



## Investigation of Organic Solvents' Effects on Kenaf (*Hibiscus cannabinus* L.) Biomass Conversion in Subcritical Water

Bahar Meryemoğlu<sup>1\*</sup>  , Arif Hasanoğlu<sup>1</sup>  , Mehtap Kurtuluş<sup>1</sup>  ,  
Sibel Irmak<sup>2</sup>  

<sup>1</sup> Department of Chemistry, Çukurova University, 01330, Adana, Turkey

<sup>2</sup> Department of Biological Systems Engineering, University of Nebraska-Lincoln, Lincoln, NE 68583, USA

**Abstract:** Kenaf biomass was hydrolyzed under subcritical water conditions in the presence of various organic solvents. The solvents tested were tetrahydrofuran (THF), acetone, xylene (mixed isomers) and methanol. The organic compounds released into hydrolysates, total organic contents, water-soluble total phenols, and the molecular weight distributions of the polysaccharides in the hydrolysates, solid residues leftover after hydrolysis and gaseous products formed during the solubilization process were determined. The results showed that organic solvents significantly enhanced the dissolution of kenaf biomass (methanol < (omp)xylene ≤ acetone ~ tetrahydrofuran). The hydrolysis percentage was found to be between 75-82% depending on the type of the solvent. Hydrolysis yield and total organic carbons released into hydrolysates highly differed when the solubilization process was performed under carbon dioxide pressure and this effect considerably varied based on the type of solvent used in hydrolysis process. The main gas product formed during hydrolysis process was carbon dioxide with ~80% composition. Morphological measurements of the solid biomass residues left after hydrolysis showed substantial degradations with increasing number of pores on the biomass surfaces.

**Keywords:** Biomass, Kenaf, Organic Solvent, Hydrolysis, Subcritical water.

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**\*Corresponding author. E-mail:** [meryemoglubahar@gmail.com](mailto:meryemoglubahar@gmail.com).

### INTRODUCTION

The need for energy has been increasing continuously as a result of the rapid increase of the world's population. There is growing interest worldwide in the utilization of renewable sources for fuels, materials, and chemicals due to the depletion of fossil resources and environmentally non-friendly nature of the synthetic products. Lignocellulosic biomass can substitute for fossil resources in the production of a wide range of value-added products such as biofuels, bioproducts, and chemicals. Lignocellulosic biomass materials are abundant, cheap, and renewable resources and their non-edible alternatives are particularly important since they do not compete with the food related raw materials in conversion into useful products (1).

The complex and rigid structures of lignocellulosic biomass require an effective pretreatment before breaking down into soluble components with the processes of hydrolysis. The methods of pretreatment can be physical, physicochemical, chemical, or biological (2). Extent of lignin and hemicellulose removals, reduction in cellulose crystallinity, and increasing the porosity of the biomass structure depend on the pretreatment method applied (1). Subcritical water is a liquid under pressure and in the temperature range of 100-374 °C. This liquid usually is used in the extraction of plants. The optimal conditions are determined by changing the pressure and temperatures to obtain the maximum efficiency of the material to be extracted. Significant changes occur in the physical and chemical properties of the water, especially in the dielectric constant ( $\epsilon$ ) under high pressure and

temperature. The extraction efficiency was equivalent to the supercritical fluid or solvent extraction yield even without reaching the critical temperature point. The extraction with subcritical water began as an alternative to supercritical fluid extraction and solvent extraction. There are many advantages to using subcritical water. Some of these are environmental friendly, inexpensive, easy to find, non-toxic, and produces no organic waste at all. Subcritical water hydrolysis is an alternative pretreatment method to break down lignocellulosic biomass by operating process temperature and pressure conditions. This method is totally environmentally friendly and uses water in as reaction medium. The maximum solubilization yields of biomass materials with this method were found to be 70-75% at 250 °C (3,4). Organic solvents play an important role in enhancing yields of the process in many applications. Several studies for biomass conversion were performed with this method using various organic solvents in the reaction medium (5-11). For this purpose, the present study was designed to solubilize lignocellulosic biomass in subcritical water by addition of non-polar (omp-xylene), aprotic polar (tetrahydrofuran, acetone) and polar protic (methanol) solvents into reaction medium. Thus, the partial solubility of lignin will be improved in this study.

