**ORIGINAL ARTICLE** 

# Cytokine Response to Acute Endurance Exercise: Regular Treadmill versus Lower Body Positive Pressure Treadmill

# Akut dayanıklılık Egzersizine Sitokin Yanıtı: Normal Koşu Bandına Karşı Alt Vücut Pozitif Basınçlı Koşu Bandı

1Muhammet Salih Kırışka ២, 1Muaz Belviranlı 匝, 1Nilsel Okudan 匝

<sup>1</sup>Selçuk University, School of Medicine, Department of Physiology, Division of Sports Physiology, Konya, Turkey, 42131

#### Correspondence

Muaz Belviranlı, Selçuk University, School of Medicine, Department of Physiology, Division of Sports Physiology, Konya, Turkey, 42131

E-Mail: mbelviranli@yahoo.com

#### How to cite ?

Kırışka MS, Belviranlı M, Okudan N. Cytokine response to acute endurance exercise: Regular treadmill versus lower body positive pressure treadmill. Genel Tip Derg. 2024;34(1):94-99.

#### ABSTRACT

Objectives: This study aimed to investigate the cytokine response to acute endurance exercise performed in the lower body positive pressure treadmill (LBPPT) and to compare it with the regular treadmill

Materials and Methods: Eleven healthy physically active men aged between 18-22 years participated in the study. All subjects performed 45 minutes of running exercise at 70% maximal oxygen consumption (VO2max) on the regular treadmill and LBPPT in random order, one week apart. Blood samples were collected at pre-exercise, immediately post-exercise, 30 min post-exercise, and 2 h post-exercise to analyze serum high sensitivity C-reactive protein (hs-CRP), tumor necrosis factor-alpha (TNF-a), and interleukin-8 (IL-8) levels. **Results:** On the regular treadmill, hs-CRP levels were higher immediately, 30 min, and 2 h post-exercise than pre-exercise. In addition, it was lower 2 h post-exercise compared with immediately, and 30 min post-exercise. No significant differences in LBPPT for hs-CRP were observed for any time in the regular treadmill. TNF-a and IL-8 levels were unchanged in response to exercise performed neither on the regular treadmill nor on the LBPPT. **Conclusion:** Acute endurance exercise induces a limited systemic inflammatory response in physically active men.

physically active men

Keywords: Cytokine response, Acute endurance exercise, Lower body positive pressure treadmill, Inflammation

ÖZ

Amaç: Bu çalışma, alt vücut pozitif basınçlı koşu bandı (lower body positive pressure treadmill; LBPPT) ile gerçekleştirilen akut dayanıklılık egzersizine verilen sitokin tepkisini araştırmayı ve normal koşu bandı ile karşılaştırmayı amaçlamıştır. Gereç ve Yöntem: Çalışmaya 18-22 yaş arası 11 sağlıklı, fiziksel olarak aktif erkek katılmıştır. Tüm denekler, bir hafta arayla rastgele sırayla normal koşu bandı ve LBPPT üzerinde Maksimum oksijen tüketimi (VO2max) değerinin % 70'inde 45 dakikalık koşu egzersizi gerçekleştirdi. Serum yüksek hassasiyetli C-reaktif proteini (hs-CRP), tümör nekroz faktör-alta (INF-a) ve interlökin-8 (IL-8) seviyelerini analiz etmek için egzersizden önce, egzersizden hemen sonra, egzersizden 30 dakika sonra ve egzersizden 2 saat sonra kan örnekleri alındı. Bulgular: Normal koşu bandında, hs-CRP seviyeleri egzersizden hemen sonra, 30 dakika ve 2 saat sonra, egzersiz öncesine göre daha yüksekti. Ek olarak, hemen ve egzersizden 30 dakika sonra ile karşılaştırıldığında egzersizden 2 saat sonra daha düşüktü. Herhangi bir zaman noktasında hs-CRP için LBPPT'de anlamlı fark gözlenmedi. Egzersizden hemen sonra ve 30 dakika sonra hs-CRP konsantrasyonu LBPPT'de normal koşu bandına göre daha düşüktü. TNF-a ve IL-8 seviyeleri, ne normal koşu bandında ne de LBPPT'de yapılan egzersize yanıt olarak değişmedi. Sonuç: Akut dayanıklılık egzersizi fiziksel olarak aktif erkeklerde sınırlı bir sistemik inflamatuvar yanıta neden olmaktadır.

