### **The relationship between ketosis and transitional nutritional diseases**

### **Hasan Atalay**

**Review Article**

Volume: 8, Issue: 3 December, 2024 Pages: 287-293

Department of Animal Nutrition and Nutritional Diseases, Faculty of Veterinary Medicine, Balıkesir University, Campus of Cagış, 10145, Altıeylül, Balıkesir, Turkey, Atalay, H. ORCID: 0000-0002-5744-7538

### **ABSTRACT**

Ketosis is a highly prevalent nutritional condition that affects fresh dairy cows during the transition period. Ketosis occurs when there is a negative energy balance. Clinical findings of ketosis include excessive loss of body condition, decreased feed consumption (especially of concentrated feed), reduced milk yield, and nervous signs. Subclinical ketosis is a serious nutritional disease that may result in displaced abomasum, decreased milk yield, poor reproductive performances, early culling of herds, and economic losses, among other adverse effects. Ketosis is linked to nutritional disorders that are commonly observed after calving including, metritis, mastitis, milk fever, lameness, retained placenta, and displaced abomasum. The two most crucial strategies for preventing ketosis are reducing negative energy balance and increasing dry matter consumption. The density of triacyl-glycerides and non-esterified fatty acids in plasma is higher after calving. Non-esterified fatty acids are oxidized to ketone bodies. A cow starts to mobilize its body fat for energy when it reaches a negative energy balance. Live weight and body condition score fall under such circumstances.

**Keywords:** dairy cattle, ketosis, negative energy balance

**DOI:** https://doi.org/10.30704/http-www-jivs-net.1555851

**To cite this article:** Atalay, H. (2024). The relationship between ketosis and transitional nutritional diseases. *Journal of Istanbul Veterinary Sciences, 8***(3), 287-293. Abbreviated Title***:* **J. İstanbul vet. sci.**

# **Introduction**

**Article History** Received: 25.09.2024 Accepted: 17.12.2024 Available online: 30.12.2024

Due to low feed consumption and high milk output, energy needs cannot be physiologically met at the start of lactation, leading to an energy deficit during this period. The body uses its fat reserves to make up for this deficit. Ketosis usually occurs in high milk producing cows at the start of lactation, when the energy balance is negative. However, it can also occur when dry matter intake is reduced and the diet lacks sufficient energy density in conditions such as displaced abomasum, mastitis, metritis, fatty liver syndrome, and retained placenta, which may reduce appetite and feed intake, increasing the risk of secondary ketosis (Atalay and Eseceli, 2015).

Ketosis is a nutritional disease characterized by liver degeneration, decrease in blood glucose concentration, and increase in ketone bodies in the blood and other body tissues particularly in the postpartum period. In a negative energy balance, the mobilization of body fat results in an increase in the

concentration of free fatty acids in various tissues. Free fatty acids in the liver are limited by the oxidation capacity of the liver. When the capacity is surpassed, free fatty acids are converted to triglycerides, which accumulate in the liver. The incidence of ketosis increases when plasma free fatty acid concentrations exceed 1000 mEq/L (Arslan and Tufan, 2010b).

## **Dry matter consumption**

Dry matter intake, milk output, and blood glucose levels have all been shown to be reduced in ketosis cows, although NEFA, BHBA, and serum total lipid levels are higher (Dann et al., 2005). Cows with subclinical ketosis had higher amounts of BHBA and NEFA, but lower levels of glucose than healthy cows (Zhang et al., 2010).

