

Bozok Journal of Science

Volume 1, No 1, Page 16-22 (2023) Research Article

Protective Effect of Taurine and Curcumin on Lung Toxicity of Bisphenol A in Rats

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Abstract

Bisphenol A (BPA) is an endocrine disruptor chemical that is frequently used in industry. Taurine is a low molecular weight organic compound in living organisms. Curcumin is a yellow colored bioactive compound of turmeric with antioxidant properties. In this study, the protective effects of taurine and curcumin on the histopathological changes, antioxidant enzyme activities [superoxide dismutase (SOD), catalase (CAT), glutathione-S-transferase (GST), glutathione peroxidase (GPx)] and malondialdehyde (MDA) levels that BPA may cause in the lung tissue of rats were investigated. Rats were divided into 7 groups. 1st group: control group, 2nd group: oil group, 3rd group: taurine (100 mg/kg day) treated group, 4th group: curcumin (100 mg/kg day) treated group, 5th group: BPA (130 mg/kg day) treated group group, 6th group: BPA plus taurine treated group, 7th group: BPA plus curcumin treated group. After the application by gavage for 4 weeks, a statistically significant increase was observed in the antioxidant enzyme activities (SOD, CAT, GST, GPx). When the groups treated with BPA plus taurine, BPA plus curcumin were compared with the group treated with BPA, a decrease was observed in the MDA levels in the lung tissues of the rats, while a statistically significant increase was observed in SOD, CAT, GST and GPx enzyme activities. In histopathological examinations, it was determined that while BPA caused cell infiltration, hemorrhage, atrophy and emphysema in the lung tissue of rats, taurine and curcumin reduced these pathological changes.

Keywords: Lung, Antioxidants, Bisphenol A, Histopathology, Oxidative stress

1. INTRODUCTION

Some substances that threaten public and environmental health are known as endocrine disruptors. These substances act on the endocrine system, control the release of hormones and cause negative effects on living things [1].

Many chemicals are classified as endocrine disruptors by acting on the endocrine system. Among these, plastics and plasticizers are the endocrine disruptors that living things are most exposed to. The very long half-life of these substances in nature causes accumulations and environmental pollution, and accordingly, they negatively affect the ecosystem and threaten the life of living things [2].

Bisphenol A (4,4'-dihydroxy-2,2-diphenylpropane, BPA) is a crystalline and white colored substance [3]. BPA is an endocrine disruptor and is frequently used in industry. It is found in various daily use products such as water pipes, electronic equipment, paper and toys [4,5]. BPA causes leaching of polycarbonate plastics, especially after exposure to heat. It has been stated that BPA leaking from plastics binds to the estrogen receptor [6]. However, there are studies that show that BPA may have negative effects on the central nervous system and immune system [7,8]. It is stated that BPA, an endocrine disruptor, disrupts the function of sex hormones, hormones secreted from the pancreas and thyroid gland, and has hepatotoxic and negative effects on the immune system [9–11].

Since BPA is also used in food contact materials such as packaging, bottle and can coatings, it may cause consumers to be exposed to BPA from their food and beverage [12–14].

Taurine is an amino acid that contains thiol and is found in many tissues such as brain, lung, liver, kidney and spleen [15]. Meat and seafood are rich in taurine [16]. Taurine is a protective and supportive antioxidant that protects cell integrity and increases body resistance [15]. Taurine has an important property that suppresses cell proliferation and directs tumor cells to apoptosis [17].

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Curcumin (diferuloylmethane) is a phenolic compound derived from turmeric (*Curcuma longa*). It is widely used as a dyestuff, spice and herbal medicine in foods [18]. Curcumin has many pharmacological properties besides its anti-inflammatory and antioxidant effects [19–21].

The aim of this study is to investigate the protective effect of taurine and curcumin, which have antioxidant properties, on the effect of BPA, which is widely used, on the lung tissue of rats. For this purpose, the histopathological effect of BPA in the lung tissue of rats, antioxidant enzyme activities [superoxide dismutase (SOD), catalase (CAT), glutathione-s-transferase (GST) and glutathione peroxidase (GPx)] and malondialdehyde levels possible changes and the protective effects of taurine and curcumin on these changes were investigated.

