

Pathogenesis & Laboratory approach to Thrombophilia

Renu Saxena*, Monica Sharma

Department of Hematology, All India Institute of Medical Sciences, New Delhi, India

Abstract. Thrombophilia is a term used for any hypercoagulable state, either inherited or acquired. The former is considered after excluding acquired predisposing causes like trauma, immobility, Dirseminated into vascular coagulation, pregnancy and anitphospholipid syndrome etc. It frequently results from interplay of genetic and acquired factors. An individual's risk for DVT would be determined by the combination of his or her baseline propensity for thrombosis and the magnitude of the acute insult. Inherited hypercoagulable states may be secondary to deficiency of natural clotting inhibitors or elevated procoagulants or increased fibrinolytic factors. Amongst these, activated protein C resistance, is the commonest underlying cause. Testing for thrombophilia is best performed in stages. Highest-yield assays (screening tests) should be performed first and, if positive, should be followed by appropriate confirmatory tests. Cornerstone of initial treatment is heparin, either unfractionated or low molecular weight, followed by oral anticoagulation.

Key words: Thrombophillia, inherited

1. Introduction

Thrombophilia is defined as an increased tendency to thrombosis. It may be acquired or inherited. The latter is considered after excluding acquired predisposing causes like trauma, immobility, DIC, pregnancy and anitphospholipid syndrome etc. Thrombosis may be arterial or venous. Venous thromboembolism (VTE) is the commonest manifestation of a thrombophilic state and approximately 25% of Thrombophilia is detected in over 50% of cases following a first clinical episode of VTE. Inherited hypercoagulable states may be secondary to deficiency of natural clotting inhibitors or elevated procoagulants or increased fibrinolytic factors.

Amongst these, *activated protein C* (APC) resistance, is the commonest underlying cause and is seen in 20-50% patients with inherited thrombophilia.

2. Prevalence of thrombophilia

Thrombophilia is lowest in unselected patient population and highest in patients with a personal and/or family history of thrombophilia. Reported prevalence of prothrombin gene mutation and factor V Leiden varies according to geographical distribution, prevalence of both of these mutations is much lower in Asian populations unlike in European populations (2:200-7:100 & 2:100-3:100 respectively) (7). In one cohort family study, the overall incidence of VTE (per 100 patient-years) was found to be 1.07 for antithrombin deficiency, 0.54 for protein C deficiency, 0.50 for protein S deficiency, and 0.30 for activated protein C resistance. Likewise for acquired causes the risk is much higher following hip or knee surgery than during pregnancy; the latter, in turn, poses a much higher risk than prolonged air travel (9).

3. Thrombosis threshold

An individual's risk for DVT would be determined by the combination of his or her baseline propensity for thrombosis and the magnitude of the acute insult. In the face of increased baseline hypercoagulability (e.g., factor V Leiden) even a relatively weak insult (e.g., blood stasis during a flight) can be sufficient to precipitate DVT (10). Likewise, in an individual

*Correspondence: Dr. Renu Saxena

Professor and Head Department of hematology All India Institute of Medical Sciences, Ansari Nagar, New Delhi-110029 India

E-mail: renusaxena@hotmail.com

with a relatively low level of baseline genetic hypercoagulability (e.g., a single mutation that is associated with a low risk of thrombosis) a relatively strong thrombogenic event (e.g., pregnancy) would be required to provoke an episode of VTE. Thus, the precipitating event in such individuals is often clinically overt. In most cases, such thrombophilic individuals never suffer VTE throughout their lifetimes, and when they do have an episode, it is unlikely to recur. In contrast, an individual with a high level of baseline genetic hypercoagulability (with multiple thrombophilic mutations or polymorphism) is at such high risk that relatively

minor acquired triggers can initiate a thrombotic episode. These triggers are therefore subclinical, giving the appearance that the patient has “idiopathic”, “spontaneous”, or “unprovoked” VTE. Furthermore, VTE in these high-risk individuals is more likely to recur (9).

4. Aetiology of thrombophilia

Hypercoagulable state or thrombophilia arises when there is an abnormality in blood coagulability which is the result of interplay of acquired and inherited factors or several inherited factors.

Table 1. Causes of Hypercoagulable state/Thrombophilia

Primary hypercoagulable states (thrombophilias)	Acquired hypercoagulable state
1. Decreased antithrombotic proteins_ Antithrombin deficiency Protein C deficiency Protein S deficiency 2. Increased prothrombotic proteins Factor V Leiden (activated protein C resistance) Prothrombin gene mutation G20210A Increased levels of factors VII, XI, IX, VIII, von Willebrand factor	1. Vascular disorders Atherosclerosis Diabetes Prosthetic materials (grafts, valves, indwelling vascular catheters) 2. Abnormal rheology Stasis (immobilization, surgery, congestive heart failure) Hyperviscosity (polycythemia vera, Waldenstrom macroglobulinemia, acute leukemia, sickle cell disease) 3. Other disorders associated with hypercoagulability Cancer (Trousseau syndrome) Cancer chemotherapeutic agents, thalidomide Oral contraceptive, estrogen therapy, selective estrogen receptor modulators Pregnancy Infusion of prothrombin complex concentrates Nephrotic syndrome Myeloproliferative disorders Paroxysmal nocturnal hemoglobinuria Inflammatory bowel disease Thrombotic thrombocytopenic purpura Disseminated intravascular coagulation Antiphospholipid antibody syndrome Heparin-induced thrombocytopenia/thrombosis

