

The Eurasia Proceedings of Science, Technology, Engineering & Mathematics (EPSTEM), 2018

Volume 2, Pages 72-80

ICRES 2018: International Conference on Research in Education and Science

Antioxidant and Antimicrobial Activities of Some Newly Synthesized 4-[1-(2,6-Dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4,5-dihydro-1*H*-1,2,4triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoates

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Abstract: In this study, 4-(3-alkyl/aryl-4,5-dihydro-1*H*-1,2,4-triazol-5-one-4-yl-azomethine)-2-methoxyphenyl benzoates were treated with 2,6-dimethylmorpholine in the presence of formaldehyde according to the Mannich reaction to synthesize six novel 4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4,5-dihydro-triazol-5on-4-yl-azomethine]-2-methoxyphenyl benzoates. The structures of synthesized six novel heterocyclic compounds were characterized by IR, ¹³C-NMR and ¹H-NMR spectroscopic methods. The novel 4-[1-(2,6dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4.5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoates were investigated in vitro antioxidant properties by using reducing power, free radical scavenging and metal chelating activity. For the measurement of the reductive ability, Fe^{3+} - Fe^{2+} transformation was investigated in the presence of compound using by the method of Oyaizu (1986). The hydrogen atoms or electrons donation ability of the synthesized compound was measured by DPPH[.] using the method of Blois (1958). The chelating effect of ferrous ions by the compound was determined according to the method of Dinis, Madeira & Almeida (1994). BHT, BHA, EDTA and α -tocopherol were used as reference antioxidant compounds. The new compounds were examined in-vitro antimicrobial properties against 6 different microorganisms (Bacillus substilis (ATCC11774), BacilluscCereus (ATCC11778), Staphylococcus aureus (ATCC6538), Escherichia coli (ATCC25922), Pseudomonas aeruginosa (ATCC27853) and Klebsiella pneumonia (ATCC4352)) by the agar well method and the obtained results were evaluated.

Keywords: Mannich base, 4,5-dihydro-1H-1,2,4-triazol-5-one, Antioxidant, Antimicrobial

Introduction

Mannich bases have applications the field medicinal chemistry, the product synthetic polymers, the petroleum industry, as products used in water treatment, cosmetics, the dyes industry, etc (Tramontini & Angiolini, 1994). Moreover, Mannich bases have some biological activities such as anticancer (Savariz et al., 2010), antibacterial

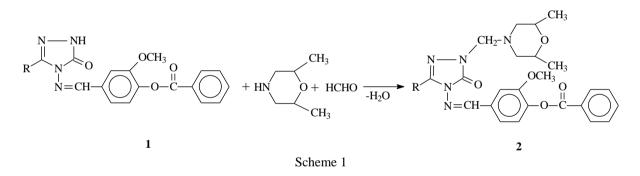
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(Maddila & Jonnalagadda, 2012), anti-inflammatory (Liu, Yu, Li, Pang, & Zhao, 2013), anti-HIV (Sriram, Yogeeswari, Dinakaran, & Sowmya, 2008), analgesic (Nithinchandra, Kalluraya, Aamir, & Shabaraya, 2012), antiviral (Chen et al., 2010), antifungal (Ozkan-Daguyan, Sahin, & Koksal, 2013), antitumor (Pati et al., 2008), antidepressant (Köksal & Bilge, 2007) and antioxidant activities (Hamama, Zoorob, Gouda, & Afsah, 2011). Antioxidants are extensively studied for their capacity to protect organism and cell from damage that is induced by the oxidative stress. A great deal of research has been devoted to the study of different types of natural and synthetic antioxidant. A large number of heterocyclic compounds, containing the 1,2,4-triazole ring, are associated with diverse biological properties such as antioxidant, anti-inflammatory, antimicrobial and antiviral activity. External chemicals and internal metabolic processes in human body or in food system might produce highly reactive free radicals, especially oxygen derived radicals, which are capable of oxidizing biomolecules by resulting in cell death and tissue damage. Oxidative damages play a significantly pathological role in human diseases. Cancer, emphysema, cirrhosis, atherosclerosis and arthritis have all been correlated with oxidative damage. Also, excessive generation of reactive oxygen species (ROS) induced by various stimuli and which exceeds the antioxidant ability of the organism leads to variety of pathophysiological processes like inflammation, diabetes, genotoxicity and cancer (McClements & Decker, 2000).

