

Eucalyptol (1.8-cineole) attenuates Gentamicin-induced liver injury

Özhan Karataş¹, Filiz Kazak², Gökhan Akçakavak^{3*}, Halil Alakuş⁴, Ahmed A.j. Jabbar⁵, Ömer Kırgız⁶, İbrahim Alakuş⁷, Bahadır Kılınç⁸, Zeynep Çelik Kenar⁹, Mehmet Tuzcu¹⁰

¹ Department of pathology, Faculty of Veterinary Medicine, Sivas Cumhuriyet University, Sivas, Türkiye

² Department of biochemistry, Faculty of Veterinary Medicine, Hatay Mustafa Kemal University, Hatay, Türkiye

³ Department of pathology, Faculty of Veterinary Medicine, Aksaray University, Aksaray, Türkiye

^{4,6,7} Department of surgery, Faculty of Veterinary Medicine, Hatay Mustafa Kemal University, Hatay, Türkiye

⁵ Department of Medical Laboratory Technology, Erbil Technical Health and Medical College, Erbil Polytechnic University, Erbil, Iraq

⁸ Department of pathology, Etlık Central Veterinary Control and Research Institute, Ankara, Türkiye

^{9,10} Department of pathology, Faculty of Veterinary Medicine, Selçuk University, Konya, Türkiye

Geliş Tarihi / Received: 13.09.2024, Kabul Tarihi / Accepted: 02.12.2024

Abstract: Gentamicin (GM), which is in the aminoglycoside antibiotic class, is frequently preferred today in the treatment of diseases caused by gram-negative bacteria. However, its significant side effects on liver and kidney functions limit its clinical usefulness. The antioxidant and anti-inflammatory medical activities of eucalyptol (EUC, 1.8-cineole) have been reported in different studies. This study aimed to evaluate the effects of EUC on GM-induced hepatotoxicity. The study groups are consisted of control (C), EUC, GM and GM + EUC, and there were 7 rats in each group. At the end of the study, the rats were euthanized under appropriate conditions and samples were collected and biochemical, histopathological and immunohistochemical analyzes were performed. It was determined that there was a important increase in serum alanine aminotransferase (ALT), aspartate transferase (AST) and gamma-glutamyl transferase (GGT) enzymes in the GM group relative to the C group ($p < 0.001$). GM application is determined that it caused histopathological damage in livers. Additionally, immunohistochemically, it caused an important increase in the expressions of 4-hydroxynonenal (4-HNE), 8-hydroxy-2'-deoxyguanosine (8-OHdG) and malondialdehyde (MDA) in rats in the GM group relative to the C group ($p < 0.001$). Eucalyptol application (GM+EUC) regulated the increase in serum ALT, AST and GGT enzymes. It also attenuated 4-HNE, 8-OHdG and MDA expressions. It attenuated the histopathological damage caused by GM application. The results of this study revealed that EUC showed antioxidative, protective and curative efficacy in GM-induced liver damage.

Keywords: eucalyptol (1.8-cineole), gentamicin, hepatotoxicity, MDA, 8-OHdG

Okaliptol (1.8-sineol) Gentamisin kaynaklı karaciğer hasarını hafifletir

Özet: Aminoglikozid antibiyotik sınıfında yer alan gentamisin (GM), gram negatif bakterilere bağlı hastalıkların tedavisinde günümüzde sıklıkla tercih edilmektedir. Ancak karaciğer ve böbrek fonksiyonları üzerine belirgin yan etkilerinin olması klinik yararlılığını sınırlamaktadır. Okaliptolün (OKA, 1.8-sineol) antioksidan ve antiinflatuar tıbbi aktiviteleri farklı çalışmalarda bildirilmiştir. Bu çalışmada GM kaynaklı hepatotoksisite üzerine OKA'nın etkilerinin değerlendirilmesi amaçlanmıştır. Çalışma grupları kontrol (K), OKA, GM ve GM + OKA'dan oluşmaktaydı ve her grupta 7 rat bulunmaktaydı. Çalışmanın sonunda ratlar uygun koşullar altında ötenazi edilerek örnekler toplandı ve biyokimyasal, histopatolojik ve immünohistokimyasal analizleri yapıldı. GM grubunda serum alanin aminotransferaz (ALT), aspartat aminotransferaz (AST) ve gama-glutamil transferaz (GGT) enzimlerinde K grubuna kıyasla anlamlı artış olduğu belirlendi ($p < 0.001$). GM uygulamasının karaciğerlerde histopatolojik hasara neden olduğu belirlendi. Ayrıca immünohistokimyasal olarak GM grubundaki ratlarda K grubuna kıyasla 4-hidroksinonenal (4-HNE), 8-hidroksi-2'-deoksiguanozin (8-OHdG) ve malondialdehit (MDA) ekspresyonlarında anlamlı artışa neden oldu ($p < 0.001$). Okaliptol uygulaması (GM+OKA) serum ALT, AST ve GGT enzimlerindeki artışı düzenledi. Ayrıca 4-HNE, 8-OHdG ve MDA ekspresyonlarını azalttı. GM uygulamasının neden olduğu histopatolojik hasarı azalttı. Bu çalışmanın sonuçları OKA'nın GM kaynaklı karaciğer hasarında antioksidan, koruyucu ve tedavi edici etkinlik gösterdiğini ortaya koydu.

