Journal of Agricultural Production (2020) 1(1): 5-7 *e*-ISSN: 2757-6620



RESEARCH ARTICLE

Investigation of Antibacterial Activity of Two Different Medicinal Plants Extracts Against Fish Pathogens

Mustafa Karga^{1*} 💿 • Osman Nezih Kenanoğlu² 💿 • Soner Bilen² 💿

¹Kastamonu University, İnebolu Vocational College, İnebolu, Kastamonu/Turkey ²Kastamonu University, Faculty of Fisheries, Department of Aquaculture, Kastamonu/Turkey

ARTICLE INFO

Article History: Received: 19.12.2020 Accepted: 24.12.2020 Available Online: 31.12.2020

Keywords: Medicinal plants Aqueous methanolic extract Antibacterial activity Minimum inhibitory concentration

ABSTRACT

Diseases are one of the leading factors affecting the sustainability of aquaculture industry. Due to the undesirable effects of the methods used in the prevention or treatment of diseases, the usability of herbal products has been investigated recently. Thus, in the present study, we aimed to investigate the effects of aqueous methanolic extracts of two different medicinal plants (*Laurus nobilis* and *Brassica nigra*) against *Vibrio anguillarium*, *Yersinia ruckeri*, *Pseudomonas putida*, and *Aeromonas hydrophila* by using minimum inhibitory concentration (MIC). The MIC values of leaf aqueous methanolic extract of *Laurus nobilis* for Aeromonas *hydrophila* and leaf aqueous methanolic extract of *Brassica nigra* for *Vibrio anguillarium* were determined as $3.125 \,\mu g \, ml^{-1}$ and $100 \,\mu g \, ml^{-1}$, respectively. The results showed that *Laurus nobilis* could be used against *Aeromonas hydrophila* and *Brassica nigra* against *Vibrio anguillarium*. Further *in vivo* studies should be conducted to evaluate the usability of these plants.

Please cite this paper as follows:

Karga, M., Kenanoğlu, O. N. and Bilen, S. (2020). Investigation of Antibacterial Activity of Two Different Medicinal Plants Extracts Against Fish Pathogens. *Journal of Agricultural Production*, 1(1): 5-7.

Introduction

Aquaculture industry is one of the most fast growing food industry in the world (Fisheries, 2018). Increased technology could give opportunities to the fish farmers to increase their production rate (Bilen et al., 2013). However, diseases seem to be most limiting factor in the industry. Increasing stoking density and the different technology usage may trigger stress conditions and this can favour occurrence and the spread of the bacterial diseases (Fazio et al., 2013).

Among the bacterial pathogens *Vibrio anguillarium*, *Yersinia ruckeri*, *Pseudomonas putida* and *Aeromonas hydrophila* are opportunistic and ubiquitous, and most commonly infect not only freshwater fish species but also marine fishes. Antibiotic have been mostly used to prevent or treat fish from those pathogens (Corum et al., 2020; Terzi et al., 2020). In some cases, vaccine usage also gives an opportunity to overcome the problem. However, antibiotics have some adverse effect on animal and the environment (Capkin et al., 2015), and vaccines are used for specific pathogens.

Medicinal plants extracts already discovered to cure the fish against diseases (Bilen and Elbeshti, 2019; Bilen et al., 2019) or protect them from many different fish diseases (Bilen et al., 2020a; Bilen et al., 2020b; Bilen et al., 2014) and even in some cases as reproductive promoter (Sonmez et al., 2019). Also, medicinal plants have growth performance and immune system activation in fishes (Amhamed et al., 2018; Bilen et al., 2020c; Elbesthi et al., 2020; Mohamed et al., 2018). Laurel (Laurus nobilis) is a tree and has been used for its astringent, healing and diuretic properties (Nayak et al., 2006). Laurel has also antimicrobial and antibacterial effects (Digrak et al., 2001). Black mustard (Brassica nigra), native to the southern Mediterranean region of Europe, which has been cultivated for thousands of years. In a previous study, addition of a medicinal plant, Brassica nigra, to Oreochromis niloticus food improves both the immune and biotransformation systems after

^{*} Corresponding author

E-mail address: mkarga@kastamonu.edu.tr

exposure to a polycyclic aromatic hydrocarbon, BaP (Abbas et al., 2016).

In the present study, we performed to demonstrate that the effects of aqueous methanolic extract of *Laurus nobilis* and *Brassica nigra* against fish pathogen such as Vibrio anguillarium, Yersinia ruckeri, Pseudomonas putida and Aeromonas hydrophila.

Materials and Methods

Plant and Preparation of the Extracts

The plants were purchased from herbalist in Kastamonu province. Aqueous methanolic extraction of the plants were performed as previously described (Bilen et al., 2016).

Table 1.	. The list of	the plants	used in the	study
----------	---------------	------------	-------------	-------

Scientific Name	Family	Vernacular Name
Laurus nobilis	Lauraceae	Laurel
Brassica nigra	Cruciferae	Black Mustard

Bacterial Strains

The plant extracts were tested against Gram negative bacteria, Vibrio anguillarium (SBVA1), Yersinia ruckeri

(SBYR1), *Pseudomonas putida* (SBPP1) and *Aeromonas hydrophila* (SBAh1) which are isolated from fish and identified using conventional and molecular methods.