Anahtar kelimeler: Sitokin yanıtı, Akut dayanıklılık egzersizi, Alt vücut pozitif basınç koşu bandı, Inflamasyon

#### Introduction

Cytokines are a large family of polypeptides and the cytokine response may depend on the several

proteins that are primarily secreted by myocytes, factors such as training volume (4), body composition adipocytes, immune cells and endothelial cells. In (5) and cardiorespiratory fitness (6). In recent years, the addition to regulating immune functions, cytokines use of new technologies and alternative exercise tools also play a role in cell proliferation, differentiation, developed for rehabilitation, treatment and sportive migration, survival and apoptosis processes. Cytokine performance has been increasing. Among these production can be modulated by a variety of stimuli devices, the lower body positive pressure treadmill such as hormonal stress, oxidative stress and exercise (LBPPT) is becoming important (7). LBPPTs like AlterG (1). Numerous studies have reported that acute are anti-gravity treadmills that can reduce the load on exercise either increases or does not affect cytokine the musculoskeletal system by up to 80% by creating levels in both sedentary subjects and endurance positive pressure around the lower body, thus allowing trained athletes (2,3). Cytokine response to acute treatment or exercise in safe conditions (8). The main exercise is dependent on the mode, intensity, and goals of this device was to decrease the stress on joints, duration (1-3). Additionally, in physically active subjects, ligaments, and tendons, and to decrease ground

Peer-Review: Double anonymized - Two External Plagiarism Checks: Yes - iThenticate Complaints: geneltip@selcuk.edu.tr Copyright & License: Authors publishing with the journal retain the copyright to their work licensed under the CC BY-NC 4.0



reaction force and the amount of load transmitted through tissues of the lower limbs (9). The result is a decrease in impact forces, an increase in stride length and flight time, and a decrease in contact time while running on the machine (7).

The LBPPT is widely used for rehabilitation purposes to support body weight (10), and its potential effects on cardiovascular and metabolic responses have also been extensively studied. It has been reported that LBPPT exercise causes less metabolic and cardiovascular load (7) compared to regular treadmill exercise at the same speed, and that peak oxygen uptake is reduced with an increase in body weight support (9-13). However, it has been reported that LBPPT is not effective in recovery from exercise-induced muscle damage or endurance exercise (14,15).

The fact that LBPPT exercise has different metabolic and cardiovascular demands compared to regular treadmill exercise of the same intensity suggests that the cytokine response may also be different. Also, to our knowledge, there is no research on cytokine responses to acute endurance exercise performed in LBPPT. The primary objective was to examine the responses of cytokines such as interleukin-8 (IL-8), tumor necrosis factor-alpha (TNF-a) and high sensitivity C-reactive protein (hs-CRP) to acute endurance exercise performed on LBPPT with partial body weight support and regular treadmill in physically active men. Our second aim was to compare cytokine responses to regular treadmill and LBPPT.

# **Materials and Methods**

# Participants

Eleven healthy male physically active men aged 18-22 years volunteered to participate. The definition of physically active included at least 1.5-2 hours of daily regular activity, 3-5 days a week, for at least 2 years.

The participants were informed and signed informed consent was obtained. The study was approved by the local Ethical Committee (approval No. 2018/452).

Before the exercise tests, all participant's body weights were measured in light clothes with an electronic scale (Philips HF-390/00, China) and their heights were measured.

# **Research Design**

This was a crossover interventional trial. All participants were invited to the laboratory 4 times. These were the familiarization session, aerobic capacity assessment, test session 1 and test session 2 with at least 1 week between visits. In Sessions 1 and 2, all subjects underwent LBPPT or regular treadmill exercise at 70% of VO2max or vice versa. Venous blood samples were collected for the analysis of IL-8, TNF-a and hs-CRP levels.

# **Exercise Protocols**

# **Measurement of Aerobic Capacity**

Aerobic capacity of the participants was estimated using the 20-m multistage shuttle run test (16,17). During

this test, subjects were asked to run continuously with a sound signal between two cones placed 20 m apart. The speed increased gradually at each stage. The test was terminated until the subject voluntarily exhausted or failed to reach one of the cones a second time before the corresponding beep. The maximum speed and maximal oxygen consumption (VO2max) reached during the test was determined using the equation proposed by Larsen et al (18).