The following processes supply the glucose and metabolic energy required during the transition period due to low dry matter consumption: hepatic

**\***Corresponding Author: Hasan Atalay

E mail: hasanatalay@balikesir.edu.tr https://dergipark.org.tr/en/pub/http-www-jivs-net



gluconeogenesis; lactic acid accumulated from (Ospina et al., 2010). The likelihood of displaced anaerobic glycolysis in muscle tissue and transported to abomasum, clinical ketosis, and metritis is 2.6–8 times, the liver; glucogenic amino acids released from protein 3–6 times, and 1–5.8 times higher, respectively, if BHBA catabolism; and glycerol released from lipolysis in levels are elevated in cows during the first two weeks adipose tissue (Arslan and Tufan, 2010a; Reynolds et following calving (Dubuc et al., 2010). Metabolic al., 2003).

to a significant decrease in dry matter consumption and levels, both of which suggest negative energy balance. an increased release of NEFA (Stockdale, 2001; Roche Increased ketone bodies in the blood raise the et al., 2009).

easily digestible non-structural carbohydrates to the system cells (Graber et al., 2010). Negative energy diet to balance energy and boost dry matter balance is linked to diseases including mastitis, metritis, consumption. Subclinical acidosis quickly progresses to and retained placenta, which are not metabolic laminitis. The cow with laminitis prefers to rest rather diseases (Grummer et al., 2004). than feed, thus dry matter consumption drops, and the negative energy balance further exacerbates (Kelley, herds in the enterprise. The occurrence of ketosis is 2014).

factors that influence milk output. Hormonal and roughage structure, pasture structure, season, and metabolic changes during the transition phase may lactation number. There is a negative correlation affect dry matter consumption (Ingvartsen and between ketosis prevalence and herd size during Andersen, 2000). Dry matter consumption decreases by lactation. It has been suggested that feeding with Total more than thirty percent in the last three weeks of Mix Ration minimizes the risk of ketosis. The highest gestation (Hayırlı et al., 2002).

nonspecific, including decreased milk production and consumption of concentrated feed has been linked to dry matter consumption as well as environmental an increased risk of ketosis due to reduced energy indifference. Abnormal behaviours like aggression, lack consumption. Cows that had only one calving showed of coordination, and chewing on nonfeed items are the lowest levels of ketosis. Cows in the second displayed by some cows. Ketone levels in milk, urine, or lactation are reported to have a higher prevalence of blood must be measured for a conclusive diagnosis. The ketosis, whereas cows in the third to seventh lactation value and accuracy of different tests have previously have the highest prevalence of ketosis. It has been been reviewed (Tatone et al., 2016).

### **Nutritional diseases**

High milk yielding cows experience decreased blood glucose levels, depleted liver glycogen stores, formation of ketone bodies from unesterified fatty acids in the body, and ketosis because of their high energy requirements during the transition period and their inability to meet these needs. Hypocalcemia, metritis, retained placenta, ketosis, and displaced abomasum may occur simultaneously. Abomasal atony is the biggest contributor to displaced abomasum. Atony is caused by RPT, ulcers, metritis, mastitis, retentio secundinarum, acidosis, elevated volatile fatty acids, and low blood calcium levels (Rukkwamsuk et al., 1999).

BHBA ˃ 1.2 mmol/ L is linked to abomasum **Hormonal change**  displacement, mastitis, metritis, and increased milk loss Insulin levels in cows have been reported to be low in cows experiencing ketosis in the first one or two after calving (Holtenius et al., 1993). Lactating cows weeks postpartum (LeBlanc, 2010). The cut-off value produce only half the insulin that non-lactating cows do

Obesity in cows during the parturition period leads glucose levels as well as high blood NEFA and BHBA Subclinical acidosis is caused by the addition of can disrupt the activities of milk and blood immune diseases are frequently associated with low blood possibility of left-displaced abomasum. Ketone bodies

Dry matter consumption is one of the most key geographical location, temperature, barn structure, Numerous clinical indicators of ketosis are during the second quarter of the year (April-June). Low It is critical to assess the prevalence of ketosis in the influenced by a variety of factors, including prevalence of ketosis was reported in cows calving stated that ketosis in the first week after calving is linked to increased abomasum displacement, metritis, and mastitis. Another study indicates a significant link among ketosis, mastitis, and clinical laminitis (Berge and Vertenten, 2014).