Anahtar kelimeler: gentamisin, hepatotoksisite, MDA, okaliptol (1.8-sineol), 8-OHdG

Introduction

One of the drug classes frequently used in the treatment of diseases today is antibiotics. Antibiotics

prevent many problems caused by infection. However, antibiotics can damage various organs such as skin, kidney, liver, brain, mouth, etc. Amino-

Yazışma adresi / Correspondence: Gökhan Akçakavak, Aksaray University Faculty of Veterinary Medicine, Aksaray, Türkiye
e-mail: gokhan.akcakavak@aksaray.edu.tr

ORCID IDs of the authors: ¹0000-0002-2778-8059 • ²0000-0002-9065-394X • ³0000-0001-5949-4752 • ⁴0000-0001-9265-2310 • ⁵0000-0001-9689-4018 • ⁶0000-0002-0222-1363 • ⁷0000-0002-2031-7035 • ⁸0000-0003-3426-2116 • ⁹0000-0002-9667-5728 • ¹⁰0000-0003-3118-1054

glycoside antibiotics have been used as antibacterial treatment for many years. Aminoglycosides are defined as a class of antibiotics that consists of at least 2 amino sugars and are used to treat gram (-) bacteria. Following treatment with aminoglycoside antibiotics, approximately 5-10% of patients may experience ototoxicity, nephrotoxicity, and hepatotoxicity side effects (Mahi-Birjand et al., 2020; Thy et al. 2023; Lang et al. 2023)

Gentamicin (GM), which is in the aminoglycoside antibiotic class, is frequently preferred today in the treatment of diseases caused by gram-negative bacteria. However, its notable side effects on liver and kidney functions limit its clinical usefulness (Yarijani et al., 2019; Babaeenezhad et al., 2021; Akcakavak et al., 2024). The liver and kidneys are particularly sensitive to drug toxicity because they play important roles in normal homeostasis in detoxification and excretion of drugs and their metabolites. Currently, the exact mechanism of GM hepato and nephrotoxicity have not been fully clarified. Different hypotheses have been proposed in many studies and oxidative stress is the most emphasized in GM hepato and nephrotoxicity (Galaly et al., 2014; Laaroussi et al., 2021). Because reactive oxygen species (ROS) and other free radicals are suggested to be one of the important mediators of GM toxicity (Banday et al., 2008). Different studies have shown that GM stimulates excessive production of ROS metabolites, affecting different processes including lipid peroxidation, protein oxidation and DNA damage, leading to necrosis and cellular damage. In addition, excessive production of free radicals causes activation of nitrosative tissue stress, modulation of the caspase family and upregulation of the inflammatory process (Morales et al., 2002; Sanchez-Gonzalez et al., 2011; Ali et al., 2020; Bulboacă et al., 2022).

Eucalyptol (EUC, 1,8-cineole), a saturated monoterpene, is obtained from botanical sources such as eucalyptus, camphor and rosemary. EUC is known as the main active ingredient of eucalyptus essential oils. EUC has a long history of use in traditional medicine. It is also frequently used in food, fragrance and cosmetics due to its taste and aroma (Galan et al., 2020; Hoch et al., 2023). EUC has a number of medical effects including antimicrobial, anti-inflammatory, antioxidant, analgesic and bronchodilator (Galan et al., 2020; Cai et al., 2021; Kazak, 2022; Akcakavak, et al., 2024, Kazak et al., 2024a; Kazak et al., 2024b). It has been reported in various studies that EUC shows highly effective free radical scavenging activity and has the ability to

defend cells from oxidative damage by neutralizing ROS (Rašković et al., 2014; Ryu et al., 2014). Ryu et al. 2014 stated that EUC significantly reduced ROS overproduction in patients with ischemic stroke. In an experimental arthritis study in rats, EUC administration was reported to reduce the levels of proinflammatory cytokines (Iqbal et al. 2024). In a lead acetate-induced liver injury study, it was stated that EUC could prevent damage by reducing inflammation and oxidative stress (Abdollahi et al. 2024). It is also reported that EUC has an inhibitory effect against lipid peroxidation (Moon et al., 2014; Hsou-na et al., 2019).

Medicinal plants have been widely used to treat many diseases since ancient times. Today, the use of herbal supplements has a special importance in the treatment of liver diseases due to their numerous properties and low side effects. EUC, a saturated monoterpene, is obtained from botanical sources such as eucalyptus, camphor and rosemary (Galan et al., 2020, Cai et al., 2021). Recently, many experimental researches are being conducted to prevent and/or treat GM-induced hepatotoxicity using different agents (Babaeenezhad et al., 2021; Mirazi et al., 2021; Ogundipe et al., 2021). The main purpose of these studies is to find new, effective and safer antioxidant compounds that can prevent and/or treat GM-induced hepatotoxicity. EUC may be an important candidate due to a number of medicinal effects such as anti-inflammatory, antioxidant, antimicrobial, analgesic. Although there are studies in the literature evaluating the effects of EUC on hepatotoxicity induced by different agents, no study evaluating its effectiveness on GM-induced hepatotoxicity was found. The aim of the present study was to evaluate the effect of EUC on GM-induced hepatotoxicity by determining histopathological damage, hepatic functional enzymes, and lipid peroxidation markers in the tissue.

Materials and methods

Animals

The material for the study consisted of 28 female Wistar Albino rats weighing 300-400 g. The rats were housed in standard plastic cages, at room temperature of 20-22°C, in 12 hours of light and 12 hours of darkness, and were fed *ad libitum*.

Experimental procedure

Rats were randomly divided into 4 groups, 7 in each group; control (C), EUC, GM, GM+EUC. GM was implemented as a single dose of 100 mg/kg intra-

peritoneal (i.p.) for 10 consecutive days of the study. EUC solution was implemented by oral gavage at 100 mg/kg for 10 consecutive days. EUC solution was prepared relative to the previously stated study (Akçakavak et al., 2024). In addition, the vehicle solution was administered to C and GM groups by oral gavage for 10 consecutive days throughout the experimental period. On the 11th day of the study, intracardiac blood was taken from the experimental animals under xylazine and ketamine (10-90 mg/kg) anesthesia, and the animals were sacrificed. Afterwards, the liver tissues of the necropsied rats were placed in 10% neutral formaldehyde solution for histopathological and immunohistochemical examinations.

