

The Reliability of Citizen Science Data: Testing A Data Set from eBird Türkiye with Benford's Law

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Abstract

Citizen science is a rapidly evolving and increasingly popular field of scientific inquiry in which data are collected by volunteers (citizens). Applications of citizen science represent a growing data resource used by researchers, nature conservationists, and public institutions. Although citizen science data can make substantial contributions within the scope of Conservation Biology—particularly to the protection and management of biological diversity, ecosystems, and natural resources—concerns regarding data quality persist, despite the expanding potential of citizen science. Ensuring that data collected through citizen science efforts are accurate and reliable is crucial for conducting valid assessments in Conservation Biology and for enabling more effective conservation practices. In this study, we sought to test the accuracy of bird observation data gathered via the citizen science platform eBird Türkiye (<http://www.ebird.org>)—which contains observation counts for 533 bird species recorded in Türkiye—by applying Benford's Law. Benford's Law is a mathematical approach developed to analyze the distribution of numbers in data sets that may not have a strictly “natural” origin. Additionally, a Chi-Square Goodness-of-Fit Test was conducted to detect differences between the observed (actual) values and the expected (Benford's Law) values. The results indicate that the tested data from the eBird Türkiye platform conform to Benford's Law.

Keywords: Benford Law, Conservation Biology, eBird, Citizen Science, Biodiversity

1. Introduction

The rapid pace of scientific advancement has led to increasingly specialized fields of expertise. Although specialization supports scientific progress, it also fosters a structure in which only experts produce knowledge, making it challenging for the broader public to grasp new developments. One response to this issue—particularly prominent in Europe—is the institutionalization and encouragement of public participation in scientific research under the banner of “Citizen Science,” which aligns with open science principles. In addition to reforming scientific methodologies, this approach seeks to enhance public engagement in scientific processes [1].

Citizen science is a method that promotes participation by non-professional individuals in scientific research. By involving volunteers, it reduces research costs, facilitates

faster project completion, strengthens communication between science and society, and increases scientific literacy. It also encourages a more collaborative form of research by contributing to participants' self-realization processes [1]. Owing to its potential for advancing scientific literacy and involving diverse segments of society in research, citizen science has grown increasingly popular in recent years [2].

Citizen science applications have emerged as a rapidly expanding data resource, widely used by researchers, nature conservationists, and public institutions [3, 4, 5, 6, 7]. The data collected by volunteers are vital, given the increased taxonomic, geographical, and temporal scope [8].

However, one of the greatest challenges in citizen science lies in ensuring data quality and the accuracy of

volunteer-collected information [2]. Despite the growing potential of citizen science, concerns regarding data reliability remain [9]. To use these data effectively in research, conservation, and policy-making, their reliability must be carefully evaluated. Data quality issues often reflect spatial, temporal, and taxonomic biases [10] or insufficient sampling [11, 12, 13]. Consequently, developing and applying methods to enhance the reliability of citizen science data is essential.

A recent Web of Science categorization of publications using the keyword “Citizen Science” reveals that Environmental Sciences has the highest number of related publications [1] as shown in Figure 1.

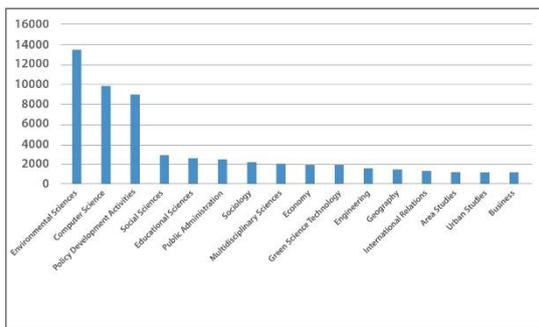


Figure 1. Citizen Science Publications by Scientific Field [1] (Reproduced from the original source).

To understand why citizen science is so significant to environmental sciences and the advantages it offers, one must first examine the concept of “Conservation Biology.” Conservation Biology is an interdisciplinary field focused on the protection and sustainable management of biological diversity, ecosystems, and natural resources. It integrates ecological, evolutionary, and socio-economic factors through various approaches to address complex issues involving the preservation of natural systems. One of its core aims is to prevent the loss of biological diversity by protecting and restoring threatened or endangered species and habitats. Simultaneously, Conservation Biology deals with the root causes of biodiversity decline, such as habitat destruction, pollution, climate change, and overexploitation of resources. This endeavor requires collaboration among scientists from different disciplines, policy makers, and other stakeholders. Conservation biologists investigate how human activities affect natural systems in order to develop effective, sustainable strategies for resource management [14].

In this context, citizen science offers critical support to conservation biology. The large-scale data collected by volunteers serve as an essential resource for monitoring biodiversity, recognizing threats, and shaping conservation strategies. Beyond aiding scientific research, citizen science also boosts public engagement in nature conservation by inviting active community

participation, thereby increasing environmental awareness and promoting scientific literacy. Consequently, citizen science not only democratizes the production of scientific knowledge but also broadens the social base for nature conservation initiatives, generating benefits for both science and society.

Numerous statistical methods have been used to evaluate the accuracy and reliability of citizen science data. In this study, we applied Benford’s Law, a mathematical technique designed to analyze the distribution of numbers in data sets potentially lacking a natural origin [15, 16]. This method tests whether a given data set follows the leading digit distribution predicted by Benford’s Law. Because Benford’s Law is a powerful tool for assessing the trustworthiness of numerical data and detecting inconsistencies, it has been applied in multiple fields, including finance and the natural sciences.

Benford’s Law has proven effective in several sectors for checking the reliability of reported natural resource harvesting data (for instance, in fishing or tropical hunting; 17) and for detecting illegal deforestation activities [18]. This approach is also employed to study naturally occurring biological data sets: prior research has demonstrated that certain biological data—such as cyanobacterial cell counts [19] pollen counts [20], genome sizes [21] and angiosperm taxon counts [22]—can align with Benford’s Law. Overall, Benford’s Law and similar digit-based tests have been recommended as simple yet effective preliminary screening methods for large, complex ecological data sets [20, 23, 24]. Given the increasing complexity and volume of ecological data—including citizen science data—Benford’s Law has significant potential for identifying anomalies or errors in naturally occurring data sets, thus providing critical insights for ecological and environmental research [25]. In particular, this method is valuable for evaluating the reliability of large-scale data sets and for deepening our understanding of ecological processes.