Triazoles are heterocyclic compounds that contain three nitrogen atoms. 1,2,4-Triazole and 4,5-dihydro-1*H*-1,2,4-triazol-5-one derivatives are reported to possess a broad spectrum of biological activities such as analgesic, antibacterial, antioxidant and antiparasitic properties (Aktas-Yokus, Yuksek, Gursoy-Kol, & Alpay-Karaoglu, 2015; Chidananda et al., 2012). Considering about the development of new hetero moieties by combining potential biological active scaffolds, an attempt was made here to obtain 1,2,4-triazoles bearing piperidine ring and to evaluate their antioxidant activity.

In the present paper, 4-(3-alkyl/aryl-4,5-dihydro-1H-1,2,4-triazol-5-one-4-yl-azomethine)-2-methoxyphenyl benzoates (1) (Koca, Yıldız, & Köçek, 2010) were treated with 2,6-dimethylmorpholine in the presence of formaldehyde according to the Mannich reaction to synthesize six novel <math>4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoates (2).



Method

Chemicals and Apparatus

Chemical reagents used in this paper were bought from Merck AG, Aldrich and Fluka. Melting points were recorded in open glass capillaries using a Stuart SMP30 melting point apparatus and were not corrected. The infrared spectra were recorded on an Alpha-P Bruker FT-IR Spectrometer. ¹H and ¹³C NMR spectra were determined in deuterated dimethyl sulfoxide with TMS as internal standard using a Bruker Avance III spectrophotometer at 400 MHz and 100 MHz, respectively.

Synthesis of 4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoates: The General Procedure

4-(3-alkyl/aryl-4,5-dihydro-1H-1,2,4-triazol-5-one-4-yl-azomethine)-2-methoxyphenyl benzoates (1) were obtained according to the literature (Koca, Yıldız, & Köçek, 2010). Then, to solution of this compound (1) (5 mmol) in absolute ethanol was added 2,6-dimethylmorpholine (6 mmol). The reaction mixture was refluxed for 4 hours. The mixture was left at room temperature for overnight. After cooling the mixture in the refrigerator,

the solid formed was obtained by filtration, washed with cold ethanol and recrystallized from ethanol. Several recrystallizations of the residue from the same solvent gave pure compounds 2 as colourless crystals.

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-methyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoate (2a)

Yield: 70.58%, m.p. 90°C. IR (KBr, v, cm⁻¹): 1741, 1695 (C=O), 1595 (C=N), 1259 (COO), 753 and 705 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 21.05-1.15 (m, 6H, 2CH₃), 3.87 (s, 3H, OCH₃;), 4.58 (m, 4H, NH₂N), 2.3-2.08, 2.33-2.37, -2.56-2.78, 2.81-3.56(m, 6H), 7.43 (d, 1H, ArH *J*=8.40Hz), 7.55-7.57 (m, 1H, ArH), 7.63-7.69 (m, 3H, ArH), 7.77 (m, 1H, ArH), 8.15-8.17 (m, 2H, ArH), 9.76(s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 10.98 (CH₃), 17.90 and 18.93 (2CH₃), 55.02 and 55.60 (2CH₂), 56.06 (OCH₃), 65.26 (NCH₂N), 71.03 (2CH), 111.66 (CH), 120.68 (CH); 123.69(CH), 128.43(C), 129.04(2CH), 129.85(2CH), 132.49(C), 134.17(CH), 141.92(C), 150.28 (C) (ArC), 143.14 (Triazole C₃), 151.39 (Triazole C₅), 153.60 (N=CH), 163.78 (COO).

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-ethyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoate (2b)

Yield: 70.14%, m.p. 122°C. IR (KBr, v, cm⁻¹): 1741, 1695 (C=O), 1595 (C=N), 1259 (COO), 753 and 705 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 1.05-1.15 (m, 6H, 2CH₃), 1.26 (t, 3H, CH₂CH₃; *J*=7.20 Hz), 2.79 (q, 2H, CH₂CH₃; *J*=7.20 Hz), 3.86 (s, 3H, OCH₃), 4.57-4.60 (m, 2H, NCH₂N), 2.02-2.06, 2.25-2.28, 2.61-2.64, 2.80-2.82, 3.51-3.56 (m, 6H), 7.43 (d, 1H, ArH; *J*=8.00 Hz), 7.54-7.56 (m, 1H, ArH), 7.63-7.67 (m, 3H, ArH), 7.77-7.79 (m, 1H, ArH), 8.15-8.17 (m, 2H, ArH), 9.75 (s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 10.05 (CH₂CH₃), 17.90 (CH<u>CH₃</u>), 18.40 (<u>CH₂CH₃</u>), 18.94 (CH<u>CH₃</u>), 55.03 and 55.63 (2CH₂), 56.05 (OCH₃), 66.29 (NCH₂N), 71.04 (2CH), 111.67; 120.57; 123.69; 128.43; 129.04 (2CH); 129.86 (2CH); 132.52; 134.18; 141.91; 150.38 (ArC), 146.87 (Triazole C₃), 151.39 (Triazole C₅), 153.64 (N=CH), 163.78 (COO).

