivity intensity in ten random microscopic fields using Image J software (National Institutes of Health, Bethesda, MD).

### Apoptosis analysis

TUNEL (Terminal-deoxynucleotidyl Transferase Mediated Nick End Labeling) as an in situ cell death detection kit (ApopTag; Millipore, Berlin, Germany) was used to detect apoptosis in kidney tissue. After deparaffinization and rehydration, the slides were treated with digoxigenin-dUTP in the presence of TdT (terminal deoxynucleotidyl transferase) for 1 h at 37°C. An Anti-digoxigenin fluorescence unconjugated antibody was used to visualize TUNEL-positive cells. Finally, the association of TUNEL-positive cells with apoptosis in the experimental groups was evaluated by identifying apoptotic cells morphologically. Cells were counted in ten randomly selected fields, and the apoptotic cell numbers were calculated by using ImageJ software (ImageJ) at the same magnification.

### Statistical analysis

The means  $\pm$  SEM (standard error of the mean) of all data were analyzed by using GraphPad Prism software version 9.0 (GraphPad Inc., San Diego, CA). One-way ANOVA with Bonferroni analysis was performed to evaluate the statistical significance of differences between the control and experimental groups in terms of TNF- $\alpha$  variables in the study. Kruskal-Wallis tests were used for comparisons of more than two groups, Dunn's test was performed for multiple comparisons for TUNEL results. P-values less than 0.05 were considered statistically significant.

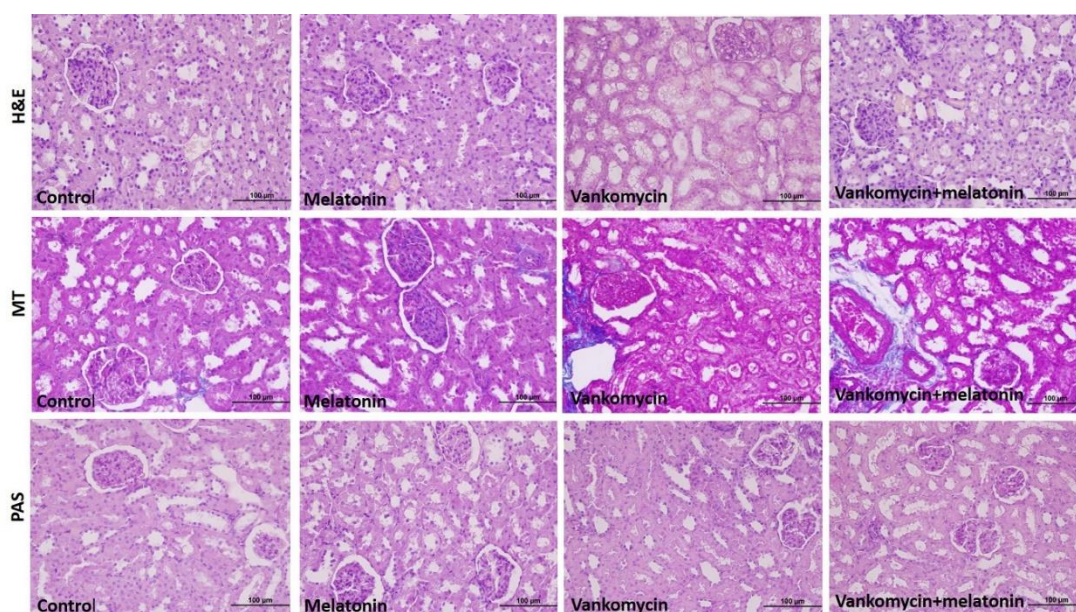
## RESULTS

The impact of vancomycin and melatonin on the function and structure of the kidney was evaluated histologically. Kidney tissues of subjects in the control and melatonin groups exhibited normal histology in the functional units such as tubules and glomerulus (Figure 1). However, degenerative changes such as abnormal necrosis, swelling of tubules, lymphocyte infiltration, and vacuolation in tubular epithelium were seen in the vancomycin treated group. In the melatonin-treated group,

damage was comparatively less because of the fact that the glomerulus was not severely disturbed and the tubules were not swollen.