## EXPERIMENTAL SECTION

### Hydrolysis of Kenaf biomass

Kenaf biomass (*Hibiscus cannabinus* L.) was hydrolyzed under subcritical water condition. The amount of 10 g of kenaf and 350 mL of water were placed into a 500 mL stainless steel high pressure reactor (Parr Model 4575 HP/HT, Parr Instrument Co., Moline, IL, USA). Then, the reactor was heated until 250 °C and pressurized with/without CO<sub>2</sub> using ISCO 260D pump (Isco Inc., Lincoln, Nebraska, USA) to 27.58 MPa for 2 h. After 2 h, the reactor was cooled up at room temperature within the reactor. The experiments with organic solvents were performed by addition of 3% of THF, acetone, xylene or methanol into reactor. After experiment, kenaf hydrolysate and solid residue were collected for analysis. The solid residue were dried at 100 °C in order to determine the percentage of hydrolysis. The experiments were performed in duplicate.

### Analysis

The solid residues leftover after hydrolysis were characterized by FTIR using ATR (Perkin Elmer Spectrum RX-I FTIR System) and SEM analysis (ZEISS SUPRA 55). The hydrolysates were analyzed by TOC (total organic carbon analyzer), UV-VIS, GC-MS, and HPLC. Total organic carbon content was determined using Tekmar Dohrmann Apollo 9000 instrument.

The compositions of volatile organic compounds in the hydrolysates were determined by a Thermo Finnigan Trace Gas Chromatograph and Mass

Spectrometer (GC-MS) using Thermo TR-5 MS capillary column (60 m x 0.25 mm ID x 0.25 mm film thickness). For this analysis, 50 mL of kenaf hydrolysate was extracted with diethyl ether and dried through a Na<sub>2</sub>SO<sub>4</sub> column. Diethylether was removed by using a rotary evaporator. The oven temperature of the GC-MS system was as follows: holding at 40 °C for 5 min; increasing the temperature from 40 °C to 280 °C with 2.5 °C/min heating rate and holding at this temperature for 10 min. Inlet temperature was 240 °C. The 70 eV and 240 °C were set as ionization voltage and ion source temperature, respectively. The 1 µL of sample was injected in splitless mode. Solvent delay was 6 min. The NIST 2002 mass spectral library was used in identification.

The molecular weight distributions of the polysaccharides in the hydrolysates were determined by gel permeation chromatography (GPC) using 4400, 9900, 21,400, 43,500, 124,000, 196,000, 277,000 and 401,000 Da dextran standards. The hydrolysates were filtered through 0.22 µm syringe filter before analysis. GPC analysis was performed by an LC-6AD Shimadzu high performance liquid chromatograph equipped with SIL-10AF Shimadzu auto injector (Shimadzu, Kyoto, Japan) and Shimadzu RID-10A refractive index detector (RID).

Total water-soluble phenolics contents of the hydrolysates were determined by Folin-Ciocalteu assay (12). Absorbance at 765 nm was recorded using a spectrophotometer (Thermo Scientific Genesys 10S UV/Vis).

## RESULTS AND DISCUSSION

### Hydrolysis yields of kenaf biomass under different water-organic solvents mixture

Kenaf samples were subjected to omp-xylene (non-polar), THF (aprotic polar), acetone (aprotic polar) and methanol (polar protic) solvents under subcritical water condition under pressure of carbon dioxide. It is known that polar aprotic solvents are good solvents for lignin solubilization.