Determination of serum biochemical parameters

Blood samples taken under anesthesia were centrifuged at 5000 rpm for 15 minutes at 4°C. The obtained sera were transferred to cryotubes and stored at -80°C. Liver function was evaluated via measurement of ALT, AST and GGT activities in serum using autoanalyzer (AU5800, Beckman Coulter, Japan) and commercial kits.

Histopathological examination

Liver tissues taken following necropsy were fixed in 10% neutral formaldehyde solution. Then, formaldehyde was removed by washing and passed through graded alcohols (70°, 80°, 90°, 100°) and xylene steps, respectively. Afterwards, the relevant liver tissues were embedded in paraffin. Sections were taken from paraffin blocks onto ground slides and stained with Hematoxylin-Eosin (H-E). Histopathological evaluation was performed semi-quantitatively in 10 different fields at x20 magnification (0; none, 1; mild, 2; moderate, 3; severe).

Immunohistochemical examination

4-5 µm sections were taken from paraffin blocks onto adhesive slides. The sections were subjected to

paraffin extraction and rehydration processes. Immunohistochemical staining was performed using a commercial kit according to the previously stated method (Akçakavak et al., 2023). 8-OHdG (8-hydroxy-2'-deoxyguanosine, Santa Cruz Biotechnology, sc-393871, 1/200 dilution, 1 hour incubation), 4-HNE (4-Hydroxynonenal, Abcam, ab46545, 1/200 dilution, 1 hour incubation), MDA (Malondialdehyde, Abcam, ab6463 1/1000 dilution, 1 hour incubation) antibodies were used as primers. 3. 3'-diaminobenzidine (DAB) was used as a chromogen and was examined under light microscopy after counterstaining with Mayers hematoxylin. Immunohistochemical scoring was performed semi-quantitatively (0; none, 1; mild, 2; moderate, 3; severe)(Akçakavak et al., 2024).

Statistical analysis

The statistical program SPSS (Inc., Chicago, USA 25.0) was used to analyse the obtained data. The biochemical, histopathological and immunohistochemical findings obtained in the study were evaluated using One-way ANOVA and Duncan's test as post-hoc test. $p < 0.05$ was used as the significance limit.

Results

Serum biochemical findings

Serum biochemical findings of the groups regarding liver function tests are presented in Table 1. The activities of the ALT, AST and GGT enzymes in the GM group were 2.1, 1.7 and 2.2. fold respectively higher than those of the C group ($p < 0.001$). The activities of the ALT, AST and GGT enzymes in the GM+EUC group significantly decreased (0.7, 0.8 and 0.7 fold, respectively) compared with the GM group, although the activities of the ALT, AST and GGT enzymes in the GM+EUC group significantly increased (1.4, 1.3 and 1.6 fold, respectively) relative to C group ($p < 0.001$).

Table 1. The impacts of EUC on the activities of ALT, AST, and GGT in rat liver, intact and with GM-induced liver injury (Mean±SE, n;7)

Liver function tests	C	EUC	GM	GM+EUC
ALT (ul/l)	35.83±1.67 ^c	39.62±2.94 ^c	75.83±1.83 ^a	52.34±2.09 ^b
AST (ul/l)	90.33±3.57 ^c	93.50±2.86 ^c	152.17±5.49 ^a	121.00±2.90 ^b
GGT (ul/l)	4.50±0.56 ^c	4.67±0.42 ^c	9.83±1.37 ^a	7.10±0.60 ^b

^{a-c} Indicates statistical significance between groups in the same line ($p < 0.001$). (ALT; alanine aminotransferase, AST; aspartate transferase, GGT; gamma-glutamyl transferase, C; Control, EUC; eucalyptol, GM; gentamicin, GM+EUC: gentamicin+eucalyptol)

Histopathological results

Histopathological statistical scores of the groups are given in Table 2. C and EUC groups showed normal histology (Figure 1.A-B). Degeneration and necrosis were detected in hepatocytes in GM and GM+EUC groups. Especially degenerative and necrotic changes were localized in the centrilobular region (Figure 1.C-E). Degeneration and necrosis scores were significantly reduced in the GM+EUC group relative

to the GM group ($p < 0.001$). In addition, bile duct proliferation, inflammatory cell infiltration and sinusoidal dilatation were seen in the GM and GM+EUC groups (Figure 1.C-E). The GM+EUC group significantly reduced the relevant disorders relative to the GM group ($p < 0.001$). Moreover, hemorrhage foci were detected in places in the GM and GM+EUC groups.

Table 2. Histopathological scores between groups (Mean+SE, n;7)

Histopathological lesion	C	EUC	GM	GM+EUC
Degeneration of hepatocytes	0.50+0.22 ^c	0.67+0.21 ^c	3.17+0.17 ^a	2.17+0.17 ^b
Necrosis of hepatocytes	0.33+0.21 ^c	0.50+0.22 ^c	2.83+0.17 ^a	1.83+0.30 ^b
Inflammatory cell infiltration	0.33+0.21 ^c	0.33+0.21 ^c	2.67+0.21 ^a	1.67+0.33 ^b
Bile duct proliferation	0.50+0.22 ^c	0.50+0.22 ^c	2.33+0.33 ^a	1.33+0.21 ^b
Sinusoidal dilatation	0.67+0.21 ^c	0.83+0.17 ^c	2.5+0.22 ^a	1.67+0.21 ^b

^{a-c} Indicates statistical significance between groups in the same line ($p < 0.001$). (C; Control, EUC; eucalyptol, GM; gentamicin, GM+EUC: gentamicin+eucalyptol)

Immunohistochemical results

The immunohistochemical scores of the groups are given in Table 3. In the C and EUC groups, immunopositivity was mild or absent for the relevant primers (8-OHdG, 4-HNE and MDA). 8-OHdG, 4-HNE and MDA immunopositivity was significantly

increased in the GM group compared to the C group ($p < 0.001$), and immunopositivity was prevalent in the centrilobular regions (Figure 2). The GM+EUC group was found to have significantly reduced immunopositivity (8-OHdG, 4-HNE and MDA) relative to the GM group ($p < 0.001$).