> Fatty liver is a major risk factor for abomasal displacement, ketosis, and immune dysfunction. In contrast, if these disorders reduce dry matter intake and induce a more severe negative energy balance, they may increase the likelihood of developing fatty liver. Obesity is a significant risk factor for developing fatty liver (Bobe et al., 2004). In the early postpartum period, fatty cows consume less dry matter than cows in proper body condition, which results in higher mobilization of body fat (Stockdale, 2001).

for subclinical ketosis in blood BHBA is 1-1.4 mmol/L (Lomax et al., 1979). Insulin resistance develops in

Holtenius, 1996). Insulin levels vary during the protein ratio at the start of lactation suggests a transition period. Low insulin levels after parturition negative energy balance. If the fat/protein ratio decrease glucose uptake in insulin-sensitive organs, exceeds 1.4, it indicates that fat is being mobilized in such as muscle and adipose tissue. This condition favors the adipose tissue and that the animal is in a negative glucose uptake in the mammary gland, which is not energy state. Milk acetone levels are commonly used to influenced by insulin action. Thus, the concentration of diagnose subclinical ketosis. If the milk acetone triglycerides in the liver rises in response to adipose concentration exceeds 0.7 mmol/L, the animal may be tissue mobilization in the body (Aschenbach et al., in ketosis. It is important to focus on the number of 2010).

glucose metabolism (Komatsu et al., 2005). NEFA indicator data. If more than 10% of the cows in the circulating freely during negative energy balance herd surpass the BHBA threshold, this is a strong contributes to milk fat formation (Bauman and Griinari, indication of a negative energy balance in the herd 2001). In a lactating cow experiencing negative energy (Serbester et al., 2012). balance, milk fat concentration tends to increase, while milk protein concentration tends to decrease. The fat type I and type II. Type I ketosis usually develops a few to protein ratio expresses negative energy intensity and weeks after parturition, when milk supply and the risk of metabolic disease. Milk samples should be mammary gland glucose demands are high. However, it evaluated within 9 to 30 days to estimate the risk is not generally associated with high hepatic fat factors for metabolic diseases (Duffield et al., 1997; concentrations. Type II occurs at or around parturition Toni et al., 2011).

adipose tissue and reduces blood glucose levels by NASEM, 2021). This classification method indicates two increasing glucose absorption into tissues. Insulin major causes of ketosis. The risk factors and causes of resistance refers to a decrease in the tissues' response type II ketosis are similar to those of fatty liver. Type I to insulin actions. As insulin resistance in adipose cows had lower blood glucose and insulin tissues increases, so does fatty acid mobilization and concentrations, as well as higher ketone concentrations the concentration of non-esterified fatty acids in the compared to healthy cows. Low insulin increases fatty blood. The body condition score used in determining acid oxidation, likely by reducing hepatocyte malonylenergy balance provides information regarding the CoA levels and decreasing the sensitivity of carnitine change in live weight. It has been stated that one unit palmitoyl-transferase 1 to malonyl-CoA. (Emery et al., change in body condition score corresponds to an 80 kg 1992; NASEM, 2021). Carnitine palmitoyltransferase 1 change in live weight. The fatty tissues that are is responsible for the transport of fatty acids from the mobilized or the live weights that are regained are cytosol to the mitochondria for oxidation, and it is located in the abdominal, intramuscular, and highly active in type I ketosis. Thus, this suggests that subcutaneous regions. Energy balance is expressed by type I ketosis is caused by an insufficient supply of the blood molecule β-Hydroxybutyric acid (BHBA). If gluconeogenic precursors. Low dry matter intake could the amount of non-esterified fatty acids entering the be the cause of limited substrate supply. Studies reveal liver exceeds the capacity for fatty acid oxidation, that increasing dietary starch postpartum lowers blood ketone bodies increase. Blood analysis should be BHBA while increasing glucose (Rabelo et al., 2005; performed in the second week of lactation and 5 hours McCarthy et al., 2015; NASEM, 2021). Propylene glycol after feeding to assess BHBA levels in the blood. The administered as a drip or bolus can lower blood BHBA BHBA blood threshold is 1.2 to 1.4 mmol/L. There is a levels (Nielsen and Ingvartsen, 2004). strong link between milk yield and energy status. When **Fatty liver** there is an energy deficiency, microbial protein Ketone bodies are produced when excess NEFA enters synthesis and the number of amino acids in mammary the liver. Subclinical ketosis may occur because not all tissue decrease, lowering the level of milk protein. A NEFA can be completely oxidized. Negative energy milk protein content of less than 2.7% relates to a balance leads to ketosis. When the body's energy negative energy value. Blood analysis should be used to reserves are mobilized, the concentrations of NEFA and determine milk protein concentration between weeks 1 ketone bodies (acetoacetate, BHBA, and acetone) and 12 of lactation, and 5 hours after feeding. Milk fat increase. Increased NEFA concentration contributes to levels increase but milk protein levels decrease during ketosis (Weber et al., 2013).

