# Effects of Temperature on Asymbiotic Seed Germination of *Himantoglossum robertianum* (Loisel.) P.Delforge

Salih PARLAK 💿

Bursa Technical University, Mimar Sinan Campus-Yildirim, Bursa, 16310, TURKİYE salih.parlak@btu.edu.tr

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#### Abstract

*Aim of study:* Despite protection by international agreements, millions of orchid tubers are harvested from their natural distribution areas each year. Of these species, *Himantoglossum robertianum* is locally threatened due to overharvesting and requires precautionary measures to ensure its protection. Reproduction of the species in an asymbiotic environment is imperative for providing ex-situ protection. There are no studies on optimum germination temperature in *H. robertianum*. This study aimed to germinate *H. robertianum* seeds in-vitro under asymbiotic conditions.

*Area of study:* The study was carried out at the Silviculture Laboratories of Bursa Technical University, Faculty of Forestry, Department of Forestry Engineering.

*Material and methods: H. robertianum* seeds were used in the study. Seeds were germinated in five replications at four different temperatures (10, 15, 20, and 25°C ( $\pm$  0.5°C). The study was conducted for 275 days under dark conditions with S1gma-Phytamax P-6668 used as the medium.

*Main results:* The highest germination was 23.8% at 20°C and germination was not obtained at 10°C. While germination was faster at 25°C in the first 18 weeks, germination accelerated at 20°C after 18 weeks.

*Highlights:* These results indicate that temperature is an important factor in the germination of *H. robertainum* seeds.

Keywords: Himantoglossum robertianum, Asymbiotic Germination, Temperature

# Himantoglossum robertianum (Loisel.) P.Delforge'un

# Asimbiyotik Tohum Çimlendirilmesinde Sıcaklığın Etkisi

# Öz

*Çalışmanın amacı:* Uluslararası sözleşmeler tarafından korunmasına rağmen, her yıl doğadan milyonlarca orkide yumrusu sökülmekte ve doğal yayılış alanları yok edilmektedir. Bu türlerden biri olan *Himantoglossum robertianum*, aşırı toplama nedeniyle yok olma tehdidi altındadır ve korunması için önlemler alınmalıdır. Ex-situ korumanın sağlanması için türlerin asimbiyotik ortamda çoğaltılması zorunludur. *H. robertianum*'da optimum çimlenme sıcaklığı ile ilgili bir çalışma bulunmamaktadır. Bu çalışmanın amacı, türün asimbiyotik koşullarda doku kültürü ortamında ve farklı sıcaklıklarda çimlenme koşullarını belirlemektir.

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*Materyal ve yöntem:* Çalışmada *H. robertianum* tohumları kullanılmıştır. Tohumlar dört farklı sıcaklıkta beş tekerrürlü olarak çimlendirilmiştir. Çimlendirme ortamı olarak Sığma-Phytamax P-6668 kullanılmış ve karanlık koşullarda 275 gün süreyle yürütülmüştür.

*Temel sonuçlar:* En yüksek çimlenme 20°C'de %23.8 olarak gerçekleşirken, 10°C'de çimlenme elde edilememiştir. İlk 18 haftada 25°C'de çimlenme hızı yüksek iken daha sonra 20°C'de çimlenme hızlanmıştır.

Araştırma vurguları: Bu sonuçlar, H. robertainum tohumlarının çimlenmesinde sıcaklığın önemli bir faktör olduğunu göstermektedir.