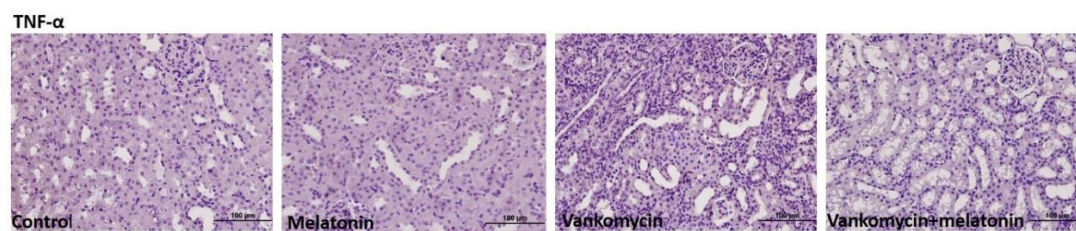
Staining with masson trichrome was used to determine collagen deposition (Figure 1). Normal connective tissue was observed in the kidney sections of the control group and melatonin treated group. It was observed that collagen fibers around the stroma area increased in the vancomycin treated group compared to the control group. Almost no fibrosis

was detected in the kidney tissue of subjects in the vancomycin+melatonin treated group. While normal proximal tubules such as prominent brush border with continued basement membrane were observed in PAS stained kidney tissue sections of the control group, severe desquamation in tubules and brush border of epithelium were seen in the vancomycin treated group. Moreover, no difference was detected between the kidney tissues of the vancomycin+melatonin treated group and the control group in terms of tubular damage.