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-benzyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoate (2c)

Yield: 70.31%, m.p. 147°C. IR (KBr, v, cm⁻¹): 1739, 1706 (C=O), 1592 (C=N), 1254 (COO), 750 and 714 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 1.06-1.15 (m, 6H, 2CH₃), 3.86 (s, 3H, OCH₃), 4.15 (s, 2H, CH₂Ph), 4.63 (s, 2H, NCH₂N), 2.03-2.08, 2.24-2.28, 2.60-2.64, 2.81-2.83, 3.51-3.56 (m,6H), 7.26-7.28 (m, 1H, ArH), 7.33-7.41 (m, 5H, ArH), 7.47-7.48 (m, 1H, ArH), 7.59-7.66 (m, 3H, ArH), 7.77-7.79 (m, 1H, ArH), 8.14-8.17 (m, 2H, ArH), 9.71 (s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 17.89 and 18.93 (2CH₃), 31.02 (CH₂Ph), 55.07 and 55.67 (2CH₂), 56.02 (OCH₃), 65.78 (NCH₂N), 71.02 (2CH), 110.87; 121.23; 123.64; <u>126.80</u>; 128.42; <u>128.50 (2CH)</u>; <u>128.83 (2CH)</u>; 129.04(2CH); 129.86 (2CH); 132.46; 134.18; <u>135.76</u>; 141.95; 150.27 (ArC), 144.92 (Triazole C₃), 151.36 (Triazole C₅), 153.00 (N=CH), 163.78 (COO).

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-p-methylbenzyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoate (2d)

Yield: 70.31%, m.p. 129°C. IR (KBr, v, cm⁻¹): 1741, 1706 (C=O), 1579 (C=N), 1252 (COO), 868 (1,4-disubstituted benzenoid ring), 748 and 711 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 1.06-1.10 (m) and 1.15 (d, *J*=6.40 Hz) (6H, 2CH₃), 2.27 (s, 3H, PhCH₃), 3.87 (s, 3H, OCH₃), 4.09 (s, 2H, CH₂Ph), 4.62 (s, 2H, NCH₂N), 2.02-2.07, 2.25-2.28, 2.60-2.64, 2.81-2.83, 3.50-3.56 (m,6H), 7.15 (d, 2H, ArH; *J*=8.00 Hz), 7.26 (d, 2H, ArH; *J*=7.60 Hz), 7.41 (d, 1H, ArH; *J*=8.00 Hz), 7.48-7.50 (m, 1H, ArH), 7.59-7.67 (m, 3H, ArH), 7.78-7.80 (m, 1H, ArH), 8.15-8.17 (m, 2H, ArH), 9.70 (s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 17.89 and 18.93 (2CH₃), 20.56 (PhCH₃), 30.63 (CH₂Ph), 55.08 and 55.68 (2CH₂), 56.01 (OCH₃), 65.76 (NCH₂N), 71.02 (2CH), 110.87; 121.24; 123.64; 128.42; <u>128.54 (2CH)</u>; 129.05(2CH) (<u>2CH</u>); 129.86 (2CH); 132.48; <u>132.63</u>; 134.18; <u>135.91</u>; 141.94; 150.26 (ArC), 145.07 (Triazol C₃), 151.36 (Triazol C₅), 152.95 (N=CH), 163.78 (COO).