Approach to thrombophilia

Clinical presentation and diagnosis

- significant percentage of women associated with thrombosis related to pregnancy or oral contraceptive use, have an inherited disorder
- inherited thrombotic disorders are usually associated with venous thromboembolism

History and physical examination

Complete history taking is an essential part of evaluation of patient with thrombosis and should include:

- age of onset
- details about the circumstances proximate to the thrombotic event like surgery, trauma, pregnancy, immobility, estrogen therapy etc.
- women should be asked about any bad obstetric history, hormone replacement therapy and use of oral contraceptive use
- details of any prior thrombotic event.

Table 2. The important pointers for inherited thrombophilia are

Most common	Less common
Thrombosis at a younger age	Recurrent pregnancy loss
Recurrent thrombosis	Pre-eclampsia – HELLP
Family history of thrombosis	Vitamin K antagonist-induced skin necrosis
Thrombosis in an unusual site	Neonatal purpura fulminans
	Heparin resistance

- detail family history as history of thrombosis in first degree relative strongly suggest a hereditary defect.
- presence of constitutional symptoms, as thrombosis may be the first manifestation of a malignancy
- presence of an underlying disease like cancer, collagen vascular disease, myeloproliferative disease, atherosclerosis, nephrotic syndrome etc
- intake of drugs like hydralazine or procainamide
- History of recurrent thrombosis in spite of oral anticoagulation which suggests presence of occult neoplasm or recurrent cancer.

Physical examination should be directed to the

- Vascular system
- Extremities
- Chest
- Heart
- Abdominal organs

Laboratory approach to thrombophilia

Whom do we investigate for inherited thrombophilia?

- Young patients with age <45 years with recurrent thrombosis
- Young patients with a single thrombotic event but has a positive family history
- Thrombosis in unusual sites
- Heparin resistance
- Warfarin-induced skin necrosis
- Thrombosis occurring with estrogen therapy or pregnancy

When to investigate for inherited thrombophilia?

Heparin therapy may be associated with upto 30% reduction in plasma level of ATIII levels over several days; warfarin produces marked reduction in functional protein C and protein S and rarely elevates the level of ATIII. Therefore testing is done when the patient has fully recovered from the acute event and is off anticoagulant therapy. Ideally these tests are carried out 2 weeks after completing initial 3-6 months of anticoagulant therapy. Normal levels during the acute phase effectively exclude deficiency of these proteins.

Laboratory tests

- Complete blood count for evidence of polycythemia vera, essential thrombocythemia, PNH and heparin-induced thrombocytopenia
- Examination of the peripheral smear for evidence of schistocytes (to rule out disseminated intravascular coagulation, thrombotic thrombocytopenic purpura / hemolytic uremic syndrome). Leucoerythroblastic picture suggests involvement of bone marrow by tumour.
- Liver and renal function tests and urinalysis: Patients with Budd-Chiari Syndrome & Nephrotic syndrome respectively.
- Baseline coagulation tests including PT, APTT for evidence of lupus anticoagulants.
- Tests for thrombophilia

Testing for thrombophilia is best performed in stages. Highest-yield assays (screening tests) should be performed first and, if positive, should be followed by appropriate confirmatory tests.

Table 3. Recommended laboratory evaluation for patients suspected of having an underlying hypercoagulable state

Activated protein C resistance	Factor V Leiden PCR
Antithrombin, protein C, and protein S activity (functional) levels	
Antigenic assays for antithrombin, protein C and/or protein S	
Screening tests for lupus anticoagulant (sensitive aPTT, aPTT mixing studies, dilute Russell viper venom time)	Anti β 2 GP1 antibodies, ELISA for Acl antibodies Confirmation: platelet neutralization procedure, Textarin/Ecarin test, platelet vesicles, DVV confirm (at least one of these)
Fasting total plasma homocysteine level	
Prothrombin G20210A mutation testing by PCR	

Protein C deficiency

- *Inherited deficiency of protein C* a vitamin K dependent anti coagulant protein, which inactivates activated FV and FVIII. Homozygous deficiency presents as life threatening neonatal thrombosis or purpura fulminans. Heterozygous deficiency is more common and is seen in 1 to 5% of general population.

Normal plasma concentration is 4 μ g / ml. The logarithm of the values for protein C antigen determined in the normal adult population is usually in the range of 70-140%. Levels <55% are likely to be associated with genetic defect. Protein C levels in newborns are 20-40% of adult levels. The estimated increased lifetime relative risk of venous thrombosis has been reported to be 31-fold. (16) Protein C activity less than 0.68 U/ml, measured at the time of thrombosis in individuals without a known hypercoagulable state, has been associated with increased rates of recurrent venous thrombosis.

