Determination of *Neospora caninum* in cattle fetuses from the Central Black Sea region using PCR

Rahşan Akpınar¹[∗] [©], Selma Kaya² [©], Coşkun Aydın³ [©], Şakir Önder Türlek⁴ [©], Semanur Çelik⁵ [©]

1,2,3,4,5 T.C. Ministry of Agriculture and Forestry, Samsun Veterinary Control Institute, Samsun, Türkiye

Geliş Tarihi / Received: 28.09.2024, Kabul Tarihi / Accepted: 31.10.2024

Abstract: Neosporosis is an infectious disease caused by the *Neospora caninum*, which leads to abortions in cattle. It causes significant economic losses in both global livestock farming and has recently become one of the leading causes of abortions. This study aims to determine the prevalence of *Neospora* infection in aborted fetuses from 136 cattle in enterprises where large-scale cattle farming is intensively practiced in the Central Black Sea Region (Amasya, Samsun, and Tokat provinces). For the diagnosis of *N. caninum*, DNA isolation was performed aborted fetuses, and these samples were subsequently analyzed using the PCR test method. According to the results obtained, *N. caninum* was found in 3 out of 136 aborted fetuses. In conclusion, this study conducted on cattle in the Central Black Sea Region detected the presence of *Neospora caninum* in aborted fetal samples at a rate of 2.2%. These findings indicate that *N. caninum* should not be overlooked in future studies involving aborted fetal samples. Conducting comprehensive research on the definitive hosts of the parasite will play a crucial role in controlling neosporosis and contribute to the development of effective strategies to prevent the spread of the disease.

Keywords: Cattle, Central Black Sea, Fetus, Neosporosis, PCR

Orta Karadeniz Bölgesindeki sığır fetüslerinde Neospora caninum'un PCR ile belirlenmesi

Özet: Neosporozis, *Neospora caninum'un* neden olduğu, sığırlarda aborta neden olan enfeksiyöz bir hastalıktır. Hastalık, büyükbaş hayvan yetiştiriciliğinde önemli ekonomik kayıplara neden olmakta ve son yıllarda abortların başlıca nedenleri arasında yer almaktadır. Bu çalışma, Orta Karadeniz Bölgesinde (Amasya, Samsun ve Tokat illeri) büyükbaş hayvancılığın yoğun olarak yapıldığı işletmelerde abort yapan 136 sığıra ait atık fetüslerde *Neospora sp. varlığını ve* prevalansını belirlemeyi hedeflemektedir. *Neospora caninum*'un teşhisi için atık fetüslerden DNA izolasyonu yapılmış ve ardından bu örnekler PCR test yöntemi ile analiz edilmiştir. Elde edilen sonuçlar doğrultusunda, 136 sığıra ait atık fetüslerden 3'ünde *N. caninum*'a rastlanmıştır. Sonuç olarak, Orta Karadeniz Bölgesindeki sığırlarda gerçekleştirilen bu çalışmada aborte fetüs numunelerinde *N. caninum*'un varlığı %2.2 oranında tespit edilmiştir. Bu bulgular, aborte fetüs numuneleri ile yapılacak gelecekteki çalışmalarda *N. caninum*'un göz ardı edilmemesi gerektiğini göstermektedir. Parazitin son konaklarıyla ilgili kapsamlı araştırmaların yürütülmesi, neosporosisin kontrolünde önemli bir rol oynayacak ve hastalığın yayılmasını engellemeye yönelik etkili stratejiler geliştirilmesine katkı sağlayacaktır.

Anahtar kelimeler: Fetüs, Neosporozis, Orta Karadeniz, PCR, Sığır

Introduction

Neosporosis is a disease caused by the protozoan *Neospora caninum*, which has a two-host heteroxenous life cycle. This disease can lead to clinical signs in various animal species, especially cattle and dogs. Neosporosis is recognized as a parasitic factor causing abortions in both wild and domestic animal species worldwide, particularly in cattle (Dubey et al. 2007; Kaltungo and Musa 2013). Transplacental infection in cattle is an important source of transmission for the parasite; however, the primary route of infection occurs through the oral intake of oocysts shed in dog feces (McAllister et al. 1998). The prevalence of *N. caninum* is attributed to the consumption of placental, aborted fetal, or uterine debris by dogs, which serves as a source of postnatal infection (Davison et al. 2001; Schares et al. 2002; Dubey 2003; Toolan 2003; Salehi et al. 2009; Goodswen et al. 2013). The presence of definitive host dogs in areas where cattle are kept and the contamination of feed and water with their feces are believed to contribute to the spread of infection (Dijkstra et al. 2002; Dubey et al. 2007).