2. MATERIAL AND METHODS

2.1 Animals and Treatment

The animal experiments in this study were approved by Gazi University Animal Experiments Local Ethics Committee (G.U.ET-14.075). 42 male Wistar rats (250-300 g) were used in the experiments. Rats were fed standard laboratory diet and water.

In this study, 7 groups were formed, with 6 rats in each group.

- 1. Control group: Rats were given 1 ml/kg of distilled water.
- 2. Oil group: Rats were given 1 ml/kg of olive oil.
- 3. Curcumin treated group: 100 mg/kg curcumin was given to rats.
- 4. Taurine treated group: Rats were given 100 mg/kg taurine.
- 5. BPA treated group: 130 mg/kg BPA was given to rats.
- 6. BPA plus curcumin treated group: 100 mg/kg of curcumin and 1 hour later 130 mg/kg of BPA was given rats.
- 7. BPA plus taurine treated group: 100 mg/kg of taurine and 1 hour later 130 mg/kg of BPA was given rats.

Curcumin and BPA were dissolved in olive oil. Taurine was dissolved in distilled water. The substances given to the rats were given daily by gavage and after 4 weeks of treated, the rats were dissected and their lung tissues were taken.

2.2 Preparation of Tissues for Biochemical Studies

Lung tissues were prepared by centrifugation after homogenization for the determination of SOD, CAT, GPx and GST enzyme activities and the MDA levels. MDA levels and antioxidant enzyme activities were determined by measuring in spectrophotometer. Protein concentration Lowry et al. [22] was determined according to the method described.

For the measurement of MDA, Ohkawa et al. [23] was measured in a spectrophotometer at 532 nm, and the results were calculated as nmol/mg protein.

For the determination of SOD enzyme activity in lung tissues, the method of Marklund and Marklund [24] was used and the results are given as U/mg protein. For the determination of CAT enzyme activity, the method determined by Aebi [25] was used and the results are given as mmol/mg protein. The method of Paglia and Valentine [26] was used for GPx activity and was given as nmol/mg protein. For the determination of GST enzyme activity, measurement was made at 340 nm and given as nmol/mg protein.

2.3 Preparation of Tissues for Light Microscopy

Tissues for light microscopy studies were fixed in Bouin's fixative. Sections of 5-7 μ thickness were taken from the lung tissues, stained with hematoxylin-eosin and examined under a microscope and photographed.

2.4 Statistical Analysis

One-way analysis of variance and Tukey test were used in Windows SPSS 23 computer program. P<0.05 was considered statistically significant.

3. RESULTS AND DISCUSSION

3.1 Evaluation of Malondialdehyde Levels

While no statistically significant difference was observed between the control, oil, taurine and curcumin groups in terms of MDA levels, a statistically significant increase was observed in the BPA-treated group compared to the control group. While a significant increase was observed in the BPA plus taurine and BPA plus curcumin treated groups compared to the control group, a statistically significant decrease was observed in the MDA level when compared to the BPA-treated rats (Figure 1).



Figure 1. Comparison of ^a control group, ^bOil treated group, ^cTaurine treated group, ^dCurcumin treated group, ^eBPA treated group and other treated groups. Mean±Standard Deviation (P<0.05)

3.2 Evaluation of Antioxidant Enzyme Activities

No statistically significant difference was observed between control, fat, taurine and curcumin groups in terms of SOD, CAT, GPx and GST enzyme activities in lung tissues. A statistically significant decrease was observed in the BPA treated groups compared to the control group. A statistically significant decrease was observed when the control group and BPA plus taurine and BPA plus curcumin groups were compared, while a statistically significant increase was detected when the BPA plus taurine and BPA plus curcumin groups were compared with the BPA-treated group (Table 1).