Besides Benford’s Law, alternative methods for assessing the accuracy of citizen science data include “Inter-Observer Agreement,” “Expert Validation,” and “Outlier Detection.” Inter-Observer Agreement compares data collected by various volunteers to measure consistency, often using statistical measures such as Cohen’s Kappa or Fleiss’s Kappa [26]. Expert Validation involves detailed reviews by specialists, who compare the volunteer-collected data with those gathered through conventional scientific techniques [26]. Meanwhile, Outlier Detection identifies data points that differ substantially from the rest of the dataset, helping pinpoint—and potentially eliminate—errors [26].

When compared with other European and Middle Eastern countries, Türkiye’s remarkable biological diversity in relation to its size positions it as one of the most

biodiverse nations in the region. Thus, Türkiye has a major need for citizen science initiatives aimed at identifying and conserving this biodiversity. Nonetheless, citizen science remains only partly developed here. Platforms such as Trakuş, AdaMerOs, and eBird Türkiye represent major strides in this domain by harnessing volunteer contributions to gather and monitor biodiversity information, highlighting the potential impact of citizen science in Türkiye. Given the country's notable biodiversity, expanding and developing citizen science activities could significantly bolster its conservation efforts.

By engaging volunteers in data collection and analysis, citizen science helps advance extensive research efforts and fosters scientific literacy. However, these data must be validated for reliable use in research, conservation, and policy-making. Although the number of global studies evaluating the reliability of citizen science data is on the rise, few such studies have been conducted in Türkiye. One validation tool is Benford's Law. In this study, we applied Benford's Law alongside a Chi-Square Goodness-of-Fit Test to a dataset containing observation counts for 533 bird species reported in Türkiye, as cataloged on the "eBird Türkiye" [27] platform. Verifying data in this manner is crucial to ensuring accuracy and reliability, which in turn enables better decision-making for biodiversity protection in the framework of Conservation Biology.

2. Materials and Methods

2.1. Study Material

We employed a data set from the citizen science platform eBird Türkiye [27], containing observation counts for 533 bird species recorded in Türkiye. Established in 2002 by the Cornell Lab of Ornithology in the United States, eBird has become the largest citizen science platform focused on bird sightings worldwide. With more than 600 million bird observations, it maintains one of the most substantial biodiversity databases, creating extensive opportunities for large-scale research on migration routes, population changes, and the effects of environmental shifts [5].

2.2. Benford Law

Benford's Law was used to assess this data set's reliability. According to the first-digit test, Benford's Law states that the leading non-zero digit of a number—or a set of values representing a given phenomenon—is not uniformly distributed. A high percentage of numbers begin with 1, followed by 2, 3, and so forth. Specifically, 30.1% of data points start with 1, 17.6% with 2, 12.5% with 3, 9.7% with 4, 6.7% with 5, 6.4% with 6, 5.1% with 7, 4.9% with 8, and 4.7% with 9, indicating that smaller leading digits arise more frequently than larger ones [15, 16]. For the first-digit test, Benford's Law is applied using the formula:

$$P(d) = \log_{10} (1 + 1/d) \quad (2.1)$$

Here it is:

P(d): The probability that the first digit of a number is the digit "d".

d: The digit representing the first digit of the number. This digit can be an integer from 1 to 9 (i.e. $d \in \{1, 2, \dots, 9\}$).

\log_{10} : Denotes the logarithm to base 10.

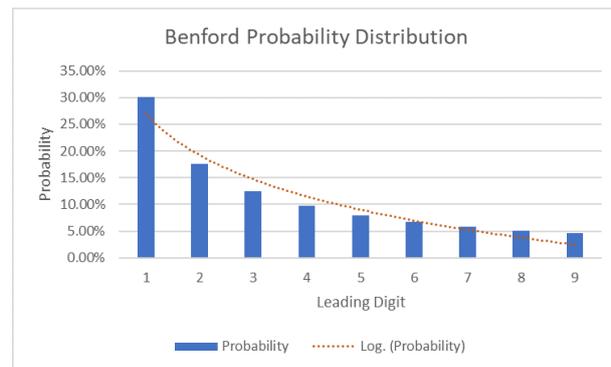


Figure 2. Probability distributions of the first digits according to Benford's Law

Note: Prepared from [28]

2.3. Data Analysis

We completed the following steps in Microsoft Excel to verify the data's conformity with Benford's Law:

2.3.1. Forming the Data Set from the Database: A .txt file was exported from the eBird database (see Appendix 1). This file was imported into MS Excel, after which bird species and their observation counts were filtered to create the final data set.

2.3.2. Extracting the First Digit: The first digit of each observation count in the data set was identified.

2.3.3. Calculating Frequencies: The observed frequency of each leading digit was recorded.

2.3.4. Calculating the Percentage Frequency: The observed frequency was divided by the total data count to obtain the percentage frequency.

2.3.5. Computing Benford Percentages: The " $=\text{LOG}_{10}(\text{number})$ " formula was used to determine each digit's expected probability under Benford's Law.

2.3.6. Calculating Expected Frequencies According to Benford's Law: Expected frequencies were derived from the total number of observations in conjunction with each digit's Benford percentage.

2.3.7. Conducting the Chi-Square Goodness-of-Fit

Test: This test identifies whether the observed variable matches a given distribution. The formula used was:

$$x^2 = \frac{(\text{Observed} - \text{Expected})^2}{\text{Expected}} \quad (2.2)$$

2.3.8. Calculating of the p-Value The sum of the values found in the Chi-Square Test of Likelihood was found. Then the p value was found as “0.19865” by using the formula “=CHI-SQUARED.DISTR.RIGHT (x; degrees of freedom)”. The degrees of freedom used in the formula is one less than the 9 variables used, i.e. the degrees of freedom is 8. The calculation was made as follows:

$$= \text{CHI-SQUARED.DISTR.RIGHT}(11,05;8)$$

*If p-value < 0.05, the data do not follow Benford's Law.

