

First Report of *Glochidia* (Mollusca: Lamellibranchiata) Infestation in Aquarium Fish (Flowerhorn) from Iran

Salar ZARRABI AHRABI^{1*}, Shahrokh SHIRAZI²

¹Department of Basic Health Science, Faculty of Health Sciences, Marmara University, İstanbul, Turkey

²Department of Veterinary Parasitology, Science and Research Branch, Islamic Azad University, Tehran, Iran

ABSTRACT

Glochidia are the larva stage of bivalve mollusks (Lamellibranchiata). These temporary obligate parasites are apparent on the fins and gills and rarely on the surface of fish in spring, summer, and autumn. In spring 2018, a flowerhorn (cichlid) with spots appearing body and gills and white small mobile organisms on the floor of an aquarium were referred to the parasitology laboratory in the School of Veterinary Medicine, Science and Research Unit, Tehran Islamic Azad University. The organisms were sampled and clear in Potas 10% fixed by Glycerin gelatin and examined with a light microscope; the samples were diagnosed as *Glochidia*.

Keywords: Bivalve, Flowerhorn, *Glochidia*

İran'dan Akvaryum Balıklarında (Flowerhorn) *Glochidia* (Mollusca: Lamellibranchiata) İstilasının İlk Raporu

ÖZ

Glochidia, Lamellibranchiata sınıfına ait çift kabuklu yumuşakçaların larva evresini temsil eder. Bu geçici zorunlu parazitler, ilkbahar, yaz ve sonbaharda balıkların yüzgeçlerinde ve solungaçlarında ve nadiren de yüzeyinde görülür. 2018 yılı ilkbaharında, bir çiçek boynuzlu (ciklet) akvaryumun içinde vücut ve solungaçlarda lekelerle birlikte akvaryum tabanında beyaz, küçük ve hareketli organizmaların görünmesi üzerine, bu durum Veteriner Fakültesi, Bilim ve Araştırma Birimi, Tahran İslam Azad Üniversitesi parazitoloji laboratuvarına yönlendirildi. Organizmalardan örnek alındı ve Potas 10%'de sabitlendi, Gliserin jel ile temizlendi ve ışık mikroskobu ile incelendi; örnekler *Glochidia* olarak tanımlandı.

Anahtar kelimeler: Çift kabuklu, Flowerhorn, *Glochidia*

To cite this article: Zarrabi Ahrabi S, Shirazi S. First Report of *Glochidia* (Mollusca: Lamellibranchiata) Infestation in Aquarium Fish (Flowerhorn) from Iran. Kocatepe Vet J. (2024) 17(1):77-80.

Submission: 20.01.2024 Accepted: 05.03.2024 Published Online: 07.03.2024

ORCID ID; SZA: 0000-0003-3543-061X, SS: 0000-0001-6819-5391

*Corresponding author e-mail: salar.zarrabi@marmara.edu.tr

INTRODUCTION

The flowerhorn cichlids are ornamental aquarium fish noted for their transparent colors, and glochidium (plural *Glochidia*) is a larval stage of some freshwater mussels, which are released from female mussels (Neves et al., 1985). These aquatic bivalve mollusks belong to the Unionidae family (Brodniewicz, 1968) and have calcareous bivalve shells, often with little hooks on their inner edge (Paperna, 1996). Freshwater bivalve mollusks exhibit diverse life cycle adaptations for parasitizing obligate hosts. The parasitic larval stage, referred to as *Glochidia*, is a crucial aspect of the life cycle of many freshwater mussels. *Glochidia* have the ability to temporarily attach to the outer surface of suitable hosts, commonly fish, and certain amphibians. This unique strategy serves a dual purpose, providing both nutrition and a means of dispersal for the parasite larvae. (Nikishchenko et al., 2022).

These larvae attach to the fish, utilizing structures like hooks, often targeting the gills or fins of the host fish. This attachment initiates a proliferative reaction in the surrounding area where they are attached (Şereflışan, 2021; Şereflışan, 2018; Arey, 1921; Paperna 1996). *Glochidia* encyst in the gill epithelium and undergo growth before eventually dropping off within a span of 10-30 days. The infestation caused by *Glochidia* is termed Glochidiosis (Gustafson and Iwamoto, 2005). *Glochidia* can bear a resemblance to trematode metacercarial cysts, and in instances of heavy infestation, they can rarely lead to mortality (Nedeau et al., 2005).

There is a need for more consensus regarding the pathogenic impact of *Glochidia* on fish. While some

researchers argue that the parasitic larval phase does not hinder recruitment and, therefore, does not substantially affect maintaining the mussel population, high concentrations of *Glochidia* are often associated with decreased swimming ability and higher mortality rates in hosts. Additionally, the relationship is classified as parasitic due to the nutrient transfer from the fish to the mussel (Ieshko et al., 2016).