# **Exercise Tests**

After determining the VO2max values of the subjects, the workload corresponding to 70% of the VO2max was determined for each subject according to the American College of Sports Medicine guidelines (19,20). Both regular and LBPPT exercise sessions started at the same hours of the day, lasted 50 minutes, and the subjects performed both exercise sessions at the same pace.

The regular treadmill exercise consisted of a fiveminute warm-up at a self-selected pace followed by a 45-minute running exercise at 70% of the previously calculated VO2max on the regular treadmill (Cosmed T150, Rome, Italy).

LBPPT exercise was performed on a AlterG Anti-Gravity treadmill (Alter-G Inc., Fremont, CA, USA). During the test, the tights necessary for the participants to settle into the treadmill were put on and their connection with the device was provided with the zipper in the tights. The device was calibrated by filling the plastic bag with air. Participants started the session with a warm-up. This included running at a self-selected pace for 5 minutes on a LBPPT without bodyweight support. After that, the device was programmed to 80% body weight support and the running started. The running speed corresponding to 70% of the participant's VO2max was reached gradually in 1 min and they were allowed to run at this speed for 45 minutes.

# **Blood sampling**

Ten milliliter venous blood samples were obtained by a certified phlebotomist pre-exercise, immediately post-exercise, 30 min post-exercise, and 2 h post-exercise to analyze cytokine levels. The samples were stored 30 min until the centrifugation for serum separation, and after that, they were centrifuged at 3000 rpm for 15 min at 4°C. The samples were stored at – 80 °C.

# **Biochemical analysis**

Serum hs-CRP, TNF-a and IL-8 concentrations were analyzed by an ELISA technique, according to the manufacturer's recommendations (Cat no: E-EL-H5134, E-EL-H0109, E-EL-H0048, respectively. Elabscience Biotechnology Co. Ltd, Wuhan, China). The intra-assay coefficient of variance (CV) was 4–6%, whereas the inter-assay CV was 5–7%. The levels of the IL-8, and TNF-a were expressed as pg/mL and levels of the hs-CRP were expressed as mg/L.

# Statistical analysis

Statistical analyses were performed using the IBM SPSS v.25.0 (Chicago, IL, USA). Normality of the variables was

assessed with the Shapiro-Wilks test. For analysis of hs-CRP, TNF-a and IL-8 data, repeated-measures ANOVA  $(2 \times 4, \text{group} \times \text{time})$  was used. Pairwise comparisons were made to explore the differences between the two groups and the four time points in the parameters in which the interaction was observed. Effect size (ES) transformations were used to assess the magnitude of change. Differences between mean values of cytokines that were observed to be statistically significant by comparing pre-exercise levels and postexercise levels and values between the two groups for each time point were converted into ESs using Cohen's d. For ESs, changes were categorized as <0.25 trivial, 0.25-0.49 small, 0.50-1.0 moderate, and >1.0 large changes (21). The negative effect size indicates that effect decreases mean of the experiment group. A p value less than 0.05 was accepted as significant.

### Results

All participants successfully completed the required exercise tests. Anthropometric and physiological characteristics of the subjects are given in Table 1.

Figure 1 shows the changes in circulating hs-CRP, TNF-a and IL-8 levels in response to regular treadmill and LBPPT exercises. Analysis of blood samples for serum hs-CRP concentrations showed a statistically significant time [F(3, 18) = 14.109, p = 0.000] and group x time [F(3, 18) = 4.464, p = 0.016] interactions. On the regular treadmill, measurements taken for serum hs-CRP immediately post- (p = 0.001), 30 min post- (p = 0.000), and 2 h post- (p = 0.038) exercises were higher than pre-exercise. The magnitude of changes between the pre- and immediately post- (ES = 2.10), pre- and 30min post- (ES = 2.14), and pre- and 2 h post-(ES = <math>1.18) exercises were large. In addition, it was lower 2 h postcompared with immediately post- (p = 0.050), and 30 min post-(p = 0.010) exercises. The magnitude of changes between the immediately post-and 2 h post-(ES = -0.91), and 30 min post- and 2 h post- (ES = -0.81)exercises were moderate. No significant differences in LBPPT for serum hs-CRP levels were observed for any time point. Serum hs-CRP concentration immediately post- (p = 0.045, ES = - 0.75) and 30 min post- (p = 0.025, ES = -0.96) exercises were lower in the LBPPT than in the regular treadmill (Figure 1A). Serum hs-CRP levels were within the normal range at all measurement points before and after exercise.