able 1	. Antioxidant	enzyme activities	s in lung	tissues of rats a	after BPA,	taurine and	curcumin administration
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Groups	SOD	САТ	GPx	GST	
Control	128+18	75 1+6 7	15 8+3 5	/3 7+3 8	
Control	120-10	/5,1±0,/	- 5,6±5,5	-5,7-5,6	
Oil	138±15	69,7±6,3	42,8±2,8	42,5±2,8	
Taurine	141±22	74,1±5,8	50,8±3,5	45,7±3,3	

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Curcumin	131±14	70,8±6,4	49,8±4,1	44,5±4,6
BPA	74±12 ^{a,b,c,d}	50,5±6,6 ^{a,b,c,d}	36,7±3,8 ^{a,b,c,d}	32,0±4,8 ^{a,b,c,d}
BPA+Taurine	$105{\pm}15$ ^{a,b,c,d,e}	55,5±7,8 ^{a,b,c,d,e}	44,5±3,2 ^{a,b,c,d,e}	38,9±3,5 ^{a,b,c,d,e}
BPA+Curcumin	$109\pm16^{a,b,c,d,e}$	60,2±7,2 ^{a,b,c,d,e}	42,5±3,7 ^{a,b,c,d,e}	40,4±2,9 ^{a,b,c,d,e}
BPA+Taurine BPA+Curcumin	105±15 ^{a,b,c,d,e} 109±16 ^{a,b,c,d,e}	55,5±7,8 ^{a,b,c,d,e} 60,2±7,2 ^{a,b,c,d,e}	44,5±3,2 ^{a,b,c,d,e} 42,5±3,7 ^{a,b,c,d,e}	38,9±3,5 ^{a,b,c,d,e} 40,4±2,9 ^{a,b,c,d,e}

Comparison of ^acontrol group, ^bOil treated group, ^cTaurine treated group, ^dCurcumin treated group, ^eBPA treated group and other treated groups. Mean \pm Standard Deviation (p<0.05)

3.3 Histopathological Evaluation

When the histological preparations of the lungs taken from the control group, fat group, taurine and curcumin-treated rats are examined under the light microscope, it is seen that the alveoli and bronchioles are in normal structure (Figure 2a). No pathological findings were found in the histological examinations of these groups. Mononuclear cell infiltration, reduction in alveolar sacs and consequent emphysema and thickening of the interalveolar septum were observed in the lungs of rats treated with BPA for 4 weeks (Figure 2b). Pathological changes were also observed in the lung tissues of the groups treated with BPA plus taurine and BPA plus curcumin. However, these changes were observed to be less in BPA-treated group compared to lung tissue.



Figure 2. Histology of lung tissues of a. Control group, b. BPA group, c. BPA+Taurine group, d. BPA+Curcumin group rats, A: Alveolar sac, *****; Cell infiltration, **‡**: Interalveolar septum H&E, X100

Endocrine disruptors affect health negatively because they cause hormonal changes in humans and animals. BPA is a compound that disrupts the function and structure of the hormonal system [4]. BPA is used as a monomer in the production of plastics. When BPA is used in packaging such as tin cans, plastic water bottles, they leak into the environment they are in depending on time and temperature [27].

It has been stated that BPA increases the risk of developing cancer [28]. BPA affects heart [29], liver [30], kidney [31] tissues, male and female reproductive systems [32] and pancreatic functions [33] is stated to cause damage. It has been observed that BPA inhibits spermatogenic cell proliferation and increases the number of apoptotic cells in experimental animals [34]. When BPA was administered subacutely to rats, it was observed that it caused pathology in the sarcoplasmic reticulum and mitochondria of myocardial cells of the rats in the study with electron microscopy. In this study, BPA caused pathological changes in the lung tissue of rats. BPA caused cell infiltration in the lungs in light microscopy examinations. This result can be accepted as an indication that BPA causes inflammation in the lungs. In addition, shrinkage and reduction in alveolar sacs were observed in this study.

