






Original research article

The effectiveness of EDDY in removing calcium hydroxide, n-acetylcysteine, and diclofenac sodium medicaments from root canals

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ABSTRACT

OBJECTIVE: To compare the removability of calcium hydroxide (Ca(OH)₂), n-acetylcysteine (NAC), and diclofenac sodium (DCF) from root canals and to assess the effectiveness of the EDDY sonic activation tip in facilitating their removal.

MATERIALS AND METHODS: Sixty-six upper central incisors were instrumented using T-Endo Must files (#40.04) and divided into six groups (n = 11) on the basis of the root canal medicament (Ca(OH)₂, NAC or DCF) and medicament removal protocol (needle or EDDY). The residual medicament was evaluated under ×20 magnification using a stereomicroscope. The Kruskal–Wallis test was used to compare the different medicament groups, whereas the Mann–Whitney U-test was employed to compare the needle irrigation and EDDY groups. The statistical significance level was set at 0.05.

RESULTS: The highest residual medicament was observed in the Ca(OH)₂ group (p < 0.05), with no significant difference between the NAC and DCF groups in the needle irrigation group (p > 0.05). In the EDDY activation group, the DCF group presented the lowest residual medicament (p < 0.05), whereas no significant difference was observed between the Ca(OH)₂ and NAC groups (p > 0.05). Compared with needle irrigation, EDDY activation significantly reduced the residual medicament for all the root canal medicaments (p < 0.05).

CONCLUSION: EDDY activation significantly reduces residual medicament. DCF resulted in less residual medicament remaining in both the needle and EDDY groups.

KEYWORDS: Calcium hydroxide; diclofenac sodium; n-acetylcysteine; root canal medicaments

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[Abstract in Turkish is at the end of the manuscript]

INTRODUCTION

Calcium hydroxide (Ca(OH)₂) is one of the most frequently used root canal medicament (RCM) for achieving effective disinfection in mechanically difficult-to-reach areas of the root canal system because of its antimicrobial properties. Ca(OH)₂ dissociates into Ca²⁺ and OH⁻ ions, creating an alkaline environment; this high pH (12.5) damages the bacterial cell membrane and protein structure¹. Another benefit of Ca(OH)₂ is its ability to neutralize bacterial lipopolysaccharides (LPS), a key virulence factor in the development of apical periodontitis²; however, there are concerns about its inability to eliminate LPS completely from the root canal³. Additionally, the buffering effect of dentin may prevent high pH levels from being fully reached in the dentinal tubules⁴ and bacteria frequently encountered in endodontic infections, such as *Enterococcus faecalis* are resistant to Ca(OH)₂⁵. Therefore, the search for alternative RCMs continues.

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N-Acetylcysteine (NAC) is an amino acid derivative with various therapeutic properties. It exhibits antioxidant activity through its thiol group, which allows it to induce glutathione synthesis and directly neutralize reactive oxygen species⁶. Additionally, NAC helps control inflammation by reducing the expression of the inflammation-inducing nuclear factor kappa B gene and decreasing the levels of proinflammatory cytokines such as interleukin-1 β , interleukin-8, and tumor necrosis factor⁷.

Diclofenac sodium (DCF), one of the most effective inhibitors of prostaglandin production via the cyclooxygenase pathway, is frequently used as a nonsteroidal anti-inflammatory drug for the treatment of acute pain and inflammation⁸. The antimicrobial and antibiofilm activity of DCF⁹ has led to its potential use as an intracanal medication.

One of the major disadvantages of Ca(OH)₂ is the difficulty in removing it from root canals. Residual Ca(OH)₂ can negatively impact the physical properties and sealing ability of root canal sealers to dentinal walls. Over time, any remaining Ca(OH)₂ in the root canals may dissolve, compromising the seal of the root canal filling and leading to bacterial infiltration^{10, 11}. The cleaning of Ca(OH)₂ from root canals remains a challenging aspect of endodontic treatment. However, the removal efficacy of promising RCMs such as NAC and DCF has not yet been examined. This study aimed to compare the removability of Ca(OH)₂, NAC, and DCF from root canals and to assess the effectiveness of a sonic activation tip, EDDY (VDW, Munich, Germany), in facilitating their removal. The null hypothesis was that no significant differences will be observed among the groups.