*If the p-value is ≥ 0.05 , the data obey Benford's Law.

According to these conditions, since the p-value is $0.19868 > 0.05$, the data obey Benford's Law.

3. Results and Discussion

In this study, the conformity of e-bird observation data in Turkey with Benford's Law was examined. The results of the analysis showed that the first digits of the observation counts for each species were distributed in a manner similar to the frequency distribution predicted by Benford's Law.

3.1. Key Findings

3.1.1. Overall Fit: A strong relationship was found between the frequency percentages of the first digits of the observation numbers (from 1 to 9) and the percentages predicted by Benford's Law. In particular, it was found that small digits such as 1, 2, and 3 appeared with a higher frequency, as predicted by Benford's Law.

Table 1 shows that the most common digit value is the number 1 (%32,8) and the least common digit value is the number 9 (%2,8) This is a situation seen in the first digit test in Benford's Law.

3.1.2. Statistical Tests: Statistical tests (e.g., chi-square test) have shown that the difference between observed and expected frequencies is not statistically significant. This confirms that the data set is generally consistent with Benford's Law.

3.1.3. Differences: Small deviations from Benford's Law predictions have been observed for some numbers. However, it has been determined that these deviations are

not significant enough to distort the overall structure of the data set.

The table shows that the absolute difference between the Benford Percentage and the percentage occurrence of the digits (frequency percentage) varies between %2,7 – 0,1. This is another indication that the phenomenon complies with Benford's Law. Looking at the graph in Figure 3, the biggest difference between the Frequency Percentage and the Benford Percentage (2,7%) is seen in the number 1. The smallest difference between the Frequency Percentage and the Benford Percentage (0,1 %) is seen in the number 8.

In short, the Frequency Percentage shows how often the first households in your dataset actually occur, while the Benford Percentage expresses how often these households should appear in a natural distribution. The difference between these two percentage indicates how well your data set fits Benford's Law and whether there are potentially any anomalies. The small numbers seen in the difference show that our data set is compliance with Benford's Law.

Figure 3 presents a bar chart of the observed frequency percentage (actual data) and the expected percentage under Benford's Law (predicted data); the close alignment between these bars illustrates consistency with Benford's Law.

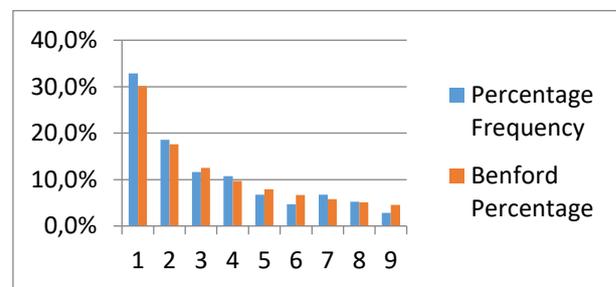


Figure 3. Conformity of Observation Counts for 533 Bird Species from eBird Türkiye to Benford's Law.

Table 1 summarizes the application of Benford's Law and the Chi-Square Goodness-of-Fit Test to the data set. As shown in Table 1, the Chi-Square Goodness-of-Fit Test produced a x^2 value of 11,05, with a p-value of 0.19865. Because this p-value is greater than 0.05, the data fit Benford's Law.

The findings show that e-bird observation data exhibit a distribution consistent with Benford's Law, which is characteristic of naturally occurring data sets. This result provides strong evidence that the data set has not been manipulated and was created as a result of a natural process.

4. Conclusion and Discussion

The reason why Benford's Law is used in the naturalness calculation of ecosystems is that its message is mathematical, that is, it is clear.

The Benford probability curve in reality means that a whole army of numbers contains “process information”. This “process information” does not explain what the numbers that go into the probability calculation are and what formulas are used to derive them. Here “process information” simply means numbers with a background.

This message of Benford's Law is as clear as its theoretical probability values [24].

What is meant here by numbers with a resume?

These numbers mean the system in other words. If the numbers that make up a whole conform to Benford's Law, there is a natural consciousness that allows that whole to exist and to continue to exist. In other words, if a whole has a natural consciousness, it is expected to obey Benford's Law. In this case, the term “system” is used instead of “whole”.

Table 1. Summary of Steps and Results in MS Excel

| Number | Frequency | Percentage Frequency | Benford Percentage | Expected Benford Frequency | Observed - Expected | (Observed - Expected) ² | Chi-Square Goodness-of-Fit Test |
|---------------|---------------|----------------------|--------------------|----------------------------|---------------------|------------------------------------|---------------------------------|
| 1 | 175 | 32,8% | 30,1% | 160,45 | 14,55 | 211,73 | 1,32 |
| 2 | 99 | 18,6% | 17,6% | 93,86 | 5,14 | 26,45 | 0,28 |
| 3 | 62 | 11,6% | 12,5% | 66,59 | -4,59 | 21,09 | 0,32 |
| 4 | 57 | 10,7% | 9,7% | 51,65 | 5,35 | 28,59 | 0,55 |
| 5 | 36 | 6,8% | 7,9% | 42,20 | -6,20 | 38,48 | 0,91 |
| 6 | 25 | 4,7% | 6,7% | 35,68 | -10,68 | 114,12 | 3,20 |
| 7 | 36 | 6,8% | 5,8% | 30,91 | 5,09 | 25,91 | 0,84 |
| 8 | 28 | 5,3% | 5,1% | 27,26 | 0,74 | 0,54 | 0,02 |
| 9 | 15 | 2,8% | 4,6% | 24,39 | -9,39 | 88,15 | 3,61 |
| Toplam | 533,00 | 1,00 | 1,00 | 533,00 | | | 11,05 |

This study examined observational data for 533 bird species from the “eBird Türkiye” citizen science platform using Benford’s Law.