Yazışma adresi / Correspondence: Rahşan Akpınar, Yeşildere mah., Atatürk Bulvarı, Alaçam Cad., Atakum-Samsun, Türkiye e-mail: rahsan.akpinar@tarimorman.gov.tr

ORCID IDs of the authors: 10000-0003-0075-9247 • 20000-0002-8934-3418 • 30000-0001-6244-414X • 40000-0001-6970-8179 • 50000-0001-5657-1273

In the diagnosis of N. caninum infection, various serological and molecular tests, along with histopathological and immunopathological examinations using light and electron microscopy, can be employed. These methods are important tools for accurately diagnosing the infection and gaining more insight into the pathogenesis of the disease (Barber et al. 1995; Lally et al. 1996; Ortega-Mora et al. 2006; Lindsay and Dubey 2020). The determination of N. caninum-specific antibodies in cattle is typically preferred through serological methods (Dubey and Schares 2006). An observed increase in antibody titers during mid-pregnancy in seropositive animals is considered an indication of reactivation of latent infection (Lindsay and Dubey 2020). Neosporosis has been associated with abortion in seropositive animals (Anderson et al. 2000; Dubey 2003; Açıcı et al. 2019). However, it has been observed that approximately 95% of calves born to seropositive cows are clinically normal despite being congenitally infected (Dubey 1999a; Quintanilla et al. 2000; Dubey 2003). The most effective method for controlling the disease is the culling of infected animals once a definitive diagnosis has been made (Reichel et al. 2013). Nevertheless, the absence of clinical signs in infected cattle other than abortion complicates the diagnosis of the disease (Barber et al. 1995).

Neosporosis represents a significant parasitic threat to cattle populations, with a high prevalence reported globally. This disease is responsible for considerable economic losses (McAllister et al. 1998; Trees et al. 1999; McAllister et al. 2000). The prevalence of N. caninum varies by region: 3.4% to 36.2% in Africa Ayinmode et al. (2017), Abdeltif et al. (2022); 37.5% to 70% in northeast Thailand Kashiwazaki et al. (2001); 0.5% to 3.9% in the Czech Republic Václavek et al. (2007), Bártová et al. (2015); 4.1% in Germany and France Dubey et al. (2007); 2.7% to 44.4% in Australia Dubey et al. (2007), Nasir et al. (2012); 23.6% to 91.2% in Brazilian Ragozo et al. (2003), Guedes et al. (2008) and 5.2% in North Dakotato Khaitsa et al. (2006). In Türkiye, serological studies have reported the seroprevalence of neosporosis in cattle to be between 2% and 37.2% (Biyıkoğlu et al. 2001; Aktaş et al. 2005; Pişkin and Ütük 2009; Kasap et al. 2020; Bulut et al. 2021; Köse et al. 2021; Kula and Gökpinar 2021). While some studies exist on the prevalence of the disease in Türkiye, they seem insufficient considering the cattle population in the country. Regularly conducted parasitological studies at specific intervals would particularly help in determining the prevalence of the disease. The

objective of this study is to ascertain the prevalence of neosporosis in abortion cases in the provinces of Amasya, Samsun, and Tokat, where extensive cattle farming is the predominant practice.

Materials and Methods

Collection of fetal samples

The samples used in this study were obtained from the abortions of cattle sent for routine diagnosis to the Samsun Veterinary Control Institute from Amasya (n=16), Samsun (n=80), and Tokat (n=40) provinces. The organ samples taken from these calves were delivered to the Parasitology Laboratory in sterile containers, numbered and stored at -20°C until analysis. The provinces from where the fetal samples were sent are shown in Figure 1.



Figure 1. The provinces from where the fetal samples were taken for analysis regarding *Neospora caninum*