Table 3. Immunohistochemical scores between groups (Mean+SE, n;7)

Primers	C	EUC	GM	GM+EUC
4-HNE	0.17+0.17 ^c	0.33+0.21 ^c	2.67+0.21 ^a	1.83+0.17 ^b
8-OHdG	0.00+0.00 ^c	0.17+0.17 ^c	2.83+0.17 ^a	2.00+0.25 ^b
MDA	0.17+0.17 ^c	0.33+0.21 ^c	2.67+0.21 ^a	1.83+0.30 ^b

^{a-c} Indicates statistical significance between groups in the same line ($p < 0.001$). (MDA; malondialdehyde, 8-OHdG; 8-hydroxy-2'-deoxyguanosine, 4-HNE; 4-Hydroxynonenal, C; Control, EUC; eucalyptol, GM; gentamicin, GM+EUC: gentamicin+eucalyptol)

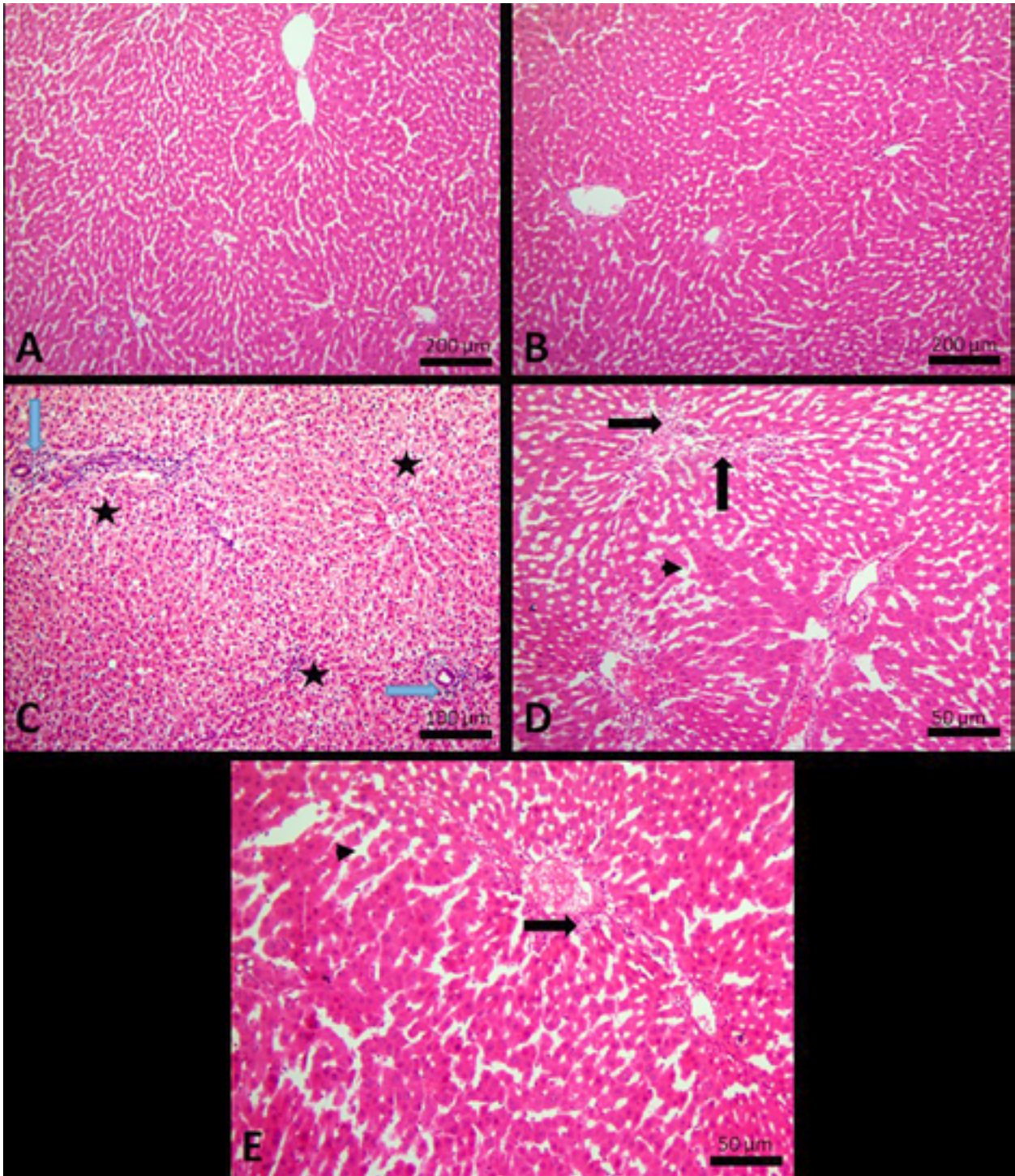


Figure 1. Histopathological examination between groups, Hematoxylin-Eosin, **A;** C group, **B;** EUC group, **C-D;** GM group, **E;** GM+EUC group. (Degeneration in hepatocytes (stars), necrosis (black arrows), inflammatory cell infiltration (blue arrows), sinusoidal dilatation (arrowheads), C; Control, EUC; eucalyptol, GM; gentamicin, GM+EUC; gentamicin+eucalyptol)