MATERIALS AND METHODS

Prior Sample Size Calculation

Six different artificial tooth groups were created using various root canal medicaments and activation techniques. To test whether there were differences in the residual medicament amounts among the groups, a priori sample size calculation was performed using two-way analysis of variance with G*Power 3.1.9.4 software (Heinrich Heine University, Dusseldorf, Germany). Based on the calculation, a total sample size of 64 was determined, considering an effect size of 0.40 (Cohen's $f = 0.40$), % α error ($\alpha = 0.05$), and 80% power ($1 - \beta = 0.80$). To ensure an equal number of samples in each group ($n = 11$), a total of 66 teeth were included in the study.

The study design was approved by the Hatay Mustafa Kemal University Non-Interventional Clinical Research Ethics Committee with Protocol Number: 30/10/2024-26.

Sample Selection

Sixty-six single-rooted, single-canal upper central

incisors were selected. Teeth presenting an open root apex, internal or external root resorption, fractures, cracks, or previously filled root canals were excluded. Dental calculus and organic debris were removed from the teeth, and the samples were stored in a 0.9% physiological saline solution until the experimental phase. All the crowns were shortened coronally to a standardized root length of 14 mm using a diamond disc under water cooling.

Chemomechanical Preparation

Access cavity preparation was performed using diamond fissure burs. The apical patency was checked using #10 and #15 K-files (Mani, Utsunomiya, Japan). Teeth with root canals larger than size #20 K-file were excluded from the study. A #15 K-file was visualized at the major apical foramen and then withdrawn by 1 mm to determine the working length (WL). Prior to root canal instrumentation, all the root apices were coated with wax. The root canals were shaped using T-Endo Must (Dentac, Istanbul, Turkey) Reciproc files (#40.04) with the crown-down preparation technique at the WL. The files were used with a pecking motion, limited to a maximum amplitude of 3 mm. After every 3 pecking motions, the file was withdrawn from the canal, and cleaned with damp gauze, and root canal irrigation was performed with 2.5% NaOCl. A 30-gauge Max-i-Probe (Dentsply Sirona, York, PA, USA) needle was used for irrigation and was placed 2 mm short of the WL. The total volume of NaOCl used during preparation was 10 ml, which was consistent for all the samples. After the preparation was complete, 5 mL of sterile saline (SS) solution was introduced into each canal. The canals were subsequently rinsed with 6 mL of 17% EDTA, 5 mL of SS, 6 mL of 2.5% NaOCl, and an additional 5 mL of SS. After the root canals were dried with absorbent paper points, the samples were randomly divided into three groups based on the RCMs: Ca(OH)₂ (Kalsin, Konya, Turkey), NAC (Aromel, Istanbul, Turkey), or DCF (Fagron, Rotterdam, Netherlands). Randomization was performed using block randomization with a block size of six to ensure equal distribution among the groups. An online software tool (www.randomizer.org) was used to implement the randomization process.

Placement of Root Canal Medicaments

All RCMs were freshly prepared by mixing with sterile saline at a 1:1 ratio. The root canals were filled to the WL with the prepared medicaments using a #30 Lentulo spiral. To confirm the complete placement of the RCMs within the canals, the wax seal was removed and the apices of the roots were examined. Once the RCMs were observed at the apical foramen, the wax was reapplied to proceed with the removal phase. The canal orifices were sealed with sterile cotton pellets, and the access cavity was temporarily sealed with Cavit-G (3M ESPE, Seefeld, Germany). The samples were then stored at 37°C in an environment with 100%

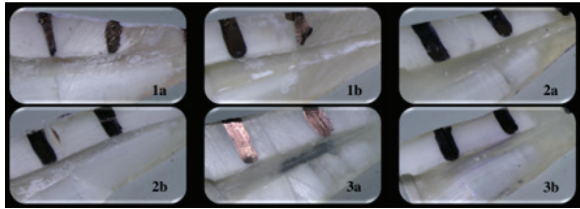


Figure 1: Representative images of the experimental groups. 1: Calcium hydroxide, 2: N-Acetylcysteine, 3: Diclofenac sodium, a: Needle Irrigation, b: EDDY

humidity for 1 week.