Anahtar Kelimeler: Himantoglossum robertianum, Asimbiyotik Çimlenme, Sıcaklık

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# Introduction

Orchidaceae is one of the largest families of flowering plants, with more than 28,000 species spanning 763 genera (Christenhusz & Byng, 2016). Seed production is high in orchids, with each capsule containing thousands or millions of small seeds, depending on the species (Arditti, 1967; Arditti & Ghani, 2000). Germination is generally difficult in temperate terrestrial orchids (Arditti et al., 1982 Rasmussen, 1995; Miyoshi & Mii, 1998), with only 0.2-0.3% of the millions of seeds in a capsule germinating in nature (Citil & Tekinşen, 2011). Since the tiny seeds have a small embryo with no endosperm, they require a mycorrhizal relationship for germination and development receiving water. in nature, nitrogen, carbohydrates, vitamins and organic compounds via mycobionts (Kauth et al., 2008). Low germination is also attributed to morphological and morphophysiological & Baskin, 2014). dormancy (Baskin Environmental conditions, such as light quality, quantity and temperature, can change the seed germination rate and duration (Lee et al., 2018).

Asymbiotic germination is ideal for studying the germination and development of orchid seeds and seedlings. It is widely used since it is simpler than symbiotic germination and is an excellent technique for studying the effects of abiotic and biotic factors on seed biology (Kauth et al., 2008). More precise results can be obtained from asymbiotic germination studies carried out under controlled conditions in the laboratory compared to symbiotic germination studies.

For many plant species, temperature is a major factor in breaking physiological seed dormancy (Rasmussen, 1995; Baskin & Baskin, 2004). However, little research has explored the effects of temperature on the germination of orchid seeds (Arditti, 1967; Kauth et al., 2008; Johnson, 2011; Calevo & Bazzicalupo, 2020). Additional research is needed to reveal the effects of temperature on germination in terrestrial temperate orchid species (Johnson, 2011).

Orchid seeds can germinate over a wide temperature range (Arditti, 1967), but maximum germination is achieved only within a narrow range between 20 and 25°C

(Rasmussen et al., 1990; Marić, 1995; Kauth et al., 2008; Lee et al., 2018). Although orchid seeds generally germinate in-vitro at constant temperatures. applying alternating temperature regimes is recommended to study germination ecology (Baskin et al. (2006; Kauth et al., 2008; McCormick et al., 2021). In a study of Western European orchids, the best germination occurred at 23°C in continuous darkness (Van Waes & Debergh, 1986; Baskin & Baskin, 2001). These studies provide valuable information regarding the role of temperature in the germination of orchid seeds (Kauth et al., 2008).

Orchid tubers can be ground into salep, a powder widely used for making hot drinks and ice cream. Salep is obtained from 30 species of terrestrial orchids belonging to Orchis, Himantoglossum, Anacamptis, Ophrys. Serapias and Dactylorhiza genera (Sezik, 1967; 1990; Tekinsen & Guner, 2009). Glucomannan and starch, as the major components used to make salep powder, are the primary indicators of the yield and quality of salep powder (Acemi et al., 2019 Teoh, 2019). Those who collect orchid tubers prefer to collect large tuberous species first. Therefore, species with large tubers tend to be harvested frequently. With a harvest rate of 39.5%, Himantoglossum robertianum is the most harvested species for salep production due to its large tubers (Molnár et al., 2017).

*Himantoglossum* is a genus of orchids native to the Canary Islands, Europe, Southwest Asia and Northern Africa that is found in dry, calcareous soils (Rasmussen, 1995; Rossi 2002; Parlak & Tutar, 2012). The plants grow 60-80 cm tall (Davis, 1984; Delfolge, 2006; Rossi, 2002; Teoh, 2016), are easy to find, and are valuable for salep production. Unfortunately, due to the overharvesting of their tubers for salep production, many species in this genus are locally threatened in Türkiye (Dulić et al, 2018; Teoh, 2019).

*In-situ* or *ex-situ* protection measures should be taken, but are limited by a lack of research on *H. robertianum* (Szendrák, 1997; Aybeke, 2013a; 2013b; 2013c; Katsalirou et al., 2017; Katsalirou et al., 2019; Calevo et al., 2020). The reproductive physiology of *H. robertianum* and germination protocols under asymbiotic conditions must be determined to ensure its *ex-situ* protection. Although the effects of temperature on germination have been studied in some orchid species (e.g., *Dactylorhiza majalis* (Rasmussen et al., 1990; Rasmussen & Rasmussen, 1991) and *Psygmorchis pusilla* (Vaz et al., 2004)), no studies on the optimal germination temperature of *H. robertianum* have been reported. Therefore, this study aims to reveal the effects of temperature on the germination of *H. robertianum*.