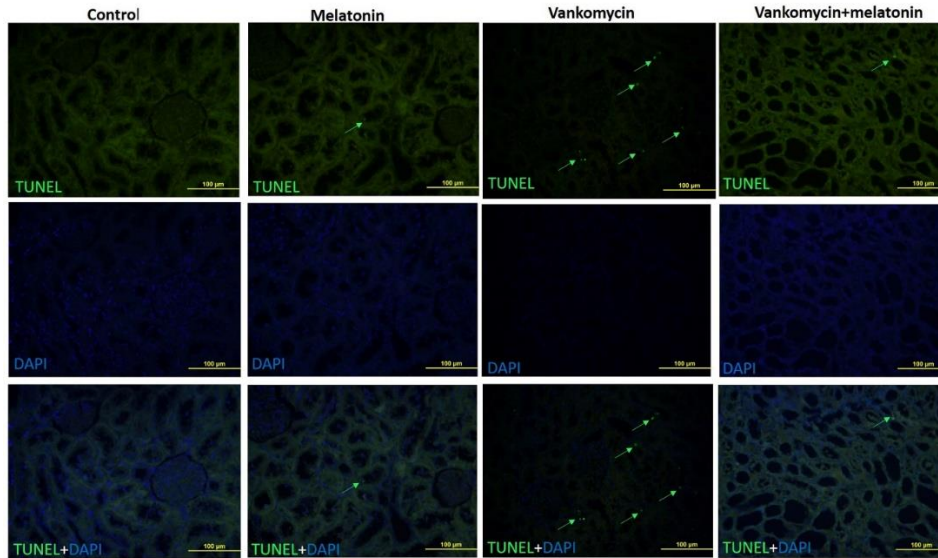


**Figure 1. Light microscopic images of the renal cortex (hematoxylin-eosin, magnification X40).**

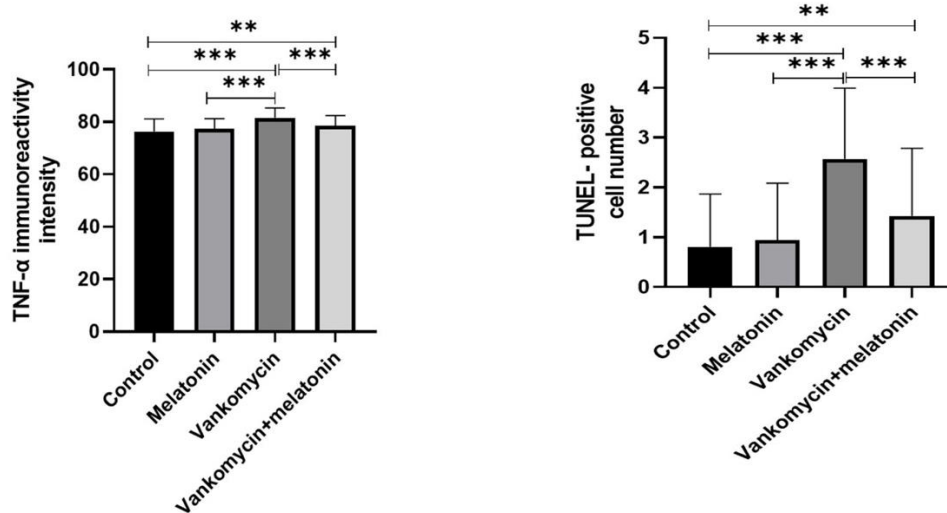
Normal glomerulus and tubules were observed both in control group and melatonin treated group, besides no cytoplasmic granulation was found in tubular. Necrosis, interstitial inflammatory infiltration and cytoplasmic granulation in tubular cells together with vacuolisation in tubular epithelium cells and tubular swelling were seen in vancomycin treated group. Less necrotic area and less tubular swelling were observed in the melatonin+vancomycin treated group. The sections staining with masson trichrome was no detected fibrosis in the kidney tissue of all experimental groups (Masson trichrome, magnification X40). Clearly visible normal brush borders were seen in proximal tubules of both control group and melatonin treated group in PAS staining sections. Besides, the basement membrane of the proximal and distal tubules and the parietal leaf of Bowman’s capsule were in normal structure. Tubular epithelial cells were seen to separate from each other or from the basement membrane, and debris accumulations in the tubular lumen were shown in vancomycin treated group. In addition, numerous atypical proximal tubules which lost their brush borders were observed. The basement membrane of the proximal and distal tubules and the parietal leaf of Bowman’s capsule were seen in normal structure in melatonin+vancomycin treated group (PAS, magnification X40).



**Figure 2. Melatonin reduces kidney inflammation caused by vancomycin in vivo.**  
Representative immunohistochemical staining of TNF- $\alpha$ .



**Figure 3. Antiapoptotic effect of melatonin treatment after vancomycin administration in vivo.**  
Representative images of apoptotic cells. The apoptotic cells were detected by TUNEL (green), and the nuclei were detected by DAPI (blue).



**Figure 4. The graphs show immunoreactivity intensity result of TNF- $\alpha$  and TUNEL-positive cell number in kidney tissue of different experimental group.**  
Data are presented as mean  $\pm$  SEM.\*p < 0.05,\*\*p < 0.01,\*\*\*p < 0.001.

To clarify the possible protective effect of melatonin on vancomycin-induced kidney injury, TNF- $\alpha$  expression in kidney tissues was analyzed by immunohistochemical methods, and the results are

shown in Figure 2. While the control and melatonin groups showed weak positive TNF- $\alpha$  expression in kidney tissue sections, the vancomycin group showed intensely positive TNF- $\alpha$  immunoreactivity in tubular

epithelial cells of kidney tissues compared to the control group. TNF- $\alpha$  immunoreactivity, on the other hand, decreased in the vancomycin+melatonin treated group compared to the vancomycin treated group (Figure 4) ( $p<0.001$ ).

In order to detect the rate of cell death in the kidney tissue sections of each group quantitatively, TUNEL staining was preferred. After staining, TUNEL positive cells were counted, and the results are shown in Figure 3. The number of TUNEL-positive cells was significantly up-regulated in the vancomycin treated group compared with the control group ( $p<0.05$ ). The TUNEL positive cell count was found to be significantly decreased in the vancomycin+melatonin treated group, compared to the vancomycin treated group (Figure 4) ( $p<0.01$ ).

## DISCUSSION

The present study shows that vancomycin causes inflammation and oxidative stress, which causes histological changes together with structural defects in the kidney. In addition, it was discovered that systemic melatonin treatment limits inflammation and pathological changes and supports functional capacity. To the best of our knowledge, this is the first study to detect melatonin's therapeutic activity on vancomycin-induced kidney injury from histological and immunohistochemical perspectives.

As an inhibitor of bacterial cell wall synthesis, vancomycin is an antibiotic extensively used to treat infections caused by methicillin resistant *Staphylococcus epidermidis* (MRSE) and *Staphylococcus aureus* (MRSA)<sup>21</sup>. Besides that, it is known to have potentially fatal side effects causing nephrotoxicity due to the excretion from the kidneys, which limits its administration and efficacy<sup>2</sup>.