CASE HISTORY

The case belongs to an aquarium fish enthusiast and owner of a pet shop who referred a case (flowerhorn) to the parasitology laboratory at the Faculty of Veterinary Medicine, Science, and Research Unit, Tehran Islamic Azad University, in spring 2018, upon observing anomalies on the fish such as spots appearing on body and gills and also white small mobile organisms on the floor of an aquarium. As the fish were alive and economically valuable, a necropsy was not feasible. Therefore, a sample was taken from the aquarium base layer for clinical diagnosis. Sample clear in Potas 10% fixed by Glycerin gelatin and examined with a light microscope, revealed the presence of *Glochidia* larvae (Klunzinger et al., 2013), and based on clinical observations, Glochidia infestation was diagnosed on the fish. No specific drug treatment was administered to the case (Smith, 2019). The recommendation included changing the aquarium substrate and water. After one month of follow-up, clinical signs of *Glochidia* infestation were no longer observed, and the fish was successfully sold by the pet shop.

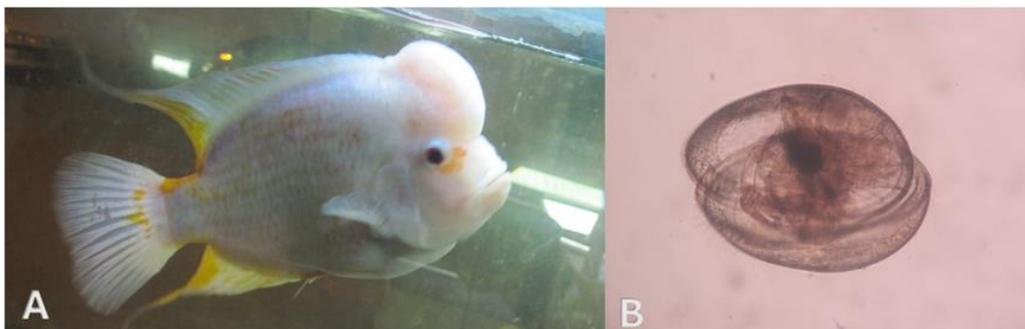


Figure 1: A- Flowerhorn aquarium fish with spots on the body, infested by *Glochidia*. B-*Glochidia* larvae were sampled from the base layer of the aquarium.

DISCUSSION

The flowerhorn fish, belonging to the Cichlid family, are man-made hybrids that do not exist in nature. They are popular ornamental fish, particularly in Southeast Asia and Iran, and are commonly kept in aquariums (Rahmati-Holasoo and Shokrpour, 2014).

Glochidia infestation is a common occurrence in freshwater fish worldwide and can be found in various aquatic environments such as rivers, lakes, mud-bottomed pools, and ponds (Lee and Mora, 2005; Parasites Paperna, 1996).

Cases of Glochidiosis have been reported in various regions, including Poland and North America (Zieritz et al., 2012). They have developed an identification key for North and Central European Unionid mussels, contributing to understanding and recognizing these parasites in different geographic areas. (Brodniewicz, 1968; Gustafson and Iwamoto, 2005; Zieritz et al., 2012). Due to the ongoing debate and lack of consensus regarding the pathogenic impact of *Glochidial* infection on fish, it is noteworthy that some researchers have reported mortalities among certain fish strains that were experimentally infested with *Glochidia* (Ieshko et al., 2016). Research indicates that elevated rates of *Glochidia* infection can enhance swimming performance and mortality in brown trout, potentially resulting in reduced performance in heavily infested fish in their natural environment. Conversely, some studies reported no mortalities or growth retardation in *Glochidia* infected trout. Comparable outcomes were observed in the experimental infestation of Atlantic salmon parr. These divergent findings underscore the intricate nature of interactions between *Glochidia* and different fish species. The observed discrepancies in the impact of *Glochidia* infection on fish may be attributed to the existence of diverse host fish strains or variations in environmental conditions, such as temperature (Ieshko et al., 2016; Taeubert and Geist, 2013). *Glochidiosis* has been documented in both wild and farmed salmonids in locations such as the Scotland River and Virginia (Hastie and Young, 2001; Neves and Widlak, 1988). According to available references, there are no reports of Glochidiosis in ornamental fish. This could be attributed to the fact that these organisms, in their adult phase, resemble stones and do not move extensively (Nedeau et al., 2005). So, it can probably be transmitted by stones that have been brought from these sources for aquariums. This is the first report of Glochidiosis of flowerhorn in Iran.

CONCLUSION

In conclusion, most reports on Glochidiosis are related to freelifving fishes, particularly in the Salmonidae family. The probability of infestation in aquarium fishes is very low. However, it is crucial to emphasize the importance of maintaining aquarium hygiene and being cautious about transferring objects from the natural environment to the aquarium.

Conflict of Interest: The authors declared there is no conflict of interest.

Author Contribution Rates: Both authors contributed equally to all aspects of the research.