According to the Chi-Square Goodness-of-Fit Test, the data conform to Benford’s Law, suggesting that the dataset has not been manipulated and can serve as a credible source for scientific research. However, while Benford’s Law offers a meaningful overview of data consistency, it does not provide an absolute guarantee of data accuracy [16].

Given the ongoing concerns about data quality in citizen science applications, this finding is quite important. Although errors, omissions, or systematic biases may be

present in data provided by volunteers, the results obtained in this study support the statistical reliability of such data. Similarly, various studies [26,29,30] have demonstrated that volunteer science data can be used reliably when evaluated using careful assessment methods.

The Benford's Law used in the study is a powerful and practical tool for analyzing natural data distributions, especially in data sets containing a large number of observations. This method has been successfully applied in the analysis of biological data (e.g., cell counts, taxonomic distributions, genome sizes) in the past. In this context, applying the Benford test to eBird Turkey data provides an important methodological contribution to

evaluating the quality of volunteer science data in our country.

However, some minor deviations were also observed during the analysis. In particular, differences were found between the observed and expected frequencies in some steps (e.g., numbers 1 and 9). Although these differences are not statistically significant, investigating the causes of such deviations in future studies may contribute to the development of improvement strategies to enhance data quality. For example, systematic biases such as the over-reporting of certain species or the re-entry of data by some observers could be responsible for these differences.

Another important contribution of the study is that a data verification method of this scale has been applied in Turkey. Although citizen science studies are becoming widespread around the world, studies testing the reliability of such data in Turkey are quite limited. This situation indicates that the study not only provides results regarding the reliability of eBird data but also has the potential to increase the scientific validity of citizen science applications in Turkey.

However, there are some limitations to this study. First, the analysis was conducted solely on the basis of the first-order distribution. For a more comprehensive evaluation of the data, second-order analysis, time series analysis, or spatial validation methods could also be used. Furthermore, it should be noted that Benford's Law alone is not sufficient for measuring the reliability of data, but it does provide an effective method for preliminary screening and general evaluation.

The number of digits of large ecological data such as the number of cells in cyanobacteria colonies [19] and pollen counts [20] are quite high. For this reason, while the second and third step test applications of Benford's Law are tested in data sets with large digit numbers of this type, it is predicted that the first step test will be sufficient considering the digit numbers of the number values in the data set in the study and the literature studies. The reason for applying the second and the third step test in Benford's Law is to detect errors and anomalies more precisely.

Our findings emphasize the need for more rigorous data validation measures to further enhance the accuracy and reliability of citizen science data. While Benford's Law serves as an effective preliminary screening tool, additional validation methods – such as expert verification, inter – observer agreement and outlier

detection – may be employed to improve data quality [25,29].

In essence, citizen science democratizes scientific knowledge production while simultaneously supporting biodiversity monitoring and conservation activities. Expanding citizen science initiatives in Türkiye could strengthen the country's research base and broaden conservation participation among the public.

Citizen science—which engages members of the public in data collection and analysis—contributes significantly to large-scale research efforts and fosters scientific literacy. Data reliability is especially critical in fields such as Conservation Biology, where accurate information can directly inform conservation measures. As one of the few studies evaluating the trustworthiness of citizen science data in Türkiye, this paper sheds light.

Benford's Law is only an indicator. A deviation from Benford's Law is a warning sign that there may be anomalies or manipulation in a data set. However, this does not necessarily mean that the data is incorrect. It simply indicates that further examination and in-depth research is required. In short, while Benford's Law is a powerful audit and analysis tool, it has important limitations that must be carefully considered in terms of its application areas and the characteristics of the data set. If these limitations are ignored, misleading results may be obtained.

In conclusion, this study proposes an applicable and valid method for evaluating the quality of data obtained from volunteer science platforms in Turkey, while also supporting the usability of this data in scientific studies. Future studies can conduct more in-depth evaluations by analyzing data sets separated according to different species groups, observers, or time periods.

5. Recommendations

Strengthening Citizen Science in Türkiye: The continued development of citizen science projects, especially in natural and environmental sciences, could greatly enhance Türkiye's research capacities.

Validating Data in Other Citizen Science Platforms: Similar assessments should be undertaken on other platforms across Türkiye, since the data generated often serve as a primary resource for biodiversity monitoring.

Integrating Citizen Science into Education: Introducing citizen science projects in the educational system can promote scientific thinking among young

people and heighten overall awareness of environmental issues.

Unlocking the Potential of Citizen Science: Citizen science approaches can address various local challenges in Türkiye, such as water quality monitoring, urban land-use mapping, and seismic event tracking. Establishing a dynamic citizen science community may offer both societal and scientific advantages, paving the way for a more robust future.

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Author's Contributions

Çetin ÇELİK: Draft and wrote the manuscript, analyzed the data and generated the result with the data.

Murat AFSAR: Draft and wrote the manuscript.

Ethics

There are no ethical issues after the publication of this manuscript.