Removal of Root Canal Medicaments

Each medicament group was divided into two subgroups based on the medicament removal protocol. Randomization will be conducted using block randomization with a block size of two to ensure equal distribution among the subgroups. The RCMs were removed from the root canals by a T-Endo Must #40.04 file and rinsed with 5 mL NaOCl. Following this, 5 mL of SS was used in the root canals. In groups without activation the medicament was removed solely using needle irrigation. Six milliliters of 17% EDTA was placed in the canals for 1 minute. This was followed by a final rinse with 5 mL of SS, 6 mL of 2.5% NaOCl, and an additional 5 mL of SS. In the activation groups, the solutions were activated using an EDDY #25/.04 tip with a sonic device, the Micron TA-200 Air-Scaler (TA-200-S2H, Micron, Japan). The tip was placed 1 mm short of the WL and activated with a 3-4 mm up-down motion. The root canals were rinsed with 2 mL of 17% EDTA solution and activated for 20 s. This cycle was repeated two more times. To neutralize the effect of EDTA, 5 mL of SS was used. After irrigation with 2 mL of 2.5% NaOCl, the solution was activated for 20 s this procedure was repeated three times. A total volume of 6 mL of EDTA and 6 mL of NaOCl were used, the activation time for each solution was 60 s (3x20 s). Finally, 5 mL of SS was used in the root canals.

Evaluation of Residual Medicament

The roots were longitudinally divided into two halves

using a precision cutting device. Only one half of each root was evaluated. Images of each half were captured under $\times 20$ magnification using a stereomicroscope (Figure 1). The digital images were imported into the ImageJ software program (ImageJ 1.47V, National Institutes of Health, Bethesda, MD, USA). The boundaries of the root canal lumen were delineated, and the canal surface area was calculated. The areas with residual RCMs were also measured and expressed as a percentage of the total canal surface area¹².

Statistical Analysis

The data obtained from different RCMs and medicament removal protocols were analyzed using SPSS software (version 22, IBM Corp., Armonk, NY, USA). Differences among the RCM groups were assessed using the Kruskal-Wallis test, followed by post-hoc Dunn's test for pairwise comparisons. Comparisons between the needle irrigation and EDDY activation groups were performed using the Mann-Whitney U test. Cohen's kappa test was used to analyze interobserver reliability between two practitioners. A p-value less than 0.05 was considered statistically significant. Significance values were adjusted for multiple comparisons using Dunn's test with Bonferroni correction.

RESULTS

The kappa value for interobserver reliability was 0.91. There were statistically significant differences among the RCMs and medicament removal protocols. Interaction between factors was not tested. With needle irrigation, the highest residual medicament percentage was observed in the $\text{Ca}(\text{OH})_2$ group ($p < 0.05$), whereas the NAC and DCF groups presented similar results ($p > 0.05$). Under EDDY irrigation, the $\text{Ca}(\text{OH})_2$ and NAC groups presented comparable results ($p > 0.05$); however, the DCF group presented the lowest percentage of residual medicament ($p < 0.05$) (Table 1). Compared with needle irrigation, EDDY activation significantly reduced the residual medicament percentages for all RCMs ($p < 0.05$) (Table 2).

Table 1: Mean, standard deviation, median and IQR values of the residual medicament percentages.

Root Canal Medicaments	Needle		EDDY	
	Mean \pm SD	Median-IQR	Mean \pm SD	Median-IQR
$\text{Ca}(\text{OH})_2$	38.39 ^A \pm 12.05	35.87 – 20.24	20.95 ^A \pm 11.19	18.03 – 21.8
NAC	19.41 ^B \pm 12.18	20.66 – 18.05	10.61 ^A \pm 4.22	12.96 – 7.81
DCF	17.63 ^B \pm 9.32	14.46 – 13.46	4.17 ^B \pm 3.22	3.71 – 6.75
P_{χ^2}	0.000		0.000	
Chi-Square	13.598		18.442	
η^2	0.387		0.548	

Superscripts with different uppercase letters indicate statistically significant differences within the same column.