# **Materials and Methods**

### Materials

*Himantoglossum robertianum* seeds were collected from plants grown by open pollination in 2019 and donated by the Ministry of Agriculture's Agricultural Research Centre in Menemen, İzmir, Türkiye. The seeds were air-dried for 2 weeks at room temperature and stored in Eppendorf® tubes at 4°C until use.

A digitally controlled germination cabinet (Lovibond TC 140 G-Liebherr. Dortmund/Austria). stereomicroscope SZ550-B-ST5-H, (Irmeco, IM Geesthacht/Germany), flow cabinet (biosafety cabinet class II), autoclave (Tomy SX-700e, Tokyo/Japan), pure water device (Elga DV 35-ELGA LabWater/UK) and hydrogen peroxide for seed sterilization (Sigma-Aldrich (Nord., 34.5-36.5%) were used. Sigma-Phytamax P-6668 medium was used for germinating the seeds in sterile 90 mm plastic petri dishes (İsolab). No modifications were made to the medium and 27.3 g/L powder were used to prepare the medium.

# Methods

Medium preparation

The natural distribution of *H. robertianum* occurs mostly on limestone bedrock and calcareous soils (Davis, 1984; Rossi, 2002). Rasmussen (1995) reported that media with a neutral or slightly alkaline pH, which resembles the conditions in their natural habitat, may be suitable for many of the European orchid species. Therefore, the media pH was adjusted to 7 with the use of 0.1 N sodium hydroxide (NaOH). After adding 7 g/L agar, the medium was autoclaved at 121°C for 60 min. Approximately 20 ml of

medium was poured into each petri dish after cooling to 65-70°C.

Sterilization, sowing and incubation of seeds

The *H. robertianum* seeds were placed in 2-ml Eppendorf® tubes and treated with 10% hydrogen peroxide for 60 min before being rinsed three times with pure sterile water. The seeds were sown into sterile plastic petri dishes with a 90 mm diameter using a curved paper clip that was disinfected in alcohol and flamed. The closure of the culture dishes affects germination (Rasmussen, 1995), so the petri dishes were wrapped with a double layer of transparent cling film. Since high germination occurs when seeds of many orchid species are incubated at constant temperatures (Arditti, 1967; Baskin & Baskin, 2014), the petri dishes were placed in a climate cabinet set at 10, 15, 20, and 25°C (± 0.5°C). The glass surface of the climate cabinet was covered with aluminium foil to provide a dark environment. There were five replicates and the average number of full seeds in the petri dishes varied between 105 and 185. The seeds were sown on May 30, 2019, and observations of protocorm development were recorded every 15-40 d for 275 d.

# Seed count and statistical analyses

Seeds were observed and counted according to Szendrák (1997) using a stereomicroscope with a magnification of 3.35 to 180x during the incubation period of 275 d. The germination stages (Figure 1) of the seeds were evaluated according to a modified method by Stewart & Kane (2006). At each counting time the germination rates were calculated using the following formula for each temperature (Eq. 1):

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Germination \ percentage \ (\%) = \frac{seed \ number \ (stages \ 3-5)}{total \ seed \ number \ (stages \ 0-5)} \times 100 \ (1)
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An arc-sin transformation was applied to the germination percentage values. The transformed data were subjected to analysis of variance (ANOVA) and the means were compared with Duncan's multiple range test using SPSS ver. 22. (IBM Corp., Armonk, NY, USA). Significance was defined at the 0.05 level.