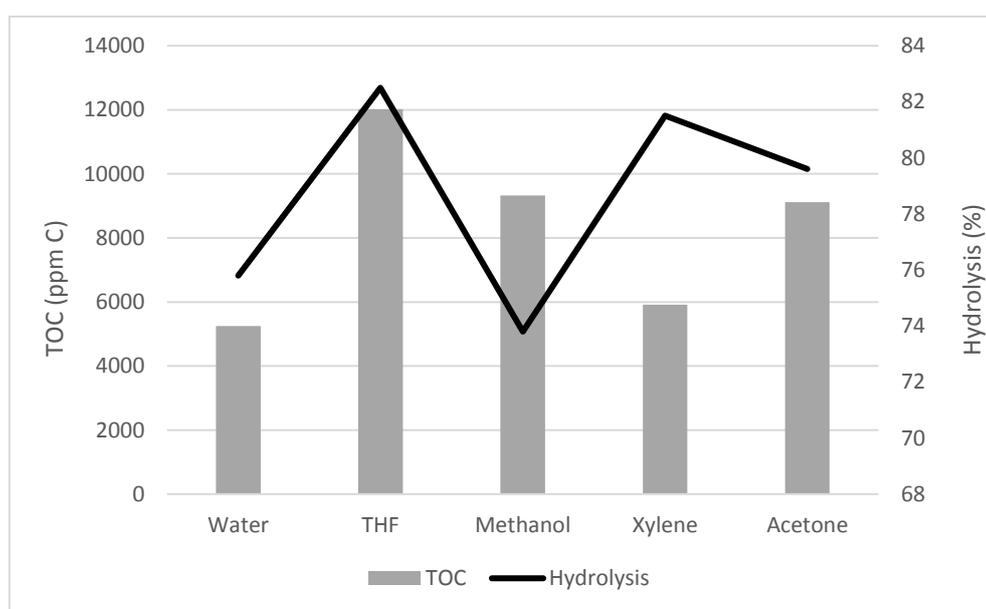
Some experiments were performed to examine the effect of the amount of solvent on the percentage hydrolysis and product distribution using THF. The 1%, 3% and 5% of THF were used. The percentage hydrolysis was increased with increasing THF amount from 1% (75,1±4,4) to 3% (82,5±3,6). When the %5 of THF was used, it was seen that the percentage hydrolysis was decreased (78,8±2,7). Similarly, the phenolics concentrations of hydrolysates were also affected by the amount of THF. According to results, the phenolics concentrations of hydrolysates increased in following order 2021,9 <2832,1 <3213,1 ppm for %1, %5 and %3 of THF respectively. It was decided that %3 organic solvent was more suitable because of the highest

percentage hydrolysis and phenolic contents, less gas formation in the hydrolysis process, more polysaccharide.

The percentage hydrolysis and total organic carbon (TOC) values obtained after these treatments were presented in Figure 1. As given in Figure 1, the percentage hydrolysis increased in the following order; methanol < omp-xylene ≤ acetone ~ tetrahydrofuran. The hydrolysis with non-polar and aprotic polar solvents was observed to be better compared to one performed with a polar protic solvent. Previous studies about solubility of lignin in the mixtures of water-organic solvents showed that our results are in harmony (13-16). The organic solvents are known to affect reaction rate, reaction mechanism, and yield and product distributions in many reaction systems. The changes in dipole moment and the hydrogen bonds between solvent and solute (biomass) can significantly change dissolution process, thermodynamic state of the reactants, activation energy of the products, and compositions of the reaction

mixture (17). It was observed that yield of hydrolysis and TOC released highly differed when the solubilization process was performed under carbon dioxide pressure and this effect considerably varied as based on the type of solvent used in hydrolysis process (Figure 1). The maximum TOC released was observed when THF solvent was used in solubilization media.

During hydrolysis process, gasification reactions can also take place. It is preferable to be gasification less but dissolution more. To determine solvent effect correctly, amount of gas formed during hydrolysis must be determined along with hydrolysis percentage and total organic carbon contents of the hydrolysates. The amount of gas mixtures formed during hydrolysis were determined by GC using TCD detector and presented in Table 1. The amount of the gas was 745 mL when acetone was used in hydrolysis in the absence of carbon dioxide pressure. The gas amounts in other solvents were ranged between 525 and 630 mL.