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-p-methoxybenzyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxybhenyl benzoate (2e)

Yield: 74.60%, m.p. 113°C. IR (KBr, v, cm⁻¹): 1740, 1706 (C=O), 1595 (C=N), 1247 (COO), 868 (1,4-disubstituted benzenoid ring), 747 and 7134 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 1.05-1.07 (m) and 1.15 (d) (6H, 2CH₃), 3.87 (s, 3H, OCH₃), 4.07 (s, 2H, CH₂Ph), 4.62 (s, 2H, NCH₂N), 2.02-2.07, 2.28-2.32, 2.60-2.64, 2.80-2.83, 3.54-3.58 (m, 6H), 6.90 (d, 2H, ArH; *J*=8.80 Hz), 7.30 (d, 2H, ArH; *J*=8.40 Hz), 7.42 (d, 1H, ArH; *J*=8.00 Hz), 7.49-7.51 (m, 1H, ArH), 7.61-7.67 (m, 3H, ArH), 7.78-7.79 (m, 1H, ArH), 8.15-8.17 (m, 2H, ArH), 9.71 (s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 17.89 and 18.93 (2CH₃), 30.16 (CH₂Ph), 56.03 (OCH₃), 65.76 (NCH₂N), 71.02 (2CH), 110.95; <u>113.96 (2CH)</u>, 121.19; 123.65; <u>127.49</u>; 128.43; 129.04 (2CH); <u>129.73 (2CH)</u>; 129.86 (2CH); 132.49; 134.48; 141.95; 150.27; <u>158.16</u> (ArC), 145.22 (Triazole C₃), 151.37 (Triazole C₅), 153.02 (N=CH), 163.79 (COO).

4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-p-chlorobenzyl-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoate (2f)

Yield: 72.13%, m.p. 127°C. IR (KBr, v, cm⁻¹): 1740, 1705 (C=O), 1596 (C=N), 1252 (COO), 868 (1,4-disubstituted benzenoid ring), 746 and 711 (monosubstituted benzenoid ring). ¹H NMR (400 MHz, DMSO-d₆): δ 1.05-1.14 (m, 6H, 2CH₃), 3.86 (s, 3H, OCH₃), 4.17 (s, 2H, CH₂Ph), 4.62 (s, 2H, NCH₂N), 2.02-2.07, 2.28-2.32, 2.62-2.66, 2.80-2.85, 3.54-3.57 (m,6H), 7.40-7.42 (m, 5H, ArH), 7.48-7.50 (m, 1H, ArH), 7.58 (m, 1H, ArH), 7.63-7.67 (m, 2H, ArH), 7.78-7.79 (m, 1H, ArH), 8.14-8.17 (m, 2H, ArH), 9.72 (s, 1H, N=CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 17.88 and 18.93 (2CH₃), 30.34 (CH₂Ph), 55.05 and 55.65 (2CH₂), 56.03 (OCH₃), 65.81 (NCH₂N), 71.02 (2CH), 110.95; 121.22; 123.65; 128.44; 129.04(2CH); 129.86 (2CH); <u>130.59 (2CH)</u>; <u>130.76 (2CH)</u>, <u>131.51</u>; 132.41; 134.18; <u>134.77</u>; 141.98; 150.26 (ArC), 144.61 (Triazole C₃), 151.37 (Triazole C₅), 153.16 (N=CH), 163.77 (COO).

Antioxidant Activity

Chemicals

Butylated hydroxytoluene (BHT), ferrous chloride, DPPH., α -tocopherol, 3- butylated hydroxyanisole (BHA), (2-pyridyl)-5,6-bis(phenylsulfonic acid)-1,2,4-triazine (ferrozine) and trichloroacetic acid (TCA) were obtained from E. Merck or Sigma.

Reducing Power

The reducing power of the compounds **2a-i** was determined using the method of Oyaizu (1986). Different concentrations of the samples (50-250 μ g/mL) in DMSO (1 mL) were mixed with phosphate buffer (2.5 mL, 0.2 M, pH = 6.6) and potassium ferricyanide (2.5 mL, 1%). The mixture was incubated at 50°C for 20 min. after which a portion (2.5 mL) of trichloroacetic acid (10%) was added to the mixture, which was then centrifuged for 10 min at 1000 x g. The upper layer of solution (2.5 mL) was mixed with distilled water (2.5 mL) and FeCl₃ (0.5 mL, 0.1%), and then the absorbance at 700 nm was measured in a spectrophometer. Higher absorbance of the reaction mixture indicated greater reducing power.