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Appendices

Appendix 1: Data Set Used in the Study

| Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı | Tür Adı | Gözlem Sayısı |
|---|---------------|------------------------------------|---------------|----------------------------------|---------------|--|---------------|-----------------------------------|---------------|---|---------------|---|---------------|
| <i>Acanthis flammea</i> | 1 | Anatidae sp. (dabbling duck sp.) | 214 | <i>Asio otus</i> | 553 | <i>Cercotrichas galactotes</i> | 543 | <i>Columba sp.</i> | 22 | <i>Emberiza caesia</i> | 718 | <i>Gallinago media</i> | 29 |
| <i>Accipiter nisus</i> | 8050 | <i>Anatidae sp. (teal sp.)</i> | 16 | <i>Astur gentilis</i> | 236 | <i>Certhia brachyactyla</i> | 1231 | <i>Coracias garrulus</i> | 1893 | <i>Emberiza calandra</i> | 19324 | <i>Gallinago sp.</i> | 1 |
| <i>Accipitridae sp. (eagle sp.)</i> | 9 | <i>Anser albifrons</i> | 2688 | <i>Athene noctua</i> | 1767 | <i>Certhia familiaris</i> | 145 | <i>Corvus corax</i> | 12480 | <i>Emberiza cia</i> | 1070 | <i>Gallinula chloropus</i> | 11805 |
| <i>Accipitridae sp. (hawk sp.)</i> | 1125 | <i>Anser anser</i> | 3631 | <i>Aythya ferina</i> | 32751 | <i>Aythya ferina</i> | 16 | <i>Corvus cornix</i> | 73733 | <i>Emberiza cineracea</i> | 62 | <i>Garrulus glandarius</i> | 20648 |
| <i>Accipitriformes/Falconiformes sp.</i> | 24 | <i>Anser cygnoides</i> | 1 | <i>Aythya fuligula</i> | 10089 | <i>Ceryle rudis</i> | 1161 | <i>Corvus frugilegus</i> | 24791 | <i>Emberiza cirius</i> | 4166 | <i>Gavia arctica</i> | 1448 |
| <i>Acridotheres tristis</i> | 2118 | <i>Anser erythropus</i> | 635 | <i>Aythya marila</i> | 6 | <i>Cettia cetti</i> | 7196 | <i>Corvus sp.</i> | 12 | <i>Emberiza citrinella</i> | 3878 | <i>Gavia stellata</i> | 15 |
| <i>Acrocephalus agricola</i> | 129 | <i>Anser serrirostris</i> | 11 | <i>Aythya nyroca</i> | 2485 | <i>Charadriidae sp.</i> | 16 | <i>Corvus sp. (crow sp.)</i> | 28 | <i>Emberiza citrinella x leucocephalus</i> | 4 | <i>Gelochelidon nilotica</i> | 2341 |
| <i>Acrocephalus arundinaceus</i> | 2351 | <i>Anser sp. (Domestic type)</i> | 16 | <i>Aythya sp.</i> | 400 | <i>Charadriiformes sp. (shorebird sp.)</i> | 45 | <i>Coturnix coturnix</i> | 389 | <i>Emberiza hortulana</i> | 1856 | <i>Geronticus eremita</i> | 1119 |
| <i>Acrocephalus melanopogon</i> | 604 | <i>Anthropoides virgo</i> | 145 | <i>Botaurus minutus</i> | 581 | <i>Charadrius hiaticula</i> | 5089 | <i>Crex crex</i> | 16 | <i>Emberiza leucocephalus</i> | 4 | <i>Glareola nordmanni</i> | 2300 |
| <i>Acrocephalus palustris</i> | 713 | <i>Anthus campestris</i> | 1707 | <i>Botaurus stellaris</i> | 187 | <i>Charadrius thinornis/Anarhynchus sp.</i> | 2 | <i>Cuculus canorus</i> | 1003 | <i>Emberiza melanocephala</i> | 1867 | <i>Glareola pratincola</i> | 3151 |
| <i>Acrocephalus palustris-scirpaceus</i> | 46 | <i>Anthus cervinus</i> | 1699 | <i>Branta leucopsis</i> | 43 | <i>Chlidonias hybrida</i> | 4825 | <i>Curruca cantillans</i> | 334 | <i>Emberiza pusilla</i> | 5 | <i>Glareola pratincola/nordmanni</i> | 2 |
| <i>Acrocephalus schoenobaenus</i> | 641 | <i>Anthus japonicus</i> | 72 | <i>Bubo bubo</i> | 37 | <i>Chlidonias leucopterus</i> | 6220 | <i>Curruca communis</i> | 3349 | <i>Emberiza rustica</i> | 12 | <i>Grus grus</i> | 11333 |
| <i>Acrocephalus scirpaceus</i> | 3468 | <i>Anthus pratensis</i> | 11995 | <i>Bucanetes githagineus</i> | 15 | <i>Chlidonias niger</i> | 563 | <i>Curruca conspiciata</i> | 16 | <i>Emberiza schoeniclus</i> | 4076 | <i>Gulosus aristotelis</i> | 6977 |
| <i>Acrocephalus scirpaceus/arundinaceus</i> | 2 | <i>Anthus pratensis/spinoletta</i> | 2 | <i>Bucanetes mongolicus</i> | 14 | <i>Chlidonias sp.</i> | 4 | <i>Curruca crassirostris</i> | 393 | <i>Emberiza sp.</i> | 87 | <i>Gymnoris xanthocolis</i> | 86 |
| <i>Acrocephalus sp.</i> | 222 | <i>Anthus richardi</i> | 22 | <i>Bucephala clangula</i> | 55 | <i>Chloris chloris</i> | 15515 | <i>Curruca curruca</i> | 4179 | <i>Eremophila alpestris</i> | 1176 | <i>Gypaetus barbatus</i> | 204 |
| <i>Actitis hypoleucos</i> | 4374 | <i>Anthus sp.</i> | 500 | <i>Burhinus oedionemus</i> | 915 | <i>Chroicocephalus geni</i> | 7384 | <i>Curruca melanocephala</i> | 4234 | <i>Eritacus rubecula</i> | 20427 | <i>Gyps fulvus</i> | 850 |
| <i>Aegithalos caudatus</i> | 9154 | <i>Anthus spinoletta</i> | 4171 | <i>Buteo buteo</i> | 91910 | <i>Chroicocephalus genes/ridibundus</i> | 115 | <i>Curruca mystacea</i> | 364 | <i>Eudromias morinellus</i> | 163 | <i>Haematopus ostralegus</i> | 1159 |
| <i>Aegolius funereus</i> | 9 | <i>Anthus trivialis</i> | 2187 | <i>Buteo lagopus</i> | 7 | <i>Chroicocephalus ridibundus</i> | 185807 | <i>Curruca nisoria</i> | 872 | <i>Falco columbarius</i> | 289 | <i>Halcyon sylvnensis</i> | 598 |
| <i>Aegypius monachus</i> | 1309 | <i>Apodidae sp.</i> | 4 | <i>Buteo rufinus</i> | 3989 | <i>Chroicocephalus ridibundus/Leithyaetus melanocephalus</i> | 71 | <i>Curruca nippeli</i> | 497 | <i>Falco eleonorae</i> | 595 | <i>Halaeletus albicilla</i> | 294 |