**Figure 1.** The percentage hydrolysis and TOC yield of kenaf biomass with different water-organic solvent mixture.

The main product in the gas mixtures was found to be carbon dioxide (78-83%) that followed by carbon monoxide (14-19%) (Table 1). Hydrogen

composition was between 2.1-2.8% in all hydrolysis experiments. The gas mixtures contained trace amount of methane (0.2%).

**Table 1.** The gas composition of kenaf biomass with different water-organic solvent mixture (CO<sub>2</sub> pressure was not applied).

Solvents	Gas Volume (mL)	Gas composition (%)			
		H <sub>2</sub>	CH <sub>4</sub>	CO <sub>2</sub>	CO
Water	630±30	2.3±0.8	0.2±0.1	78.5±1.1	19.0±1.1
Water+ 3 % THF	525±35	2.5±0.1	0.2±0.0	80.6±0.5	16.7±0.8
Water+ 3 % Methanol	650±30	2.8±0.2	0.3±0.1	80.9±0.4	16.0±0.6
Water+ 3 % Acetone	745±15	2.7±0.3	0.3±0.1	82.7±1.2	14.3±0.4
Water+ 3 % Omp-Xylene	670±20	2.1±0.3	0.2±0.1	81.0±2.1	16.7±0.9

Based on these results, we can conclude that THF is the best organic solvent for hydrolysis process resulting the highest hydrolysis percentage (82.5%) and TOC (> 10,500 ppm) values by producing the least gas volume (525 mL).

### Products

The products in the hydrolysates were characterized by GC-MS, UV-Vis and GPC analytical techniques. Table 2 shows the compounds formed after hydrolysis of kenaf biomass in different water-organic solvent mixtures. Carbon dioxide was used as a pressurizing gas in the hydrolysis process. Differences in the viscosity, the dielectric constant, polarity of the solvents (18,19) affected the solubilization of biomass.

In many studies, lignocellulosic material was treated with organic solvents and as a result of this treatment, most of lignin was removed (20-22). And also, higher hemicellulose conversion was obtained with use of organic solvent such as dimethylformamide (DMF) in the processes (23).

Although these studies were conducted, the comparison of organic solvents on kenaf subcritical water hydrolysis has not been reported yet. The use of organic solvents in hydrolysis partially hydrolyzes the lignin bonds and lignin-carbohydrate bonds, and solid residues leftover after hydrolysis mainly consists of cellulose and hemicellulose. In this study, the organic solvent removes the lignin from the lignocellulosic material but most of the hemicellulose sugars are also dissolved in this process. Because of that the compounds were mainly phenolics which were released from lignin fraction of biomass. Water-soluble phenolic composition of the hydrolysates varies depending on the solvent used in experiments (Table 3). The phenolics concentrations were highest in water-xylene mixture. The total phenolics in this solvent mixture were found to be 3601.1 mg/L. On the other hand, water-THF mixture yielded the lowest phenolics in the hydrolysis (Table 3). From our work, it was found that the use of organic solvents in hydrolysis process provided enhancing the product distribution as well as hydrolysis yield.