$p < 0.05$ indicates statistical significance. Significance values were adjusted for multiple comparisons using Dunn's test with Bonferroni correction.

χ^2 : Kruskal Wallis test.

Table 2: Mean rank values of the residual medicament percentages.

	Needle	EDDY	P*	Z	r
Root Canal Medicaments	Mean Rank	Mean Rank			
Ca(OH) ₂	15.18 ^a	7.82 ^b	0.008	-2.659	0.57
NAC	14.36 ^a	8.64 ^b	0.039	-2.069	0.44
DCF	16.36 ^a	6.64 ^b	0.000	-3.513	0.74

Superscripts with different lowercase letters indicate statistically significant differences within the same row.

$p < 0.05$ indicates statistical significance.

*:Mann-Whitney U test.

DISCUSSION

The intricate root canal system may not be accessible for conventional irrigation procedures, and Ca(OH)₂ may remain within the root canals¹³. No current strategy is able to completely clean Ca(OH)₂ medicament before root canal obturation. To achieve better results, a combination of chemical irrigation and mechanical activation is recommended for its removal^{14, 15}. In this study, the effectiveness of conventional irrigation methods and the EDDY activation system in the removal of different RCMs was investigated, and the null hypothesis was rejected.

The smear layer impedes the penetration of RCMs into deeper regions of the root canal system, thereby diminishing their therapeutic efficacy¹⁶. Therefore, following canal preparation, we used NaOCl-EDTA-NaOCl sequentially to remove the smear layer.

Mobarakeh *et al.*¹⁷ reported no difference in the efficacy of removing pure and injectable forms of Ca(OH)₂ from root canals. To follow a similar protocol, all RCMs were mixed with sterile saline, and the prepared medicaments were delivered into the root canals using a Lentulo spiral in this study.

In previous studies, the retention time of RCMs within root canals has frequently been reported to be one week^{12, 14}. Keskin *et al.*¹⁸ reported that the intracanal duration of triple antibiotic paste does not influence its removal. However, the impact of retention time on the removal of the RCMs used in this study remains unclear. A period of one week was chosen, considering the application of Ca(OH)₂ under clinical conditions¹⁹. We prepared RCMs by mixing powder with sterile saline at a 1:1 ratio to achieve a paste-like consistency suitable for placement into canals.

The minimum inhibitory concentration (MIC) of NAC against *E. faecalis* has been reported as 1.56 mg/mL²⁰ and 100 mg/mL NAC has been shown to completely disrupt mature multispecies endodontic biofilms²¹. The MIC of DCF against *E. faecalis* has been reported as 50 µg/mL²², and DCSs at 5% and 2.5% have demonstrated greater antimicrobial effects than DCSs at 1.25%²³. Further research is needed to investigate the effects of the concentration and type of transporter for NAC and DCF on the removal of these medicaments.

The most commonly used methods for cleaning Ca(OH)₂ involve mechanical shaping of the root canal at the WL using the master apical file, combined with extensive irrigation using NaOCl and EDTA^{24, 25}. Despite the implementation of this approach, no sample was observed without residual medicament in this study.

Needle irrigation resulted in the highest residual medicament in the Ca(OH)₂ group, with no difference between NAC and DCF. Under EDDY irrigation, Ca(OH)₂ and NAC were similar, but DCF had the lowest residue. It is not possible to directly compare our study results, as the removal of NAC or DCF as root canal medicaments from root canals has not been previously investigated. However, the hydrophilic nature of NAC, resulting from its ability to form direct bonds with methyl groups, has been reported to facilitate its easy elimination from the body⁷. DCF is not biodegradable²⁶; however, it is a water-soluble substance that easily accumulates in water²⁷, which may account for its low residue levels.