Many chemical substances cause the formation of free radicals in cells. The system that renders free radicals harmless or eliminates them in the cell is called the antioxidant enzyme system [35]. Reactive oxygen species formed in cells as a result of toxic substances act on fatty acids in membrane structures, causing a hydrogen atom to be detached from the methylene group and initiation of lipid peroxidation. One of the indicators of lipid peroxidation is the change in MDA level. Because MDA is the end product of lipid peroxidation [36]. In this study, BPA caused an increase in the amount of MDA by causing lipid peroxidation in the lung tissue. Taurine and curcumin caused a decrease in the increase in the amount of MDA caused by BPA in the lung tissue.

Enzymes such as SOD, CAT, GPx and GST, which are in charge of the antioxidant enzyme system in the cell, operate to render the reactive oxygen species/free radicals formed in the cell harmless [37]. Substances with cytotoxic properties, especially chemical substances, can suppress the antioxidant enzyme system and cause oxidative stress [15]. As a result, reactive oxygen species, which are formed as a result of various activities in the cell, bind to various molecules in the cell, causing disruption of their functions and ultimately deterioration of cell hemostasis [38]. When BPA was administered orally to rats subacutely, it was observed that it suppressed antioxidant enzyme activities in liver, kidney, heart and testis tissues. Taurine and curcumin reduced the effect of BPA [30–33]. In this study, BPA suppressed the antioxidant enzyme system in the lung tissue, and taurine and curcumin partially prevented this negative effect caused by BPA [39].

4. CONCLUSION

As a result, subacute oral BPA administration caused histopathological changes, lipid peroxidation and suppression of the antioxidant enzyme system in the lungs of rats, and taurine and curcumin reduced the negative effects of BPA. Therefore, when using plastic material products, consumers should pay attention to the plastic numbers, and they should prefer products with reliable plastic numbers in the use of other materials, especially foodstuffs.

ACKNOWLEDGEMENT

We would like to thank Dr. F. G. Apaydın for helping us with our work. This study was presented as a poster at the 23rd National Biology Congress, Gaziantep, 2016. This study was produced from the MSc Dissertation prepared by Elif Atabay.

AUTHOR'S CONTRIBUTIONS

The authors contributed equally.

CONFLICTS OF INTEREST

There is no conflict of interest.

RESEARCH AND PUBLICATION ETHICS

The author declares that this study complies with Research and Publication Ethics.

