

Baseline Evaluation of Causative Agents and Their Susceptibility Patterns of Late-Onset Blood Stream Infections in A NICU; Gram-Negative Domination

Bir Yenidoğan Yoğun Bakım Ünitesinde Geç Başlangıçlı Kan Akımı Enfeksiyonlarına Neden Olan Ajanlar ve Antibakteriyel Duyarlılıklarının Değerlendirilmesi; Gram Negatifler Baskın
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Abstract

Background Blood stream infections (BSI) are the most common hospital-acquired infections in neonatal intensive care units (NICU). Despite the improvement of medical capabilities with neonatal care, diagnosis of BSI in neonates is still problematic. Blood cultures are main confirmatory tool for neonates with clinical signs and symptoms of BSI. They are also useful for determination of pathogen and their antimicrobial susceptibilities for surveillance purposes. In this retrospective study, we aim to determine the causative bacteria of late-onset neonatal sepsis and their antimicrobial susceptibilities to create a baseline data for active surveillance and empiric antimicrobial regimens.

Materials and Methods: The study was conducted at NICU during three-year period from opening date 2013 to 2015. We retrospectively evaluated the blood cultures of the neonates with the proven diagnosis of late-onset blood stream infection.

Results: Total number of 1245 blood culture samples of 516 neonates with suspected blood stream infection was evaluated during the three-year study period. Among 516 neonate included in the study, 35 of them (6.8%) suffered BSI. Causative agent of BSI was reported in 2.8% (n=35) of all the samples. When the results of the blood cultures were evaluated through the years, we did not determine any significant difference (p>0.05). Gram-negative bacteria caused late-onset neonatal sepsis cases (88.9%) more commonly than gram positive ones (17.1%). *K. pneumoniae* was identified as the most predominant bacterium (45.7%), followed by *E. cloaca* (11.4%) and other enteric bacilli. Coagulase negative staphylococci and *Enterococcus faecalis* were the Gram-positives determined in 17.1% of the cases. Carbapenems for the Gram-negative bacteria and glycopeptides for the Gram-positive bacteria were the most susceptible antimicrobials as expected.

Discussion and Conclusion: The study was conducted at NICU during three-year period from opening date 2013 to 2015. We retrospectively evaluated the blood cultures of the neonates with the proven diagnosis of late-onset blood stream infection.

Key words: Neonate, Blood stream infections, blood cultures, surveillance, empirical treatment.

Özet

Giriş: Kan akışı enfeksiyonları (KAE), yenidoğan yoğun bakım ünitelerinde (YYBÜ) hastane kaynaklı enfeksiyonlardır. Yenidoğan bakımında tıbbi tekniklerdeki gelişmeye rağmen yeni doğanlarda KAE tanısı hala sorunludur. Kan kültürleri, BSI'nın klinik bulgular ve belirtileri olan yenidoğanlar için temel doğrulayıcı yöntemlerdir. Kan kültürleri ayrıca patojenlerin ve bunların süreyans amaçlı antimikrobiyal duyarlılıklarının belirlenmesinde de yararlıdır. Bu retrospektif çalışmada, aktif gözetim ve ampirik antimikrobiyal rejimler için başlangıç verileri oluşturmak için geç başlangıçlı yenidoğan sepsis etkeni bakterileri ve bunların antimikrobiyal yatkınlıklarını belirlemeyi amaçladık.

Materyal ve Metod: Çalışma, YYBÜ'de 2013'ten 2015'e kadar üç yıllık süre boyunca gerçekleştirildi. Geç retinoptero gelişimi tanısı ile yenidoğanların kan kültürlerini retrospektif olarak değerlendirdik.

Bulgular: Üç yıllık çalışma süresi boyunca Kan akımı şüphesi bulunan 516 yenidoğanın toplam 1245 kan kültür örneği değerlendirildi. Çalışmaya dahil edilen 516 yenidoğanın 35'inde (% 6.8) KAE vardı. BSI'nın etken ajanı tüm örneklerin % 2.8'inde (n = 35) bildirildi. Kan kültürlerinin sonuçları yıllara göre değerlendirildiğinde anlamlı fark tespit etmedik (p>0.05). Gram negatif bakteriler geç başlangıçlı yenidoğan sepsis vakalarına (% 88.9) gram pozitiflere (% 17.1) göre daha sık neden oldu. *K. pneumoniae* en baskın bakteri (% 45.7), ve ardından *E. cloaca* (% 11.4) diğer enterik basil olarak tanımlandı. Olguların % 17.1'inde koagülaz negatif stafilokoklar ve *Enterococcus faecalis* gram pozitif olarak bulundu. Beklenildiği gibi Gram negatif bakteriler için karbapenemler ve Gram pozitif bakteriler için glikopeptitler en duyarlı antimikrobiyal maddelerdi.