Free Radical Scavenging Activity

Free radical scavenging effect of the compounds **2a-f** was estimated by DPPH⁻, by the method of Blois (1958). Briefly, 0.1 mM solution of DPPH⁻ in ethanol was prepared, and this solution (1 mL) was added to sample solutions in DMSO (3 mL) at different concentrations (50-250 μ g/mL). The mixture was shaken vigorously and allowed to stand at room temperature for 30 min. Then the absorbance was measured at 517 nm in a spectrophometer. Lower absorbance of the reaction mixture indicated higher free radical scavenging activity. The DPPH⁻ concentration (mM) in the reaction medium was calculated from the following calibration curve and determined by linear regression (R: 0.997):

Absorbance = $0.0003 \times \text{DPPH}^{-} - 0.0174$

The capability to scavenge the DPPH radical was calculated using the following equation:

DPPH. scavenging effect (%) = $(A_0 - A_1/A_0) \times 100$

where A_0 is the absorbance of the control reaction and A_1 is the absorbance in the presence of the samples or standards.

Metal chelating activity

The chelating of ferrous ions by the compounds **2a-f** and references was measured according to the method of Dinis et al., (1994). Briefly, the synthesized compounds (30–60 µg/mL) were added to a 2 mM solution of FeCl₂ (0.05 mL). The reaction was initiated by the addition of 5 mM ferrozine (0.2 mL), and then the mixture was shaken vigorously and left standing at room temperature for 10 min. After the mixture had reached equilibrium, the absorbance of the solution was measured at 562 nm in a spectrophotometer. All tests and analyses were run in triplicate and averaged. The percentage of inhibition of ferrozine–Fe²⁺ complex formation was given by the formula: % inhibition = $(A_0 - A_1 / A_0) \times 100$, where A_0 is the absorbance of the control, and A_1 is the absorbance in the presence of the samples or standards. The control did not contain compound or standard.

Antimicrobial Activity

All bacterial and yeast strains were obtained from the company of Microbiological Environmental Protection Laboratories (France) and were as follows: *Bacillus Substilis* (ATCC 11774), *Bacillus Cereus* (ATCC 11778), *Pseudomonas aeruginosa* (ATCC 27853), *Klebsiella pneumonia* (ATCC 4352), *Staphylococcus aureus* (ATCC 6538) and *Escherichia coli* (ATCC 25922). Simple susceptibility screening test using agar well diffusion method was used (Perez, Pauli, & Bazerque, 1990, Ahmad, Mehmood, & Mohammed, 1998). All the newly synthesized compounds were weighed and dissolved in dimethylsulphoxide (DMSO) to prepare extract stock solution of 1 mg/ml.

Each microorganism was suspended in Mueller-Hinton Broth and diluted to 106 colony forming unit (cfu) per ml. They were "flood-inoculated" onto the surface of Mueller Hinton Agar and then dried. Five-millimeter diameter wells were cut from the agar using a sterile cork-borer, and 250–5000 μ g/50 μ l of the chemical substances were delivered into the wells. The plates were incubated for 18 h at 35 °C. Antimicrobial activity was evaluated by measuring the zone of inhibition against the test organism. Ampicillin, neomycin and streptomycin were standard antibacterial and antifungal agents, DMSO was used as solved control.

Results and Discussion

In this study, the structures of six new 4-[1-(2,6-dimethylmorpholin-4-yl-methyl)-3-alkyl(aryl)-4,5-dihydro-triazol-5-on-4-yl-azomethine]-2-methoxyphenyl benzoates (**2a-f**) were characterized with IR, ¹H NMR and ¹³C NMR spectral data.

Antioxidant Activity

The antioxidant capacities of nine newly synthesized compounds **2a-f** were determined. Different processes have been used to identify antioxidant capacities. The processes used in the paper are clarified below:

Reducing Power

The reducing power of the compounds 2 was determined. The reductive capabilities of compounds are assessed by the extent of conversion of the $\text{Fe}^{3+}/\text{ferricyanide}$ complex to the $\text{Fe}^{2+}/\text{ferrous}$ form. The reducing powers of the compounds were observed at different concentrations, and results were compared with BHA, BHT and α -

tocopherol. The reducing capacity of a compound may serve as a significant indicator for its potential antioxidant activity (Meir, Kanner, Akiri, & Philosoph-Hadas, 1995). The antioxidant activity of a putative antioxidant has been attributed to various mechanisms, among which are prevention chain initiation, binding of transition metal ion catalyst, decomposition of peroxides, prevention of continued hydrogen abstraction, reductive capacity and radical scavenging (Yildirim, Mavi, & Kara, 2001). In this study, all of the amounts of the compounds showed lower absorbance than blank. Hence, no activities were observed to reduce metal ions complexes to their lower oxidation state or to take part in any electron transfer reaction. In other words, compounds did not show the reductive activities.