| <i>Aerospiza Tachyspiza/Accipiter/Astur sp.</i> | 118 | <i>Apus affinis</i> | 225 | <i>Buteo sp.</i> | 64 | <i>Ciconia ciconia</i> | 560426 | <i>Corsorius cursor</i> | 53 | <i>Falco naumanni</i> | 2225 | <i>Hieraaetus pennatus</i> | 2766 |
| <i>Alaemon alaudipes</i> | 16 | <i>Apus apus</i> | 31512 | <i>Casina moschata</i> | 17 | <i>Ciconia nigra</i> | 15269 | <i>Cyanistes caeruleus</i> | 12328 | <i>Falco naumanni/innunculus</i> | 63 | <i>Himantopus himantopus</i> | 17257 |
| <i>Alauda arvensis</i> | 12120 | <i>Apus apus pallidus</i> | 868 | <i>Calandrella brachydactyla</i> | 3033 | <i>Ciconia sp.</i> | 11 | <i>Cygnus atratus</i> | 1 | <i>Falco peregrinus</i> | 784 | <i>Hippoboscus icterina</i> | 42 |
| <i>Alauda heinei</i> | 382 | <i>Apus pallidus</i> | 3914 | <i>Calcarus lapponicus</i> | 52 | <i>Cinclus cinclus</i> | 265 | <i>Cygnus columbianus</i> | 14354 | <i>Falco sp.</i> | 114 | <i>Hippoboscus languida</i> | 222 |
| <i>Alauda rufescens</i> | 391 | <i>Apus Tachymarptis sp.</i> | 295 | <i>Calidris alba</i> | 918 | <i>Circus albus</i> | 17565 | <i>Cygnus columbianus/cygnus</i> | 28 | <i>Falco sp. (small falcon sp.)</i> | 5 | <i>Hippoboscus olivetorum</i> | 109 |
| <i>Alauda rufescens/heinei</i> | 1 | <i>Aquila chrysaetos</i> | 460 | <i>Calidris alpina</i> | 24860 | <i>Circus aeruginosus</i> | 10207 | <i>Cygnus cygnus</i> | 680 | <i>Falco subbuteo</i> | 1811 | <i>Hippoboscus sp.</i> | 15 |
| <i>Alaudidae sp.</i> | 63 | <i>Aquila fasciata</i> | 19 | <i>Calidris canutus</i> | 522 | <i>Circus cyaneus</i> | 614 | <i>Cygnus olor</i> | 3060 | <i>Falco tinnunculus</i> | 6199 | <i>Hirundinidae sp.</i> | 707 |
| <i>Alcedo atthis</i> | 3764 | <i>Aquila heliaca</i> | 484 | <i>Calidris falcinellus</i> | 220 | <i>Circus macrourus</i> | 211 | <i>Cygnus sp.</i> | 10 | <i>Falco vespertinus</i> | 970 | <i>Hirundo rustica</i> | 121192 |
| <i>Alectoris chukar</i> | 2358 | <i>Aquila nipalensis</i> | 82 | <i>Calidris ferruginea</i> | 2611 | <i>Circus macrourus/pygargus</i> | 19 | <i>Delichon urbicum</i> | 22147 | <i>Ficedula albicollis</i> | 1307 | <i>Hirundo rustica x Delichon urbicum</i> | 2 |
| <i>Alopochen aegyptiaca</i> | 4 | <i>Aquila sp.</i> | 20 | <i>Calidris minuta</i> | 15342 | <i>Circus pygargus</i> | 721 | <i>Dendrocopos leucotos</i> | 179 | <i>Ficedula hypoleuca</i> | 455 | <i>Hydrobatas pelagicus</i> | 22 |
| <i>Amandava amandava</i> | 1 | <i>Ardea alba</i> | 11820 | <i>Calidris pugnax</i> | 30337 | <i>Circus sp.</i> | 55 | <i>Dendrocopos major</i> | 651 | <i>Ficedula hypoleuca/albicollis/semitorquata</i> | 18 | <i>Hydrocoleus minutus</i> | 2486 |
| <i>Ammonomanes deserti</i> | 14 | <i>Ardea cinerea</i> | 35126 | <i>Calidris sp.</i> | 228 | <i>Cisticola juncidis</i> | 378 | <i>Dendrocopos major/syriacus</i> | 21 | <i>Ficedula parva</i> | 862 | <i>Hydroprogne caspia</i> | 811 |
| <i>Ammodramus priscogularis</i> | 298 | <i>Ardea cinerea/purpurea</i> | 1 | <i>Calidris sp. (peep sp.)</i> | 507 | <i>Clamator glandarius</i> | 170 | <i>Dendrocopos sp.</i> | 59 | <i>Ficedula semitorquata</i> | 146 | <i>Ichthyophaga aedonii</i> | 248 |
| <i>Anarhynchus alexandrinus</i> | 7109 | <i>Ardea ibis</i> | 6203 | <i>Calidris temminckii</i> | 210 | <i>Clanga clanga</i> | 364 | <i>Dendrocopos syriacus</i> | 4980 | <i>Ficedula sp.</i> | 11 | <i>Ichthyophaga ichthyophaga</i> | 1875 |
| <i>Anarhynchus asiaticus</i> | 7 | <i>Ardea purpurea</i> | 3967 | <i>Calonectris diomedea</i> | 506 | <i>Clanga pomarina</i> | 194711 | <i>Dendrocopos medius</i> | 579 | <i>Francolinus francolinus</i> | 718 | <i>Ichthyophaga melanocephalus</i> | 6259 |
| <i>Anarhynchus leschenaultii</i> | 144 | <i>Ardea sp.</i> | 7 | <i>Caprimulgus aegyptius</i> | 1 | <i>Clanga pomarina/clanga</i> | 103 | <i>Dryobates minor</i> | 925 | <i>Fringilla coelebs</i> | 85856 | <i>Iduna caligata</i> | 3 |
| <i>Anas acuta</i> | 8831 | <i>Ardeidae sp.</i> | 11 | <i>Caprimulgus europaeus</i> | 428 | <i>Coccothraustes coccothraustes</i> | 1264 | <i>Dryocopus martius</i> | 81 | <i>Fringilla montifringilla</i> | 2469 | <i>Iduna pallida</i> | 3801 |
| <i>Anas crecca</i> | 172593 | <i>Ardeola ralloides</i> | 4580 | <i>Carduelis carduelis</i> | 48687 | <i>Coloeus monedula</i> | 110839 | <i>Egretta garzetta</i> | 25515 | <i>Fringillidae sp.</i> | 28 | <i>Irania putilaris</i> | 411 |
| <i>Anas platyrhynchos</i> | 71682 | <i>Arenaria interpres</i> | 437 | <i>Carpodacus erythrinus</i> | 897 | <i>Columba livia</i> | 105231 | <i>Egretta/Ardea sp.</i> | 46 | <i>Fulica atra</i> | 443372 | <i>Jynx torquilla</i> | 398 |
| <i>Anatidae (duck sp.)</i> | 1968 | <i>Argya altostris</i> | 458 | <i>Carpospiza brachyactyla</i> | 104 | <i>Columba oenas</i> | 783 | <i>Elanus caeruleus</i> | 169 | <i>Galerida cristata</i> | 22237 | <i>Ketupa zeylonensis</i> | 1 |
| <i>Anatidae sp.</i> | 40 | <i>Asio flammeus</i> | 42 | <i>Cecropis rufula</i> | 9280 | <i>Columba palumbus</i> | 23238 | <i>Emberiza buchanani</i> | 30 | <i>Gallinago gallinago</i> | 5720 | <i>Laniidae sp.</i> | 63 |



