

## A New Record of Reduced Chromosome Number in Tenebrionidae (Coleoptera)

Dirim ŞENDOĞAN

Ege University, Faculty of Science, Department of Biology, İzmir, Türkiye  
ORCID ID: Dirim ŞENDOĞAN: <https://orcid.org/0000-0003-2191-2124>

Received: 12.05.2023

Accepted: 29.06.2023

Published online: 30.06.2023

Issue published: 30.06.2023

**Abstract:** The karyological features of *Akis subtricotata* was determined for the first time with conventional and silver nitrate staining. The diploid number  $2n=16$  and meioformula  $7+neoXY$  represents a deviation from the modal karyotype of Coleoptera. The pericentromeric heterochromatin was detected with both Giemsa and silver nitrate staining. In addition to determining a single possible NOR on prophase I nuclei,  $AgNO_3$  revealed that several telomeric regions of mitotic metaphase chromosomes were slightly more argyrophilic.

**Keywords:** Karyotype, NOR, neoXY, *Akis subtricotata*, heterochromatin.

### Tenebrionidae'de (Coleoptera) İndirgenmiş Kromozom Sayısına Yeni Bir Kayıt

**Öz:** *Akis subtricotata*'nın karyolojik özellikleri geleneksel ve gümüş nitrat boyama ile ilk kez belirlenmiştir.  $2n=16$  diploid sayısı ve  $7+neoXY$  formülü, Coleoptera takımının model karyotipinden bir sapma temsil etmektedir. Perisentromerik heterokromatin hem Giemsa hem de gümüş nitrat boyamaları saptanmıştır. Profaz I nükleuslarında olası tek bir NOR belirlenimin yanı sıra,  $AgNO_3$  birkaç mitotik metafaz kromozomunun telomerik bölgelerinin biraz daha arjirofilik olduğunu ortaya çıkartmıştır.

**Anahtar kelimeler:** Karyotip, NOR, neoXY, *Akis subtricotata*, heterokromatin.

#### 1. Introduction

Tenebrionidae is the seventh largest family of Coleoptera with more than 20.000 described species (Fattorini, 2000; Bouchard et al., 2005; Lillig et al., 2012; Tezcan et al., 2012; Slipinsky et al., 2011, McKenna & Farrell, 2009, Iwan & Löbl, 2020). Beetles belonging to the family Tenebrionidae occupy a great array of diverse habitats and show considerable species diversity mainly in arid and semiarid environments. Due to having a worldwide distribution and comprising several agriculturally and economically important species, Tenebrionidae has been the focus of evolutionary biology (Papadopoulou et al., 2009, 2010; Condamine et al., 2013; Lamb & Bond, 2013; Kergoat et al., 2014), biogeography (Juan et al., 1995, 1996a, 1996b; Rees et al., 2001) and ecology studies (Los Santos et al., 2000; Carrara & Flores, 2012; Fattorini & Ulrich, 2012; Fattorini, 2013). However, cytogenetic studies and chromosomal information about the group are still insufficient to represent this diverse family. Possessing small chromosomes is one of the reasons that makes conventional banding techniques and fluorescence labelling difficult and therefore cytogenetic studies scarce (Dutrillaux et al., 2006).

While the chromosome number in Tenebrionidae ranges between 14 and 38, the most prominent diploid number within the family is  $2n=20$ . Considering the studied species, there is a tendency of a decrease in chromosome number in Pimelinae subfamily. However, the species with increased chromosome number are members of the subfamily Tenebrioninae (Juan & Petitpierre, 1991). Even though it has been reported in two genera (*Akis* and *Morica*) so far,  $2n=16$  is the second

most common record of reduced diploid number in Pimelinae (Blackmon & Demuth, 2015). *Akis* is a genus which comprises approximately 34 species in the palearctic region yet only four species (*Akis acumita*, *A. bacarozzo*, *A. bremeri* and *A. discoidea*) have been studied cytogenetically (Juan & Petitpierre, 1991). All four species possess the meioformula of  $7+neoXY$ . It is stated that neoXY system is derived from autosome-gonosome translocations (Schneider et al., 2006; Dutrillaux & Dutrillaux, 2009; Lira-Neto et al., 2012) and it has been frequently reported in Coccinellidae, Chrysomelidae and Scarabaeidae families (Blackmon & Demuth, 2015).