Minimum Inhibitory Concentration (MIC) Determination with Broth Microdilution Method

The antibacterial activity of the plants aqueous methanolic extracts were determined using sterile 300 µl 96-well plates as previously described (Wiegand et al., 2008) with small modifications. Briefly, all plant extracts were diluted up to 3200 µg ml⁻¹ starting from 1.5625 µg ml⁻¹ (3200 µg ml⁻¹, 1600 µg ml⁻¹, 800 µg ml⁻¹, 400 µg ml⁻¹, 200 µg ml⁻¹, 100 µg ml⁻¹, 50 µg ml⁻¹, 25 µg ml⁻¹, 12.5 µg ml⁻¹, 6.25 µg ml⁻¹, 3.125 µg ml⁻¹ and 1.5625 µg ml⁻¹). 150 µl plant extract and same amount of the bacterial suspension each contains 1×10^8 CFU was mixed by pipetting. For control, only bacterial suspension and the only methanolic extraction of the plant were prepared and added to 96-well plates. The plates were then placed in the incubator and kept at 25 °C for 48 hours. Each bacteria and all concentrations were studied in triplicate.

Results

The results of the study were given in Table 2.

Table 2. Antimicrobial activity of aqueous methanolic extracts of the medicinal plants used in the study

Diant Species	Bacterial Strains				
Plant species	VA	YR	PP	АН	
Laurus nobilis (µg ml ⁻¹)	1600	3200		3.125	
Brassica nigra (µg ml·1)	100	3200		800	

VA: Vibrio anguillarium; YR: Yersinia ruckeri; PP: Pseudomonas putida; AH: Aeromonas hydrophila.

The MIC value of laurel aqueous methanolic extract showed the strongest activity against *Aeromonas hydrophila*. Black mustard aqueous methanolic extract showed the strongest activity against *Vibrio anguillarium*. Both of the plant extracts exhibit no activity against *Pseudomonas putida*. Also the activity of the plants against *Yersinia ruckeri* was very weak (3200 µg ml⁻¹).

Discussion

Medicinal herbs have many different chemical complex and substance and novel mechanism of the plans haven't been explained yet. In the present study effectiveness of the laurel and black mustard were demonstrated against *A. hydrophila* and *V. anguillarium*.

Kamaraj et al. (2012) have reported that the methanolic extract of the A. indica showed strong activity against Klebsiella pneumonia. Also, strong antimicrobial activity of Cotinus coggyria was determined against Bacillus cereus, Bacillus subtilis, Staphylococcus aureus, Micrococcus luteus, Escherichia coli, Enterobacter aerogenes, Proteus vulgaris, Pseudomonas aeruginosa, Pseudomonas putida, Salmonella typhimurium, Salmonella typhi, Hanseniaspora guilliermondii, Rhodotorula rubra, Kluyveromyces fragilis, Kluyveromyces marxianus and Debaryomyces hansenii (Dulger et al., 2009). Dulger et al. (2005) showed that several extracts and fractions of some Hypericum species have antimicrobial activity against bacterial pathogens.

In conclusion, this study highlights *Laurus nobilis* has antimicrobial activity against *Aeromonas hydrophila* and *Brassica nigra* has similar activity against *Vibrio anguillarium*. *In vivo* studies should be conducted using these plants extracts for the fish.

References

- Abbas, W. T., Awad, E., & Abdel-Rahman, E. H. (2016). Effect of Black Mustard (*Brassica nigra*) on the Interaction between Immune and Biotransformation Systems of Nile Tilapia (*Oreochromis niloticus*) Exposed to Benzo-a-Pyrene. Journal of Fisheries and Aquatic Science, 11(1): 56.
- Amhamed, I. D., Mohamed, G. A., Almabrok, A. A., Altief, T. A. S., & Bilen, S. (2018). Efficacy of dietary *Chenopodium album* extract on some health parameters, digestive enzymes and growth performance in juvenile *Cyprinus carpio*. Alınteri Zirai Bilimler Dergisi, 33(2): 165-176.