DPPH Radical Scavenging Activity

Free radical scavenging effect of the compounds **2** was estimated by DPPH radical model. The effect of antioxidants on DPPH radical scavenging was thought to be due to their hydrogen donating ability (Baumann, Wurn, & Bruchlausen, 1979). DPPH is a stable free radical and accepts an electron or hydrogen radical to become a stable diamagnetic molecule (Soares, Dinis, Cunha, & Almeida, 1997). The reduction capability of DPPH radicals was determined by decrease in its absorbance at 517 nm induced by antioxidants. BHT, BHA and α -tocopherol were used as a reference to antioxidant compounds. Compounds **2a-f** did not show any ability.

Iron Binding Capacity

The chelating of ferrous ions by the compounds **2** and references was measured. Ferrozine can quantitatively form complexes with Fe^{2+} . In the presence of chelating agents, the complex formation is disrupted with the result that the red colour of the complex is decreased. Measurement of colour reduction therefore allows estimation of the chelating activity of the coexisting chelator (Yamaguchi, Ariga, Yoshimura, & Nakazawa, 2000). The transition metals ions play an important role as catalysts of oxidative process, leading to formation of hydroxyl radicals and hydroperoxide decomposition reaction via Fenton chemistry (Halliwell, 1996). The production of these radicals may lead to lipid peroxidation, protein modification and DNA damage. Chelating agents are effective as secondary antioxidants because they potentially inhibit the metal-dependent processes thereby stabilizing the oxidized form of the metal ion (Finefrock, Bush, & Doraiswamy, 2003). Iron binding activities of the compounds **2**, BHT, BHA and α -tocopherol are shown in Figure 2.

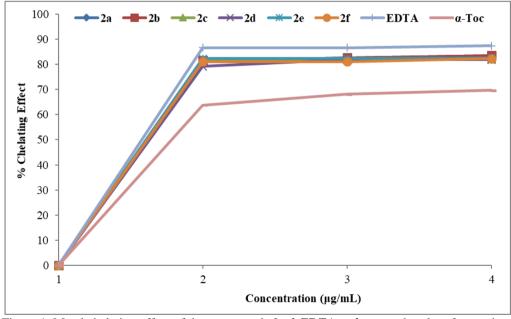


Figure 1. Metal chelating effect of the compounds 2a-f, EDTA and α -tocopherol on ferrous ions

In the current paper, high iron binding capacity of synthesized compounds would be beneficial in retarding metal-chelating oxidation. The data acquired from Figure 1 disclose that the metal chelating effects of all the

compounds (2) were significant and concentration-dependent. The metal chelating effect of the compounds and references decreased in order of EDTA > $2a > 2b = 2e > 2c = 2d > 2f > \alpha$ -tocopherol which were 86.6, 82.7, 82.3, 82.3, 81.9, 81.9, 81.1, 68.1 (%), at the 30 µg/mL, respectively.

Antimicrobial Activity

The microbiological results are summarized in Table I. Microbiology results are promising; all the compounds showed very good antimicrobial activity against to Bacillus subtilis (ATCC-11774) and Bacillus cereus (ATCC-11778). However, not all compounds showed any activity against other microorganisms.

Code	Compounds	Bs	Bc	Pa	Кр	Sa	Ec
1	2a	18	21	-	-	-	-
2	2b	21	23	-	-	-	-
3	2c	22	20	-	-	-	-
4	2d	18	16	-	-	-	-
5	2e	19	17	-	-	-	-
6	2f	21	20	-	-	-	-
А	Ampicillin X3261	33	36	36	35	37	34
Ν	Neomycin X3385	17	17	17	16	13	16
S	Streptomycin X3385	12	12	12	11	21	10

Table 1. Antimicrobial activity of the compounds 2.

Bs: Bacillus subtilis (ATCC-11774), Bc: Bacillus cereus (ATCC-11778), Pa: Pseudomonas aeruginosa (ATCC-27853), Kp: Klebsiella pneumoniae (ATCC-4352) Sa: Staphylococcus aureus (ATCC-6538), Ec: Escherichia coli (ATCC-25922), Amp.: Ampicillin (X3261), Neo.: Neomycin (X3360), Str.: Streptomycin (X3385).