In this study, *Akis subtricotata*, a new record from Türkiye (Keskin & Yağmur, 2008), has been analyzed cytogenetically in order to corroborate the reduced diploid number reported from the genus. Furthermore, we provide the first cytogenetic information about the species by analyzing mitotic and meiotic spreads using conventional and differential staining.

#### 2. Material and Methods

Adult *A. subtricotata* specimens that were collected from the Harran Ruins, Şanlıurfa brought alive to our laboratories in Ege University (İzmir). Mitotic and meiotic plates were obtained from the male gonads applying the microspreading (Chandley et al., 1994) and splashing (Murakami & Imai, 1974) methods. The slides were stained with 4% Giemsa solution for 20 minutes for conventional staining. The silver impregnation method (Patkin & Sorokin, 1983) was applied in order to determine the possible NOR regions.

The chromosome spreads were photographed and

analyzed with Zeiss Axioscope light microscope using ZEN software. The male karyotype and chromosomal measurements were made with Image J software (Rasband, 1997-2015) and Levan plugin (Sakamoto & Zacaro, 2009).

### 3. Results

Spermatogonial plates of *A. subtricornata* showed  $2n=16$  diploid number with meioformula  $7+neoXY$ . The karyotype is composed of 2 subtelocentric, 2 submetacentric, and 3 metacentric pairs of autosomes. The submetacentric neoX is the second largest chromosome of *A. subtricornata* while the neoY is subtelocentric (Fig. 1). In metaphase I the heteromorphic bivalent can be determined as neoXY (Fig. 2a). Giemsa-stained prophase nuclei presented dark stained pericentromeric regions (Fig. 2b).

Silver nitrate revealed a single possible NOR (Fig. 3a) and predominantly stained the pericentromeric regions of the chromosomes at prophase I (Fig. 3b). In silver-stained mitotic metaphase plates, telomeric regions of at least 3 pairs of chromosomes gave relatively high argyrophilic signals along with centromeric regions (Fig. 4).



Figure 1. Karyotype of *Akis subtricornata* (bar = 5µm).

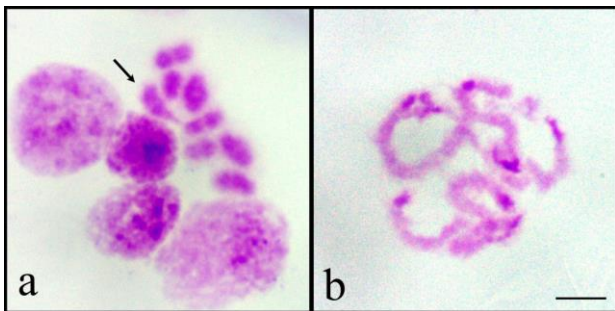


Figure 2. Giemsa stained a) metaphase I, b) prophase I stages (arrow indicates neoXY bivalent, bar = 5µm).

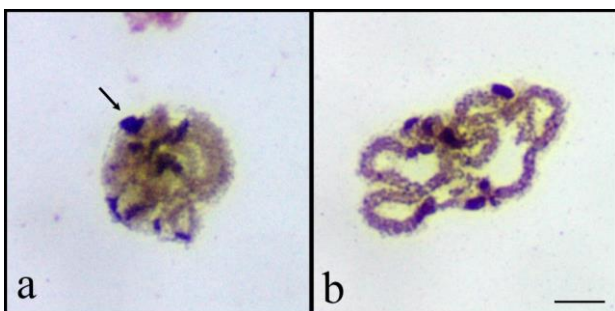


Figure 3. Silver nitrate stained prophase I nuclei a) possible NOR showed with arrow, b) heterochromatin distribution (bar = 5µm).