This study aimed to determine the prevalence of Neospora infection in aborted fetal tissues (lung, heart, liver, spleen, and stomach contents) from 136 cattle that were aborted in the provinces of Amasya, Samsun, and Tokat, located in the Central Black Sea Region. Approximately 25 mg tissue pieces (lung, heart, liver, spleen, and stomach contents) were taken from each fetal sample and transferred to 7 mL cryo-tubes, followed by the addition of 3 mL of PBS. The samples were homogenized in an automatic homogenization device (Bead Ruptor Elite, Bead Mill Homogenizer, SKU 19-042E, OMNI International, USA) at 7000 rpm for 1 minute. Following homogenization, the samples were centrifuged at +4°C at 4000 rpm for 10 minutes, and 100 µL of the supernatant was taken for DNA extraction according to the manufacturer's protocol (Genomic DNA Mini Kit/Invitrogen). Specific primers for Neospora

caninum, Np6/Np21 primers (5'-GGG TGT GCG TCC AAT CCT GTA AC-3', 5'-CTC GCC AGT CAA CCT ACG TCT TCT-3') were used for PCR (Kamali et al. 2014). The PCR mixture was prepared in a total volume of 25 μL, consisting of 2.5 μL Dream Taq buffer, 0.5 μL dNTP Mix (10 mM), 0.8 µL of each primer (10 pmol), 0.4 μ L Dream Taq DNA polymerase (5 U/ μ L), and 15 μ L sterile distilled water, with 5 μ L of template DNA added to reach a total volume of 20 µL. The PCR mixture was placed in a thermal cycler with the following amplification conditions: an initial denaturation at 94°C for 7 minutes, followed by 35 cycles consisting of denaturation at 94°C for 1 minute, annealing at 60°C for 1 minute, and extension at 72°C for 1 minute. The process concluded with a final extension step at 72°C for 7 minutes. Subsequently, the amplified PCR products were subjected to electrophoresis in a 1% agarose gel stained with 0.05% ethidium bromide (5 mg/mL) at 90 V and 100 mA for 50 minutes, and the presence of DNA bands was visualized under UV light using a gel imaging device. A PCR product showing a band of 337 bp was considered positive.

Statistical analysis

The frequencies of aborted fetal samples from 136 cattle between 2018 and 2020 were analyzed and summarized in a frequency table 1.

Results

As a result of this study, 136 aborted fetal samples from cattle were analyzed using Polymerase Chain Reaction (PCR), and *Neospora caninum* was detected in 3 samples (2.2%). Of the 80 samples from Samsun, 2 (2.5%) were positive, and of the 40 samples from Tokat, 1 (2.5%) was positive. No *N. caninum* was found in any of the 16 samples from Amasya. The number of samples collected by province and the positivity status are presented in Table 1.

Table 1. Distribution of cattle abortion materials in this study according to provinces and results.

Provinces	Taken samples	Positive	Negative
Amasya	16	0	16
Samsun	80	2	78
Tokat	40	1	39
Total	136	3	133

The PCR image of the *N. caninum* abortion samples is shown in Figure 2.



Figure 2: For *Neospora caninum*, PCR analysis of the samples showed specific banding at 337 bp. M: Molecular weight marker (100 bp ladder), PC: Positive control, NC: Negative control, 1,8: Positive samples, 2-7: Negative samples

Discussion and Conclusion

Neosporosis is one of the most important parasitic causes of cattle abortions, widely observed both globally and in Türkiye, leading to significant economic losses. N. caninum is a parasite with a broad host spectrum and can cause infections in many domestic and wild animals, particularly cattle. This situation increases the impact of N. caninum on both livestock farming and natural ecosystems, resulting in serious economic losses. The diversity of hosts facilitates the spread of the parasite and complicates its control. Therefore, the prevention and management of N. caninum infections are crucial for animal health and productivity (McAllister et al. 1998; Dubey 1999b; Dubey et al. 2007; Sentürk et al. 2020). Congenital infections associated with N. caninum can lead to abortions, stillbirths, and the birth of clinically or subclinically infected calves at different stages of pregnancy. This situation poses a significant problem for cattle breeding and causes economic losses. Since the timing of these infections has a decisive impact on animal health and productivity, careful monitoring and management are required (Innes et al. 2007). One of the main reasons for abortions caused by N. caninum is the presence of definitive host dogs on farms. The presence of these dogs in the same environment as cattle facilitates the contamination of feed and water sources with feces from canids, thereby promoting the spread of infection (Dubey et al. 2007; Kaltungo and Musa 2013).

Abortions related to neosporosis can occur in any season of the year (Anderson et al. 1991; Moen and Wouda 1995; Thurmond et al. 1995). Numerous studies using various serological methods have been conducted on cattle in different countries around the world. The prevalence of *N. caninum* has varied, with rates of 56.9% in Argentina Campero et al. (1998), 0.5% to 3.9% in the Czech Republic Václavek et al. (2007), Bártová et al. (2015); 12.5% in Wales and England Davison et al. (1999); 36.8% in Spain Quintanilla-Gozala et al. (1999), 15.6% in Poland Cabaj et al. (2000), and 10.7% in Sudan (Ibrahim et al. 2012). In Türkiye, a study by Eşki and Ütük covering N. caninum seroprevalence research up to 2018 reported an average prevalence of 13.06% (1023/7830) in cattle. Similarly, a study conducted by Demir et al. in 2020 reviewed all serological studies on cattle in Türkiye and reported an average seroprevalence of 14.7% (1672/11,373) for N. caninum.