Conclusion

In the present study, new Mannich bases derivatives with 1,2,4-triazole moiety (**2a-f**) were designed and synthesized. Their structures were identified using IR, ¹H NMR and ¹³C NMR spectral data. The entire target compounds were also investigated for their antioxidant and antimicrobial potential. All of the compounds demonstrate a marked ability for metal chelating activity. The data reported with regard to the observed metal chelating activities of the studied compounds could prevent redox cycling. The results may also give several advices for the improvement of new triazole-based therapeutic target. Also, The synthesized compounds showed very good antimicrobial activity against to *Bacillus subtilis* and *Bacillus cereus*.

References

- Ahmad I., Mehmood Z., & Mohammed F., (1998). Screening of some Indian Medicinal Plants for Their Antimicrobial properties. Journal of Ethnopharmacology, 62, 183-193.
- Aktas-Yokus, O., Yuksek, H., Gursoy-Kol, O., & Alpay-Karaoglu, S. (2015). Synthesis and biological evaluation of new 1,2,4-triazole derivatives with their potentiometric titrations. *Medicinal Chemistry Research*, 24(7), 2813–2824. doi: 10.1007/s00044-015-1334-8
- Baumann, J., Wurn, G., & Bruchlausen, V. (1979). Prostaglandin synthetase inhibiting O₂ radical scavenging properties of some flavonoids and related phenolic compounds. *Naunyn-Schmiedebergs Archives of Pharmacology*, 308, R27.
- Blois, M. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*, *181*(4617), 1199–1200. http://doi.org/10.1038/1811199a0
- Chen, D., Zhai, X., Yuan, Q. H., Luo, J., Xie, S. C., & Gong, P. (2010). Synthesis and in vitro anti-hepatitis B virus activity of 1H-benzimidazol-5-ol derivatives. *Chinese Chemical Letters*, 21(11), 1326-1329. doi: 10.1016/j.cclet.2010.05.011
- Chidananda, N., Poojary, B., Sumangala, V., Kumari, N. S., Shetty, P., & Arulmoli, T. (2012). Facile synthesis, characterization and pharmacological activities of 3,6-disubstituted 1,2,4-triazolo[3,4-b][1,3,4]thiadiazoles and 5,6-dihydro-3,6-disubstituted-1,2,4-triazolo[3,4-b][1,3,4]thiadiazoles. *European Journal of Medicinal Chemistry*, 51, 124–136. http://doi.org/10.1016/j.ejmech.2012.02.030