Serum TNF-a concentrations showed no statistically significant time [F(3, 18) = 0.774, p = 0.524] or group x time [F(3, 18) = 0.968, p = 0.429] effect. Analysis of blood samples for serum IL-8 levels demonstrated a statistically significant time [F(3, 18) = 3.810, p = 0.028] but no group x time [F(3, 18) = 0.968, p = 0.429] interaction (Figure 1B). In the LBPPT, serum IL-8 concentration was higher 2 h post-compared to pre- exercise (p = 0.015). The magnitude of changes was large (ES = 1.21). No significant differences were observed in the regular treadmill for any time point (Figure 1C).

#### Discussion

This study showed a statistically significant increase in

the serum hs-CRP concentration following the acute endurance exercise performed on the regular treadmill but not on the LBPPT. Additionally, serum TNF-a levels were unchanged in response to exercise performed neither on the regular treadmill nor on the LBPPT, while serum IL-8 levels were higher 2 h post-exercise than pre-exercise in response to exercise performed on the LBPPT. This study is novel because to our knowledge, there is no study investigating the systemic cytokine response to acute endurance exercise performed in the LBPPT and comparing it with the regular treadmill.

Although various studies examined hs-CRP responses to exercises of different intensities and durations such as marathon (22), ultra-marathon (23), half-marathon (24), triathlon (25) and walking exercise (26), there are only a few reports regarding the acute endurance exercise (27). Our results confirmed that acute endurance exercise leads to elevation in systemic hs-CRP concentration. In our study, the post-exercise increment in hs-CRP after acute endurance exercise was significant and large in magnitude (ES = 2.10, 2.14, and 1.18 immediately post-exercise, 30 min postexercise, and 2 h post-exercise, respectively). This hs-CRP response to exercise seems to be proportional to the duration and intensity of the exercise as well as muscle injury (3) and it increased between 122% and 2000% immediately after the marathon (22), while it increased by 81% after acute endurance exercise (27), which is in line with our findings. However, no significant changes in hs-CRP levels were observed after 30 min of walking on a treadmill at the 50% VO2max (26). Additionally, Jatene et al (28) claimed that serum hs-CRP levels should be an internal indicator of the exercise load.

CRP is a widely used biomarker of inflammation, and it is considered an indicator of the likelihood of cardiovascular disease. Elevated levels of CRP have been independently associated with the progression of atherosclerosis in humans (29,30). The hs-CRP provides a more sensitive and accurate measurement of CRP at lower concentration levels. In this study, the concentrations of hs-CRP significantly increased in the regular treadmill phases when compared to baseline level. The mechanism in which regular treadmill leads to an increase in hs-CRP may be related to the activation of a pro-inflammatory signaling pathway that is regulated by toll-like receptors (TLRs) and nuclear factor kappa-B (NF-KB) (31). TLRs, one of the innate immune components, are stimulated by exercise and induce the production of inflammatory cytokines. In addition, the NF-KB signaling pathway can be activated in a redox-sensitive manner during muscular contraction, presumably due to increased oxidant production (32).

Our study data show that hs-CRP concentrations were lower at 2 h post-exercise than immediately post-exercise and 30 min post-exercise, respectively. The 2 h post-exercise decrement in hs-CRP levels was significant but moderate in magnitude (ES = - 0.91, - 0.83; immediately post-exercise, and 30 min postexercise, respectively). Previous studies have shown

Characteristics	Mean ± SD
Age (years)	19.54 ± 0.89
Height (m)	1.75 ± 0.06
Weight (kg)	69.10 ± 6.66
VO <sub>2max</sub> (mL/kg/min)	57.90 ± 2.06
Running speed (km/h)	11.1 ± 0.4

Data are presented as mean  $\pm$  standard deviation (SD).  $\rm VO_{2max}$  maximal oxygen consumption ( $\rm VO_{2max}).$ 