Globally, PCR diagnostic studies for Neospora caninum in aborted cattle fetuses have been conducted. Sager et al. (2001) reported 21% positivity in 58 out of 242 samples in Sweden, while Sadrebazzaz et al. (2004) found N. caninum in 33% of 12 aborted fetuses in Iran. In Brazil, Cabral et al. (2009) detected N. caninum in 6.7% of 105 aborted fetuses, and Suteu et al. (2012) found 38.9% positivity in 21 aborted fetal samples. Macedo et al. (2017) detected N. caninum DNA in 38.8% of 14 tissue samples from 35 aborted fetuses. In Türkiye, PCR studies on aborted calf fetuses have reported varying results; Özkaraca et al. (2017) found N. caninum in 25.49% of 102 aborted fetuses in Elazığ, while Açıcı et al. (2019) reported a 49.4% positivity rate in 44 aborted fetuses from 89 farms using Real-time PCR. In Senel (2022) doctoral thesis, which investigated N. caninum in the Marmara Region, DNA from brain, heart, liver, lung, spleen, and kidney tissues of 84 aborted samples revealed N. caninum DNA in 26.19% of cases. Additionally, a study by Irehan et al. (2022) detected N. caninum in 8 out of 30 aborted fetuses using Real-time PCR, with only two of these also testing positive by conventional PCR. The lower positivity rate found in our study (2.2%) compared to the 49.4% reported by Acici et al. (2019) could be attributed to the lower detection rate of conventional PCR. Furthermore, the lower prevalence of *N. caninum* in the Central Black Sea Region compared to previous studies may be explained by the high level of integrated farming practices in the provinces where this study was conducted and the minimization of contact between cattle and uncontrolled dogs.

The type of cattle farming systems and management strategies are significant risk factors influencing the prevalence of N. caninum. Studies have shown that the seroprevalence of N. caninum is lower in integrated farms compared to rural family farms. It has been noted that cattle in rural family farms are more exposed to uncontrolled dogs, and those cattle that come into contact with these dogs carry a higher risk of infection (Öcal et al. 2014; Noori et al. 2019). The variability in results from PCR studies for *N. caninum* diagnosis conducted globally and in our country may stem from differences in regions, cattle breeds and rearing conditions, sample sizes, types and quantities of examined tissues, parasitic load, the presence of risk factors associated with N. caninum, and the different tests used. Therefore, the presence of definitive host dogs in areas where cattle are located, which can contaminate feed and water with their feces, is thought to play a significant role in the spread of infection (Dijkstra et al. 2002; Dubey et al. 2007).

In conclusion, understanding the relationship between intermediate and definitive hosts of Neospora caninum and implementing preventive measures is of great importance. Additionally, since calves born from N. caninum infections can transmit the infection from generation to generation, and due to the lack of effective treatment or vaccines, it is considered essential to conduct herdwide screenings and remove infected animals from the herd. Informing veterinarians and farmers about infections that cause abortion in cattle is crucial for combating these infections and, consequently, for the national economy. Furthermore, the use of double or triple test combinations in diagnosing N. *caninum* has been significantly evaluated for accurate diagnosis.

Ethical Statement: This study was conducted with permission from the Local Ethics Committee for Animal Experiments of the Samsun Veterinary Control Institute, under the letter dated 07.07.2022 with reference number 19572899/031-65 (Decision no: 2022/5).

Financial support and conflict of Interest: As the diagnosis and control of Neospora disease, which is the subject of this study, is among the official duties of Samsun Veterinary Control Institute, it was financed by the Republic of Türkiye Ministry of Agriculture and Forestry. The authors declare that they have no conflict of interest. *The article "Views and opinions expressed are however those of the author(s) onlyand do not necessarily reflect those of the Republic of Türkiye Ministry of Agriculture and

Forestry. Neither the the Republic of Türkiye Ministry of Agriculture and Forestry can be held responsible for". This text is included in accordance with the recommendation of our Ministry.