peripheral tissues during early lactation (Holtenius and lactation due to a negative energy balance. A high fat/ The mammary gland does not require insulin for the average value when interpreting metabolic samples that surpass the threshold values rather than

Insulin inhibits the mobilization of fatty acids from often more difficult to treat than type I (Herdt, 2000; Holtenius and Holtenius (1996) classified ketosis as and is typically linked with fatty liver. Type II ketosis is

Fat accumulates in the liver when fatty acid intake exceeds the liver's ability to oxidize or eliminate fatty fatty acids butyric acid, propionic acid, and acetic acid. acids, which occurs when blood concentrations of The liver uses acetic acid as a source of acetyl stored fatty acids increase (Bobe et al., 2004; Grummer, coenzyme A to make ATP. The rumen absorbs butyric 2008). Ketosis almost always develops when cows have acid, which is then transformed into ketone bodies moderate (5 to 10% of liver wet weight raised as (BHBA). The liver uses propionic acid in the process of triacylglycerides) to severe (more than 10% increased gluconeogenesis (Reynolds et al., 1988). as triacylglycerides) fatty liver. Plasma NEFA concentrations in healthy cows are low (less than 0.2 parturition, insulin level was found to be consistent mEq/L) until a few days before parturition, increases to with liver lipid concentrations in cows fed a high energy as high as 0.8 mEq/L during parturition, remain high for diet compared to those on a restricted diet. However, several days, and then gradually fall (Bertics et al., because it was higher, this level was found to be 1992; NASEM, 2021). Cows with a higher risk of inconsistent with the NEFA and BHBA levels (Dann et postpartum metabolic disorders often have plasma al., 2006). Compared to cows fed a high energy NEFA concentrations higher than 0.5 mEq/L prepartum intensive diet, more significant and relevant results and higher than 1.0 mEq/L postpartum (LeBlanc et al., were found in dry matter consumption and energy 2005; NASEM, 2021).

NEFA concentrations in the blood (Emery et al., 1992) dry period, cows fed a restricted energy diet eliminated and NEFA taken up by the liver can be esterified or negative energy balance and achieved a balanced oxidized (Drackley, 1999). Triglycerides, the primary energy state more quickly than those fed an intensive result of esterification, can be absorbed or retained as diet. Additionally, it was noted that cows overfed part of very low density lipoprotein. Compared to other during the far-off dry period following parturition species, ruminants absorb triglycerides at a very slow showed signs of health issues (Dann et al., 2006, Kelley, pace (Kleppe et al., 1988; Pullen et al., 1990). As a 2014). Cows tend to consume more feed during the dry result, fatty acid esterification and triglyceride buildup period to gain fat, which provides energy after occur in the liver when it takes up excessive levels of parturition. hepatic NEFA.

### **Negative energy balance**

When milk production begins, the cow's energy need increases by around threefold compared to the period before parturition (Van Dorland et al., 2009). As milk production begins, glucose and metabolic energy needs increase two to threefold by the 21st day postpartum, compared to the 21st day prepartum (Drackley et al., 2001).