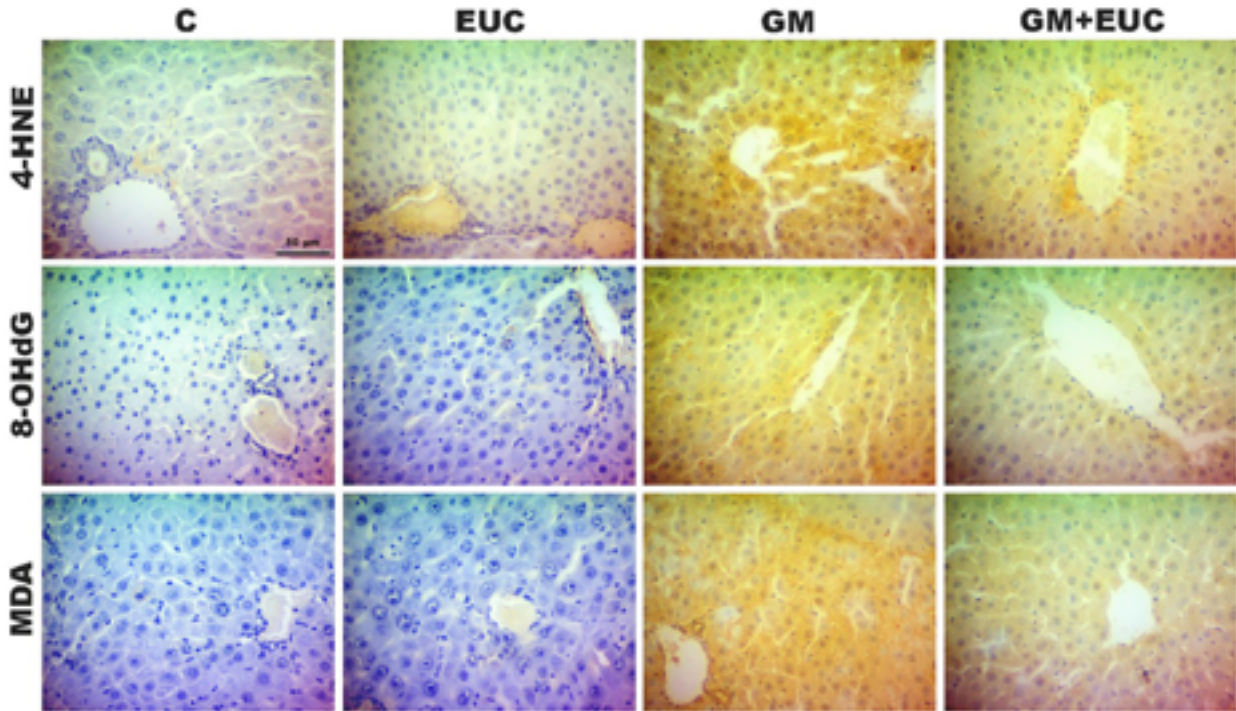


Figure 2. Immunohistochemical examination between groups (DAB), (**4-HNE**; 4-Hydroxynonenal, **8-OHdG**; 8-hydroxy-2'-deoxyguanosine, **MDA**; malondialdehyde, **C**; Control, **EUC**; eucalyptol, **GM**; gentamicin, **GM+EUC**; gentamicin+eucalyptol)

Discussion

Although the effects of many phytochemical components on GM-induced liver toxicity have been evaluated, no study evaluating the effect of EUC on hepatotoxicity has been found. Present study, we aimed to evaluate the effects of EUC (100 mg/kg, 10 consecutive days) on liver damage induced by GM (100 mg/kg, 10 consecutive days) in rats. The current biochemical, histopathological and immunohistochemical findings showed that GM administration caused liver hepatotoxicity and that the changes related to hepatotoxicity could be prevented with simultaneous administration of EUC with GM.

Oxidative stress is a condition that occurs when the balance between free radicals and antioxidants in normal homeostasis is disrupted. Antioxidants are defined as protective systems that protect cells against the damaging effects of free radicals. It is known that suppression of the antioxidant system for various reasons can lead to a wide variety of pathological conditions, including liver damage. Gentamicin reduces the activity of antioxidant enzymes such as CAT, GPx and SOD in cells, causing the redox system to deteriorate and the formation

of free radicals (Đuračková, 2010; Randjelovic et al.2017; Laaroussi et al., 2021; Bulboacă et al., 2022).

AST is present in both hepatocyte cytosol and hepatocyte mitochondria while ALT exists prevalently in the hepatocyte cytosol. The release of these enzymes from hepatocytes is largely due to hepatocellular injury (Zoppini et al., 2016). The enzyme GGT is also associated with hepatitis and biliary tract obstruction (Ibraheem et al., 2021). Literature revealed that GM may directly lead to hepatotoxicity in rats through oxidative stress and apoptosis and finally cause the elevation of the serum activities of ALT, AST and GGT. It has been found that GM increased ALT, AST and GGT activities in rats (Yarjani et al., 2019; Babaenezhad et al., 2021; Ibraheem et al., 2021; Bulboacă et al., 2022). In keeping with such findings, the present study demonstrated that GM caused liver damage as shown by important increases in serum ALT, AST and GGT activities. Thus, the elevation of ALT, AST and GGT in GM administered animals can imply the injury of liver cytoarchitecture and hepatic cell integrity, linked to microsomal membrane fluidity, mitochondrial dysfunction, and free radical generation. In addition, the activities of the ALT, AST and GGT enzymes in serum of the

groups that received GM and EUC importantly reduced, which indicates the protective effect of EUC against hepatotoxicity induced by GM.