Although studies suggests that oxidative stress<sup>22,23</sup>, inflammation, and apoptosis may play a role in the pathology of vancomycin induced renal toxicity<sup>2</sup>, the exact mechanism of vancomycin-induced nephrotoxicity is not fully understood. In many countries, in addition to chemical drugs, the use of natural antioxidants, which reduce complications with fewer side effects, low toxicity, and affordable prices, are being investigated. By deactivating ROS, these antioxidants are used to protect against drug-induced toxicity, induce endogenous antioxidant production, and renovate homeostatic balance<sup>24</sup>. The alleviation of vancomycin-induced nephrotoxicity is dependent on enhancing the clinical success of

vancomycin therapy. In this study, we examined vancomycin-induced acute renal injury and its related underlying mechanism. On the other hand, we examined vancomycin-induced apoptosis and its inhibition by melatonin, a natural antioxidant.

Melatonin is one of the hormones secreted by the pineal gland, which has immunomodulatory activity and an antioxidant ability to scavenge free radicals<sup>25,26</sup>. Besides being a chief hormone of the pineal gland, melatonin is a pleiotropic molecule that plays a role in regulation of the circadian system, seasonal reproduction, and regulation of sleep, exhibiting significant immunoregulatory, antiinflammatory, and antioxidant properties<sup>11</sup>. In addition, it has recently been shown to have a strong antioxidant capacity<sup>27</sup>.

As in a similar study<sup>13</sup>, interstitial inflammations were found in histopathological examinations of kidney tissues from subjects treated with vancomycin. This study support that vancomycin causes nephrotoxicity, by histopathology examination of kidney tissues. However, we observed various features related to renal injury, such as necrosis, leukocyte infiltration, and obliteration of renal tubules, which were significantly reversed with melatonin treatment. We detected that melatonin reduced the tubular necrosis and interstitial fibrosis due to vancomycin administration. This further confirms that melatonin has potent antioxidant and anti-inflammatory effects on oxyradicals. Under certain pathophysiological conditions, melatonin reduces the level of pro-inflammatory cytokines by diminishing free-radical-mediated damage; therefore, it contributes to the decrease of pro-inflammatory cytokines and the increase of anti-inflammatory cytokines<sup>28</sup>.

In the present study, we showed that, vancomycin treatment leads to elevated expression of the TNF- $\alpha$  and this correlates with tissue damage. On the other hand, this increase in immunoreactivity of inflammation biomarker, TNF- $\alpha$  due to vancomycin-induction significantly abrogated by melatonin. These results were supported by similar findings that were reported previously<sup>29,30</sup>. Melatonin administration resulted in a dramatic reduction in TNF- $\alpha$  expression in the current study, as evidenced by the recovery of renal tubule parenchyma.

Apoptosis, or programmed cell death, which occurs with the death receptor pathway (exogenous) or the chondrial pathway (endogenous), clears damaged

cells and maintains homeostasis. Accumulating evidence has shown that apoptosis may be one of the main factors leading to kidney injury<sup>31,32,33</sup>. Melatonin increases cell survival by reducing oxidative stress, endoplasmic reticulum stress, apoptosis, and mitochondrial fission by activating various signaling pathways<sup>34</sup>. Due to its antioxidant properties, melatonin is considered an antiapoptotic medication<sup>35</sup> and can modulate the apoptosis process<sup>36</sup>. The TUNEL assay was used for DNA fragmentation analysis which presented cell degeneration through the presence or absence of the apoptosis cascade<sup>37</sup>. The present study revealed that quantitative data of the TUNEL assay was significantly reduced in the melatonin treated group in tubules, compared with the control group. This finding revealed that melatonin is one of the most robust biochemical components that can reduce DNA degradation independent of the apoptotic pathway. Although we reported a therapeutic effect of melatonin on vancomycin-induced kidney damage, our study still has some limitations. As a limitation of this study, the kidney function tests could be done, but we have not been able to provide enough urine samples from rats because of technical deficiency. Analyzing the long-term effect of melatonin on the degenerative effects of vancomycin on the kidney with more samples might be an interesting topic for future study.