More than fifty percent of cows are thought to have at least one subclinical illness throughout the transition phase (Overton and Waldron, 2004). The negative energy picture is shaped by the imbalance between stagnant dry matter consumption during the transition period and increased energy demand during lactation. The relationship between energy expended through milk production during lactation, the mobilization of body fat, and energy intake from feed defines the condition known as ketosis (Collard et al., 2000; Mulligan et al., 2006). The transition period for highyielding dairy cattle is extremely challenging. During this time, the animal frequently enters a negative energy state. A cow with a negative energy balance requires more energy than it can receive from feed (De vries et al., 1999). Dry matter consumption during this time is most likely determined by NEFA oxidation in the liver (Allen et al., 2009).

Microorganisms in the rumen produce the volatile

The uptake of NEFA by the liver is proportional to off dry period. It was reported that during the far-off During the far off dry period, 4 to 6 weeks before balance in cows fed a restricted diet throughout the far

> Cows that are in negative energy balance which can happen when their dry matter intake decreases during the prepartum phase and almost always happens during the first few weeks of lactation have higher plasma NEFA levels because their energy needs are high and their intake is low, which causes their body reserves to be mobilized. Most cows in the early stages of lactation have some degree of hepatic fat accumulation since NEFA levels are elevated in most of them during the peripartum period (Jorritsma et al., 2001; Bobe et al., 2004; NASEM, 2021).

> Ketosis, also known as hyperketonaemia, occurs when an excess of long chain fatty acids is oxidized by B -oxidation. Cows with a severe negative energy balance after calving release an excessive amount of long-chain fatty acids. Ketone bodies (B-hydroxybutyric acid and acetoacetate) are the byproducts of B-oxidation, and when they accumulate in the blood, clinical symptoms can appear. When plasma insulin levels are low and glucagon levels are high, fatty acid release from adipose tissue and subsequent B-oxidation are triggered (Holtenius and Holtenius, 1996; NASEM, 2021).

## **Conclusion**

The transition period includes the three weeks preceding parturition and the three weeks following parturition. Almost all high-milk-yielding cows

experience a negative energy balance throughout the Bobe, G., Young, J. W., & Beitz, D. C. (2004). Invited transition period. This negative energy balance results from an increased need for energy following parturition, which is caused by a decrease in dry matter intake and the onset of milk secretion. In a negative Collard, B. L., Boettcher, P. J., Dekkers, J. M., Petitclerc, energy balance, the cow mobilizes body fat to meet its NEFA (Non-Esterified Fatty Acids) requirements. The cow with a negative energy balance attempts to correct the imbalance by boosting NEFA and BHBA (Beta Hydroxy Butyric Acid) levels. Because the cow is in Dann, H. M., Morin, D. E., Bollero, G. A., Murphy, M. R., negative energy balance during ketosis, ration energy cannot be used to make up the difference. Thus, conducting a ration study during ketosis might not yield useful results.