References

- Abdeltif B, Tennah S, Derdour SY, Temim A, Boufendi H, Ghalmi F, (2022). The first study on seroprevalence and risk factors of *Neospora caninum* infection in pregnant local cows from Northeast Algeria. *Vet World*. 15(2), 442-448. doi: 10.14202/ vetworld.2022.442-448.
- Açıcı M, Bölükbaş CS, Pekmezci GZ, Gürler H, Genç O, Gürler AT, Kaya S, Umur Ş, (2019). A diagnostic survey of *Neospora caninum* infection in aborted fetuses in the Middle Black Sea Region and Sivas Province. *Turkey. Turk J Vet Anim Sci.* 43(6), 761-766. doi: 10.3906/vet-1908-16.
- Aktaş M, Şaki CE, Altay K, Şimşek S, Ütük AE, Köroğlu E, Dumanlı N, (2005). Doğu Anadolu Bölgesinin Bazı İllerinde Bulunan Sığırlarda Neospora caninum'un Araştırılması. Türkiye Parazitol Derg. 29(1), 22-25.
- Anderson ML, Blanchard PC, Barr BC, Dubey JP, Hoffman RL, Conrad PA, (1991). *Neospora*-like protozoan infection as a major cause of abortion in California dairy cattle. *J Am Vet Med Assoc.* 198(2), 241-244.
- Anderson ML, Andrianarivo AG, Conrad, PA, (2000). Neosporosis in cattle. *Anim Reprod Sci*. 60-61(2), 417-431.doi: 10.1016/ S0378-4320(00)00117-2.
- Ayinmode A, Akinseye V, Schares G, Cadmus S, (2017). Serological survey of toxoplasmosis, neosporosis and brucellosis among cattle herds in Oyo state, SouthWestern Nigeria. *Afr J Infect Dis.* 11(2), 95–101. doi: 10.21010/ajid.v11i2.13.
- Barber JS, Holmdahl OJM, Owen MR, Guy F, Uggla A, Trees, AJ, (1995).Characterization of the first European isolate of *Neospora caninum* (Dubey, Carpenter, Speer, Topper and Uggla). *Parasitol.* 111(5), 563-568.
- Bártová E, Sedlak K, Budíková M, (2015). A study of *Neospora caninum* and *Toxoplasma gondii* antibody seroprevalence in healthy cattle in the Czech Republic. *Ann Agric Environ Med.* 22(1), 32-4. doi: 10.5604/12321966.1141365.
- Bıyıkoğlu G, Aksoy E, Bozkır M, Küçükayan U, Ertürk A, (2001). İç Anadolu Bölgesi sığırlarında *Neospora caninum*'un varlığının araştırılması. 12. Ulusal Parazitoloji Kongresi, Elazığ, Türkiye.
- Bulut G, Ün H, Sanioğlu G, Camkerten İ, (2021). The İncidence of *Neospora Caninum* in Dairy Cows with Abortion and İnfertility Problems in Aksaray Providence. *Kocatepe Vet J.* 14(4), 520-524. doi: 10.30607/kvj.1009265.
- Cabaj W, Choromanski L, Rodgers S, Moskwa B, Malczewski A, (2000). *Neospora caninum* infections in aborting dairy cows in Poland. *Acta Parasitol*. 45(2), 113-114.
- Cabral AD, Camargo CN, Galleti NTC, Okuda LH, Pituco EM, Del Fava C, (2009). Diagnosis of *Neospora caninum* in bovine fetuses by histology, immunohistochemistry, and nested-PCR. *Rev Bras Parasitol Vet*. 18(4), 14-19. doi: 10.4322/ rbpv.01804003.
- Campero CM, Anderson ML, Conosciuto G, Odriozola H, Bretschneider G, Poso MA, (1998). *Neospora caninum*-associated abortion in a dairy herd in Argentina. *Vet Rec.* 143(8), 228-229.
- Davison HC, Otter A, Trees AJ, (1999). Significance of *Neospora caninum* in British dairy cattle determined by estimation of seroprevalence in normally calving cattle and aborting cattle. *Int J Parasitol.* 29(8), 1189–1194. doi: 10.1016/S0020-7519(99)00094-6