### 4. Discussion

$2n=20$  is considered to be the modal karyotype for Coleoptera as it is the conserved diploid number among its species.  $Xy_p$  sex determination system and meta-submetacentric morphology of the chromosomes are two other conserved features of the order. However, different chromosome numbers and morphologies resulting from

chromosomal rearrangements like translocations, fusions, and fissions have been reported in various families and subfamilies (Smith & Virkki, 1978; Petitpierre et al., 1991; Cabral-de-Mello et al., 2008; Lira-Neto et al., 2012). The most common reduced diploid number is  $2n=18$  since it only requires the fusion of two autosomal pairs. It is followed by  $2n=16$  which is mostly reported in Chrysomelidae and Coccinellidae families. In Tenebrionidae, this diploid number is only present in the tribe Akidini and it represents 2.4% of the studied Tenebrionids (Blackmon & Demuth, 2015).

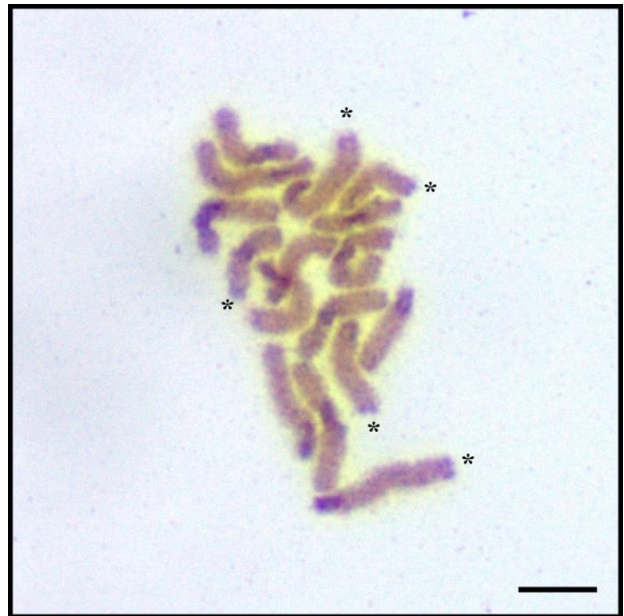


Figure 4. Silver stained mitotic metaphase plate shows argyrophilic telomeric regions (asterisks indicate telomeric heterochromatin, bar = 5µm).

Sex chromosome morphologies and determination systems also vary among the families of Coleoptera. While  $Xy_p$  is the most common system; XO, neoXY, and multiple sex chromosomes have been reported within the order (Pons, 2004; Karagyan et al., 2012; Lira-Neto et al., 2012; Blackmon & Demuth, 2015). In addition to being common in Coccinellidae, Chrysomelidae, and Scarabaeidae, neoXY system has been reported in Tenebrionidae as well (Blackmon & Demuth, 2015). This sex determination system along with the reduced diploid number is present in all studied Akidini representatives (Juan & Petitpierre, 1991).

This study demonstrated that *A. subtricornata* corroborated with the  $2n=16$  reduced chromosome number and neo-sex determination system of the tribe. In addition to reduced diploid number, chromosome morphology is quite deviated from Coleoptera that frequently has metacentric and submetacentric chromosomes (Petitpierre, 1996). In *A. subtricornata* two autosomal pairs along with the neoY chromosome were determined as subtelocentric. It is evident that multiple chromosomal rearrangements such as Robertsonian translocations, pericentric inversions, and fusions are at work in the genome evolution of the genus.

The heterochromatin in Coleoptera can be detected on pericentromeric regions (Rozek, 1998; Pons et al., 2004; Bione et al., 2005; Lachowska et al., 2005; Şendoğan & Alpagut Keskin, 2016; Çalışan & Alpagut-Keskin, 2023) as

well as on telomeric regions (Bione et al., 2005; Colomba et al., 2006; Dutrillaux & Dutrillaux, 2009; Şendoğan et al., 2019). In *A. subtricotata* pericentromeric blocks were demonstrated with silver nitrate and Giemsa staining. On the other hand, high argyrophilic signals were also detected on several chromosomal ends. These dark stained chromosomal regions can be associated with the heterochromatin as they represent more condensed and predominantly stained areas of the chromosomes. Silver nitrate staining also revealed a possible NOR regions on prophase nuclei. It is stated that silver particles highlight the nucleolar protein around the rDNA and; thus, determine the transcriptionally active NOR (Medina et al., 1983; Jordan, 1987; Vitturi et al., 1999; Kavalco & Pazza, 2004; Dutrillaux et al., 2007).