Tartışma ve Sonuç: Bu çalışmada yenidoğanların kan kültürü örneklerinden ağırlıklı olarak ve dikkat çekici derecede Gram-negatif bakteriler izole edilmiştir. Bu çalışmadan elde edilen antimikrobiyal yatkınlık test sonuçlarına göre, daha dirençli izolatlar için karbapenem püskürtmeyle birlikte alternatif olarak amikasin ve piperasilin-tazobaktam kullanımı ampirik tedaviyi planlamak için şimdilik mantıklı bir yaklaşım gibi gözükmektedir.

Anahtar kelimeler: Yenidoğan, Kan akımı enfeksiyonları, kan kültürleri, sürvelans, ampirik tedavi

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Introduction

Blood stream infections (BSI) are the most common hospital-acquired infections in neonatal intensive care units (NICU). All premature infants possess the risk for blood stream infection at a rate of 21-43%¹. Despite all the improvements and the interventions related with neonatal care are not enough to diminish the rates of morbidity and mortality related to the BSI in NICU^{2,3}. Patients followed in a NICU are very vulnerable to hospital-acquired infections since they do not have a mature and fully functional immune system and their poorly developed skin/mucous barriers do not prevent transmission of potentially pathogenic infectious agents. Very low birth weight and low gestational age are determined as additive risk factors for infections in neonates. Other documented risk factors are previous exposure to broad-spectrum antimicrobials, extreme invasive procedures, male sex, use of steroids and intravenous lipids^{4,5}.

BSI in neonates are mainly classified into two as early-onset neonatal sepsis and late-onset neonatal sepsis. This classification aids to differentiate potential pathogens. Early-onset neonatal sepsis is defined as sepsis occurring in the first 24-72 hours after the birth^{6,7}. Group B streptococci and *Escherichia coli* are common cause of early-onset neonatal sepsis, which are transmitted vertically. Neonatal sepsis arising after 24-72 hours after the birth is called as late-onset neonatal sepsis. Gram-negative enteric bacilli (mainly *Klebsiella pneumoniae* and *Escherichia coli*) and Gram-positive cocci (mainly coagulase-negative staphylococci) are commonly isolated from the patients with late-onset neonatal sepsis. *Candida* species are also increasingly isolated in neonates with gestational age lower than 28 weeks and very low birth weight^{4,5,8,9}.

Despite the improvement of medical capabilities with neonatal care, diagnosis of BSI in neonates is still problematic. Blood parameters are not dependable to set accurate diagnosis. Blood cultures are main confirmatory tool for neonates with clinical signs and symptoms of BSI¹⁰. Blood cultures are also useful for determination of pathogen and their antimicrobial susceptibilities for surveillance purposes. Possible pathogens may differ for each NICU and their distribution and resistance patterns may change over time. Appropriate empiric antimicrobial treatment schemes can only be

suggested by evaluating the data from blood cultures. In this retrospective study, we aim to determine the causative bacteria of late-onset neonatal sepsis and their antimicrobial susceptibilities to create a baseline data for active surveillance and empiric antimicrobial regimens.

Materials and Methods

Setting-Neonatal intensive care unit

NICU of Sakarya University Training and Research Hospital is a level III NICU and has 12 incubators and cradles. It was first begun serving in the year 2013. There are 3 neonatologist, 2 assistant doctors, 25 nurses and 5 other staff working. Approximately 172/year newborns are admitted to the unit. The unit serves not only the obstetric clinic of the hospital but also serves as a regional center for critically ill neonates from the other hospitals in the city and neighboring cities.

Patients

The study was conducted at NICU during three-year period from opening date 2013 to 2015. We retrospectively evaluated the blood cultures of the neonates with the proven diagnosis of late-onset blood stream infection. Patients' data were gathered from the NICU files, laboratory and hospital records.

Determination of the isolates

BacT/ALERT® PF Plus bottles were used for sampling blood from the suspected neonates. The bottles were incubated in BacT/Alert 3DTM automated blood culture system (bioMerieux, Marcy l'Etoile, France). Incubation period was set as 5 days. Subsequent cultures from positive samples were inoculated onto the Tryptic Soy agar containing 5% sheep blood and Eosin Methylene Blue agar, and chocolate agar plates. The plates were incubated at 35° C for 18-24 hours. VITEK® 2 automated system (bioMerieux, Marcy l'Etoile, France) was used for identification and antimicrobial susceptibility testing. When the same microorganism was isolated from the consecutive samples of the same individual patient, only one isolate was included in the study. The results were interpreted according to the Clinical and Laboratory Standards Institute (CLSI) 2015 criteria.