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|--|---------------|--------------------------------|---------------|-------------------------------|---------------|----------------------------------|---------------|------------------------------------|---------------|-------------------------------------|---------------|--------------------|---------------|
| Lanius collurio | 12817 | Melospittacus undulatus | 1 | Oenanthe sp. | 43 | Phylloscopus humei | 70 | Pterocles orientalis | 1013 | Sterna hirundo | 5938 | Turdus sp. | 101 |
| Lanius excubitor | 97 | Mergellus albellus | 14 | Oenanthe xanthopyrmya | 24 | Phylloscopus inornatus | 4 | Pytonoprogne rupestris | 3678 | Sterna sp. | 1 | Turdus torquatus | 343 |
| Lanius isabellinus | 14 | Mergus merganser | 18 | Oriolus oriolus | 1364 | Phylloscopus nitidus | 201 | Puffinus yelkouan | 88982 | Sterninae sp. | 100 | Turdus viscivorus | 3780 |
| Lanius minor | 12358 | Mergus serrator | 1235 | Otis tarda | 231 | Phylloscopus orientalis | 283 | Pycnonotus leucotis | 71 | Sternula albifrons | 2535 | Tyto alba | 248 |
| Lanius nubicus | 1285 | Merops apiaster | 51541 | Otus brucei | 26 | Phylloscopus proregulus | 26 | Pycnonotus xanthopygus | 7346 | Streptopelia decaocto | 34292 | Upupa epops | 5617 |
| Lanius senator | 2014 | Merops bicoloroides | 4 | Otus scops | 641 | Phylloscopus sibilatrix | 374 | Pyrrhocorax graculus | 2334 | Streptopelia orientalis | 1 | Vanellus gregarius | 21 |
| Larinae sp. | 1435 | Merops persicus | 1752 | Oxyura leucocephala | 6135 | Phylloscopus indiarum | 1131 | Pyrrhocorax pyrrhocorax | 4871 | Streptopelia sp. | 12 | Vanellus indicus | 177 |
| Larus argentatus | 48 | Microcarbo pygmaeus | 19761 | Pandion haliaetus | 367 | Phylloscopus sp. | 72 | Pyrrhula pyrrhula | 1149 | Streptopelia turtur | 4718 | Vanellus leucurus | 17 |
| Larus arcticus | 23660 | Milvus migrans | 12707 | Panurus biarmicus | 2356 | Phylloscopus trochilus | 3784 | Rallus aquaticus | 1752 | Strigiformes sp. | 11 | Vanellus spinosus | 6586 |
| Larus cachinnans | 1716 | Milvus milvus | 44 | Paridae sp. | 44 | Phylloscopus trochilus/collybita | 2 | Recurvirostra avosetta | 7717 | Strix aluco | 373 | Vanellus vanellus | 29376 |
| Larus cachinnans/michahellii | 102 | Milvus milvus/migrans | 2 | Parus major | 28122 | Pica pica | 44830 | Regulus ignicapilla | 1096 | Sturnus vulgaris | 391248 | Xenus cinereus | 15 |
| Larus cachinnans/michahellii/armenicus | 1695 | Monticola saxatilis | 340 | Passer domesticus | 118523 | Picidae sp. | 11 | Regulus regulus | 1993 | Sylvia atricapilla | 8003 | Zapornia parva | 227 |
| Larus canus | 463 | Monticola solitarius | 1170 | Passer domesticus/hispanicus | 649 | Picus canus | 121 | Regulus regulus/ignicapilla | 7 | Sylvia borin | 1038 | Zapornia pusilla | 13 |
| Larus fuscus | 1378 | Montifringilla nivalis | 1156 | Passer domesticus/montanus | 24 | Picus viridis | 1000 | Remiz pendulinus | 2653 | Sylvia/Curruca sp. | 77 | | |
| Larus marinus | 39 | Morus bassanus | 5 | Passer hispaniolensis | 31230 | Platalea leucorodia | 7426 | Rhodopechys sanguineus | 334 | Tachybaptus ruficollis | 27473 | | |
| Larus michahellii | 186373 | Motacilla alba | 32703 | Passer moabiticus | 486 | Plectrophenax nivalis | 38 | Rhodospiza obsoleta | 1252 | Tachymarpis melba | 42792 | | |
| Larus sp. | 2898 | Motacilla cinerea | 3104 | Passer montanus | 15613 | Plegadis falcinellus | 19263 | Riparia riparia | 29302 | Tachyspiza badia | 8 | | |
| Limosa lapponica | 99 | Motacilla citreola | 266 | Passer sp. | 71 | Pluvialis apricaria | 2127 | Rissa tridactyla | 57 | Tachyspiza brevipes | 8142 | | |