In conclusion, the diploid number ( $2n=16$ ) and meioformula ( $7+neoXY$ ) of *A. subtricotata* was demonstrated for the first time in this study. Karyological findings of the species resembled those of other Akidini yet the sex determination system and reduced diploid number are what make this species intriguing. It is necessary to increase the cytogenetic studies on beetles in order to broaden the data available. Further comparative studies are important in order to understand the karyotype evolution in beetles. Completely understanding the karyotype evolution of the group needs further comparative studies.

**Acknowledgment:** The author would like to thank Dr Bekir KESKİN for his help with collection of the specimens and Nurşen ALPAGUT KESKİN for her revision of the English language of the manuscript. The author would also like to thank to Dr Beril GÜNDOĞAN for her contribution on the laboratory studies.

**Ethics committee approval:** Ethics committee approval is not required for this study.

**Conflict of interest:** The author declares that there is no conflict of interest.

## References

- Blackmon, H., & Demuth, J.P. (2015) Coleoptera Karyotype Database. *The Coleopterists Bulletin*, 69(1), 174-175.
- Bione, E., Camparoto, M. L., & Simões, Z. L. P. (2005). A study of the constitutive heterochromatin and nucleolus organizer regions of *Isocoprini inhiata* and *Diabroctis mimas* (Coleoptera: Scarabaeidae, Scarabaeinae) using C-banding, AgNO<sub>3</sub> staining and FISH techniques. *Genetics and Molecular Biology*, 28, 111-116. <https://doi.org/10.1590/S1415-47572005000100019>
- Bouchard, P., Lawrence, J.F., Davies, A.E., & Newton, A.F. (2005). Synoptic Classification of the World Tenebrionidae (Insecta: Coleoptera) With a Review of Family-Group Names. *Annales Zoologici*, 55(4), 499-530.
- Cabral-de-Mello, D.C., Oliveira, S.G., Ramos, I.C.G. and Moura, R.C. (2008). Chromosome differentiation patterns in Scarabaeinae subfamily (Coleoptera: Scarabaeidae). *Micron*, 39, 1243-1250
- Carrara, R. & Flores, G.E. (2012). Endemic tenebrionids (Coleoptera: Tenebrionidae) from the Patagonian steppe: a preliminary identification of areas of micro-endemism and richness hotspots. *Entomological Science*, 16, 100-111.
- Chandley, A.C., Speed, R.M. & Ma, K. (1994). Meiotic Chromosome Preparation. In: Gosden, J.R. (ed) *Chromosome Analysis Protocols. Methods in Molecular Biology™*, vol 29. Humana Press. <https://doi.org/10.1385/0-89603-289-2-27>
- Colomba, M., Vitturi, R., Libertini, A., Gregorini, A., & Zunino, M. (2006). Heterochromatin of the scarab beetle, *Bubas bison* (Coleoptera: Scarabaeidae) II. Evidence for AT-rich compartmentalization and a high amount of rDNA copies. *Micron*, 37, 47-51. <https://doi.org/10.1016/j.micron.2005.06.004>
- Condamine, F.L., Soldati, L., Clamens, A.L., Rasplus, J.Y. & Kergoat, G.J. (2013). Diversification patterns and processes of wingless endemic insects in the Mediterranean Basin: historical biogeography of the genus *Blaps* (Coleoptera: Tenebrionidae). *Journal of Biogeography*, 40, 1899-1913.
- Çalışan, U., & Alpogut-Keskin, N. (2023). Cytogenetic analysis on *Turkonalassus quercanus* Keskin, Nabozhenko et Alpogut-Keskin, 2017 (Coleoptera: Tenebrionidae: Helopini). *Caucasian Entomological Bulletin*, 19(1), 15-21. <https://doi.org/10.23885/181433262023191-1521>
- Dutrillaux, A.M., Xie, H., & Dutrillaux, B. (2007). Nucleolus and chromosome relationships at pachynema in four Scarabaeoidea (Coleoptera) species with various combinations of NOR and sex chromosomes. *Chromosome Research*, 15, 417-427. <https://doi.org/10.1007/s10577-007-1133-2>