## **References**

- Allen, M. S., Bradford, B. J., & Oba, M. (2009). Boardinvited review: The hepatic oxidation theory of the control of feed intake and its application to ruminants*. Journal of Animal Science, 87*(10), 3317- 3334.
- Arslan, C., & Tufan, T. (2010a). Geçiş dönemindeki süt ineklerinin beslenmesi I. Bu dönemde görülen fizyolojik, hormonal, metabolik ve immonolojik değişiklikler ile beslenme ihtiyaçları. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi, 16*(1), 151- 158.
- Arslan, C., & Tufan, T. (2010b). Geçiş dönemindeki süt ineklerinin beslenmesi II. Bu dönemde görülen metabolik hastalıklar ve besleme ile önlenmesi. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi, 16*(1), 159-166.
- Aschenbach, J. R., Kristensen, N. B., Donkin, S. S., Hammon, H. M., & Penner, G. B. (2010). Gluconeogenesis in dairy cows: the secret of making sweet milk from sour dough. *IUBMB Life, 62*(12), 869 -877.
- Atalay, H., & Eseceli, H. (2015). Doğum sonrası yüksek verimli sığırlarda ketozis. *Balıkesir ili Damızlık Sığır Yetiştiricileri Birliği Yayınları, 7*(27), 37-39.
- Bauman, D. E., & Griinari, J. M. (2001). Regulation and nutritional manipulation of milk fat: low-fat milk syndrome. *Livestock Production Science, 70*(1-2), 15- 29.
- Berge, A. C., & Vertenten, G. (2014). A field study to determine the prevalence, dairy herd management systems, and fresh cow clinical conditions associated with ketosis in European dairy herds. *Journal of Dairy Science, 97*(4), 2145-2154.
- Bertics, S. J., Grummer, R. R., Cadorniga-Valino, C., & Stoddard, E. E. (1992). Effect of prepartum dry matter intake on liver triglyceride concentration and early lactation. *Journal of Dairy Science, 75*(7), 1914- 1922.
- review: pathology, etiology, prevention, and treatment of fatty liver in dairy cows. *Journal of Dairy Science, 87*(10), 3105-3124.
- D., & Schaeffer, L. R. (2000). Relationships between energy balance and health traits of dairy cattle in early lactation. *Journal of Dairy Science, 83*(11), 2683 -2690.
- & Drackley, J. K. (2005). Prepartum intake, postpartum induction of ketosis, and periparturient disorders affect the metabolic status of dairy cows. *Journal of Dairy Science, 88(*9), 3249-3264.
- Dann, H. M., Litherland, N. B., Underwood, J. P., Bionaz, M., D'angelo, A., McFadden, J. W., & Drackley, J. K. (2006). Diets during far-off and close-up dry periods affect periparturient metabolism and lactation in multiparous cows. *Journal of Dairy Science, 89*(9), 3563-3577.
- Duffield, T. F., Kelton, D. F., Leslie, K. E., Lissemore, K. D., & Lumsden, J. H. (1997). Use of test day milk fat and milk protein to detect subclinical ketosis in dairy cattle in Ontario. *Canadian Veterinary Journal, 38* (11), 713.
- De Vries, M. J., Van Der Beek, S., Kaal-Lansbergen, L. M. T. E., Ouweltjes, W., & Wilmink, J. B. M. (1999). Modeling of energy balance in early lactation and the effect of energy deficits in early lactation on first detected estrus postpartum in dairy cows. Journal *of Dairy Science, 82*(9), 1927-1934.
- Drackley, J. K. (1999). ADS Foundation Scholar Award Biology Of Dairy Cows During The Transition Period: The Final Frontier. *Journal of Dairy Science, 82*(11), 2259-2273.
- Drackley, J. K., Overton, T. R., & Douglas, G. N. (2001). Adaptations of glucose and long-chain fatty acid metabolism in liver of dairy cows during the periparturient period. *Journal of Dairy Science, 84*, E100-E112.
- Dubuc, J., Duffield, T. F., Leslie, K. E., Walton, J. S., & LeBlanc, S. J. (2010). Risk factors for postpartum uterine diseases in dairy cows. *Journal of Dairy Science, 93*(12), 5764-5771.
- Emery, R. S., Liesman, J. S., & Herdt, T. H. (1992). Metabolism of long chain fatty acids by ruminant liver. *Journal of Nutrition, 122*, 832-837.