Ethics Committee Information: The current study does not require ethics approval.

REFERENCES

- Arey, L. B. (1921). An experimental study on *glochidia* and the factors underlying encystment. *Journal of Experimental Zoology*, 33(2), 462-499. <https://doi.org/10.1002/jez.1400330209>
- Brodniewicz, I. (1968). On *glochidia* of the genera Unio and Anodonta from the Quaternary fresh-water sediments of Poland. *Acta Palaeontologica Polonica*, 13(4), 619-628.
- Gustafson, R. G., & Iwamoto, E. M. (2005). A DNA-based Identification Key to Pacific Northwest Mussel *Glochidia*: Importance to Salmonid and Mussel Conservation. *Northwest Science*, 79(4), 233.
- Hastie, L. C., & Young, M. R. (2001). Freshwater pearl mussel (*Margaritifera margaritifera*) glochidiosis in wild and farmed salmonid stocks in Scotland. *Hydrobiologia*, 445, 109-119. <https://doi.org/10.1023/A:1017588222480>
- Ieshko, E., Geist, J., Murzina, S., Veselov, A., Lebedeva, D., & Ziuganov, V. (2016). The characteristics of the infection of juvenile Atlantic salmon with *glochidia* of the freshwater pearl mussel in rivers of Northwest Russia. *Knowledge and Management of Aquatic Ecosystems*(417), 6. <http://dx.doi.org/10.1051/kmae/2015039>
- Klunzinger, M. W., Thomson, G. J., Beatty, S. J., Morgan, D. L., & Lymbery, A. J. (2013). Morphological and morphometrical description of the glochidia of *Westralunio carteri* Iredale, 1934 (Bivalvia: Unionoida: Hyriidae). *Molluscan Research*, 33(2), 104-109. <https://doi.org/10.1080/13235818.2013.782791>
- Lee, H. H., & Mora, D. A. (2005). Survival of glochidia of freshwater mussels, *Pyganodon grandis* (Mollusca: Unionidae), in vitro in the United States. *World aquaculture*, 36 (4), 56-61.
- Nedeau, E. J., Smith, A. K., & Stone, J. (2005). Freshwater mussels of the Pacific Northwest. US Fish and Wildlife Service.
- Neves, R. J., Weaver, L. R., & Zale, A. V. (1985). An evaluation of host fish suitability for glochidia of *Villosa vanuxemi* and *V. nebulosa* (Pelecypoda: Unionidae). *American Midland Naturalist*, 113(1), 13-19. <https://doi.org/10.2307/2425343>
- Neves, R. J., & Widlak, J. C. (1988). Occurrence of glochidia in stream drift and on fishes of the upper North Fork Holston River, Virginia. *American Midland Naturalist*, 119(1), 111-120. <https://doi.org/10.2307/2426059>
- Nikishchenko, V. E., Sayenko, E. M., & Dyachuk, V. A. (2022). First immunodetection of sensory and nervous systems of parasitic larvae (*glochidia*) of freshwater bivalve *Nodularia douglasiae*. *Frontiers in Physiology*, 13, 624. <https://doi.org/10.3389/fphys.2022.879540>
- Paperna, I. (1996). Parasites infections and diseases of fishes in Africa: an update (No. 31). Food and Agriculture Organization of the United Nations (FAO).
- Rahmati-Holasoo, H., & Shokrpour, S. (2014). Unilateral testicular hypoplasia in flowerhorn fish (hybrid cichlid). *Bulletin of the European Association of Fish Pathologists*, 35(1), 21-25.

Smith, S. A. (2019). Fish Diseases and Medicine. Taylor and Francis Group.

Şereflişan, H. (2018). Determination of Host Fish Suitability for *Unio terminalis delicatus* (Bivalvia: Unionidae) From Gölbaşı Lake in Turkey. Journal of Advances in VetBio Science and Techniques, 3(3), 15-22. <https://doi.org/10.31797/vetbio.423361>

Şereflişan, H. (2021). Host Selection of *Potomida semirugata* (Unionidae: Bivalvia) in Reproduction Strategy. Aquatic Sciences and Engineering, 36(3), 109-115. <https://doi.org/10.26650/ASE2021744926>

Taubert, J.-E., & Geist, J. (2013). Critical swimming speed of brown trout (*Salmo trutta*) infested with freshwater pearl mussel (*Margaritifera margaritifera*) glochidia and implications for artificial breeding of an endangered mussel species. Parasitology Research, 112, 1607-1613. <https://doi.org/10.1007/s00436-013-3314-6>

Zieritz, A., Gum, B., Kuehn, R., & Geist, J. (2012). Identifying freshwater mussels (Unionoida) and parasitic *glochidia* larvae from host fish gills: a molecular key to the North and Central European species. Ecology and Evolution, 2(4), 740-750. <https://doi.org/10.1002/ece3.220>