- Dutrillaux, A.M., & Dutrillaux, B. (2009). Chromosome polymorphism and fanconi-like instability in the scarabaeid beetle *Macraspis tristis* from guadeloupe. *Cytogenetic and Genome Research*, 125, 142-148. <https://doi.org/10.1159/000227839>
- Dutrillaux, A.M., Moulin, S., & Dutrillaux, B. (2006). Use of meiotic pachytene stage of spermatocytes for karyotypic studies in insects. *Chromosome Research*, 14, 549-557. <https://doi.org/10.1007/s10577-006-1052-7>
- Fattorini, S. (2000). Dispersal, vicariance and refuges in the Anatolian Pimeliinae (Coleoptera, Tenebrionidae): remarks on some biogeographical tenets. *Biogeographia*, XXI(December), 355-398. <https://doi.org/10.21426/B6110103>
- Fattorini, S. (2013) Tenebrionid beetle distributional patterns in Italy: multiple colonisation trajectories in a biogeographical crossroad. *Insect Conservation and Diversity*, 7, 144-160. <https://doi.org/10.1111/icad.12042>
- Fattorini, S. & Ulrich, W. (2012). Drivers of species richness in European Tenebrionidae (Coleoptera). *Acta Oecologica*, 43, 22-28. <https://doi.org/10.1016/j.actao.2012.05.003>
- Iwan, D. & Löbl, I. (2020). Catalogue of Palaearctic Coleoptera, Vol. 5, Tenebrionoidea, Revised and Updated Second Edition. Koninklijke Brill NV, Leiden, 945p.
- Jordan, J. (1987). At the heart of the nucleolus. *Nature*, 329, 489-490.
- Juan, C., & Petitpierre, E. (1991). Chromosome numbers and sex determining systems in Tenebrionidae. *Advances in Coleopterology*, 167-176.
- Juan, C., Oromi, P., & Hewitt, G.M. (1995). Mitochondrial DNA phylogeny and sequential colonization of Canary Islands by darkling beetles of the genus *Pimelia* (Tenebrionidae). *Proceedings of the Royal Society B: Biological Sciences*, 261(1361), 173-180.
- Juan, C., Ibrahim, K.M., Oromi, P., & Hewitt, G.M. (1996a). Mitochondrial DNA sequence variation and phylogeography of *Pimelia* darkling beetles on the island of Tenerife (Canary Islands). *Heredity*, 77, 589-598. <https://doi.org/10.1038/hdy.1996.186>
- Juan, C., Oromi, P. & Hewitt, G.M. (1996b) Phylogeny of the genus *Hegeter* (Tenebrionidae, Coleoptera) and its colonization of the Canary Islands deduced from cytochrome oxidase I mitochondrial DNA sequences. *Heredity*, 76, 392-403.
- Karagyan, G., Lachowska, D., & Kalashian, M. (2012). Karyotype analysis of four jewel-beetle species (Coleoptera, Buprestidae) detected by standard staining, C-banding, AgNOR-banding and CMA3/DAPI staining. *Comparative Cytogenetics*, 6(2), 183-197. <https://doi.org/10.3897/CompCytogen.v6i2.2950>
- Kavalco, K. F., & Pazza, R. (2004). A rapid alternative technique for obtaining silver-positive patterns in chromosomes. *Genetics and Molecular Biology*, 27(2), 196-198.
- Kergoat, G.J., Soldati, L., Clamens, A.L., Jourdan, H., Jabbour-Zahab, R., Genson, G., Bouchard, P., & Condamine, F.L. (2014). Higher level molecular phylogeny of darkling beetles (Coleoptera: Tenebrionidae). *Systematic Entomology*, 39(3), 486-499. <https://doi.org/10.1111/syen.12065>
- Keskin, B., & Yağmur, E.A. (2008). A new record for the Tenebrionidae fauna of Turkey: *Akis subtricotata* Redtenbacher, 1850 (Coleoptera: Tenebrionidae). *Zoology in the Middle East*, 43, 113-114. <https://doi.org/10.1080/09397140.2008.10638278>
- Lachowska, D., Roz, M., & Holecova, M. (2005). C-banding karyotype and NORs analyse in eight species of Barypeithes Duval from Central Europe (Coleoptera, Curculionidae, Entiminae). *Caryologia*, 58(3), 274-280.
- Lamb, T. & Bond, J.E. (2013). A multilocus perspective on phylogenetic relationships in the Namib darkling beetle genus *Onymacris* (Tenebrionidae). *Molecular Phylogenetics and Evolution*, 66,757-765.