GM is often prescribed to treat bacterial conjunctivitis, sepsis, endocarditis, and infections caused by gram-negative bacteria (Chen et al., 2014). Although it has many beneficial effects, it increases ROS levels by inhibiting enzymatic and non-enzymatic antioxidants in the liver. Thus, in addition to increasing oxidative stress, it causes liver damage by causing damage to lipids, nucleic acids and cellular proteins in the membrane (Laaroussi et al., 2021; Ogundipe et al., 2021). Galan et al. (2014) detected histopathologically degeneration, steatosis, congestion, inflammatory cell infiltration and bile duct proliferation in liver injury induced by GM in rats. In another study examining liver damage due to GM, histopathological findings included steatosis, congestion, inflammatory cell infiltration, sinusoid dilatation, and an increase in the number of Kupfer cells (Al-Khamas et al. 2020). In another study, degeneration, necrosis and sinusoid dilatation were detected histopathologically in GM-induced liver damage (Wijayanti et al., 2023). When the histopathological findings of the current study were evaluated, it was seen that the histopathological changes observed in the GM group were consistent with the findings of previous studies. It is thought that the finding of degeneration and necrosis in the centrilobular region (Figure 1), especially in the GM group, may be because of the increase in ROS levels for GM application and the subsequent oxidative stress.

It was determined that GM-induced histopathological damage was reduced when EUC was administered together with GM. It has been reported in various studies that EUC shows highly effective free radical scavenging activity and has the ability to protect cells from oxidative damage by neutralizing ROS (Rašković et al., 2014; Galan et al., 2020; Akcakavak et al., 2024). EUC is suggested that it exhibits its antioxidant and anti-inflammatory medical activities through manipulations on Nrf2 and NF- κ B pathways. Research in the literature reports that EUC causes upregulation of the Nrf2 transcription factor and downregulation of the NF- κ B pathway, thus exhibiting strong antioxidant and anti-inflammatory bioactivities (Cai et al., 2021; Venkataraman et al., 2023; Akcakavak et al., 2024; Iqbal et al., 2024). The current study shows that EUC administration can prevent the upregulation of oxidative stress and inflammatory processes that occur with GM administration and thus reduce liver damage. In addition, the fact that EUC down-regulated pro-inflammatory

cytokine levels in previous studies further strengthened our idea (Akcakavak et al., 2023; Akcakavak et al., 2024; Iqbal et al., 2024).

Excessive formation of ROS due to chemical toxicity causes oxidative damage to DNA. This situation contributes to the formation of 8-OHdG, which is the most crucial effect of DNA damage. 8-OHdG has been commonly assessed as a biomarker of oxidative DNA damage in recent years (Graille et al., 2020; Akcakavak et al., 2023). Another oxidative reaction induced by excessive ROS levels is lipid peroxidation. 4-HNE and MDA are known as the cytotoxic end products of lipid peroxidation (Yang et al., 2003; Ayala et al., 2014). In the literature, it is reported that causes upregulation of 4-HNE, 8-OHdG and MDA in different GM-induced toxicity studies (Aycan-Ustyol et al., 2017; Cui et al., 2019; Mohammed et al., 2019; Ince et al., 2020). In the present study, higher expressions of 4-HNE, 8-OHdG and MDA were detected in the GM group relative to the control groups and were consistent with the literature (Figure 2). Present findings showed that GM (100 mg/kg, 10 consecutive days) administration caused oxidative DNA damage and lipid peroxidation. GM and EUC administration provided protection against GM-induced hepatotoxicity by reducing the expressions of 4-HNE, 8-OHdG and MDA relative to the GM only group.

In conclusion, it has been determined that GM application causes complications characterized by degeneration and necrosis in hepatocytes, inflammatory cell infiltration, bile duct hyperplasia and sinusoidal dilatation, and this structural and cellular damage in the liver causes an increase in serum liver enzymes. The current study shows that EUC administered simultaneously with GM plays a protective/curative role in reducing GM-induced liver damage by suppressing liver function tests (AST, ALP, GGT), histopathological changes and 8-OHdG, 4-HNE and MDA expressions. These results suggest that EUC may be a promising candidate for clinical drug development in GM-induced liver damage.

Financial support and conflict of interest: There is no person/organization that financially supports the work and the authors do not have any interest-based relationship.

Ethics committee for the use of experimental animals and other ethical committee decisions and permissions: Hatay Mustafa Kemal University Animal Experiments Local Ethics Committee approved with the decision number 2024/06-07.