This study has detected the potential protective effects of melatonin against vancomycin-induced renal injury, which is a main disadvantage of vancomycin treatment. Besides that, this study also identified the anti-inflammatory characteristics of melatonin. The antioxidant activity of melatonin is crucial in attenuating the effects of oxy radicals produced due to vancomycin treatment. Renoprotective effects of melatonin seem to be associated with decreased TNF- $\alpha$  expression. Moreover, it has also inhibited activation of the apoptotic process, which suppresses renal TUNEL expression. Although the protective effects of melatonin on renal damage caused by inflammatory disease are a current topic<sup>38,39</sup>, there is limited evidence about the role of melatonin on vancomycin-related renal injury in the literature. According to our histological examination, we found impairments in the brush borders of renal tubules, swelling in glomeruli, tubular necrosis, peritubular capillary congestion, and an increase in the renal corpuscle area. Regarding our results, the cellular and extracellular injuries of the kidneys may be the result

of inflammation triggered directly or indirectly by proinflammatory cytokines such as TNF- $\alpha$ . TUNEL activation is considered an unrecoverable final event before cell death, which is used as a marker of apoptosis.

In this experimental study, we also examined the possible apoptotic effects induced by the vancomycin on the kidney with TUNEL staining. In conclusion, melatonin showed beneficial effects in preventing vancomycin-induced nephrotoxicity. Nevertheless, the results of this study may pave the way for the search for optimal therapeutic agents against vancomycin-induced toxicity. Further evaluation is required to understand the role of melatonin in nephrotoxicity caused by vancomycin.

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**Yazar Katkıları:** Çalışma konsepti/Tasanımı: ÖÖG; Veri toplama: ÖCM, DB; Veri analizi ve yorumlama: DB, ÖCM, MK; Yazı taslağı: ÖÖG; İçeriğin eleştirel incelenmesi: GC, DB, ÖCM; Son onay ve sorumluluk: ÖÖG, ÖCM, DB, MK, SSP, GC; Teknik ve malzeme desteği: DB, MK, ÖCM; Süpervizyon: ÖÖG; Fon sağlama (mevcut ise): yok.

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**Author Contributions:** Concept/Design : ÖÖG; Data acquisition: ÖCM, DB; Data analysis and interpretation: DB, ÖCM, MK; Drafting manuscript: ÖÖG; Critical revision of manuscript: GC, DB, ÖCM; Final approval and accountability: ÖÖG, ÖCM, DB, MK, SSP, GC; Technical or material support: DB, MK, ÖCM; Supervision: ÖÖG; Securing funding (if available): n/a.

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**Peer-review:** Externally peer-reviewed.

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## REFERENCES

1. Basarslan F, Yilmaz N, Ates S, Ozgur T, Tutanc M, Motor VK et al. Protective effects of thymoquinone on vancomycin-induced nephrotoxicity in rats. *Hum Exp Toxicol.* 2012;31:726–33.
2. Humanes B, Jado JC, Camano S, López-Parra V, Torres AM, Álvarez-Sala LA et al. Protective effects of cilastatin against vancomycin-induced nephrotoxicity. *Biomed Res Int.* 2015;2015:704382.
3. Álvarez R, López Cortés LE, Molina J, Cisneros JM, Pachón J. Optimizing the clinical use of vancomycin. *Antimicrob Agents Chemother.* 2016;60:2601–9.
4. Martin JH, Norris R, Barras M, Roberts J, Morris R, Doogue M et al. Therapeutic monitoring of vancomycin in adult patients: a consensus review of the American Society of health system pharmacists, the infectious diseases society of America, and the society of infectious diseases pharmacists. *Clin Biochem Rev.* 2010;31:21–4.