- Davison HC, Guy CS, McGarry JW, Guy F, Williams DJL, Kelly DF, Trees AJ, (2001). Experimental studies on the transmission of *Neospora caninum* between cattle. *Res Vet Sci.* 70(2), 163-168. doi: 10.1053/rvsc.2001.0457.
- Demir AP, Ekşi F, Ütük AE, (2020). Estimating the total economic costs of *Neospora caninum* infections in dairy cows in Turkey. *Trop Anim Health Prod.* 52(6), 3251–3258. doi: 10.1007/ s11250-020-02351-1.
- Dijkstra TH, Barkema HW, Eysker M, Hesselink JW, Wouda W, (2002). Natural transmission routes of *Neospora caninum* between farm dogs and cattle. *Vet Parasitol.* 105(2), 99-104. doi: 10.1016/S0304-4017(02)00010-9.
- Dubey JP, (1999a). Neosporosis in cattle: biology and economic impact. J Am Vet Med Assoc. 214(89), 1160-1163. doi: 10.2460/javma.1999.214.08.1160.
- Dubey JP, (1999b). Recent advances in *Neospora* and neosporosis. *Vet Parasitol.* 84(3-4), 349–367. doi:10.1016/s0304-4017(99)00044-8
- Dubey JP, (2003). Review of *Neospora caninum* and neosporosis in animals. *Korean J Parasitol.* 41(1), 1-16. doi: 10.3347/kjp.2003.41.1.1.
- Dubey JP, Schares G, (2006). Diagnosis of bovine neosporosis. *Vet Parasitol.* 140(1-2), 1–34. 10.1016/j.vetpar.2006.03.035.
- Dubey JP, Schares G, Ortega-Mora LM, (2007). Epidemiology and control of neosporosis and *Neospora caninum*. *Clin Microbiol Rev.* 20(2), 323-367. doi: 10.1128/CMR.00031-06.
- Eşki F, Ütük AE, (2018). Detection of anti-*Neospora caninum* antibodies in cattle in Adana province of Turkey. *Van Vet J.* 29(2), 93-99.
- Goodswen SJ, Kennedy PJ, Ellis JT, (2013). A review of the infection, genetics, and evolution of *Neospora caninum*: from the past to the present. *Infect Genet Evol.* 13, 133-150. doi: 10.1016/j.meegid.2012.08.012.
- Guedes MHP, Guimarães AM, Rocha CMBM, Hirsch C, (2008). Frequência de anticorpos anti-*Neospora caninum* em vacas e fetos provenientes de municípios do sul de Minas Gerais. *Rev Bras Parasitol Vet.* 17(4), 189-194. doi:10.1590/S1984-29612008000400004.
- Innes EA, Bartley PM, Maley SW, Wright SE, Buxton D, (2007). Comparative hostparasite relationships in ovine toxoplasmosis and bovine neosporosis and strategies for vaccination. *Vaccine*. 5495-5503, 2007. doi: 10.1016/j.vaccine.2007.02.044.
- İbrahim AME, Elfahal AM, Hussein ARME, (2012). First report of Neospora caninum infection in cattle in Sudan. Trop Anim Health Prod. 44(4), 769-772. doi: 10.1007/s11250-011-9963-5.
- İrehan B, Sönmez A, Atalay MM, Ekinci Aİ, Çelik F, Durmuş N, Çiftci AT, Şimşek S, (2022). Investigation of *Toxoplasma gondii*, *Neospora caninum* and *Tritrichomonas foetus* in abortions of cattle, sheep and goats in Turkey: Analysis by real-time PCR, conventional PCR and histopathological methods. *Comp Immunol Microbiol Infect Dis.* 89, 101867. doi:10.1016/j. cimid.2022.101867.
- Kaltungo BY, Musa IW, (2013). A review of some protozoan parasites causing infertility in farm animals. *ISRN Trop Med*, 2013(1), 1-6. doi: 10.1155/2013/782609.
- Kamali A, Seifi HA, Movassaghi AR, Razmi GR, Naseri Z, (2014). Histopathological and molecular study of *Neospora caninum* infection in bovine aborted fetuses. *Asian Pac J Trop Biomed.* 4(12), 990-994. doi: 10.12980/APJTB.4.201414B378
- Kasap S, Ertunc S, Temizel EM, ŞŞentürk S, (2020). A study of *Neospora caninum* antibody seroprevalence in dairy cows in Turkey. J Hellenic Vet Med Soc. 71(1), 2018-2020. doi: 10.12681/jhvms.22950.