Figure 1. Germination stages of asymbiotically cultured *Himantoglossum robertianum in vitro* (adapted from Stewart & Kane, 2006). 0) Ungerminated seed, testa intact; 1) embryo whitening and swelling; 2) enlarged embryo, testa ruptured (35 days after sowing); 3) appearance of rhizoids indicates germination (at least 1 mm); 4) appearance of promeristem; and 5) emergence of the first leaf (seedling).

# **Results and Discussion**

Seeds began to swell within 2 weeks (stage 1) of sowing and germinated to form protocorms within the 35 d after sowing, as shown in Figure 1, stage 2. The study was completed once germination ceased at 275 d. Homogeneous germination did not occur in *H. robertianum* seeds and some of the seeds did not germinate until the end of the study. Rasmussen (1995) reported that germination of *H. robertianum* is sporadic, even when seeds are sown from green capsules, and very

slow, taking up to 10 months. At temperatures both above and below the optimal range, some viable seeds remain dormant.

Temperature was a significant factor in the germination of *H. robertianum* seeds (Table 1). Duncan's test revealed significant differences among all four temperatures (Table 2). The best germination rate was obtained at 20°C, followed by 25°C and 15°C (Table 2; Figure 2). At 10°C, germination was 0%.

Tuote 1. Thai jois of variance for minimuto grossian rober nariant germination						
	Sum of Squares	df	Mean Square	F	Sig.	
Among groups	1964.032	3	654.677	103.252	0.000	
Within groups	88.768	14	6.341			
Total	2052.800	17				

Table 1. Analysis of variance for Himantoglossum robertianum germination

Table	2.	Duncan	test	results	for	germination	rates	of	Himantoglossum	robertianum	by
tempe	ratu	re.									

A		
Temperature (°C)	N	Subset for $alpha = 0.05$
10	5	$0.0000 \pm 0.00000^{a}$
15	5	$11.7531 \pm 1.72935^{\rm b}$
25	4	$17.9284 \pm 2.71872^{\circ}$
20	4	$28.9898 \pm 4.26736^{d}$
Sig.		1.000

Means followed by the same letter indicate no statistical difference at the 5% level.



Figure 2. Seeds germinating at 20°C (left) and 25°C (right) 210 days after sowing.

As with many other species, orchid seeds germinate over a wide temperature range, but maximum germination is achieved only within a narrow range (Kauth et al., 2008). Arditti (1967) reported that orchid seeds germinate best at 20-25°C. For most orchid species, the temperature yielding the highest germination rates is between 22 and 25°C, although some germinate best below 20°C. (Rasmussen, 1995). Although the first germination of H. robertianum (Szendrák, 1997; Calevo et al., 2017) was reported to occur 60 d after sowing, the first protocorm formation began after 35 d in our study. The highest germination rate (23.8%) was observed under dark conditions at 20°C after 275 d (Figure 2). The results of our study are consistent with those of previous studies. Calevo et al., (2017; 2020) found germination rates of 23% and 46.1% for H. robertianum after 180 days at 26±1°C. Tsutsumi et al. (2011) found the highest germination rate at 20°C under dark conditions for Liparis fujisanensis, L. koreojaponica and L. kumokiri. Roca (1984) determined that temperatures of ~20°C favoured in vitro development of alpine plants. Gümüş et al. (2017) found that at the lower temperature, the highest germination rates (20.34-24.41%) were observed for Dactylorhiza nieschalkiorum. On the other hand, Özkoç &

Dalci (1994) obtained the highest germination rate (25.1%) for Orchis laxiflora Lam in Knudson C medium, which does not contain inorganic nitrogen. The optimum temperature range for Dactylorhiza majalis seeds appears to be between 23 and 24.5°C. Germination rates for D. majalis decreased at temperatures below 15°C and above 27°C. These rates were 42% and 21%, respectively, under symbiotic asymbiotic conditions at 23.6°C and (Rasmussen et al., 1990; Rasmussen & Rasmussen, 1991). The best germination rate for *Cattleya purpurata* was 46.5 $\pm$ 6.4% in  $\frac{1}{2}$ MS medium and 26.3±4.3% in KC medium (Bazzicalupo et al., 2021). In another study, Calevo & Bazzicalupo (2020) determined that temperature changes play an important role in the germination of Orchis patens and recommended the application of variable temperature instead of constant temperature for the germination of European orchid species. Arditti (1967) reported that seeds of many orchid species germinate at high rates when incubated at constant temperatures between 20 and 25°C. Van Waes & Debergh (1986) obtained the highest germination rate in 21 of 23 European orchids in a BM1 environment, at 23°C in constant darkness. These studies show that the parameters of orchid seed germination are species-specific (Kaut et al., 2008).