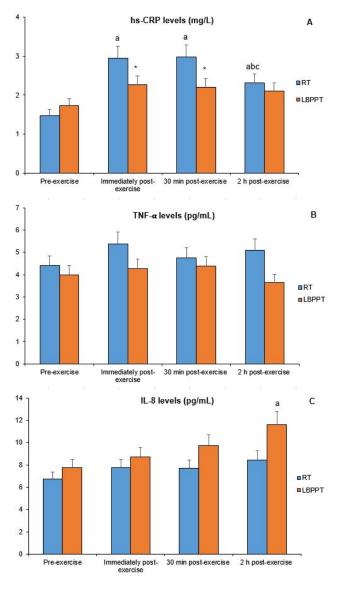


Figure 1. Systemic cytokine concentrations in response to running on the regular treadmill and lower body positive pressure treadmill

Data are presented as mean ± standard deviation (SD). RT: regular treadmill, LBPPT: lower body positive pressure treadmill, hs-CRP: high-sensitivity C-reactive protein, TNF-a: tumor necrosis factor-alpha, IL-8: interleukin-8. °p < 0.05 compared to pre-exercise, °p < 0.05 compared to 30 min post-exercise, °p < 0.05 compared to 30 min post-exercise, °p < 0.05 compared to regular treadmill.

that recovery time takes 2-6 days after a marathon race, 48 hours after a football match (28), and 5 days after a 3-stage trial running race (33). These data indicate that recovery time after exercise also depends on the intensity of the exercise.

In this study, serum hs-CRP concentrations did not change in response to acute endurance exercise performed on LBPPT. In addition, hs-CRP levels immediately post-exercise and 30 min post-exercise were lower than exercise performed on a regular treadmill and the effects size of the decrement was - 0.75, and - 0.96, respectively. These data indicate that the exercise performed on the LBPPT causes less systemic cytokine response than the exercise performed on the regular treadmill at the same speed. Previous studies have shown that the metabolic cost of running with body weight support in LBPPT is lower than running at the same intensity on a regular treadmill, and that exercising on such treadmills can reduce metabolic cost as body weight decreases (11,12).

In this study, serum TNF-a levels changed in response to exercise performed neither on the regular treadmill nor on the LBPPT, while serum IL-8 levels were higher 2 h post-exercise than pre-exercise in response to exercise performed on the LBPPT. In addition, there was no significant difference between the normal treadmill and LBPPT in terms of TNF-a and IL-8. It has been reported that the acute responses of the TNF-a and IL-8 to exercise depends on the intensity and duration of effort (34). Studies have shown that circulating TNF-a and IL-8 levels only increase in response to muscle damage during very intense or strenuous exercises such as marathon running, but do not change in response to acute endurance exercises (35,36). In this context, it was reported that IL-8 and TNF-a levels increased immediately after a marathon race (37,38). In addition, Gokbel et al. (39) reported that repeated bouts of supramaximal exercise did not affect plasma TNF-a levels in sedentary subjects. To our knowledge there is no study investigating the TNF-a and IL-8 response to acute endurance exercise performed in the LBPPT and comparing it with the regular treadmill. However, West et al (14) compared the effect of LBPPT with cycling and stretching exercises on TNF-a levels in the recovery period from acute exhaustive exercise and showed that all three exercise types had similar effects and there was no difference between the aroups.

Although these results provide new information, we recognize that there are some limitations to our study design. One of the limitations inherent in this study was the lack of funding to conduct more biochemical tests that could provide more information on indicators of immune function such as macrophages, neutrophils and eosinophils. The second limitation was that, due to technical difficulties, exercise intensity was not measured with markers of exercise intensity such as heart rate, oxygen consumption or Borg scale during the LBPPT exercise, and only exercised at 80% body weight support. Although it has been shown in previous studies that exercises performed with different body weight support cause different metabolic responses, we could not test this situation in our study. The third limitation of our study is the relatively small and only male sample size included in the study. Despite the appropriate sample size calculation, further randomized controlled trials with larger sample sizes are needed to address the limitations of the current study. Another limitation of our study is that we only examined the time up to 2 hours after exercise. To observe changes in cytokine levels in more detail, more samples are needed over a wider time period. Future studies should analyze these factors over a broader time period.

#### Conclusion

In this study, in response to regular treadmill exercise and LBPPT, there was only a significant change in hs-CRP levels, but no changes in TNF-a and IL-8 levels. This suggests that acute endurance exercise causes a limited systemic inflammatory response in physically active men. However, more comprehensive studies involving more diverse populations are required in the future to clarify the potential effects of LBPPT.