- Kashiwazaki Y, Pholpark S, Charoenchai A, Polsar C, Teeverapanya S, Pholpark M, (2001). Postnatal neosporosis in dairy cattle in northeast Thailand. *Vet Parasitol.* 94(3), 217-20. doi:10.1016/ s0304-4017(00)00358-7.
- Khaitsa ML, Barigye R, Dyer NW, Doetkott DM, Foster JR, (2006). Serologic and other diagnostic evidence of *Neospora caninum* presence in North Dakota beef herds. *The Bovine Practitioner*. 40(1), 51–56. doi: 10.21423/bovine-vol40no1p51-56.
- Köse O, Adanır R, Kocamüftüoğlu M, Çetin Y, (2021). Investigation of *Neospora caninum* Seroprevalence and Association with Reproductive Problems in Cows in Burdur Province of Turkey. *Iran J Parasitol.* 16(3), 386-393. doi:10.18502/ijpa.v16i3.7091.
- Kula D, Gökpınar S, (2021). Seroprevalence of Neospora caninum and Besnoitia besnoiti in Cattle in Oğuzlar Region. Türkiye Parazitol Derg. 45(2), 108-112. doi: 10.4274/tpd.galenos.2020.7075.
- Lally NC, Jenkins MC, Dubey JP, (1996). Evaluation of two *Neospora caninum* recombinant antigens for use in an ELISA for the diagnosis of bovine neosporosis. *Clin Diagn Lab Immunol.* 3(3), 275-279. doi: 10.1128/cdli.3.3.275-279.1996
- Lindsay DS, Dubey JP, (2020). Neosporosis, toxoplasmosis, and sarcocystosis in ruminants: an update. *Vet Clin Food Anim Pract. 36*(1), 205-222. doi: 10.1016/j.cvfa.2019.11.004
- Macedo CABD, Macedo MFSBD, Miura AC, Taroda A, Cardim ST, Innes EA, Katzer F, Cantón GJ, Chianini F, Headley SA, Garcia JL, (2017). Occurrence of abortions induced by *Neospora caninum* in dairy cattle from Santa Catarina, southern Brazil. *Rev Bras Parasitol Vet*. 26(3)-292-298. doi: 10.1590/S1984-29612017051
- McAllister MM, Dubey JP, Lindsay DS, Jolley WR, Wills RA, Mc-Guire AM, (1998). Rapid communication:Dogs are definitive hosts of *Neospora caninum*. *Int J Parasitol*. 28(9), 1473-1478. doi: 10.1016/S0020-7519(98)00138-6.
- McAllister MM, Björkman C, Anderson-Sprecher R, Rogers DG, (2000). Evidence of point-source exposure to *Neospora caninum* and protective immunity in a herd of beef cows. J Am Vet Med Assoc. 217(6), 881–887. doi:10.2460/javma.2000.217.881
- Moen AR, Wonda, W, (1995). Field experiences with bovine *Neospora* abortion in Dutch dairy herds. Proceedings. Symposium *Neospora* Abortus Bij Het Rund, Morra 2, Drachten, pp.11-17.
- Nasir A, Lanyon SR, Schares G, Anderson ML, Reichel MP, (2012). Sero-prevalence of *Neospora caninum* and *Besnoitia besnoiti* in South Australian beef and dairy cattle. *Vet Parasitol.* 186(3-4), 480-5. doi: 10.1016/j.vetpar.2011.11.032.
- Noori M, Rasekh M, Ganjali M, Nourollahi Fard SR, (2019). Seroprevalence of Neospora caninum Infection and Associated Risk Factors in Cattle of Sistan Areas, Southeastern Iran in 2016. *Iran J Parasitol.* 14(2), 340-346.
- Ortega-Mora L, Fernández-García A, Gómez-Bautista M, (2006). Diagnosis of bovine neosporosis: recent advances and perspectives. *Acta Parasitol.* 51(1), 1-14. doi: 10.2478/s11686-006-0001-0
- Öcal N, Atmaca HT, Albay MK, Deniz A, Kalender H, Yildiz K, Kul O, (2014). A new approach to *Neospora* caninum infection epidemiology: neosporosis in integrated and rural dairy farms in Turkey. *Turk J Vet Anim Sci.* 38(2), 161-168. doi: 10.3906/ vet-1307-11.
- Özkaraca M, Irehan B, Parmaksiz A, Ekinci AI, Çomaklı S, (2017). Determination of *Neospora caninum* and *Toxoplasma gondii* in aborted bovine foetuses by duplex PCR, immunohistochemistry and immunofluorescence methods. *Med Weter*. 73(6), 346-351. doi: 10.21521/mw.5707.
- Quintanilla-Gozalo A, Pereira-Bueno J, Tabare's E, Innes EA, Gonza'lez-Paniello R, Ortega-Mora LM, (1999). Seropreva-

lence of Neospora caninum infection in dairy and beef cattle in Spain. *Int J Parasitol.* 29(8), 1201–1208. doi: 10.1016/ s0020-7519(99)00084-3.