- Lillig, M., Barthes, H.B., & Mifsud, D. (2012). An identification and informative guide to the Tenebrionidae of Malta (Coleoptera). *Bulletin of the Entomological Society of Malta*, 5, 121–160.
- Lira-Neto, A.C., Silva, G.M., Moura, R.C., & Souza, M.J. (2012). Cytogenetics of the darkling beetles *Zophobas aff. confusus* and *Nyctobates gigas* (Coleoptera, Tenebrionidae). *Genetics and Molecular Research*, 11(3), 2432–2440. <https://doi.org/10.4238/2012.June.15.5>
- Los Santos, A., Gomez-Gonzalez, L.A., Alonso, C., Arbelo, C.D. & Nicolas, J.P. (2000). Adaptive trends of darkling beetles (Col. Tenebrionidae) on environmental gradients on the island of Tenerife. *Journal of Arid Environment*, 45, 85–98.
- Medina, F.J., Risueno, M.C., Sanchez-Pina, M.A., Fernandez-Gomez, M.E. (1983). A study of nucleolar silver staining in plant cells. The role of argyrophilic proteins in nucleolar physiology. *Chromosoma*, 88, 149–155.
- Mckenna, D. & Farrell B.D. (2009). Beetles (Coleoptera), The Timetree of Life. In: S.B. Hedges and S. Kumar (ed). Oxford University Press, New York, NY, 278–289.
- Murakami, A. & Imai, H. (1974) Cytological evidence for holocentric chromosomes of the silkworms, *Bombyx mori* and *B. mandarina*, (Bombycidae, Lepidoptera). *Chromosoma*, 47, 167–178.
- Papadopoulou, A., Anastasiou, I., Keskin, B., & Vogler, A.P. (2009). Comparative phylogeography of tenebrionid beetles in the Aegean archipelago: The effect of dispersal ability and habitat preference. *Molecular Ecology*, 18(11), 2503–2517. <https://doi.org/10.1111/j.1365-294X.2009.04207.x>
- Papadopoulou, A., Anastasiou, I., & Vogler, A.P. (2010). Revisiting the Insect Mitochondrial Molecular Clock: The Mid-Aegean Trench Calibration. *Molecular Biology and Evolution*, 27(7), 1659–1672. <https://doi.org/10.1093/molbev/msq051>
- Patkin, E.L. & Sorokin, A.V. (1984) Nucleolus-Organizing Regions Chromosomes in Early Embryogenesis of Laboratory Mice. *Bulletin of Experimental Biology and Medicine* (USSR), 96, 92–94 p.
- Petitpierre, E., Juan, C., & Alvarez-Fuster, A. (1991). Evolution of chromosomes and genome size in Chrysomelidae an Tenebrionidae. *Advances in Coleopterology*, 129–144.
- Petitpierre, E. (1996). Molecular cytogenetics and taxonomy of insects, with particular reference to the Coleoptera. *International Journal of Insect Morphology and Embryology*, 25(1–2), 115–134. [https://doi.org/10.1016/0020-7322\(95\)00024-0](https://doi.org/10.1016/0020-7322(95)00024-0)
- Pons, J. (2004). Evolution of diploid chromosome number, sex-determining systems, and heterochromatin in Western Mediterranean and Canarian species of the genus *Pimelia* (Coleoptera: Tenebrionidae). *Journal of Zoological Systematics and Evolutionary Research*, 42, 81–85. <https://doi.org/10.1046/j.1439-0469.2003.00247.x>