References

- Abdollahi M, Asle-Rousta M, Mahmazi S. (2024) Protective effect of 1, 8-cineole (eucalyptol) against lead-induced liver injury by ameliorating oxidative stress and inflammation and modulating TLR4/MyD88/NF- κ B signaling. *Iranian J Basic Med Sci*, 27(10), 1293-1299, doi: 10.22038/ijbms.2024.78448.16964.
- Akcakavak G, Kazak F, Karatas O, Alakus H, Alakus I, Kirgiz O, Celik Z, Yilmaz Devenci MZ, Ozdemir O, Tuzcu M. (2024) Eucalyptol regulates Nrf2 and NF- κ B signaling and alleviates gentamicin-induced kidney injury in rats by downregulating oxidative stress, oxidative DNA damage, inflammation, and apoptosis. *Toxicol Mech Methods*, 34(4), 413-422. doi: 10.1080/15376516.2023.2297234.
- Akcakavak G, Kazak F, Yilmaz Devenci MZ. (2023) Eucalyptol Protects against Cisplatin-Induced Liver Injury in Rats. *Biol Bulletin*, 50(5), 987-994. doi: 10.1134/s106235902360085x
- Ali FE, Hassanein EH, Bakr AG, El-Shoura EA, El-Gamal DA, Mahmoud AR, Abd-Elhamid TH. (2020) Ursodeoxycholic acid abrogates gentamicin-induced hepatotoxicity in rats: Role of NF- κ B-p65/TNF- α , Bax/Bcl-xl/Caspase-3, and eNOS/iNOS pathways. *Life Sci*, 254, 117760. doi: 10.1016/j.lfs.2020.117760
- Al-Khamas AJ, Kadhim ZH, Al-Charak AG, Faris JK. (2020). Biochemical and histological study of rat liver toxicity induced by gentamicin and protective action of berberine. *Plant Arch (09725210)*, 20(1), 3073-3078
- Ayala A, Muñoz MF, Argüelles S. (2014) Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid med cell longev*, 2014(1), 360438.
- Aycan-Ustyol E, Kabasakal M, Bekpınar S, Alp-Yıldırım Fİ, Tepe O, Giris M, Ozluk Y, Unlucerci Y, Uydes-Dogan BS, Uysal M (2017) Vascular function and arginine and dimethylarginines in gentamicin-induced renal failure: a possible effect of heme oxygenase 1 inducer hemin. *Can J physiol pharmacol* 95(12), 1406-1413. doi: 10.1139/cjpp-2016-0578.
- Babaeenezhad E, Nouryazdan N, Nasri M, Ahmadvand H, Sarabi MM. (2021) Cinnamic acid ameliorate gentamicin-induced liver dysfunctions and nephrotoxicity in rats through induction of antioxidant activities. *Heliyon*, 7(7). e07465. doi: 10.1016/j.heliyon.2021.e07465
- Banday AA, Farooq N, Priyamvada S, Yusufi AN, Khan F. (2008) Time dependent effects of gentamicin on the enzymes of carbohydrate metabolism, brush border membrane and oxidative stress in rat kidney tissues. *Life Sci*, 82(9-10), 450-459. doi: 10.1016/j.lfs.2007.11.014.
- Bulboacă AE, Porfire AS, Rus V, Nicula CA, Bulboacă CA, Bolboacă SD. (2022) Protective effect of liposomal epigallocatechin-gallate in experimental gentamicin-induced hepatotoxicity. *Antioxidants*, 11(2), 412. doi: 10.3390/antiox11020412
- Cai ZM, Peng JQ, Chen Y, Tao L, Zhang YY, Fu LY, Long QD, Shen XC. (2021) 1, 8-Cineole: A review of source, biological activities, and application. *J Asian Nat prod res*, 23(10), 938-954. doi: 10.1080/10286020.2020.1839432.
- Chen C, Chen Y, Wu P, Chen B. (2014) Update on new medicinal applications of gentamicin: evidence-based review. *J Formos Med Assoc* 113(2), 72-82. doi: 10.1016/j.jfma.2013.10.002.
- Cui J, Tang L, Hong Q, Lin S, Sun X, Cai G, Bai XY, Chen X. (2019) N-acetylcysteine ameliorates gentamicin-induced nephrotoxicity by enhancing autophagy and reducing oxidative damage in miniature pigs. *Shock*, 52(6), 622-630. doi: 10.1097/SHK.0000000000001319.
- Đuračková, Z. (2010). Some current insights into oxidative stress. *Physiologic res*, 59(4), 459-469. doi:10.33549/physiol-res.931844.
- Galaly S, Ahmed O, Mahmoud A. (2014) Thymoquinone and curcumin prevent gentamicin-induced liver injury by attenuating oxidative stress, inflammation and apoptosis. *J Physiol Pharmacol*, 65(6), 823-832.
- Galan DM, Ezeudu NE, Garcia J, Geronimo CA, Berry NM, Malcolm B J. (2020) Eucalyptol (1, 8-cineole): an underutilized ally in respiratory disorders? *J Essent oil res*, 32(2), 103-110.
- Graille M, Wild P, Sauvain JJ, Hemmendinger M, Guseva Canu I, Hopf NB. (2020) Urinary 8-OHdG as a biomarker for oxidative stress: a systematic literature review and meta-analysis. *Int J Mol Sci* 21(11), 3743. doi: 10.3390/ijms21113743
- Hoch CC, Petry J, Griesbaum L, Weiser T, Werner K, Ploch M, Verschoor A, Multhoff G, Dezfouli AB, Wollenberg B. (2023) 1, 8-cineole (eucalyptol): A versatile phytochemical with therapeutic applications across multiple diseases. *Biomed Pharmacother*, 167, 115467.
- Hsouna AB, Dhibi S, Dhifi W, Mnif W, Hfaiedh N. (2019) Chemical composition and hepatoprotective effect of essential oil from *Myrtus communis* L. flowers against CCL 4-induced acute hepatotoxicity in rats. *RSC advances*, 9(7), 3777-3787. doi:10.1039/c8ra08204a
- Ibraheem ZO, Farhan SS, Al Sumaidae A, Al Sufi L, Bashir A, Balwa A, Basir R. (2021) Liver functions in combined models of the gentamicin induced nephrotoxicity and metabolic syndrome induced by high fat or fructose diets: a comparative study. *Toxicol Res*, 37, 221-235. doi: 10.1007/s43188-020-00059-w.
- Ince S, Kucukkurt I, Demirel HH, Arslan-Acaroz D, Varol N. (2020) Boron, a trace mineral, alleviates gentamicin-induced nephrotoxicity in rats. *Biol Trace elem res*, 195(2), 515-524. doi: 10.1007/s12011-019-01875-4.
- Iqbal U, Malik A, Sial NT, Uttra AM, Rehman MF, Mehmood MH. (2024) Molecular insights of Eucalyptol (1, 8-Cineole) as an anti-arthritis agent: in vivo and in silico analysis of IL-17, IL-10, NF- κ B, 5-LOX and COX-2. *Inflammopharmacology*, 32(3), 1941-1959. doi: 10.1007/s10787-024-01465-4.
- Kazak F, Akcakavak G, Alakus I, Alakus H, Kirgiz O, Karatas O, Devenci MZY, Coskun P. (2024a) Proanthocyanidin alleviates testicular torsion/detorsion-induced ischemia/reperfusion injury in rats. *Tissue and Cell*, 89, 102459. doi: 10.1016/j.tice.2024.102459
- Kazak F, Devenci MZY, Akcakavak G. (2024b) Eucalyptol alleviates cisplatin-induced kidney damage in rats. *Drug chem toxicol*, 47(2), 172-179. doi: 10.1080/01480545.2022.2156530.
- Kazak F. (2022) A bioactive compound: eucalyptol. *Livre de Lyon, Lyon, France*, pp.125-138.
- Laaroussi H, Bakour, M, Ousaad D, Ferreira-Santos P, Genisheva Z, El Ghouzi A, Aboulghazi A, Teixeira JA, Lyoussi B. (2021) Protective Effect of Honey and Propolis against Gentamicin-Induced Oxidative Stress and Hepatorenal Damages. *Oxid med cell longev*, 2021(1), 9719906. doi: 10.1155/2021/9719906.
- Lang M, Carvalho A, Baharoglu Z, Mazel D. (2023). Aminoglycoside uptake, stress, and potentiation in Gram-negative bacteria: new therapies with old molecules. *Microbiol Mol Biol Rev*. 87(4), e00036-00022.
- Mahi-Birjand M, Yaghoubi S, Abdollahpour-Alitappeh M, Keshkaran Z, Bagheri N, Pirouzi A, Khatami M, Sineh Sepehr K, Peymani P, Karimzadeh I. (2020) Protective effects of pharmacological agents against aminoglycoside-induced nephrotoxicity: A systematic review. *Expert Opin Drug Safety*, 19(2), 167-186. doi: 10.1080/14740338.2020.1712357