**Table 2.** GC-MS analysis of kenaf hydrolysates in different water- organic solvent mixtures

Retention time (min)	Compound Name	MS fragments used in identification (m/z)
15.69	Furfural	96 (M <sup>+</sup> ), 95, 68, 67, 51
18.7	Corylon (2-hydroxy-3-methyl-2-cyclopenten-1-one)	112 (M <sup>+</sup> ), 97, 83, 41, 27
22.3	2-methoxyphenol	124 (M <sup>+</sup> ), 109, 95, 81, 53
28.06	3-methyl-1,2-cyclopentadione	112 (M <sup>+</sup> ), 97, 83, 69, 55
31.23	4-methyl phenol	110, 108 (M <sup>+</sup> ), 107, 79, 53
36.8	2-6-dimethoxyphenol (Syringol)	154 (M <sup>+</sup> ), 139, 96, 65, 51
40.27	Hydroxymethylfurfural	126, 97(M <sup>+</sup> ), 81, 69, 53
47.21	2-methoxy- 3-methyl hydroxy quinone	154 (M <sup>+</sup> ), 139, 93, 68, 65
49.81	Vanillin	152, 151 (M <sup>+</sup> ), 123, 109, 81
51.5	4-hydroxy-3,5-dimethoxybenzaldehyde (syringyl aldehyde)	182 (M <sup>+</sup> ), 181, 111, 93, 65
56.25	7-acetyl-2,3,4,5,6,7-hexahydrobenzofuran-4-one	180, 138, 137 (M <sup>+</sup> ), 122, 94
67.0	1,2-diphenyl propan-2-one	210, 192 (M <sup>+</sup> ), 168, 167, 123
79.59	2 2'-methylenebis 6-(1 1-dimethylethyl)-4-methyl-phenol	340, 284, 177 (M <sup>+</sup> ), 161, 149

**Table 3.** Water-soluble phenolic contents and polysaccharide distribution of kenaf hydrolysates

Solvents	Phenolic content ( mg/L)	Polysaccharides Mp, Da
Water	1431.1	6975; 25
Water+ 3 % THF	3213.1	6075; 3332
Water+ 3 % Methanol	3415.3	68477; 22390; 24
Water+ 3 % Acetone	3562.8	58519; 19705; 25
Water+ 3 % omp-Xylene	3601.1	70496; 23116; 23

The results showed that organic solvents affect the hydrolysis and molecular mass distribution of the polysaccharides in the hydrolysates. Kenaf biomass had three fractions of polysaccharides after hydrolysis (Table 3). These fractions are also associated with phenolic fragments in the biomass. Type of the organic solvent used in hydrolysis process was the main factor on the

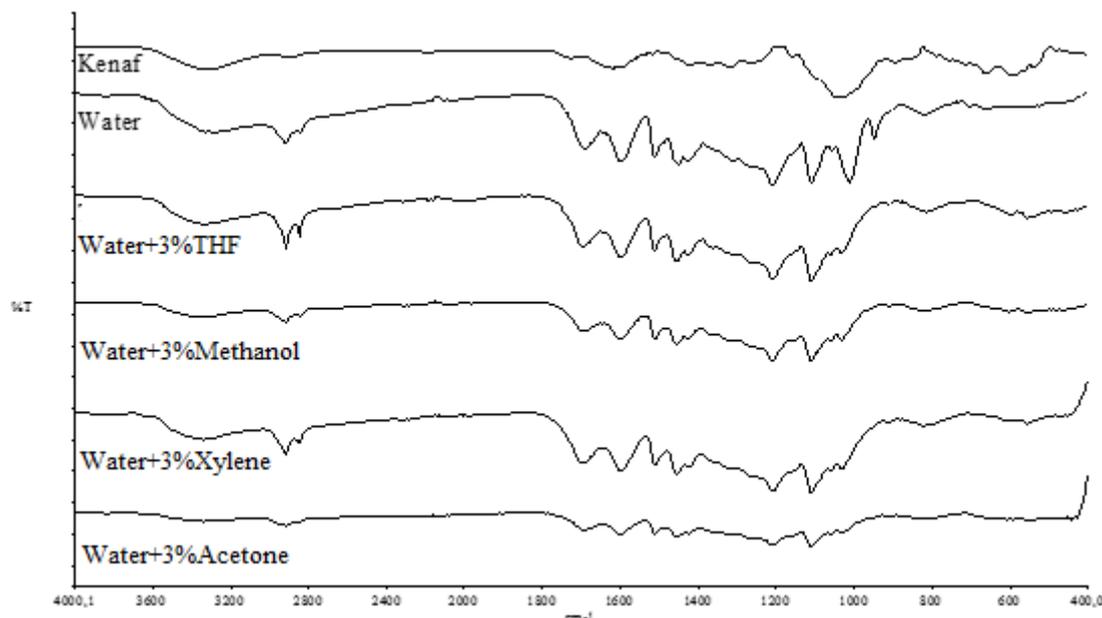
differences of polysaccharides distributions (Table 3).