- Bilen, S., & Elbeshti, H. T. A. G. (2019). A new potential therapeutic remedy against *Aeromonas hydrophila* infection in rainbow trout (*Oncorhynchus mykiss*) using tetra, *Cotinus coggygria*. Journal of Fish Diseases, 42(10): 1369-1381.
- Bilen, S., Filogh, A. M., Ali, A. B., Kenanoğlu, O. N., & Zoral, M. A. (2020a). Effect of common mallow (*Malva sylvestris*) dietary supplementation on growth performance, digestive enzyme activities, haemotological and immune responses of common carp (*Cyprinus carpio*). Aquaculture International, 28(1): 73-84.
- Bilen, S., Ispir, S., Kenanoglu, O. N., Taştan, Y., Güney, K., & Terzi, E. (2020b). Effects of Greek juniper (*Juniperus excelsa*) extract on immune responses and disease resistance against *Yersinia ruckeri* in rainbow trout (*Oncorhynchus mykiss*). Journal of Fish Diseases.
- Bilen, S., Karga, M., Altunoğlu, Ç., Yasemin, Ulu, F., & Biswas, G. (2020c). Immune Responses and Growth Performance of the Aqueous Methanolic Extract of *Malva sylvestris* in *Oncorhynchus mykiss*. Marine Science and Technology Bulletin, 9(2): 159-167.
- Bilen, S., Kızak, V., & Gezen, A. M. (2013). Floating Fish Farm Unit (3FU). Is it an Appropriate Method for Salmonid Production? Marine Science and Technology Bulletin, 2(1): 9-13.
- Bilen, S., Sirtiyah, A. M. A., & Terzi, E. (2019). Therapeutic effects of beard lichen, Usnea barbata extract against Lactococcus garvieae infection in rainbow trout (Oncorhynchus mykiss). Fish & Shellfish Immunology, 87: 401-409.
- Bilen, S., Soydaş, E., & Bilen, A. (2014). Effects of methanolic extracts of nettle (*Urtica dioica*) on non-specific immune response of gold fish (*Carassius auratus*). Alınteri Zirai Bilimler Dergisi, 27: 24-29.
- Bilen, S., Ünal, S., & Güvensoy, H. (2016). Effects of oyster mushroom (*Pleurotus ostreatus*) and nettle (*Urtica dioica*) methanolic extracts on immune responses and resistance to *Aeromonas hydrophila* in rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 454: 90-94.
- Capkin, E., Terzi, E., & Altinok, I. (2015). Occurrence of antibiotic resistance genes in culturable bacteria isolated from Turkish trout farms and their local aquatic environment. Diseases of Aquatic Organisms, 114(2): 127-137.
- Corum, O., Terzi, E., Corum, D. D., Kenanoglu, O. N., Bilen, S., & Uney, K. (2020). Pharmacokinetic/pharmacodynamic integration of marbofloxacin after oral and intravenous administration in rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 514: 734510.
- Digrak, M., Alma, M. H., & Ilçim, A. (2001). Antibacterial and antifungal activities of Turkish medicinal plants. Pharmaceutical Biology, 39(5): 346-350.
- Dulger, B., Gonuz, A., Bilen, S., & Jäger, A. (2005). Antimicrobial studies on three Hypericum species from Turkey. South African Journal of Botany, 71(1): 100-103.

- Dulger, B., Hacioglu, N., & Bilen, S. (2009). Antimicrobial activity of *Cotinus coggyria* from Turkey. Asian Journal of Chemistry, 21(5): 4139-4140.
- Elbesthi, R. T. A., Özdemir, K. Y., Taştan, Y., Bilen, S., & Sönmez, A. Y. (2020). Effects of ribwort plantain (*Plantago lanceolata*) extract on blood parameters, immune response, antioxidant enzyme activities, and growth performance in rainbow trout (*Oncorhynchus mykiss*). Fish Physiology and Biochemistry, 46(4): 1295-1307.
- Fazio, F., Marafioti, S., Filiciotto, F., Buscaino, G., Panzera, M., & Faggio, C. (2013). Blood hemogram profiles of farmed onshore and offshore gilthead sea bream (*Sparus aurata*) from Sicily, Italy. Turkish Journal of Fisheries and Aquatic Sciences, 13(3): 415-422.
- Fisheries, F. (2018). Aquaculture Department. The state of world fisheries and aquaculture. Rome: FAO; 2010.
- Kamaraj, C., Rahuman, A. A., Siva, C., Iyappan, M., & Kirthi, A. V. (2012). Evaluation of antibacterial activity of selected medicinal plant extracts from south India against human pathogens. Asian Pacific Journal of Tropical Disease, 2: S296-S301.
- Mohamed, G. A., Amhamed, I. D., Almabrok, A. A., Barka, A. B. A., Bilen, S., & Elbeshti, R. T. (2018). Effect of celery (*Apium graveolens*) extract on the growth, haematology, immune response and digestive enzyme activity of common carp (*Cyprinus carpio*). Marine Science and Technology Bulletin, 7(2): 51-59.
- Nayak, S., Nalabothu, P., Sandiford, S., Bhogadi, V., & Adogwa, A. (2006). Evaluation of wound healing activity of *Allamanda cathartica*. L. and *Laurus nobilis*. L. extracts on rats. BMC Complementary and Alternative Medicine, 6(1): 12.
- Sonmez, A., Ozdemir, R., Bilen, S., & Kadak, A. (2019). Effect of Ginseng Root (*Araliaceae sp.*) Extracts on Sperm Quality Parameters and Reproductive Performance in Rainbow Trout (*Oncorhynchus mykiss*). The Israeli Journal of Aquaculture-Bamidgeh, IJA_71.2019.1570.
- Terzi, E., Corum, O., Bilen, S., Kenanoglu, O. N., Atik, O., & Uney, K. (2020). Pharmacokinetics of danofloxacin in rainbow trout after different routes of administration. Aquaculture, 520: 734984.
- Wiegand, I., Hilpert, K., & Hancock, R. E. (2008). Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. Nature Protocols, 3(2): 163.