#### Acknowledgements

The authors would like to thank the participants for volunteering the in time and efforts. This study was produced from the MSc thesis of Muhammet Salih Kırışka

#### **Conflict of Interest**

None of the authors has any conflict of interest to disclose.

#### Author contributions

MSK, MB and NO conceived and designed the study. MSK and MB performed the experiments. MB wrote the paper. NO reviewed and edited the manuscript. All authors read and approved the manuscript for publication

#### References

1.Cannon JG. Inflammatory cytokines in nonpathological states. News Physiol Sci 2000;15:298-303.

2.Nieman DC, Henson DA, Smith LL, Utter AC, Vinci DM, Davis JM, et al. Cytokine changes after a marathon race. J Appl Physiol 2001;91:109-14.

3.Nieman DC, Konrad M, Henson DA, Kennerly K, Shanely RA, Wallner-Liebmann SJ. Variance in the acute inflammatory response to prolonged cycling is linked to exercise intensity. J Interferon Cytokine Res 201232:12-7.

4.Rämson R, Jürimäe J, Jürimäe T, Mäestu J. The influence of increased training volume on cytokines and ghrelin concentration in college level male rowers. Eur J Appl Physiol 2008;104:839-46.

5.Beavers KM, Brinkley TE, Nicklas BJ. Effect of exercise training on chronic inflammation. Clin Chim Acta 2010;411:785-93.

6. Jürimäe J, Tillmann V, Purge P, Jürimäe T. Body composition, maximal aerobic performance and inflammatory biomarkers in endurance-trained athletes. Clin Physiol Funct Imaging 2017;37:288-92.

7.De Heer HD, Kline JR, Charley B. Anti-Gravity treadmill training for prevention and rehabilitation of running injuries. In: Harrast MA (editor). Clinical Care of the Runner. Elsevier, 2020:113-30.

8.Cutuk A, Groppo ER, Quigley EJ, White KW, Pedowitz RA, Hargens AR. Ambulation in simulated fractional gravity using lower body positive pressure: cardiovascular safety and gait analyses. J Appl Physiol 2006;101:771-7.

9.Raffalt PC, Hovgaard-Hansen L, Jensen BR. Running on a lower-body positive pressure treadmill: VO2max, respiratory response, and vertical ground reaction force. Res Q Exerc Sport 2013;84:213-22.

10.Patil S, Steklov N, Bugbee WD, Goldberg T, Colwell CW Jr, D'Lima DD. Anti-gravity treadmills are effective in reducing knee forces. J Orthop Res 2013;31:672-9.

11.Farina KA, Wright AA, Ford KR, Wirfel LA, Smoliga JM. Physiological and biomechanical responses to running on lower body positive pressure treadmills in healthy populations. Sports Med 2017;47:261-75.

12.Barnes KR, Janecke JN. Physiological and biomechanical responses of highly trained distance runners to lower-body positive pressure treadmill running. Sports Med Open 2017;3:41.

13.Gojanovic B, Cutti P, Shultz R, Matheson GO. Maximal physiological parameters during partial body-weight support treadmill testing. Med Sci Sports Exerc 2012;44:1935-41.

14.West AD, Cooke MB, LaBounty PM, Byars AG, Greenwood M. Effects of G-trainer, cycle ergometry, and stretching on physiological and psychological recovery from endurance exercise. J Strength Cond Res 2014;28:3453-61.

15.Cooke MB, Nix CM, Greenwood LD, Greenwood MC. No differences between Alter G-trainer and active and passive recovery strategies on isokinetic strength, systemic oxidative stress and perceived muscle soreness after exercise-induced muscle damage. J Strength Cond Res 2018;32:736-47.

16.Léger LA, Lambert J. A maximal multistage 20-m shuttle run test to predict VO2 max. Eur J Appl Physiol Occup Physiol 1982;49:1-12.

17.Léger LA, Mercier D, Gadoury C, Lambert J. The multistage 20 metre shuttle run test for aerobic fitness. J Sports Sci 1988;6:93-101.

18.Larsen GE, George JD, Alexander JL, Fellingham GW, Aldana SG, Parcell AC. Prediction of maximum oxygen consumption from walking, jogging, or running. Res Q Exerc Sport 2002;73:66-72.

19.Pescatello LS, Arena R, Riebe D, Thompson P (Editors). ACSM's guidelines for exercise testing and prescription, 9th ed.; Wolters Kluwer/Lippincott Williams & Wilkins Health: Philadelphia, PA, USA, 2014; p. 169-73.