### Characterization of kenaf samples after hydrolysis

Chemical changes that occurred in the structure of kenaf biomass were determined by taking FTIR spectra of the samples before and after

hydrolysis. Kenaf biomass spectra before and after treatments were given in Figure 2. The 3340-3350  $\text{cm}^{-1}$  in the original kenaf infrared spectrum was assigned to O-H vibrations. This band belongs to hydroxyl groups located together by hydrogen bonds between the repeating units in the cellulose matrix. The band between 2890-

2920  $\text{cm}^{-1}$  belongs to the C-H stretching. The 1742  $\text{cm}^{-1}$  band is due to stretching of the carbonyl group in the hemicellulose structures (24). The functional group C = O was observed at 1600-1650  $\text{cm}^{-1}$ . This band takes place in the lower region of 1700  $\text{cm}^{-1}$  because of water absorption in the structure (25,26).

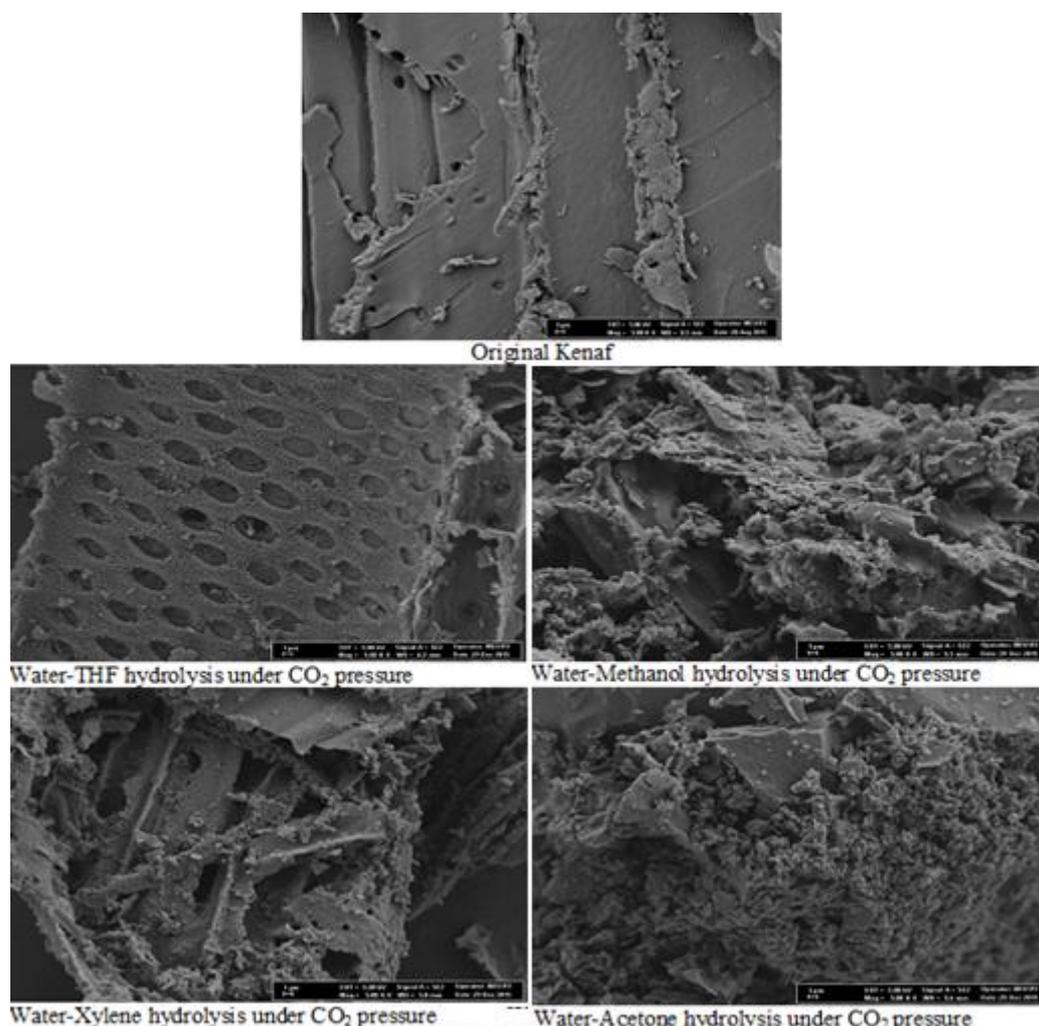


**Figure 2.** FTIR spectra of kenaf after hydrolysis with different water- organic solvent mixtures.

The weak bands seen around 1500-1514  $\text{cm}^{-1}$  belong to lignin (25,26). The C-H vibrations in the cellulose at 1429  $\text{cm}^{-1}$  and C-O-C antisymmetric vibration in glycosidic band at 1161  $\text{cm}^{-1}$  were observed. The 1370  $\text{cm}^{-1}$  is assigned to stretching of C-H peaks in cellulose (26). The band at 1160-1000  $\text{cm}^{-1}$  is due to the C-O vibrations. The bands that belong to typical xylan hemicellulose structures were observed between 1175 and 1000  $\text{cm}^{-1}$  (25,26). The weak band around 1109  $\text{cm}^{-1}$  indicates stress of glucose ring in cellulose and the presence of a band at 890  $\text{cm}^{-1}$  in original kenaf attributed to  $\beta$ -glycosidic bonds in the cellulose structure (27,28).