REFERENCES

- A. Roncaglioni, N. Piclin, M. Pintore, and E. Benfenati, "Binary Classification Models for Endocrine Disrupter Effects Mediated through the Estrogen Receptor," SAR QSAR Environ. Res., vol. 19, pp. 697–733, Oct. 2008, doi:10.1080/10629360802550606.
- [2] R. Garcia Morales, M. Rodríguez Delgado, K. Gomez Mariscal, C. Orona Navar, C. Hernandez Luna, E. Torres, R. Parra, D. Cárdenas-Chávez, J. Mahlknecht, and N. Ornelas-Soto, "Biotransformation of Endocrine-Disrupting Compounds in Groundwater: Bisphenol A, Nonylphenol, Ethynylestradiol and Triclosan by a Laccase Cocktail from *Pycnoporus Sanguineus* CS43," Water Air Soil Pollut., vol. 226, no. 8, pp. 251, Jul. 2015, doi: 10.1007/s11270-015-2514-3.
- [3] R. C. Gupta, Biomarkers in Toxicology. 1st ed. Academic Press, Cambridge, Massachusetts, United States, pp. 459, Jan. 2014.
- [4] S. Flint, T. Markle, S. Thomson, and E. Wallace, "Bisphenol A Exposure Effects and Policy; A Wildlife Perspective," J. Environ. Manage., vol. 104, pp. 19–34, Apr. 2012, doi:10.1016/j.jenvman.2012.03.021.
- [5] Y. Huang, C. Wong, J. Zheng, H. Bouwman, R. Barra, B. Wahlstrom, L. Neretin, and M. Hong, "Bisphenol A (BPA) in China: A Review of Sources, Environmental Levels, and Potential Human Impacts," Environ. Int., vol. 42, pp. 91–99, Jul. 2012, doi: 10.1016/j.envint.2011.04.010.
- [6] A.V. Krishnan, P. Stathis, S. F. Permuth, L. Tokes, and D. Feldman, "Bisphenol-A: An Estrogenic Substance is Released from Polycarbonate Flasks during Autoclaving," Endocrinology, vol. 132, no. 6, pp. 2279–2286, Jul. 1993, doi: 10.1210/en.132.6.2279.
- [7] J. Lopez-Cervantes and P. Paseiro-Losada, "Determination of Bisphenol A in, and Its Migration from, PVC Strechfilm Used for Food Packaging," Food Addit. Contam., vol. 20, no. 6, pp. 596–606, 2003, Jul. doi: 10.1080/0265203031000109495.
- [8] Y. B. Wetherill, L. N. Fisher, A. Staubach, M. Danielsen, R. de Vere White, and K. Knudsen, "Xenoestrogen Action in Prostate Cancer: Pleiotropic Effects Dependent on Androgen Receptor Status," Cancer Res., vol. 65, no. 1, pp. 54–65, Jan. 2005, doi:10.1158/0008-5472.54.65.1.
- [9] J. D. Meeker, A. M. Calafat, and R. Hauser, "Urinary Bisphenol A Concentration in Relation to Serum Thyroid and Reproductive Hormones in Men from an Infertility Clinic," Environ. Sci. Technol., vol. 44, no. 4, pp. 1458–1463, Feb. 2010, doi: 10.1021/es9028292.
- [10] L. F. Doherty, J. G. Bromer, Y. Zhou, T. S. Aldad, and H. S. Taylor, "In Utero Exposure to Diethylstilbestrol (DES) or Bisphenol-A (BPA) Increases EZH2 Expression in the Mammary Gland: An Epigenetic Mechanism Linking Endocrine Disruptors to Breast Cancer," Horm. Cancer, vol. 1, no. 3, pp. 146–155, Jun. 2010, doi:10.1007/s12672-010-0015-9.
- [11] E. Clayton, M. Todd, J. B. Dowd, and A. E. Aiello, "The Impact of Bisphenol A and Triclosan on Immune Parameters in the U.S. Population, NHANES 2003-2006," Environ. Health Perspect., vol. 119, no. 3, pp. 390–396, Mar. 2011, doi: 10.1289/ehp.1002883.
- [12] T. Yoshida, M. Horie, Y. Hoshino, and H. Nakazawa, "Determination of Bisphenol A in Canned Vegetables and Fruitby High Performance Liquid Chromatography," Food Addit. Contam., vol. 18, no. 1, pp. 69–75, Jan. 2001, doi: 10.1080/026520301446412.
- [13] Y. Niu, J. Zhang, Y. Wu, and B. Shao, "Analysis of Bisphenol A and Alkylphenols in Cereals by Automated On-Line Solid-Phaseextraction and Liquid Chromatography Tandem Massspectrometry," J. Agric. Food Chem., vol. 60, no. 24, pp. 6116– 6122, Jun. 2012, doi: 10.1021/jf301401k.
- [14] K. C. Makris, S. S. Andra, A. Jia, L. Herrick, C. A. Christophi, S. A. Synder, and R. Hauser, "Association between Water Consumption from Polycarbonate Containers and Bisphenol A intake during Harsh Environmental Conditions in Summer," Environ. Sci. Technol., vol. 47, no. 7, pp. 3333–3343, Apr. 2013, doi: 10.1021/es304038k.
- [15] Y. Sunucu Karafakioğlu, "Antioxidants and Taurine as an Antioxidant," Kocatepe Vet. J., vol. 3, no. 1, pp. 55–61, Mar. 2010.
- [16] T. Ito, S.W. Schaffer, and J. Azuma, "The Potential Usefulness of Taurine on Diabetes Mellitus and Its Complications," Amino Acids, vol. 42, no. 5, pp. 1529–1539, Mar. 2012, doi:10.1007/s00726-011-0883-5.
- [17] F. Klamt, and E. Shacter, "Taurine Chloramine, an Oxidant Derived from Neutrophils, Induces Apoptosis in Human B Lymphoma Cells through Mitochondrial Damage," J. Biol. Chem., vol. 280, no. 22, pp. 21346–21352, Jun. 2005, doi: 10.1074/jbc.M501170200.
- [18] B. B. Aggarwal, C. Sundaram, N. Malani, and H. Ichikawa, "Curcumin: The Indian Solid Gold," Adv. Exp. Med. Biol., vol. 595, pp. 1–75, 2007, doi: 10.1007/978-0-387-46401-5_1.