Studies have demonstrated that the antibacterial activity of NAC remains unaffected by the presence of dentin and is effective against both biofilm and planktonic forms of endodontic pathogens, including *E. faecalis*²⁰. Jariyamana *et al.*²⁸ reported that NAC is effective in addressing inflammation associated with LPS. Furthermore, the combination of Ca(OH)₂ and NAC has been found to be more effective against *Escherichia coli*'s LPS than either NAC or Ca(OH)₂ alone. The antibacterial efficacy of DCF against *E. faecalis* is superior to that of Ca(OH)₂²², and the addition of DCF to Ca(OH)₂ enhances its antimicrobial effects against *E. faecalis* biofilms²⁹. DCF exhibits bactericidal activity by disrupting bacterial DNA synthesis³⁰. Additionally, it has been reported to effectively suppress peptidoglycan biosynthesis, biofilm formation, and the expression of genes and proteins associated with bacterial virulence^{9,31,32}. Adl *et al.*³³ evaluated the antibacterial activity against *E. faecalis* and reported that DCF displayed similar effectiveness to that of Ca(OH)₂ and NAC at a depth of 100 µm. However, at a depth of 200 µm, DCF demonstrated superior efficacy to that of Ca(OH)₂ and NAC. The authors attributed the high efficacy of DCF to its resistance to dentin. We believe that the affinity of DCF for water may contribute to its enhanced effectiveness in deeper dentinal tubules.

Although NAC and DCF are easier to remove from root canals than $\text{Ca}(\text{OH})_2$ is, their effects on the mechanical properties of dentin remain uninvestigated and require further examination.

Compared with needle irrigation, EDDY activation significantly decreased the residual percentage of all RCMs. The superiority of EDDY over needle irrigation in the removal of RCMs from simulated immature tooth models has been demonstrated³⁴. Similarly, Güven *et al.*³⁵ reported that EDDY significantly improved the removal of $\text{Ca}(\text{OH})_2$ from artificially created grooves in the apical region of root canals, whereas Silva *et al.*³⁶ highlights its significant effect in removing $\text{Ca}(\text{OH})_2$ from simulated internal resorption cavities. The EDDY (VDW, Munich, Germany) sonic activation device is designed to increase the effectiveness of irrigation solutions. It features a flexible polyamide tip with a size of 25/04 and operates at a frequency of 6000 Hz. This allows the irrigation solution to mechanically reach canal irregularities that are otherwise difficult to access³⁷.

The root canal configuration may affect the effectiveness of intracanal medicament removal³⁸. Although root canals with a Type 1 Vertucci configuration were selected for this study, the lack of standardization in the canal morphology of the samples represents a significant limitation. Therefore, the removal of the RCMs was calculated proportionally for each sample. Different methods, such as scanning electron microscopy³⁶, or digital photography under a stereomicroscope³⁹, as used in this study—have been employed to evaluate RCMs within the root canal system. However, these techniques require longitudinal sectioning of the root to assess the internal surfaces. As a result, they only allow for the evaluation of the superficial layer of $\text{Ca}(\text{OH})_2$ and make it difficult to accurately quantify the amount of medicament remaining within the canal system. In contrast, micro-computed tomography (micro-CT) enables quantitative analysis of the three-dimensional volume of $\text{Ca}(\text{OH})_2$ remnants. Furthermore, the non-invasive nature of micro-CT allows the same specimens to be evaluated both before and after irrigation procedures, providing 3D superimposed images and more accurate quantitative measurements⁴⁰.

CONCLUSION

Compared with needle irrigation EDDY activation significantly reduces the residual percentage of all RCMs. While the $\text{Ca}(\text{OH})_2$ group presented the highest residual medicament percentage with needle irrigation, the DCF group presented the lowest residual medicament percentage with EDDY activation.