Statistical analyses

Comparisons between groups were made with Chi-square or Fisher's exact test for categorical variables. A p value <0,05 was considered as significant. Comparisons between the blood culture results and the years were analysed. Commercial statistical software SPSS version 22.0 (SPSS Inc., Chicago, IL, USA) was used to perform statistical evaluations.

Results

Total number of 1245 blood culture samples of 516 neonates with suspected blood stream infection was evaluated during the three-year study period. The mean gestational age of patients with late-onset neonatal sepsis was 26.2 weeks and the gestational ages ranged between 24 and 30 weeks. Among 516 neonate included in the study, 35 of them (6.8%) suffered BSI. Blood cultures yielded no growth in 93.5% (n=1165) of the whole cultures included in the study. Contamination (microbiological and clinical contamination) was determined in 3.6% (n=45) of the blood cultures. Causative agent of BSI was reported in 2.8% (n=35) of the samples. When the results of the blood cultures were evaluated through the years, we did not determine any significant difference.

Gram-negative bacteria caused late-onset neonatal sepsis cases (88.9%) more commonly than gram positive ones (17.1%). *K. pneumoniae* was identified as the most predominant bacterium (45.7%), followed by *E. cloaca* (11.4%) and other enteric bacilli. Coagulase negative staphylococci and *Enterococcus faecalis* were the Gram-positives determined in 17.1% of the cases. All the pathogens, isolated as the causatives of the late-onset neonatal sepsis, were listed in Table 2.

The majority of the susceptible antimicrobials were determined to be carbapenems for the Gram-negative bacteria and glycopeptides for the Gram-positive bacteria as expected according to the antimicrobial susceptibility testing results (Table 3) Ampicillin, Amoxicillin-clavulonate and cefazolin were the least effective antimicrobials for the Gram-negative bacteria. Extended spectrum beta-lactamase (ESBL) production, which limited the use of certain cephalosporins as treatment choice, was determined in 15 out of 29 (51.7%) of the Gram-negatives. No significant resistance

was detected for the Gram-positive bacteria expect for an isolate of *S. haemolyticus*, which was resistant to ceftazidime-representative antibiotic for resistance to penicillins, Beta lactam/beta lactamase inhibitor combinations, cepheems, and carbapenems.

Discussion

BSIs are the most common cause of morbidity and mortality among neonates and it is estimated that neonatal BSIs cause for more than one million deaths annually throughout the world⁶. Approximately 95 % of the neonates followed in a NICU receive empirical antimicrobials and antimicrobials are being the most prescribed drugs in NICU^{11,12}. Among these patients only 1-5% of them has a culture-proven BSI⁶. In most NICU units empirical antimicrobial treatment schemes mainly depend on experience of health care provider, data from other age groups and international guides, however such guidance may not represent the actual status of the pathogens. Each institute should have dependable epidemiological data about causative agents of BSI to set appropriate empirical treatment schemes.

We determine 35 culture-proven late onset BSI attacks in NICU during three-year period. The frequency of neonatal BSI in NICU is determined as 6.8%. Wide ranges of frequencies from 1.8% to 39.8% regarding to neonatal BSI are given in various papers¹³. Although not significant, percentage of reports resulted as contamination tend to have a gradual decrease throughout the years. When sampling difficulties are taken into consideration, decrease in contamination rates is interpreted as a favorable condition.

Gram-negative bacteria predominantly and remarkably are isolated from the blood culture samples of the neonates in this study (82.9%). In developed countries gram positives, especially coagulase negative *Staphylococcus* spp., *Enterococcus* spp. and *S. aureus* are commonly isolated as the agents of BSI in neonates⁷. However, members of *Enterobacteriaceae* cause more BSI compared to the Gram-positives in developing countries^{7,14}. We do not identified any fungal agent from the blood samples, despite the fact that the high ratio of Gram-negatives and high prevalence of ESBL producers force the use of carbapenems, which in turn may cause fungal colonization and infection.

Table 1. Evaluation of the blood culture results according to years in neonatal intensive care unit.

Result of the Blood Cultures	2013		2014		2105		Total		p
	n	%	n	%	n	%	n	%	
No Growth	551	92.0	353	94.6	261	95.6	1165	93.6	>0.05
Contamination	28	4.7	13	3.5	4	1.5	45	3.6	
Blood stream infection	20	3.3	7	1.9	8	2.9	35	2.8	
Total Blood Cultures evaluated	599	100	373	100	273	100	1245	100	

Table 2. Distribution of pathogens causing blood stream infections in patients followed in neonatal intensive care unit.