The band at 1742  $\text{cm}^{-1}$  in the original kenaf sample disappeared after hydrolysis since acetyl, uronic, and ferulic ester bonds in hemicellulose fraction were completely broken (Figure 2). The cleavages of ester bonds caused releases of phenolic compounds (27, 26). The guaiacyl aromatic C-O band stretchings in lignin structure could be seen at 1510  $\text{cm}^{-1}$  (29). These bands became prominent in the non-hydrolyzed kenaf

treated with any solvents. The 1460  $\text{cm}^{-1}$  and 1320  $\text{cm}^{-1}$  show absorption bands of syringyl ring in lignin structure (29-30). The morphological changes in kenaf samples after hydrolysis were also examined by taking SEM images (Figure 3). The original kenaf structure was straight and it consisted of thin fibrils. The SEM image of non-hydrolyzed kenaf indicated that dense and compact outer surface were reduced by treatments. The formations of a number of pores having a few micrometer diameters were observed after hydrolysis process with solvents (Figure 3). Substantial degradation with increasing number of pores in the rigid structure of kenaf increased after hydrolysis with omp-xylene, acetone and methanol. THF hydrolysis resulted in more degradation in the kenaf structure. The cell wall structure was completely fragmented after hydrolysis in xylene. Some droplets which are known to be lignin based fragments released after lignin degradation were observed as a result of hydrolysis of the lignocellulosic material (33-35).



**Figure 3.** SEM images of kenaf samples after hydrolysis in different water- organic solvent mixture.

## CONCLUSION

Use of organic solvents in the subcritical water hydrolysis process increased the dissolution of kenaf biomass. However, product distribution in the hydrolysates did not considerably change. Non-polar (omp-xylene) and aprotic polar (THF, acetone) solvents enhanced the hydrolysis more than polar protic (methanol) solvent. The hydrolysis process with organic solvents generated soluble polysaccharides which could be utilized for production of various value-added products including biofuels and bioproducts by thermochemical and biochemical conversion processes.

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