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REFERENCES

1. Mohammadi Z, Dummer PM. Properties and applications of calcium hydroxide in endodontics and dental traumatology. *Int Endod J* 2011;8:697-730.
2. Adl A, Motamedifar M, Shams MS, Mirzaie A. Clinical investigation of the effect of calcium hydroxide intracanal dressing on bacterial lipopolysaccharide reduction from infected root canals. *Aust Endod J* 2015;1:12-6.
3. Bedran NR, Nadelman P, Magno MB, de Almeida Neves A, Ferreira DM, Braga Pintor AV, et al. Does Calcium Hydroxide Reduce Endotoxins in Infected Root Canals? Systematic Review and Meta-analysis. *J Endod* 2020;11:1545-58.
4. Haapasalo HK, Sirén EK, Waltimo TM, Ørstavik D, Haapasalo MP. Inactivation of local root canal medicaments by dentine: an in vitro study. *Int Endod J* 2000;2:126-31.
5. van der Waal SV, van der Sluis LW, Özok AR, Exterkate RA, van Marle J, Wesselink PR, et al. The effects of hyperosmosis or high pH on a dual-species biofilm of *Enterococcus faecalis* and *Pseudomonas aeruginosa*: an in vitro study. *Int Endod J* 2011;12:1110-7.
6. Sadowska AM, Manuel YKB, De Backer WA. Antioxidant and anti-inflammatory efficacy of NAC in the treatment of COPD: discordant in vitro and in vivo dose-effects: a review. *Pulm Pharmacol Ther* 2007;1:9-22.
7. Siemer K, Husari A, Vach K, Tomakidi P, Hellwig E, Schulz SD, et al. N-Acetylcysteine modulates the effects of composites on human gingival keratinocytes. *Dent Mater* 2021;4:597-611.
8. Dheeraj M, Johar S, Mahajan P, Jandial T, Chauhan S, Gupta K. Comparative Evaluation of the Efficacy of Two Modes of Delivery of Diclofenac for the Management of Post-Endodontic Pain: A Randomized Controlled Clinical Trial. *J Pharm Bioallied Sci* 2023;Suppl 2:S1283-s6.
9. Riordan JT, Dupre JM, Cantore-Maty SA, Kumar-Singh A, Song Y, Zaman S, et al. Alterations in the transcriptome and antibiotic susceptibility of *Staphylococcus aureus* grown in the presence of diclofenac. *Ann Clin Microbiol Antimicrob* 2011;30.
10. Kim SK, Kim YO. Influence of calcium hydroxide intracanal medication on apical seal. *Int Endod J* 2002;7:623-8.
11. Hosoya N, Kurayama H, Iino F, Arai T. Effects of calcium hydroxide on physical and sealing properties of canal sealers. *Int Endod J* 2004;3:178-84.
12. Taşdemir T, Celik D, Er K, Yildirim T, Ceyhanli KT, Yeşilyurt C. Efficacy of several techniques for the removal of calcium hydroxide medicament from root canals. *Int Endod J* 2011;6:505-9.
13. van der Sluis LW, Wu MK, Wesselink PR. The evaluation of removal of calcium hydroxide paste from an artificial standardized groove in the apical root canal using different irrigation methodologies. *Int Endod J* 2007;1:52-7.
14. Donnermeyer D, Wyrsch H, Bürklein S, Schäfer E. Removal of Calcium Hydroxide from Artificial Grooves in Straight Root Canals: Sonic Activation Using EDDY Versus Passive Ultrasonic Irrigation and XPendo Finisher. *J Endod* 2019;3:322-6.
15. Keskin C, Keleş A, Sarıılmaz Ö. Efficacy of glycolic acid for the removal of calcium hydroxide from simulated internal Resorption cavities. *Clin Oral Investig* 2021;7:4407-13.
16. Wang Z, Shen Y, Haapasalo M. Effect of smear layer against disinfection protocols on *Enterococcus faecalis*-infected dentin. *J Endod* 2013;11:1395-400.
17. Nasab Mobarakeh NM, Taheri A, Rahmanian H, Jafarpour D, Rahmanian S. Effect of Various Irrigating Devices on the Removal of Two Different Forms of Calcium Hydroxide from Internal Resorption Cavities. *Int J Dent* 2020:8881177.
18. Keskin C, Güler DH, Sarıılmaz E. Effect of intracanal time of triple antibiotic paste on its removal from simulated immature roots using passive ultrasonic irrigation and XP-endo Finisher. *J Dent Res Dent Clin Dent Prospects* 2018;4:288-93.
19. Law A, Messer H. An evidence-based analysis of the antibacterial effectiveness of intracanal medicaments. *J Endod* 2004;10:689-94.
20. Quah SY, Wu S, Lui JN, Sum CP, Tan KS. N-acetylcysteine inhibits growth and eradicates biofilm of *Enterococcus faecalis*. *J Endod* 2012;1:81-5.