- Mirazi N, Baharvand, F, Moghadasali R, Nourian A, Hosseini A. (2021) Human umbilical cord blood serum attenuates gentamicin-induced liver toxicity by restoring peripheral oxidative damage and inflammation in rats. *Basic Clin Pharm Toxicol*, 128(2), 268-274. doi: 10.1111/bcpt.13502
- Mohammed M, Aboulhoda B, Mahmoud R. (2019) Vitamin D attenuates gentamicin-induced acute renal damage via prevention of oxidative stress and DNA damage. *Hum exp toxicol*, 38(3), 321-335.
- Moon HK, Kang P, Lee HS, Min SS, Seol GH. (2014) Effects of 1, 8-cineole on hypertension induced by chronic exposure to nicotine in rats. *J Pharm Pharmacol* 66(5), 688-693. doi: 10.1111/jphp.12195.
- Morales AI, Buitrago JM, Santiago JM, Fernández-Tagarro M, López-Novoa JM, Pérez-Barriocanal F. (2002) Protective effect of trans-resveratrol on gentamicin-induced nephrotoxicity. *Antioxid Redox Signal*, 4(6), 893-898. doi: 10.1089/152308602762197434.
- Ogundipe OJ, Akinpelu OF, Oyerinde A, Oluwakemi OR. (2021) Ocimum gratissimum (Linn) leaves extract attenuates oxidative stress and liver injury in gentamicin-induced hepatotoxicity in rats. *Egypt J Basic Appl Sci*, 8(1), 146-155.
- Rašković A, Milanović I, Pavlović N, Čebović T, Vukmirović S, Mikov M. (2014) Antioxidant activity of rosemary (*Rosmarinus officinalis* L.) essential oil and its hepatoprotective potential. *BMC complement alter med*, 14, 1-9.
- Randjelovic P, Veljkovic S, Stojiljkovic N, Sokolovic D, Ilic I. (2017) Gentamicin nephrotoxicity in animals: Current knowledge and future perspectives. *EXCLI journal*, 16, 388.
- Ryu S, Park H, Seol GH, Choi IY. (2014) 1, 8-Cineole ameliorates oxygen-glucose deprivation/reoxygenation-induced ischaemic injury by reducing oxidative stress in rat cortical neuron/glia. *J Pharm Pharmacol*, 66(12), 1818-1826. doi: 10.1111/jphp.12295.
- Sanchez-Gonzalez PD, Lopez-Hernandez FJ, Perez-Barriocanal F, Morales AI, Lopez-Novoa JM. (2011) Quercetin reduces cisplatin nephrotoxicity in rats without compromising its anti-tumour activity. *Nephrol Dialysis Transplant*, 26(11), 3484-3495. doi: 10.1093/ndt/gfr195.
- Thy M, Timsit JF, de Montmollin E. (2023) Aminoglycosides for the treatment of severe infection due to resistant gram-negative pathogens. *Antibiotics*, 12(5), 860.
- Venkataraman B, Almarzooqi S, Raj V, Bhongade BA, Patil RB, Subramanian VS, Attoub S, Rizvi TA, Adrian TE, Subramanya SB. (2023) Molecular docking identifies 1, 8-Cineole (Eucalyptol) as a novel PPAR γ agonist that alleviates colon inflammation. *Int J Mol Sci*, 24(7), 6160. doi: 10.3390/ijms24076160.
- Yang Y, Sharma R, Sharma A, Awasthi S, Awasthi Y. (2003) Lipid peroxidation and cell cycle signaling: 4-hydroxynonenal, a key molecule in stress mediated signaling. *Acta Biochim Pol*, 50(2), 319-336.
- Wijayanti H, Fadhilah Y, Yuniarti W, Lukiswanto B, Arimbi A, Suprihati E, Kurnijasanti R. (2023). Protective effect of Moringa oleifera leaves extract against gentamicin induced hepatic and nephrotoxicity in rats. *Iraqi J Vet Sci*. 37(1), 129-135.
- Yarjani ZM, Najafi H, Shackebaei D, Madani SH, Modarresi M, Jassemi SV. (2019) Amelioration of renal and hepatic function, oxidative stress, inflammation and histopathologic damages by Malva sylvestris extract in gentamicin induced renal toxicity. *Biomed Pharmacother*, 112, 108635. doi: 10.1016/j.biopha.2019.108635.
- Zoppini G, Cacciatori V, Negri C, Stoico V, Lippi G, Targher G, Bonora E. (2016) The aspartate aminotransferase-to-alanine aminotransferase ratio predicts all-cause and cardiovascular mortality in patients with type 2 diabetes. *Medicine*, 95(43), e4821. doi: 10.1097/MD.0000000000004821.