In our study, no seed germination was observed at 10°C (Figure 2). Low temperatures delayed germination and development in a study of *Bletia purpurea* (Johnson & Kane, 2011). Rasmussen (1995) also pointed out that low temperatures may prevent seeds from germinating immediately after dispersal. The seeds of *Liparis* spp. did not germinate at all at 5°C and had the lowest germination at 10°C (Tsutsumi et al., 2011).

In this study, although the germination of seeds was higher at 25°C for the first 18 weeks, the germination at 20°C continued to increase after 18 weeks (Figure 3).



Figure 3. Germination rates of *Himantoglossum robertianum* at 10, 15, 20, and  $25^{\circ}C (\pm 0.5^{\circ}C)$ .

Germination started early at 25°C, and the protocorms that were initially yellow in colour turned brown over time. As a result of the drying of 7 protocorms germinating at 15°C and 8 protocorms at 25°C, the germination graph shows a decreasing trend (Fig. 4). Pierce and Belotti (2011) reported that this occurs because protocorms are left in the medium for too long. Stoutamire (1974) and Neiland (1994) reported that this is common and possibly caused by inappropriate cultural conditions. Additionally, phenolic compounds may exudate during seedling growth and cause browning. This might be intensified by light, high temperatures and oxidizing substances, such as iron, but is reduced by transferring the cultures to a dark environment or by frequent subcultivation of new media (Harbeck, 1968; Haas, 1977; Ponert et al., 2011).



Figure 4. Protocorms that germinate early (A) and begin to turn brown (B)

# Conclusion

Excessive harvesting of orchids tubers from their natural habitats for salep production threatens orchid species survival. Harvesters prefer H. robertianum because of its large tubers and easy identification, and fragmenting its natural limiting distribution areas. Isolation and habitat fragmentation can cause problems, such as insufficient pollination and seed formation. Therefore, it is important to be able to culture this species to prevent it from being overharvested from its natural environment. In this study, germination trials of H. robertianum were carried out at 10, 15, 20, and 25°C ( $\pm 0.5$ °C). The best germination was 23.8% at 20°C, while no germination was observed at 10°C. Temperature is significant in the germination of this endangered species. More germination trials should be conducted at temperatures of 20±3°C to narrow the optimal temperature range. Testing the effects of changing temperature regimes on seed germination would also benefit conservation efforts for *H. robertianum*.

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#### **Ethics Committee Approval**

N/A

#### **Peer-review**

Externally peer-reviewed.

#### **Author Contributions**

The author confirms sole responsibility for the following: study conception and design, laboratory studies, data collection, analysis and interpretation of results, and manuscript preparation.

#### **Conflicts of interest**

The author declares that they have no conflict of interest.