- Pons, J., Bruvo, B., Petitpierre, E., Plohl, M., Ugarkovic, D., & Juan, C. (2004). Complex structural features of satellite DNA sequences in the genus *Pimelia* (Coleoptera: Tenebrionidae): Random differential amplification from a common “satellite DNA library.” *Heredity*, 92(5), 418–427. <https://doi.org/10.1038/sj.hdy.6800436>
- Rasband, W.S. (1997-2015). ImageJ, U.S. National Institutes of Health, Bethesda, Maryland, USA, <http://imagej.nih.gov/ij/>
- Rees, D.J., Emerson, B.C., Oromi, P. & Hewitt, G.M. (2001). The diversification of the genus *Nesotes* (Coleoptera: Tenebrionidae) in the Canary Islands: evidence from mtDNA. *Molecular Phylogenetics and Evolution*, 21, 321–326.
- Rozeq, M. (1998). C. bands and NORs on chromosomes in four species of the genus *Trechus* Clairv. (Coleoptera, Carabidae). *Caryologia*, 51, 189–194.
- Sakamoto, Y. & Zacaro, A.A. (2009) LEVAN, an ImageJ plugin for morphological cytogenetic analysis of mitotic and meiotic chromosomes, Initial version, An open source Java plugin distributed over the Internet from <http://rsbweb.nih.gov/ij/>
- Schneider, M. C., Almeida, M. C., Rosa, S. P., Costa, C., & Cella, D. M. (2006). Evolutionary chromosomal differentiation among four species of *Conoderus* Eschscholtz, 1829 (Coleoptera, Elateridae, Agrypninae, Conoderini) detected by standard staining, C-banding, silver nitrate impregnation, and CMA 3/DA/DAPI staining. *Genetica*, 128, 333–346. <https://doi.org/10.1007/s10709-006-7101-5>
- Slipinski, S.A., Leschen, R.A.B. & Lawrence J.F. (2011). Order Coleoptera Linnaeus, 1758, Animal Biodiversity: an Outline of Higher-Level Classification and Survey of Taxonomic Richness. In: Z.-Q. Zhang (ed). *Zootaxa*, 3148, 203–208.
- Smith, S.G. & Virkki, N. (1978). Animal Cytogenetics, Coleoptera. Berlin, Gebruder Borntraeger.
- Şendoğan, D., & Alpagut-Keskin, N. (2016). Karyotype and sex chromosome differentiation in two *Nalassus* species (Coleoptera, Tenebrionidae). *Comparative Cytogenetics*, 10(3), 371–385. <https://doi.org/10.3897/CompCytogen.v10i3.9504>
- Şendoğan, D., Gündoğan, B., Nabozhenko, M.V., Keskin, B., & Alpagut Keskin, N. (2019). Cytogenetics of *Accanthopus velikensis* (Piller et Mitterpacher, 1783) (Tenebrionidae: Helopini). *Caryologia*, 72(3), 97–103. <https://doi.org/10.13128/caryologia-771>
- Tezcan, S., Keskin, B. & Anlaş, S. (2012). Notes on the Tenebrionidae (Coleoptera) Fauna Collected by Hibernation Trap-Bands and Pitfall Traps in Bozdağlar Mountain, Western Turkey. *Munis Entomology & Zoology*, 7(1), 583–591.
- Vitturi, R., Colomba, M.S., Barbieri, R., & Zunino, M. (1999). Ribosomal DNA location in the scarab beetle *Thorectes intermedius* (Costa) (Coleoptera: Geotrupidae) using banding and fluorescent in-situ hybridization. *Chromosome Research*, 7(1), 255–260. <https://doi.org/10.1023/A:1009270613012>