- [19] A. B. Kunnumakkara, P. Anand, and B. B. Aggarwal, "Curcumin Inhibits Proliferation, Invasion, Angiogenesis and Metastasis of Different Cancers through Interaction with Multiple Cell Signaling Proteins," Cancer Lett., vol. 269, no. 2, pp. 199–225, Oct. 2008, doi: 10.1016/j.canlet.2008.03.009.
- [20] M. S. Lin, Y. H. Lee, W.T. Chiu, and K.S. Hung, "Curcumin Provides Neuroprotection after Spinal Cord Injury," J. Surg. Res., vol. 166, no. 2, pp. 280–289, Apr. 2011, doi: 10.1016/j.jss.2009.07.001.
- [21] H.S. Abdeen, M. E. El-Houseini, M. El-Sherbiny, R. Tabashy, and A. Salah, "Ex Vivo Assessment of the Protective Effect of Curcumin and Taurine Against Human Hepatocarcinogenesis," J. Basic Appl. Zool., vol. 66, no. 4, pp. 180–187, Aug. 2013, doi:.org/10.1016/j.jobaz.2013.01.005.
- [22] O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. J. Randall, "Protein Measurement with the Folin Phenol Reagent," J. Biol. Chem., vol. 193, no. 1, pp. 265–275, Nov. 1951, doi: 10.1016/S0021-9258(19)52451-6.
- [23] H. Ohkawa, N. Ohishi, and K. Yagi, "Assay for Lipid Peroxides in Animal Tissues by Thiobarbituric Acid Reaction," Anal. Biochem., vol. 95, no. 2, pp. 351–358, Jun. 1979, doi: 10.1016/0003-2697(79)90738-3.
- [24] S. Marklund, and G. Marklund, "Involvement of the Superoxide Anion Radical in the Autoxidation of Pyrogallol and a Convenient Assay for Superoxide Dismutase," Eur. J. Biochem., vol. 47, no. 3, pp. 469–474, Sep. 1974, doi: 10.1111/j.1432-1033.1974.tb03714.x.
- [25] H. Aebi, "Catalase in Vitro," Meth. Enzymol., vol. 105, pp. 121–126, 1984, doi: 10.1016/s0076-6879(84)05016-3.
- [26] D. E Paglia and W. N. Valentine, "Studies on the Quantative and Qualitative Characterization of Erythrocyte Glutathione Peroxidase," J. Lab. Med., vol. 70, no. 1, pp. 158–165, Jul. 1967.
- [27] S. S. Karnam, R. C. Ghosh, S. Mondal, and M. Mondal, "Evaluation of Subacute Bisphenol- A Toxicity on Male Reproductive System," Vet. World, vol. 8, no. 6, pp. 738–744, Jun. 2015, doi: 10.14202/vetworld.2015.738-744.
- [28] S. Jenkins, N. Raghuraman, I. Eltoum, M. Carpenter, J. Russo, and C. A. Lamartiniere, "Oral Exposure to Bisphenol A Increases Dimethylbenzanthracene-Induced Mammary Cancer in Rats," Environ. Health Perspect., vol. 117, no. 6, pp. 910– 915, Jun. 2009, doi:10.1289/ehp.11751.
- [29] F. G. Apaydin, A. Aslanturk, M. Uzunhisarcikli, H. Baş, S. Kalender, and Y. Kalender, "Histopathological and Biochemical Studies on the Effect of Curcumin and Taurine against Bisphenol A Toxicity in Male Rats," Environ. Sci. Pollut. Res., vol. 26, no. 12, pp. 12302–12310, Mar. 2019, doi: 10.1007/s11356-019-04578-4.