- Graber, M., Kohler, S., Kaufmann, T., Doherr, M. G., Bruckmaier, R. M., & Van Dorland, H. A. (2010). A field study on characteristics and diversity of gene expression in the liver of dairy cows during the transition period. *Journal of Dairy Science, 93*(11), 5200-5215.
- Grummer, R. R., Mashek, D. G., & Hayırlı, A. (2004). Dry LeBlanc, S. J., Leslie, K. E., & Duffield, T. F. (2005). matter intake and energy balance in the transition period. Veterinary Clinics: *Food Animal Practice, 20* (3), 447-470.
- Grummer, R. R. (2008). Nutritional and management strategies for the prevention of fatty liver in dairy cattle. *Veterinary Journal, 176*(1), 10-20.
- Hayırlı, A., Grummer, R. R., Nordheim, E. V., & Crump, P. M. (2002). Animal and dietary factors affecting feed intake during the prefresh transition period in 3443.
- Herdt, T. H. (2000). Ruminant adaptation to negative energy balance: Influences on the etiology of ketosis and fatty liver. *Veterinary Clinics of North America: Food Animal Practice, 16*(2), 215-230.
- Holtenius, P., Olsson, G., & Björkman, C. (1993). Periparturient concentrations of insulin glucagon and ketone bodies in dairy cows fed two different levels of nutrition and varying concentrate/roughage ratios. *Journal of Veterinary Medicine Series A, 40(1‐ 10)*, 118-127.
- Holtenius, P., & Holtenius, K. (1996). New aspects of ketone bodies in energy metabolism of dairy cows: a review. *Journal of Veterinary Medicine Series A, 43(1 ‐10)*, 579-587.
- Ingvartsen, K. L., & Andersen, J. B. (2000). Integration of metabolism and intake regulation: a review focusing on periparturient animals. *Journal of Dairy Science, 83*(7), 1573-1597.
- Jorritsma, R., Jorritsma, H., Schukken, Y. H., Bartlett, P. C., Wensing, T. H., & Wentink, G. H. (2001). Prevalence and indicators of post partum fatty infiltration of the liver in nine commercial dairy herds in The Netherlands. *Livestock Production Science, 68*(1), 53-60.
- Kelley, A. W. (2014). Physiological impacts and Overton, T. R., & Waldron, M. R. (2004). Nutritional lactational performance of dairy cows fed Brown midrib corn silage during dry period through early to midlactation, *Master of Science (MS),* Utah State University, US, digitalcommons. usu. edu/etd/ Pullen, D. L., Liesman, J. S., & Emery, R. S. (1990). A index.9.html
- Kleppe, B. B., Aiello, R. J., Grummer, R. R., & Armentano, L. E. (1988). Triglyceride accumulation and very low density lipoprotein secretion by rat and goat hepatocytes in vitro. Journal *of Dairy Science, 71*(7), 1813-1822.
- Komatsu, T., Itoh, F., Kushibiki, S., & Hodate, K. (2005). Changes in gene expression of glucose transporters in lactating and nonlactating cows. *Journal of Animal Science, 83*(3), 557-564.
- Metabolic predictors of displaced abomasum in dairy cattle. Journal of Dairy Science, 88(1), 159-170.
- Leblanc, S. (2010). Monitoring metabolic health of dairy cattle in the transition period. *Journal of Reproduction and Development, 56*, 29-35.
- Lomax, M. A., Baird, G. D., Mallinson, C. B., & Symonds, H. W. (1979). Differences between lactating and non -lactating dairy cows in concentration and secretion rate of insulin. *Biochemical Journal, 180*(2), 281-289.
- Holsteins. *Journal of Dairy Science, 85*(12), 3430- McCarthy, M. M., Yasui, T., Ryan, C. M., Mechor, G. D., & Overton, T. R. (2015). Performance of earlylactation dairy cows as affected by dietary starch and monensin supplementation. *Journal of Dairy Science, 98*(5), 3335-3350.
	- Mulligan, F. J., O'grady, L., Rice, D. A., & Doherty, M. L. (2006). A herd health approach to dairy cow nutrition and production diseases of the transition cow. *Animal Reproduction Science, 96*(3-4), 331-353.
	- NASEM 2021 (*National Academies of Science, Engineering, and Medicine) Nutrient Requirements of Dairy Cattle, 8th ed*., Washington, DC.,US, National Academies Press