Isolate	n	%
Gram Negatives	29	82.9
Gram Positives	6	17.1
Total	35	100
Klebsiella pneumoniae	16	45.7
Enterobacter cloaca	4	11.4
Escherichia coli	2	5.7
Klebsiella oxytoca	2	5.7
Pseudomonas aeruginosa	2	5.7
Serratia marcescens	2	5.7
Enterobacter aerogenes	1	2.9
Enterococcus faecalis	2	5.7
Staphylococcus epidermidis	2	5.7
Staphylococcus haemolyticus	2	5.7
Total	35	100

Table 3. Antibiotic susceptibility test results of isolates causing blood stream infections in neonatal patients.

Gram-negative	Number of susceptible isolates												
	n	AMP	CZ	GN	AK	AMC	CFX	CRO	TZP	IMP	MEM	FEP	ESBL
Bacteria													
Klebsiella pneumoniae	16	0	9	3	16	8	5	5	15	16	16	6	12
Enterobacter cloaca	4	0	2	3	3	0	1	1	2	4	4	2	1
Escherichia coli	2	0	1	1	1	1	1	1	2	2	2	1	1
Klebsiella oxytoca	2	0	2	1	1	1	0	1	2	2	2	2	1
Pseudomonas aeruginosa	2	0	0	2	2	0	0	2	1	2	2	2	-
Serratia marcescens	2	2	2	2	2	0	1	1	2	2	2	2	-
Enterobacter aerogenes	1	0	1	1	1	0	1	1	1	1	1	1	-
Gram-positive	Number of susceptible isolates												
Bacteria	n	P	AMP	FOX	E	CIP	GN		DA	SXT	LNZ	VA	TEC
Enterococcus faecalis	2	NA	2	NA	0	2	2	2	2	NA	2	2	2
Staphylococcus epidermidis	2	0	NA	0	2	2	1	NA	2	2	2	2	2
Staphylococcus haemolyticus	2	0	NA	1	0	2	1	NA	2	2	2	2	2

AMP: Ampicillin, KZ: Cefazolin, GN: Gentamicin, AK: Amikacin, AMC: Amoxicillin-clavulonate, CFX: Cefuroxime, CRO: Ceftriaxone, CIP: Ciprofloxacin, IMP: Imipenem, MEM: Meropenem, FEP: Cefepime, SXT: trimethoprim-sulfamethoxazole, NF: Nitrofurantoin, P: Penicillin, FOX: Cefoxitin, E: Erythromycin, HLAR: High level aminoglycoside resistance, FOS: Fosfomycin, LNZ: Linezolid, VA: Vancomycin, TEC: Teicoplanin

According to the antimicrobial susceptibility testing results obtained from this study, alternately use of amikacin and piperacillin-tazobactam with sparing the carbapenems for more resistant isolates appears to be a logical approach for planning empirical treatment for the time being. This will avoid the selection pressure and development of carbapenem resistant isolates, which are not uncommon in our hospital and country. In the last three years experienced two *K. pneumoniae* epidemics, one of which was occurred by NDM-1 producing, carbapenem resistant *K. pneumoniae*^{15,16}. Vancomycin, the life-saving drug against Gram-positive bacteria, can be added to the empirical treatment in case of infections with suspected such pathogens. However, despite staying in the range of susceptible values, increase in MIC levels of vancomycin for the *Staphylococcus* spp. is a worrying concern recently¹⁷. The development of resistance is inevitable in case of inappropriate use of broad-spectrum and last choice antimicrobials^{17,18}. Antibiotic stewardship policies, effective implementation of infection control measures and continuous education will slow down the unavoidable upcoming era of pan-resistant bacteria.

In conclusion, it was determined that Gram-negative bacteria are the main cause of BSI in our newly opened level III NICU. Then, we have provided a baseline data about the causative agents and have proposed an empirical antimicrobial treatment scheme which is alternately use of amikacin and piperacillin-tazobactam, when needed, in combination with vancomycin. The knowledge of causative agents of BSI and their antimicrobial susceptibility properties is one of the corner stones of the efforts to decrease the BSIs in NICU and is one of the major components of neonatal infection surveillance programs to discover the epidemiology of the disease. Further efforts to identify BSI in a timely manner and close following of NICU patients for surveillance purposes are the future obligations to be done.

Conflict of Interest:

The authors declare that they have no conflict of interest.