5. Van Hal SJ, Paterson DL, Lodise TP. Systematic review and metaanalysis of vancomycin-induced nephrotoxicity associated with dosing schedules that maintain troughs between 15 and 20 milligrams per liter. *Antimicrob Agents Chemother.* 2013;57:734–44.
6. Arimura Y, Yano T, Hirano M, Sakamoto Y, Egashira N, Oishi R. Mitochondrial superoxide production contributes to vancomycin-induced renal tubular cell apoptosis. *Free Radic Biol Med.* 2012;52:1865–73.
7. Gupta A, Biyani M, Khaira A. Vancomycin nephrotoxicity: myths and facts. *Neth J Med.* 2011;6:379–83.
8. Filippone EJ, Kraft WK, Farber JL. The nephrotoxicity of vancomycin. *Clin Pharmacol Ther.* 2017;102:459–69.
9. Reiter RJ, Rosales-Corral S, Tan DX, Jou MJ, Galano A, Xu B. Melatonin as a mitochondria-targeted antioxidant: One of evolution's best ideas. *Cell Mol Life Sci.* 2017;74:3863–81.
10. Tan DX, Manchester LC, Liu X, Rosales-Corral SA, Acuna-Castroviejo D, Reiter RJ. Mitochondria and chloroplasts as the original sites of melatonin synthesis: a hypothesis related to melatonin's primary function and evolution in eukaryotes. *J Pineal Res.* 2013;54:127–38.
11. Reiter RJ, Mayo JC, Tan DX, Sainz RM, Alatorre-Jimenez M, Qin L. Melatonin as an antioxidant: under promises but over delivers. *J Pineal Res.* 2016;61:253–78.
12. Tahan V, Atug O, Akin H, Eren F, Tahan G, Tarcin O et al. Melatonin ameliorates methionine- and choline-deficient diet-induced nonalcoholic steatohepatitis in rats. *J Pineal Res.* 2009;46:401–7.
13. Uckun Z, Guzel S, Canacankatan N, Yalaza C, Kibar D, Coskun Yilmaz B. Potential protective effects of naringenin against vancomycin-induced nephrotoxicity via reduction on apoptotic and oxidative stress markers in rats. *Drug Chem Toxicol.* 2020;43:104–111.
14. Keskin-Aktan A, Akbulut KG, Yazici-Mutlu Ç, Sonugur G, Ocal M, Akbulut H. The effects of melatonin and curcumin on the expression of SIRT2, Bcl-2 and Bax in the hippocampus of adult rats. *Brain Res Bull.* 2018;137:306–10.
15. Han YS, Kim SM, Lee JH, Jung SK, Noh H, Lee SH. Melatonin protects chronic kidney disease mesenchymal stem cells against senescence via PrP(C)-dependent enhancement of the mitochondrial function. *J Pineal Res.* 2019;66:e12535.
16. Mercantepe F, Mercantepe T, Topcu A, Yilmaz A, Tumkaya L. Protective effects of amifostine, curcumin, and melatonin against cisplatin-induced acute kidney injury. *Naunyn Schmiedebergs Arch Pharmacol.* 2018;391:915–31.
17. Potić M, Ignjatović I, Ničković VP, Živković JB, Krdžić JD, Mitić JS et al. Two different melatonin treatment regimens prevent an increase in kidney injury marker-induced by carbon tetrachloride in rat kidneys. *Can J Physiol Pharmacol.* 2019;97:422–8.
18. Kobroob A, Peerapanyasut W, Chattipakorn N, Wongmekiat O. Damaging effects of bisphenol a on the kidney and the protection by melatonin: emerging evidences from in vivo and in vitro studies. *Oxid Med Cell Longev.* 2018;2018:3082438.
19. Opie LH, Lecour S. Melatonin has multiorgan effects. *Eur Heart J Cardiovasc Pharmacother.* 2016;2:258–265.
20. Bayram A, Erkan GN, Talih G, Baskol G, Deniz K, Yildiz K et al. The alpha-2 receptor agonist dexmedetomidine attenuates vancomycin-induced acute kidney injury. *Bratisl Lek Listy.* 2019;120:429–433.
21. Umstätter F, Domhan C, Hertlein T, Ohlsen K, Mühlberg E, Kleist C et al. Vancomycin resistance is overcome by conjugation of polycationic peptides. *Angew Chem Int Ed Engl.* 2020;59:8823–27.
22. Marvin JL, Levine BJ, Papas M, Rosini JM. An evaluation of the incidence of nephrotoxicity after a loading dose of vancomycin in patients with severe renal impairment. *J Emerg Med.* 2019;56:701–8.
23. Sabler IM, Berkovitch M, Sandbank J, Kozer E, Dagan Z, Goldman M et al. Exposure to hyperbaric oxygen intensified vancomycin-induced nephrotoxicity in rats. *PLoS One.* 2016;11:e0152554.
24. Brewer MS. Natural antioxidants: sources, compounds, mechanisms of action, and potential applications. *Compr Rev Food Sci Food Saf.* 2011;10:221–47.
25. Cai J, Yang J, Chen X, Zhang H, Zhu Y, Liu Q et al. Melatonin ameliorates trimethyltin chloride-induced cardiotoxicity: the role of nuclear xenobiotic metabolism and Keap1-Nrf2/ARE axis-mediated pyroptosis. *Biofactors.* 2022;48:481–97.
26. Liu XJ, Wang YQ, Shang SQ, Xu S, Guo M. TMT induces apoptosis and necroptosis in mouse kidneys through oxidative stress-induced activation of the NLRP3 inflammasome. *Ecotoxicol Environ Saf.* 2022;230:113167.
27. Hardeland R. Aging, melatonin, and the pro- and anti-inflammatory networks. *Int J Mol Sci.* 2019;20:1223.
28. Chitimus DM, Popescu MR, Voiculescu SE, Panaitescu AM, Pavel B, Zagrean L et al. Melatonin's impact on antioxidative and anti-inflammatory reprogramming in homeostasis and disease. *Biomolecules.* 2020;10:1211.
29. Dutta S, Saha S, Mahalanobish S, Sadhukhan P, Sil PC. Melatonin attenuates arsenic induced nephropathy via the regulation of oxidative stress and inflammatory signaling cascades in mice. *Food Chem Toxicol.* 2018;118:303–16.
30. D'Angelo G, Chimenz R, Reiter RJ, Gitto E. Use of melatonin in oxidative stress related neonatal diseases. *Antioxidants (basel).* 2020;9:477.
31. Li J, Zhang W, Zhou P, Tong X, Guo D, Lin H. Selenium deficiency induced apoptosis via