21. Choi YS, Kim C, Moon JH, Lee JY. Removal and killing of multispecies endodontic biofilms by N-acetylcysteine. *Braz J Microbiol* 2018;1:184-8.
22. Salem-Milani A, Balaei-Gajan E, Rahimi S, Moosavi Z, Abdollahi A, Zakeri-Milani P, et al. Antibacterial Effect of Diclofenac Sodium on *Enterococcus faecalis*. *J Dent (Tehran)* 2013;1:16-22.
23. Ferrer-Luque CM, Baca P, Solana C, Rodríguez-Archilla A, Arias-Moliz MT, Ruiz-Linares M. Antibiofilm Activity of Diclofenac and Antibiotic Solutions in Endodontic Therapy. *J Endod* 2021;7:1138-43.
24. Alturaiki S, Lamphon H, Edrees H, Ahlquist M. Efficacy of 3 different irrigation systems on removal of calcium hydroxide from the root canal: a scanning electron microscopic study. *J Endod* 2015;1:97-101.
25. Ma JZ, Shen Y, Al-Ashaw AJ, Khaleel HY, Yang Y, Wang ZJ, et al. Micro-computed tomography evaluation of the removal of calcium hydroxide medicament from C-shaped root canals of mandibular second molars. *Int Endod J* 2015;4:333-41.
26. Du J, Xu K, Yang X, Dong Z, Zhao L. Removal of diclofenac sodium from aqueous solution using different ionic liquids functionalized tragacanth gum hydrogel prepared by radiation technique. *Int J Biol Macromol* 2024;Pt 1:130758.
27. He X, Chang C, Yang L, Cai Y, Wang Q. Construction of bismuth based MOF for efficient removal of sodium diclofenac via adsorption and photocatalysis. *J Environ Sci (China)* 2025:14-24.
28. Jariyamana N, Chuveera P, Dewi A, Leelapornpisid W, Ittichaicharoen J, Chattipakorn S, et al. Effects of N-acetyl cysteine on mitochondrial ROS, mitochondrial dynamics, and inflammation on lipopolysaccharide-treated human apical papilla cells. *Clin Oral Investig* 2021;6:3919-28.
29. de Freitas RP, Greatti VR, Alcalde MP, Cavenago BC, Vivan RR, Duarte MA, et al. Effect of the Association of Nonsteroidal Anti-inflammatory and Antibiotic Drugs on Antibiofilm Activity and pH of Calcium Hydroxide Pastes. *J Endod* 2017;1:131-4.
30. Dastidar SG, Ganguly K, Chaudhuri K, Chakrabarty AN. The antibacterial action of diclofenac shown by inhibition of DNA synthesis. *Int J Antimicrob Agents* 2000;3:249-51.
31. Zhang S, Qu X, Tang H, Wang Y, Yang H, Yuan W, et al. Diclofenac Resensitizes Methicillin-Resistant *Staphylococcus aureus* to β -Lactams and Prevents Implant Infections. *Adv Sci (Weinh)* 2021;13:2100681.
32. Silva A, Silva PM. Non-antibiotic compounds: the activity of the NSAID diclofenac on bacteria—a review. *Int J Curr Microbiol Appl Sci* 2018:340-51.
33. Adl A, Motamedifar M, Malekzadeh P, Sedigh-Shams M. Disinfection of dentinal tubules with diclofenac sodium and N-Acetylcysteine compared with calcium hydroxide as intracanal medicaments against *Enterococcus faecalis*. *Aust Endod J* 2022;3:386-91.
34. Usta SN, Erdem BA, Gündoğar M. Comparison of the removal of intracanal medicaments used in regenerative endodontics from root canal system using needle, ultrasonic, sonic, and laser-activated irrigation systems. *Lasers Med Sci* 2024;1:27.
35. Güven Y, Ali A, Arslan H. Efficiency of Endosonic Blue, Eddy, Ultra X and Endoactivator in the removal of calcium hydroxide paste from root canals. *Aust Endod J* 2022;1:32-6.
36. Marques-da-Silva B, Alberton CS, Tomazinho FSF, Gabardo MCL, Duarte MAH, Vivan RR, et al. Effectiveness of five instruments when removing calcium hydroxide paste from simulated internal root resorption cavities in extracted maxillary central incisors. *Int Endod J* 2020;3:366-75.
37. Swimberghe RCD, De Clercq A, De Moor RJG, Meire MA. Efficacy