	- Nielsen, N. I., & Ingvartsen, K. L. (2004). Propylene glycol for dairy cows: A review of the metabolism of propylene glycol and its effects on physiological parameters, feed intake, milk production and risk of ketosis. *Animal Feed Science and Technology, 115*(3- 4), 191-213.
	- Ospina, P. A., Nydam, D. V., Stokol, T., & Overton, T. R. (2010). Association between the proportion of sampled transition cows with increased nonesterified fatty acids and β-hydroxybutyrate and disease incidence, pregnancy rate, and milk production at the herd level. *Journal of Dairy Science, 93*(8), 3595-3601.
	- management of transition dairy cows: strategies to optimize metabolic health. Journal *of Dairy Science, 87*, E105-E119.
	- species comparison of liver slice synthesis and secretion of triacylglycerol from nonesterified fatty acids in media. *Journal of Animal Science, 68(*5), 1395-1399.
	- Rabelo, E., Rezende, R. L., Bertics, S. J., & Grummer, R. R. (2005). Effects of pre-and postfresh transition diets varying in dietary energy density on metabolic status of periparturient dairy cows. *Journal of Dairy Science, 88*(12), 4375-4383.
- Reynolds, C. K., Huntington, G. B., Tyrrell, H. F., & Toni, F., Vincenti, L., Grigoletto, L., Ricci, A., & Reynolds, P. J. (1988). Net portal-drained visceral and hepatic metabolism of glucose, L-lactate, and nitrogenous compounds in lactating Holstein cows. *Journal of Dairy Science, 7*1(7), 1803-1812.
- J., & Beever, D. E. (2003). Splanchnic metabolism of dairy cows during the transition from late gestation through early lactation. *Journal of Dairy Science, 86* (4), 1201-1217.
- Relationship between overfeeding and overconditioning in the dry period and the problems of high producing dairy cows during the postparturient period. *Veterinary Quarterly, 21*(3), 71-77.
- Roche, J. R., Friggens, N. C., Kay, J. K., Fisher, M. W., Body condition score and its association with dairy cow productivity, health, and welfare. *Journal of Dairy Science, 92*(12), 5769-5801.
- Serbester, U., Çınar, M., & Hayırlı, A. (2012). Sütçü ineklerde negatif enerji dengesi ve metabolik indikatörleri. *Kafkas Üniversitesi Veteriner Fakültesi Dergisi, 18*(4).
- Stockdale, C. R. (2001). Body condition at calving and the performance of dairy cows in early lactation under Australian conditions: a review. *Australian Journal of Experimental Agriculture, 41*(6), 823-839.
- Tatone, E. H., Gordon, J. L., Hubbs, J., LeBlanc, S. J., DeVries, T. J., & Duffield, T. F. (2016). A systematic review and meta-analysis of the diagnostic accuracy of point-of-care tests for the detection of hyperketonemia in dairy cows. *Preventive Veterinary Medicine, 130*, 18-32.
- Schukken, Y. H. (2011). Early lactation ratio of fat and protein percentage in milk is associated with health, milk production, and survival. *Journal of Dairy Science, 94*(4), 1772-1783.
- Reynolds, C. K., Aikman, P. C., Lupoli, B., Humphries, D. Van Dorland, H. A., Richter, S., Morel, I., Doherr, M. G., Castro, N., & Bruckmaier, R. M. (2009). Variation in hepatic regulation of metabolism during the dry period and in early lactation in dairy cows. *Journal of Dairy Science, 92*(5), 1924-1940.
- Rukkwamsuk, T., Kruip, T. A. M., & Wensing, T. (1999). Weber, C., Hametner, C., Tuchscherer, A., Losand, B., Kanitz, E., Otten, W., ... & Hammon, H. M. (2013). Hepatic gene expression involved in glucose and lipid metabolism in transition cows: Effects of fat mobilization during early lactation in relation to milk performance and metabolic changes. *Journal of Dairy Science, 96*(9), 5670-5681.
	- Stafford, K. J., & Berry, D. P. (2009). Invited review: Zhang, Z., Li, X., Wang, H., Guo, C., Gao, L., Liu, L., ... & Liu, G. (2011). Concentrations of sodium, potassium, magnesium, and iron in the serum of dairy cows with subclinical ketosis. *Biological Trace Element Research, 144*, 525-528.