- Quintanilla-Gozalo A, Pereira-Bueno J, Seijas-Carballedo A, Costas E, Ortega-Mora, LM, (2000). Observational studies in *Neospora caninum* infected dairy cattle: relationship infection-abortion and gestational antibody fluctuations. *Int J Parasitol. 30*, 900-906.
- Pişkin Ç, Ütük AE, (2009). Prevalence of *Neospora caninum* in cows with stillbirth and abortion. *Etlik Vet Mikrobiyol Derg.* 20, 23-26.
- Ragozo, A.M.A., Paula, V.S.O., Souza, S.L.P., Bergamaschi, D.P., Gennari, S.M. (2003). Ocorrência de anticorpos anti-*Neospora caninum* em soros bovinos procedentes de seis estados brasileiros. *Rev Bras Parasitol Vet*. 12(1), 33-37.
- Reichel MP, Ayanegui-Alcérreca MA, Gondim LF, Ellis JT, (2013). What is the global economic impact of *Neospora caninum* in cattle–the billion dollar question. *Int J Parasitol.* 43(2), 133-142. doi: 10.1016/j.ijpara.2012.10.022.
- Sadrebazzaz A, Haddadzadeh H, Esmailnia K, Habibi G, Vojgani M, Hashemifesharaki R, (2004). Serological prevalence of *Neospora caninum* in healthy and aborted dairy cattle in Mashhad, Iran. *Vet Parasitol.* 124(3-4), 201-204. doi: 10.1016/j. vetpar.2004.06.027.
- Sager H, Fischer I, Furrer K, Strasser M, Waldvogel A, Boerlin P, Audigé L, Gottstein B, (2001). A Swiss case–control study to assess *Neospora caninum*-associated bovine abortions by PCR, histopathology and serology. *Vet Parasitol.* 102(1-2), 1-15. doi: 10.1016/S0304-4017(01)00524-6.
- Salehi N, Haddadzadeh H, Ashrafihelan J, Shayan P, Sadrebazzaz A, (2009). Molecular and pathological study of bovine aborted fetuses and placenta from Neospora caninum infected dairy cattle. *Iranian J Parasitol.* 4(3), 40-51.
- Schares G, Bärwald A, Staubach C, Söndgen P, Rauser M, Schröder R, Peters M, Wurm R, Selhors T, Conraths, FJ, (2002). p38-avidity-ELISA: examination of herds experiencing epidemic or endemic *Neospora caninum*-associated bovine abortion. *Vet Parasitol.* 106(4), 293-305. doi: 10.1016/S0304-4017(02)00103-6.
- Şenel M, (2022). Marmara Bölgesindeki ruminant atık fötuslarında *Toxoplasma gondii* ve *Neospora caninum*'un moleküler, patolojik yöntemlerle araştırılması ve etkenlerin moleküler karakterizasyonu. Doktora Tezi, Kafkas Üniversitesi Sağlık Bilimleri Enstitüsü, Kars.
- Şentürk S, Temizel EM, Kasap S, (2020). Bir Buzağıda Klinik Kongenital Neosporozis. *Türkiye Parazitol Derg.* 44(2), 109-11. doi: 10.4274/tpd.galenos.2020.6666.
- Şuteu O, Paştiu A, Györke A, Cozma V, (2012). Molecular detection of *Neospora caninum* abortion in dairy cattle from different historical regions of Romania. *Sci Parasitol*. 13(4), 159-62.
- Thurmond MC, Anderson ML, Blanchard PC, (1995). Secular and seasonal trends of Neospora abortion in California dairy cows. J Parasitol. 81(3), 364-367.
- Trees AJ, Davison HC, Innes EA, Wastling JM, (1999). Towards evaluating the economic impact of bovine neosporosis. Int J Parasitol. 29(8), 1195-1200. doi: 10.1016/S0020-7519(99)00093-4.
- Toolan DP, (2003). *Neospora caninum* abortion in cattle- a clinical perspective. *Irish Vet J.* 56(8), 404–410.
- Václavek P, Sedlák K, Hůrková L, Vodrážka P, Šebesta R, Koudela B, (2007). Serological survey of *Neospora caninum* in dogs in the Czech Republic and a long-term study of dynamics of antibodies. *Vet. Parasitol.* 143(1), 35-41. doi: 10.1016/j.vetpar.2006.07.020