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#### References

- Acemi, A., Çobanoğlu, Ö., Türker-Kaya, S. (2019) FTIR-based comparative analysis of glucomannan contents in some tuberous orchids and effects of pre-processing on glucomannan measurement. *Journal of Science Food and Agriculture*, 99, 3681-3686.
- Arditti, J. & Ghani, A.K.A. (2000). Numerical and physical properties of orchid seeds and their biological implications. *New Phytologist*, 145, 367-421.
- Arditti, J. (1967). Factors affecting the germination of orchid seed. *The Botanical Review*, 33, 1-97.
- Arditti, J. Clements, M.A.; Fast, G., et al. (1982). Orchid seed germination and seedling culture: A manual. In: Orchid Biology: Reviews and Perspectives (Vol II), Cornell University Press, Ithaca, New York, USA.
- (2013a). Morphological Aybeke, M. and histochemical investigations on Himantoglossum robertianum (Loisel.) P. (Orchidaceae) Plant Delforge seeds. **Systematics** and Evolution, DOI 10.1007/s00606-013-0862-2.
- Aybeke, M. (2013b). Embryo and protoplast isolation from *Barlia robertiana* seeds (Orchidaceae). *American Journal of Plant Sciences*, 4, 1-8. doi:10.4236/ajps.2013.46A001.
- Aybeke, M. (2013c). Maceration techniques on Barlia (Orchidaceae) seeds. International Research Journal of Plant Sciences, 4(4), 94-96.
- Baskin, C.C. & Baskin, J.M. (2001). Seeds Ecology, Biogeography, and Evolution of Dormancy and Germination. Academic Press, San Diego, USA.
- Baskin, C.C. & Baskin, J.M. (2004). Determining dormancy-breaking and germination requirements from the fewest seeds, In: Guerrant EO, Havens K, Maunder M (Eds) Ex Situ Plant Conservation: Supporting Species Survival in the Wild. Island Press, Washington, DC, USA.
- Baskin, C.C. & Baskin, J.M. (2014). Seeds: ecology, biogeography, and evolution of dormancy and germination. Elsevier, Kentucky, USA.
- Baskin, C.C., Thompson, K. & Baskin, J.M. (2006). Mistakes in germination ecology and how to avoid them. *Seed Science Research*, 16, 165-168.
- Bazzicalupo, M., Calevo, J., Adamo, M., Giovannini, A., Copetta, A. & Cornara, L. (2021). Seed micromorphology, *in vitro* germination, and early-stage seedling morphological traits of *Cattleya purpurata*

(Lindl. & Paxton) Van den Berg. Horticulturae 7, 480. https://doi.org/10.3390/ horticulturae7110480.

- Calevo, J. & Bazzicalupo, M. (2020). Less is more: low-cost in vitro propagation of an endangered Italian orchid. *Nature Conservation Research*.
  5. 10.24189/ncr.2020.043.
- Calevo, J., Copetta, A., Marchioni, I., Bazzicalupo, M., Pianta, M., Shirmohammadi, N. & Giovannini, A. (2020). The use of a new culture medium and organic supplement to improve in vitro early stage development of five orchid species. *Plant Biosystems* - An International Journal Dealing with All Aspects of Plant Biology, 1-9.doi:10.1080/11263504.2020.1840454.
- Calevo, J., Giovannini, A., Cornara, L. & Peccenini, S. (2017). Asybiotic seed germination of hand-pollinated terrestrial orchids. Proc. VI Int. Symp. on Production and Establishment of Micropropagated Plants, *Acta Horticulture*, 1155. ISHS 2017. DOI 10.17660/ActaHortic.2017.1155.61.
- Christenhusz, M.J.M. & Byng, J.W. (2016). The number of known plants species in the world and its annual increase. *Phytotaxa* 261(3), 201-217.
- Citil, O.B. & Tekinsen, K.K. (2011). A comparative study on fatty acid composition of salep obtained from some orchidaceae species. *Chemistry of Natural Compounds*, 46(6), 943-945.
- Davis, P.H. (1984). *Flora of Turkey and East Aegean Islands*. Vol.8. Edinburgh at the University Press. UK.
- Delfolge, P. (2006). Orchids of Europe, North Africa, and the Middle East. 3 rd edition, Timber Press Inc, Oregon USA.
- Dulić, J., ljubojević, M., Prlanović, I., Barać, G., Narandžić, T. & Ognjanov, V. (2018). Germination and protocorm formation of *Ophrys sphegodes* MILL. – *In Vitro* protocol for a rare orchid species, *Contemporary Agriculture*, 67(3-4), 196-201.
- Gümüş, C., Ellialtıoğlu, Ş.Ş. & Eman, Ş.B. (2017). Studies for obtaining the protocorms and plantlets in Orchis pinetorum, Anacamptis pyramidalis, and Dactylorhiza nieschalkiorum under in vitro conditions. International Journal of Forestry and Horticulture, (IJFH), 3(3), 28-36. http://dx.doi.org/10.20431/2454-9487.0303005.
- Haas, N.F. (1977). Asymbiotische vermehrung Europäischer erdorchideen I. Dactylorhiza sambucina (L.). Soó. Die Orchidee, 28, 27-31.
- Harbeck, M. (1968). Versuche zut Samenvermehrung einiger Dactylorhiza-