of sonically, ultrasonically and laser-activated irrigation in removing a biofilm-mimicking hydrogel from an isthmus model. *Int Endod J* 2019;4:515-23.

38. Donnermeyer D, Dust PC, Schäfer E, Bürklein S. Comparative Analysis of Irrigation Techniques for Cleaning Efficiency in Isthmus Structures. *J Endod* 2024;5:644-50.e1.

39. Keskin C, Sariyılmaz E, Sariyılmaz Ö. Efficacy of XP-endo Finisher File in Removing Calcium Hydroxide from Simulated Internal Resorption Cavity. *J Endod* 2017;1:126-30.

40. Yoo YJ, Kim JH, Hwang JJ, Sigurdsson A, Kim HC. Efficacy of Different Irrigation Needles and Ultrasonic Activation on Calcium Hydroxide Removal: A Micro-CT Study Using 3D-Printed Endodontic Models. *J Endod* 2024;10:1478-83.

EDDY'nin kalsiyum hidroksit, n-asetilsistein ve diklofenak sodyum ilaçlarını kök kanallarından uzaklaştırmadaki etkinliği

ÖZET

AMAÇ: Bu çalışmanın amacı kalsiyum hidroksit ($\text{Ca}(\text{OH})_2$), N-asetilsistein (NAC) ve diklofenak sodyumun (DCF) kök kanallarından uzaklaştırılabilirliğini karşılaştırmak ve bu süreçte EDDY sonik aktivasyon ucunun etkinliğini değerlendirmektir.

GEREÇ VE YÖNTEMLER: Altmış altı üst santral kesici diş, T-Endo Must eğeleri (#40.04) kullanılarak şekillendirildi ve kök kanal medikamenti ($\text{Ca}(\text{OH})_2$, NAC veya DCF) ile medikament uzaklaştırma protokolüne (iğne veya EDDY) göre altı gruba (n = 11) ayrıldı. Rezidüel medikament, stereomikroskop altında $\times 20$ büyütme ile değerlendirildi. Farklı medikament gruplarını karşılaştırmak için Kruskal-Wallis testi, iğne irrigasyonu ve EDDY gruplarını karşılaştırmak için ise Mann-Whitney U testi kullanıldı. İstatistiksel anlamlılık düzeyi 0.05 olarak belirlendi. **Bulgular:** İğne irrigasyonu grubunda en fazla rezidüel medikament $\text{Ca}(\text{OH})_2$ grubunda gözlemlendi ($p < 0.05$); NAC ve DCF grupları arasında anlamlı bir fark bulunmadı ($p > 0.05$). EDDY aktivasyonu grubunda en düşük rezidüel medikament miktarı DCF grubunda gözlemlendi ($p < 0.05$), ancak $\text{Ca}(\text{OH})_2$ ve NAC grupları arasında anlamlı bir fark yoktu ($p > 0.05$). İğne irrigasyonuna kıyasla, EDDY aktivasyonu tüm kök kanal medikamentleri için rezidüel madde miktarını anlamlı derecede azalttı ($p < 0.05$).

SONUÇ: EDDY aktivasyonu rezidüel medikament miktarını önemli ölçüde azaltmaktadır. DCF, hem iğne hem de EDDY gruplarında daha az medikament kalıntısına neden olmuştur.

ANAHTAR KELİMELER: Diklofenak sodyum; kalsiyum hidroksit; kök kanalı ilaçları; n-asetilsistein.