- [30] M. Uzunhisarcikli and A. Aslanturk, "Hepatoprotective Effects of Curcumin and Taurine against Bisphenol A-Induced Liver Injury in Rats," Environ. Sci. Pollut. Res., vol. 26, pp. 37242–37253, Dec. 2019, doi:10.1007/s11356-019-06615-8.
- [31] A. Aslanturk and M. Uzunhisarcikli, "Protective Potential of Curcumin or Taurine on Nephrotoxicity Caused by Bisphenol A," Environ. Sci. Pollut. Res., vol. 27, pp. 23994–24003, Apr. 2020, doi: 10.1007/s11356-020-08716-1.
- [32] S. Kalender, F. G. Apaydin, and Y. Kalender, "Testicular Toxicity of Orally Administrated Bisphenol A in Rats and Protective Role of Taurine and Curcumin," Pak. J. Pharm. Sci. vol. 32, no. 3, pp. 1043–1047, May. 2019.
- [33] P. Alonso-Magdalena, S. Morimoto, C. Ripoll, E. Fuentes, and A. Nadal, "The Estrogenic Effect of Bisphenol A Disrupts Pancreatic B-Cell Function in Vivo and Induces Insulin Resistance," Environ. Health Perspect., vol. 114, no. 1, pp. 106–112, Jan. 2006, doi:10.1289/ehp.8451.
- [34] L. Yin, Y. Dai, Z. Cui, X. Jiang, W. Liu, F. Han, A. Lin, J. Cao, and J. Liu, "The Regulation of Cellular Apoptosis by the ROS-Triggered PERK/EIF2α/chop Pathway Plays a Vital Role in Bisphenol A-Induced Male Reproductive Toxicity," Toxicol. Appl. Pharmacol., vol. 314, pp. 98–108, Jan. 2017, doi: 10.1016/j.taap.2016.11.013.
- [35] F. G. Uzun and Y. Kalender, "Protective Effect of Vitamins C and E on Malathion-Induced Nephrotoxicity in Male Rats," Gazi Univ. J. Sci., vol. 24, no. 2, pp. 193–201, Apr. 2011.
- [36] H. Baş, and Y. Kalender, "Nephrotoxic Effects of Lead Nitrate Exposure in Diabetic and Nondiabetic Rats: Involvement of Oxidative Stress and the Protective Role of Sodium Selenite," Environ. Toxicol., vol. 31, no. 10, pp. 1229–1240, Oct. 2016, doi: 10.1002/tox.22130.
- [37] H. Baş, F. G. Apaydın, S. Kalender, and Y. Kalender, "Lead Nitrate and Cadmium Chloride Induced Hepatotoxicity and Nephrotoxicity: Protective Effects of Sesamol on Biochemical Indices and Pathological Changes," J. Food Biochem., vol. 45, no. 7, e13769, Jul. 2021, doi: 10.1111/jfbc.13769.
- [38] C. Adiguzel and Y. Kalender, "Bendiocarb-Induced Nephrotoxicity in Rats and the Protective Role of Vitamins C and E," Environ. Sci. Pollut. Res., vol. 27, no. 6, pp. 6449–6458, Feb. 2020, doi: 10.1007/s11356-019-07260-x.
- [39] E. Atabay, "Protective Role of Taurine and Curcumin on Lung Toxicity of Bisphenol A in Rats," MSc Dissertation, Gazi University, Ankara, Türkiye, 63s. 2017.