- Dinis, T. C. P., Madeira, V. M. C., & Almeida, L. M. (1994). Action of phenolic derivatives (acetaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid peroxidation and as proxyl radical scavengers. Archives of Biochemistry and Biophysics, 315(1), 161–169. http://doi.org/10.1006/abbi.1994.1485
- Finefrock, A. E., Bush, A. I., & Doraiswamy, P. M. (2003). Current status of metals as therapeutic targets in Alzheimer's disease. *Journal of the American Geriatrics Society*. 51(8), 1143–1148. http://doi.org/10.1046/j.1532-5415.2003.51368.x
- Halliwell, B. (1996). Antioxidants: The Basics-what they are and how to evaluate them. Advances in *Pharmacology*, 38(C), 3–20. http://doi.org/10.1016/S1054-3589(08)60976-X
- Hamama, W. S., Zoorob, H. H., Gouda, M. A., & Afsah, E. M. (2011). Synthesis and antimicrobial and antioxidant activities of simple saccharin derivatives with N-basic side chains. *Pharmaceutical Chemistry Journal*, 45(2), 118–124. doi: 10.1007/s11094-011-0573-3
- Koca E., Yıldız Ç., & Köçek N. (2010). 3-m-Klorobenzil-4-amino-4,5-dihidro-1H-1,2,4-triazol-5-on ve Bazı Türevlerinin Sentezi. Bitirme Tezi, Kakkas Ünv. Fen Edebiyat Fakültesi Kimya Bölümü, Kars.
- Köksal, M., & Bilge, S. S. (2007). Synthesis and antidepressant-like profile of novel 1-aryl-3-[(4benzyl)piperidine-1-yl]propane derivatives. Archiv Der Pharmazie, 340(6), 299–303. doi: 10.1002/ardp.200700028
- Liu, D., Yu, W., Li, J., Pang, C., & Zhao, L. (2013). Novel 2-(E)-substituted benzylidene-6-(N-substituted aminomethyl)cyclohexanones and cyclohexanols as analgesic and anti-inflammatory agents. *Medicinal Chemistry Research*, 22(8), 3779–3786. doi: 10.1007/s00044-012-0362-x
- Maddila, S., & Jonnalagadda, S. B. (2012). New class of triazole derivatives and their antimicrobial activity. *Letters in Drug Design & Discovery*, 9(7), 687–693. doi: 10.2174/157018012801319526.
- Meir, S., Kanner, J., Akiri, B., & Philosoph-Hadas, S. (1995). Determination and involvement of aqueous reducing compounds in oxidative defense systems of various senescing leaves. *Journal of Agricultural* and Food Chemistry, 43(7), 1813–1819.
- McClements, D., & Decker, E. (2000). Lipid oxidation in oil-in-water emulsions: Impact of molecular environment on chemical reactions in heterogeneous food systems. *Journal of Food Science*, 65(8), 1270–1282. http://doi.org/10.1111/j.1365-2621.2000.tb10596.x
- Nithinchandra, Kalluraya, B., Aamir, S., & Shabaraya, A. R. (2012). Regioselective reaction: Synthesis, characterization and pharmacological activity of some new Mannich and Schiff bases containing sydnone. *European Journal of Medicinal Chemistry*, 54, 597–604. doi: 10.1016/j.ejmech.2012.06.011
- Oyaizu, M. (1986). Studies on products of browning reaction. Antioxidative activities of products of browning reaction prepared from glucosamine. *The Japanese Journal of Nutrition and Dietetics*, 44(17), 307–315. http://doi.org/10.5264/eiyogakuzashi.44.307
- Ozkan-Daguyan, I., Sahin, F., & Koksal, M. (2013). Synthesis, Characterization and Antimicrobial Activity of Novel 3,5-Disubstituted-1,3,4-oxadiazole-2-ones. *Revista de Chimie -Bucharest- Original Edition-*, 64(5), 534–539.
- Pati, H. N., Das, U., Kawase, M., Sakagami, H., Balzarini, J., De Clercq, E., & Dimmock, J. R. (2008). 1-Aryl-2-dimethylaminomethyl-2-propen-1-one hydrochlorides and related adducts: A quest for selective cytotoxicity for malignant cells. *Bioorganic & Medicinal Chemistry*, 16(10), 5747–5753. doi: 10.1016/j.bmc.2008.03.060
- Perez C., Pauli M., & Bazerque P., (1990). An Antibiotic Assay by the Well Agar Method. Acta Biologia et Medicine Experimentalis, 15,113-115.
- Savariz, F. C., Formagio, A. S. N., Barbosa, V. A., Foglio, M. A., Carvalho, J. E. de, Duarte, M. C. T., ... Sarragiotto, M. H. (2010). Synthesis, antitumor and antimicrobial activity of novel 1-substituted phenyl-3-[3-alkylamino(methyl)-2-thioxo-1,3,4-oxadiazol-5-yl] β-carboline derivatives. *Journal of the Brazilian Chemical Society*, 21(2), 288–298. doi: 10.1590/S0103-50532010000200014.
- Soares, J. R., Dinis, T. C. P., Cunha, A. P., & Almeida, L. M. (1997). Antioxidant activities of some extracts of Thymus zygis. Free Radical Research, 26(5), 469–478. http://doi.org/10.3109/10715769709084484
- Tramontini, M., & Angiolini, L. (1994). Mannich Bases: Chemistry and Uses. CRC Press.
- Sriram, D., Yogeeswari, P., Dinakaran, M., & Sowmya, M. (2008). Synthesis, anti-HIV and antitubercular activities of nelfinavir diester derivatives. *Biomedicine and Pharmacotherapy*, 62(1), 1–5. doi: 10.1016/j.biopha.2007.08.002
- Yamaguchi, F., Ariga, T., Yoshimura, Y., & Nakazawa, H. (2000). Antioxidative and anti-glycation activity of garcinol from Garcinia indica fruit rind. *Journal of Agricultural and Food Chemistry*, 48(2), 180–185. http://doi.org/10.1021/jf990845y
- Yildirim, A., Mavi, A., & Kara, A. A. (2001). Determination of antioxidant and antimicrobial activities of Rumex crispus L. extracts. *Journal of Agricultural and Food Chemistry*, 49(8), 4083–4089.

http://doi.org/10.1021/jf0103572

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