20.Görgens SW, Eckardt K, Jensen J, Drevon CA, Eckel J. Exercise and regulation of adipokine and myokine production. Prog Mol Biol Transl Sci 2015;135:313-36.

21.Rhea MR. Determining the magnitude of treatment effects in strength training research through the use of the effect size. J Strength Cond Res. 2004;18:918-20.

22.Weight LM, Alexander D, Jacobs P. Strenuous exercise: Analogous to the acute-phase response? Clin Sci (Lond) 1991;81:677-83.

23.Fallon KE. The acute phase response and exercise: the ultramarathon as prototype exercise. Clin J Sport Med 2001;11:38-43.

24.Jürgenson J, Serg M, Kampus P, Kals J, Zagura M, Zilmer K, et al. Effect of half-marathon running on arterial stiffness and blood biomarkers in high-level and recreational male athletes. J Sports Sci Med 2021,548-56.

25.Taylor C, Rogers G, Goodman C, Baynes RD, Bothwell TH, Bezwoda WR, et al. Hematologic, iron-related, and acute-phase protein responses to sustained strenuous exercise. J Appl Physiol 1987;62:464-9.

26.Markovitch D, Tyrrell RM, Thompson D. Acute moderate-intensity exercise in middle-aged men has neither an anti- nor proinflammatory effect. J Appl Physiol. 2008;105:260-5.

27.Shojaei EA, Jafari A, Farajov A. Effect of acute moderate aerobic cycling on systemic inflammatory responses in young untrained men. Sci Sports 2011;26:298-302.

28.Jatene P, Dos Santos GS, Portella DL. C-Reactive protein serum levels as an internal load indicator of sprints in competitive football

matches. Int J Sports Med 2019;40:762-7.

29. Chen J, Huang L, Song M, Yu S, Gao P, Jing J. C-reactive protein upregulates receptor for advanced glycation end products expression and alters antioxidant defenses in rat endothelial progenitor cells. J Cardiovasc Pharmacol 2009;53:359-67.

30.Zhou Y, Han W, Gong D, Man C, Fan Y. Hs-CRP in stroke: A metaanalysis. Clin Chim Acta 2016;453:21-7.

31.Lee SJ, Kim J. Inflammation and insufficient or disordered sleep. Korean J Clin Lab Sci 2015;47:97–104.

32.Ji LL, Gomez-Cabrera MC, Steinhafel N, Vina J. Acute exercise activates nuclear factor (NF)-kappaB signaling pathway in rat skeletal muscle. FASEB J 2004;18:1499-506.

33.Giovanelli N, Lazzer S, Cauci S. Muscle damage and inflammatory status biomarkers after a 3-stage trail running race. J Sports Med Phys Fitness 2020;60:1486-92.

34.Nieman DC, Wentz LM. The compelling link between physical activity and the body's defense system. J Sport Health Sci 2019;8:201-17.

35.Nieman DC, Davis JM, Henson DA, Walberg-Rankin J, Shute M, Dumke CL, et al. Carbohydrate ingestion influences skeletal muscle cytokine mRNA and plasma cytokine levels after a 3-h run. J Appl Physiol 2003;94:1917-25.

36.Nieman DC, Davis JM, Henson DA, Gross SJ, Dumke CL, Utter AC, et al. Muscle cytokine mRNA changes after 2.5 h of cycling: influence of carbohydrate. Med Sci Sports Exerc 2005;37:1283-90.

37.Suzuki K, Nakaji S, Yamada M, Liu Q, Kurakake S, Okamura N, et al. Impact of a competitive marathon race on systemic cytokine and neutrophil responses. Med Sci Sports Exerc 2003;35:348-55.

38.Santos VC, Sierra AP, Oliveira R, Caçula KG, Momesso CM, Sato FT, et al. Marathon race affects neutrophil surface molecules: Role of inflammatory mediators. PLoS One. 2016;11:e0166687.

39.Gökbel H, Okudan N, Gül I, Belviranli M, Gergerlioğlu HS, Başaral MK. Effects of repeated bouts of supramaximal exercise on plasma adiponectin, interleukin-6, and tumor necrosis factor-a levels in sedentary men. J Strength Cond Res 2012;26:1675-9.