Arten. Jehresbericht der naturwissenschaftlichen Vereins in Wuppertal, 21/22, 112-118.

- Johnson, T.R. (2011). Developing a model of orchid seed germination: *In vitro* studies of the threatened Florida species *Bletia purpurea*, Dissertation, University of Florida.
- Johnson, T.R. & Kane, M. (2011). Effects of temperature and light on germination and early seedling development of the pine pink orchid (*Bletia purpurea*). https://doi.org/10.1111/j.1442-1984.2011.00347.x.
- Katsalirou, E., Gerakis, A. & Haldas, X. (2019). Optimal scarification times for seeds of two Mediterranean orchids. *European Journal of Environmental Sciences*, 9(1), 47-52 https://doi.org/10.14712/23361964.2019.6.
- Katsalirou, E., Gerakis, A., Haldas, X. & Deconninck, G. (2017). Optimal disinfection times for seeds of Mediterranean orchids propagated on nutrient media. *European Journal of Environmental Sciences*, 7(2), 119-124.

https://doi.org/10.14712/23361964.2017.10.

- Kauth, P.J., Dutra, D., Johnson, T.R., Stewart, S.L., Kane, M.E. & Vendrame, W. (2008). Techniques and applications of *in vitro* orchid seed germination, *Floriculture, Ornamental, and Plant Biotechnology*: Advances and Topical Issues, Volume V Chapter: Techniques and applications of in vitro orchid seed germination, Global Science Books, UK.
- Lee, Y.I., Chee, E. & Yeung, T. (2018). Orchid Propagation: From Laboratories to Greenhouses-Methods and Protocols, , Springer Science+Business Media, LLC, part of Springer Nature.
- Marić, M.M. (1995). *Plant tissue culture*. Publishing house Draganić, Croatia.
- McCormick, M., Burnett, R. & Whigham, D. (2021). Protocorm-Supporting fungi are retained in roots of mature *Tipularia discolor* orchids as mycorrhizal fungal diversity increases. *Plants*. 10, 1251. https://doi.org/ 10.3390/plants10061251.
- Miyoshi, K. & Mii, M. (1998). Stimulatory effects of sodium and calcium hypochlorite, prechilling and cytokinins on the germination of *Cypripedium macranthos* seed in vitro. *Physiologia Plantarum*, 102, 481-486.
- Molnár, A.V., Nagy, T., Löki, V., Süveges, K., Takács, A., Bódis, J. & Tökölyi, J. (2017). Turkish graveyards as refuges for orchids against tuber harvest. *Ecology and Evolution*, 1–8, DOI: 10.1002/ece3.3562.