- mitochondrial pathway caused by Oxidative Stress in porcine gastric tissues. *Res Vet Sci.* 2022;144:142-8.
32. Wang Z, Wu J, Hu Z, Luo C, Wang P, Zhang Y et al. Dexmedetomidine alleviates lipopolysaccharide-induced acute kidney injury by inhibiting p75NTR-mediated oxidative stress and apoptosis. *Oxid Med Cell Longev.* 2020;2020:5454210.
33. Yingjie K, Haihong Y, Lingwei C, Sen Z, Yuanting D, Shasha C et al. Apoptosis repressor with caspase recruitment domain deficiency accelerates ischemia/reperfusion (I/R)-induced acute kidney injury by suppressing inflammation and apoptosis: the role of AKT/mTOR signaling. *Biomed. Pharmacother.* 2019;112:108681.
34. Nduhirabandi F, Lamont K, Albertyn Z, Opie LH, Lecour S. Role of toll-like receptor 4 in melatonin-induced cardioprotection. *J Pineal Res.* 2016;60:39-47.
35. Pi H, Xu S, Reiter RJ, Guo P, Zhang L, Li Y et al. SIRT3-SOD2-mROS-dependent autophagy in cadmium-induced hepatoxicity and salvage by melatonin. *Autophagy* 2015;11:1037-51.
36. Rodriguez C, Martín V, Herrera F, García-Santos G, Rodríguez-Blanco J, Casado-Zapico S et al. Mechanisms involved in the pro-apoptotic effect of melatonin in cancer cells. *Int J Mol Sci.* 2013;14:6597-6613.
37. Negoescu A, Guillermet C, Lorimier P, Brambilla E, Labat-Moleur F. Importance of DNA fragmentation in apoptosis with regard to TUNEL specificity. *Biomed Pharmacother.* 1998;52:252-8.
38. Leibowitz A, Volkov A, Voloshin K, Shemesh C, Barshack I, Grossman E. Melatonin prevents kidney injury in a high salt diet-induced hypertension model by decreasing oxidative stress. *J Pineal Res.* 2016;60:48-54.
39. Shi S, Lei S, Tang C, Wang K, Xia Z. Melatonin attenuates acute kidney ischemia/reperfusion injury in diabetic rats by activation of the SIRT/Nrf2/HO-signaling pathway. *Biosci Rep.* 2019;39:BSR20181614.