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Ethical Committee Approval:

Sakarya University Non-Invasive Ethics Committee, 050.01.04.87
Declaration: All the authors declare that they have obeyed the rules in "Helsinki Declaration", "Good Medical Practice Guidelines", and "Good Laboratories Practice Guidelines".

References

1. Stoll BJ, Hansen NI, Adams-Chapman I, et al. National Institute of Child Health and Human Development Neonatal Research Network. Neurodevelopmental and growth impairment among extremely low birth-weight infants with neonatal infection. *JAMA* 2004; 292(19): 2357–65.
2. Legeay C, Bourigault C, Lepelletier D, Zahar J. Prevention of healthcare-associated infections in neonates: room for improvement. *J of Hosp Infect* 2015; 89(4): 319-323.
3. Manzoni P, Rizzollo S, Decembrino L, Ruffinazzi G, Rossi Ricci A, Farina D, et al. Recent advances in prevention of sepsis in the premature neonates in NICU. *Early Hum Dev* 2011; 87 :31-33.
4. Jacqz-Aigrain E, Zhao W, Sharland M, van den Anker J. Use of antibacterial agents in the neonate: 50 years of experience with vancomycin administration. *Semin Fetal Neonatal Med* 2013; 18: 28-34
5. Cipolla D, Giuffrè M, Corsello G, Mammina C. Prevention of nosocomial infections and surveillance of emerging resistances in NICU. *J Matern Fetal Neonatal Med* 2011; 24 :23-26.
6. Tziella C, Borghesi A, Pozzi M, Stronati M. Neonatal infections due to multi-resistant strains: Epidemiology, current treatment, emerging therapeutic approaches and prevention. *Clin Chim Acta* 2015; 451: 71-78
7. Cailles B, Vergnano S, Kortsalioudaki C, Heath P, Sharland M. Review: The current and future roles of neonatal infection surveillance programmes in combating antimicrobial resistance. *Early Hum Dev* 2015; 91: 613-618
8. Resende D, Peppe A, dos Reis H, Abdallah V, Ribas R, Gontijo Filho P. Original article: Late onset sepsis in newborn babies: epidemiology and effect of a bundle to prevent central line associated bloodstream infections in the neonatal intensive care unit. *Braz J Infect Dis* 2015; 19: 1952-57.
9. Cohen-Wolkowicz M, Moran C, Benjamin DK, et al. Early and late onset sepsis in late preterm infants. *Pediatr Infect Dis J* 2009; 28: 1052-7.
10. Delanghe J, Speeckaert M. Translational research and biomarkers in neonatal sepsis. *Clin Chim Acta* 2015; 451: 46-64
11. Clark RH, Bloom BT, Spitzer AR, Gerstmann DR. Empiric use of ampicillin and cefotaxime, compared with ampicillin and gentamicin, for neonates at risk for sepsis is associated with an increased risk of neonatal death. *Pediatrics* 2006; 117: 67–74.
12. Depani SJ, Ladhani S, Heath PT, et al. The contribution of infections to neonatal deaths in England and Wales. *Pediatr Infect Dis J* 2011; 30: 345–7.
13. Yalaz M, Cetin H, Akisu M, Kultursay N, Aydemir S, Tunger A. Neonatal nosocomial sepsis in a level-III NICU: Evaluation of the causative agents and antimicrobial susceptibilities. *Turk J Of Pediatr* 2006; 48(1): 13-18.
14. Santos R, Tristram D. A Practical Guide to the Diagnosis, Treatment, and Prevention of Neonatal Infections. *Pediatr Clin North Am* 2015; 62: 491-508.
15. Koroglu M, Ozbek A, Demiray T, Hafizoglu T, Guclu E, Durmaz R, et al. Investigation of clonal relationships of *K. pneumoniae* isolates from neonatal intensive care units by PFGE and rep-PCR. *J of Infect In Dev Count* 2015; 9(8): 829-836.
16. Karabay O, Altindis M, Koroglu M, Aydemir A, Karatuna O, Erdem A. The carbapenem-resistant Enterobacteriaceae threat is growing: NDM-1 epidemic at a training hospital in Turkey. *Ann Clin Microbiol Antimicrob* 2016;15(1):1-6
17. van den Anker J. How to optimize the evaluation and use of antibiotics in neonates. *Early Hum Dev* 2014; 90(1): 10-12
18. Cohen-Wolkowicz M, Poindexter B, Bidegain M, Weitkamp JH, Schelonka RL, Randolph DA, et al.; Meropenem Study Team. Safety and effectiveness of meropenem in infants with suspected or complicated intra-abdominal infections. *Clin Infect Dis* 2012; 55: 1495–1502.