| Limosa limosa | 8908 | Motacilla flava | 28620 | Passeriformes sp. | 1844 | Pluvialis fulva | 17 | Saxicola maurus | 1050 | Tachyspiza brevipes/Accipiter nisus | 3 | | |
| Limosa sp. | 7 | Motacilla flava/tschutschensis | 2 | Pastor roseus | 17377 | Pluvialis squatarola | 7615 | Saxicola rubetra | 3582 | Tadorna ferruginea | 91380 | | |
| Linaria cannabina | 14223 | Motacilla sp. | 190 | Pelecanus crispus | 4965 | Podiceps auritus | 78 | Saxicola rubicola | 4498 | Tadorna tadorna | 15947 | | |
| Linaria flavirostris | 430 | Muscicapa sp. | 8 | Pelecanus onocrotalus | 40275 | Podiceps cristatus | 37633 | Saxicola rubicola/maurus | 26 | Tetraoallus caspius | 141 | | |
| Locustella fluviatilis | 22 | Muscicapa striata | 4890 | Pelecanus sp. | 60 | Podiceps grisegena | 91 | Scolopax rusticola | 79 | Tetrax tetrax | 3 | | |
| Locustella luscinioides | 341 | Neophron percnopterus | 1000 | Perdix perdix | 266 | Podiceps nigricollis | 4957 | Serinus pusillus | 737 | Thalasseus benzenensis | 45 | | |
| Locustella naevia | 31 | Netta rufina | 10096 | Periparus ater | 5268 | Podiceps sp. | 10 | Serinus serinus | 6331 | Thalasseus sandvicensis | 8673 | | |
| Loxia curvirostra | 573 | Numenius arquata | 3884 | Pernis apivorus | 12912 | Podicipedidae sp. | 8 | Sitta europaea | 1322 | Thinornis dubius | 4204 | | |
| Lullula arborea | 4579 | Numenius phaeopus | 293 | Pernis ptilorhynchus | 1 | Poecile lugubris | 812 | Sitta krueperi | 1366 | Tichodroma muraria | 40 | | |
| Luscinia luscinia | 485 | Numenius phaeopus/arquata | 2 | Petronia petronia | 5883 | Poecile palustris | 299 | Sitta neumayer | 2840 | Tringa erythropus | 3019 | | |
| Luscinia luscinia/megarhynchos | 16 | Numida meleagris | 1 | Phalacrocoracidae sp. | 165 | Porphyrio poliocephalus | 749 | Sitta sp. | 4 | Tringa glareola | 8316 | | |
| Luscinia megarhynchos | 3464 | Nycticorax nycticorax | 7115 | Phalacrocorax carbo | 113833 | Porphyrio sp. (swamphen sp.) | 30 | Sitta tephronota | 258 | Tringa nebularia | 5427 | | |
| Luscinia svecica | 2216 | Oena capensis | 24 | Phalacrocorax carbo/Gulosus | 336 | Porzana porzana | 49 | Somateria mollissima | 2 | Tringa ochropus | 5113 | | |
| Lymnocyrtus minimus | 82 | Oenanthe cyprica | 80 | Phalaropus fulicarius lobatus | 2 | Prinia lepida | 2145 | Spatula clypeata | 24849 | Tringa sp. | 19 | | |
| Lyrurus mikosiewiczi | 87 | Oenanthe deserti | 29 | Phalaropus lobatus | 212 | Prunella collaris | 74 | Spatula querquedula | 30509 | Tringa stagnatilis | 1410 | | |
| Mareca penelope | 28129 | Oenanthe finschii | 943 | Phasianus colchicus | 540 | Prunella modularis | 1151 | Spatula querquedula/Anas crecca | 150 | Tringa totanus | 10826 | | |
| Mareca strepera | 3459 | Oenanthe isabellina | 3372 | Phoenicurus minor | 1 | Prunella ocularis | 114 | Spilopelia senegalensis | 10817 | Troglodytes troglodytes | 5528 | | |
| Marmarotetta angustirostris | 15 | Oenanthe lugens | 2 | Phoenicopus roseus | 359938 | Psittacula eupatria | 3263 | Spinus spinus | 19225 | Turdus atropularis | 3 | | |
| Melanitta fusca | 164 | Oenanthe melanoleuca | 2585 | Phoenicurus ochruros | 6358 | Psittacula eupatria/krameri | 16 | Stercorarius parasiticus | 131 | Turdus iliacus | 1603 | | |
| Melanitta nigra | 87 | Oenanthe monacha | 57 | Phoenicurus phoenicurus | 4167 | Psittacula krameri | 8672 | Stercorarius parasiticus/pomarinus | 9 | Turdus merula | 32522 | | |
| Melanocorypha bimaculata | 150 | Oenanthe oenanthe | 6707 | Phoenicurus sp. | 1 | Psittacula sp. | 714 | Stercorarius pomarinus | 13 | Turdus philomelos | 11318 | | |
| Melanocorypha calandra | 14059 | Oenanthe pleschanka | 7 | Phylloscopus collybita | 17489 | Pterocles alchata | 2 | Stercorarius sp. (jaeger sp.) | 1 | Turdus pilaris | 5089 | | |