- Neiland, M.R.M. (1994). *Reproductive Ecology of British and Mediterranean Orchids*. Dissertation, University of Aberdeen.
- Özkoç, I. & Dalcı, M. (1994). Germination of the seeds of *Orchis laxiflora* Lam. (Orchidaceae) through asymbiotic culture techniques. *Turkish Journal of Botany*, 18, 46-464.
- Parlak, S. & Tutar, M. (2012). Some soil properties of the most collected salep orchids in Karaburun Peninsula. Turkey 2. Orchids and Sahlep Workshop, April 25 to 26, 2012, İzmir.
- Pierce, S., & Belotti, J. (2011). The Conservation of Terrestrial Orchids: from the Alps to the Po Plain of Lombardy. Parco delle Orobie Bergamasche and the Centro Flora Autoctona della Regione Lombardia. The Native Flora Centre, Italy.
- Ponert, J. Vosolsobě, S., Kmecová, K., & Lipavská, H. (2011). European orchid cultivation-from seed to mature plant. *European Journal of Environmental Sciences*, 1(2), 95-107.
- Rasmussen, H.N. & Rasmussen, F.N. (1991). Climactic and seasonal regulation of seed plant establishment in *Dactylorhiza majalis* inferred from symbiotic experiments *in vitro*. *Lindleyana*, 5, 221-227.
- Rasmussen, H.N. (1995). *Terrestrial orchids, from seed to mycotrophic plant*. Cambridge University Press, New York.
- Rasmussen, H.N., Anderson, T.F. & Johansen, B. (1990). Temperature sensitivity of *in vitro* germination and seedling development of *Dactylorhiza majalis* (Orchidaceae) with and without a mycorrhizal fungus. *Plant, Cell and Environment*, 13, 171-177.
- Roca, W.M. (1984). Cassava. In WR Sharp DA Evans, PV Ammirato and Y Yamada (eds.) *Handbook of Plant Cell Culture*, 2. Macmillan, New York.
- Rossi, W. (2002). *Orchidee d'Italia*. Quad. Cons. Natura, 15. Bologna, Min. Ambiente - Ist. Naz. Fauna Selvatica, Italy.
- Sezik, E. (1967). Turkiye'nin Salepgilleri Ticari Salep Çeşitleri ve Özellikle Muğla Salebi Üzerinde Araştirmalar. Dissertation, İstanbul Universitesi.
- Sezik, E. (1990). Turkiye'nin orkideleri. *Bilim ve Teknik*, 269, 5-8.
- Stewart, S.L. & Kane, M.E. (2006). Asymbiotic seed germination and in vitro seedling development of *Habenaria macroceratitis* (Orchidaceae), a rare Florida terrestrial orchid. *Plant Cell, Tissue and Organ Culture*, 86, 147-158.
- Stoutamire, W.P. (1974). Terrestrial orchid seedlings. In: Withner CL, ed. The orchids:

scientific studies. John Wiley & Sons, New York.

- Szendrák, E. (1997). Asymbiotic İn Vitro Seed Germination, Micropropagation and Scanning Electron Microscopy of Several Temperate Terrestrial Orchids (Orchidaceae). Dissertation, University of Nebraska.
- Tekinsen, K.K. & Guner, A. (2009). Chemical composition and physicochemical properties of tubera salep produced from some Orchidaceae species. *Food Chemistry*, 121, 468-471.
- Teoh, E.S. (2016). *Medicinal Orchids of Asia*. Springer, Singapore.
- Teoh, E.S. (2019). Orchids as Aphrodisiac, Medicine or Food. Springer Nature, Switzerland.
- Tsutsumi, C., Miyoshi, K., Yukawa, T. & Kato, M. (2011). Responses of seed germination and protocorm formation to light intensity and temperature in epiphytic and terrestrial *Liparis* (Orchidaceae). *Botany*, 89, 84-848, doi:10.1139/B11-066.
- Van Waes, J.M. & Debergh, P.C. (1986). In vitro germination of some Western European orchids. *Physiologia Plantarum*, 67(2), 253-261, https://doi.org/10.1111/j.1399-3054.1986.tb02452.x.
- Vaz, A.P., Figueiredo-Ribeiro Rd, R. & Kerbauy, G.B. (2004). Photoperiod and temperature effects on in vitro growth and flowering of *P. pusilla*, an epiphytic orchid. *Plant physiology and biochemistry*. PPB, 42(5), 41-415. https://doi.org/10.1016/j.plaphy.2004.03.008.