Protein S deficiency

- Protein S exists in the plasma in two forms: bound to C4b-binding protein (60% of total protein S) and free (40% of total). The total protein S antigen in normal adults is 23 μ g/ml. Levels increase with increasing age.

Up to 25% of patients with protein S deficiency may experience arterial thrombosis including stroke (17). The estimated lifetime increased relative risk of thrombosis has been reported to be as high as 36-fold for protein S deficiency. *Type I protein S deficiency* is a quantitative disorder in which protein S functional activity, total antigen, and free antigen levels are equally reduced to approximately 50% of normal. *Type IIa protein S deficiency* with preserved normal levels of total Protein S. but a selective deficiency of free Protein S and reduced Protein S functional activity. *Type IIb protein S*

deficiency, the levels of both total and free protein S antigen are normal with reduced Protein S functional activity. Apparent Type IIb protein S deficiency has been seen in patients with APC-R.

In infections and other stress situation, C4b levels increase as C4b is an acute phase reactant. Therefore, since more protein S binds to C4b, the free Protein S become falsely low. Thus if one gets low levels of Protein S, in a patient, it is important to exclude underlying infections & repeat the test after about 1-2 weeks for clearing of infection, before labeling it as inherited Protein S deficiency.

Diagnosis of protein S and C deficiency

- Patients should not be diagnosed as protein S deficient until APC-R is excluded.
- Patients with heterozygous protein C & S deficiency have normal PT and aPTT values whereas those of homozygous protein C deficiency have abnormal coagulation tests consistent with DIC.

Pro C^R Global test (Dade Behring, Maburg, Germany) is an effective screening test for protein C and protein S deficiency. It is a coagulometric test based on activation of protein C of test sample by Protein C activating protease from venom *Agkistrodon contortrix*. The resulting prolongation of the clotting time then reflects the functioning of the Protein C anticoagulant pathway [13]. The time taken for clot formation is determined as PCAT (Protein C Activity dependent Clotting Time). This assay is recommended as screening for F V Leiden related APCR, Protein C, Protein S & AT III deficiencies.

Protein C assay Functional (chromogenic) & antigenic (ELISA) assays are available.

The *Protein S assay* both functional and immunologic assays are available. Functional assay may be PT-or aPTT-based, measuring inhibition of factor Va by APC this measures free

protein S activity. ELISA measures total protein S levels.

AT III deficiency

- *Deficiency of AT III*, which inactivates thrombin, is found in 0.2% general population. It has an autosomal dominant inheritance and constitutes 5-10% of DVT patients.
- Type I of inherited ATIII deficiency is a result of reduced synthesis of

biologically normal protease inhibitor molecule and type II results from molecular defect within the protease inhibitor. Diagnosis is made by immunologic(ELISA) and functional assays.

Functional assays assess quantitative as well as qualitative abnormality of ATIII and can be done either by coagulation or by chromogenic methods.

Table 4. Causes of acquired deficiencies in antithrombin III, protein C, and protein S

Antithrombin III	Protein C	Protein S
Neonatal period	Neonatal period	
Pregnancy		
Liver disease	Liver disease	Liver disease
DIC	DIC	DIC
Nephrotic syndrome	Chemotherapy (CMF)	
Major surgery		Inflammatory states
Acute thrombosis	Acute thrombosis	Acute thrombosis
Treatment with heparin, L-asparaginase	Treatment with warfarin, L-asparaginase	Treatment with warfarin, L-asparaginase

The normal range of ATIII is between 0.75 and 1.25 IU/ml in adults; in ATIII deficiency, it is less than 0.7 IU/ml. Healthy newborns have concentrations lower than the in adults (0.6 to 0.8 IU/ml) and reach adult level by age 6 months. In ATIII deficient neonates, the level has been found to be less than 0.3 IU/ ml.

A variety of pathophysiologic conditions can reduce the concentration of ATIII, Protein C and Protein S in the blood, which one should keep in mind while evaluating their deficiencies.(TABLE 4)

Activated protein C resistance (APC-R)

Activated Protein C Resistance (APC-R) is the most common laboratory abnormality in patients with history of thromboembolism. Factor V Leiden accounts for 92% of cases of APC-R; rest 8% is due to pregnancy, OCP use, antiphospholipid antibody syndrome, plasma glucosylceramide deficiency and other factor V mutations. Heterozygous carriers of FVL have a 2- to 10-fold increased lifetime relative risk of developing VTE. This is further increased by pregnancy (9-fold), OCP (36-fold) and HRT (13-16-fold) (8). Homozygous carriers are estimated to have an 80-fold increased lifetime relative risk of VTE. (19).

Specific clinical associations with APC-R:

- Recurrent miscarriages: 20% of second trimester pregnancy loss is associated with APC-R
- Children with thrombosis
- Cerebral venous thrombosis
- Myocardial infarction in young women; risk increases to 30-fold if associated with smoking

Laboratory evaluation of APCR is done by clotting assays based on inhibition of factor Va by APC and prolongation of clotting time. In individuals with APC resistance, there is very little prolongation of APTT since FVa is not inactivated due to mutation in FV cleavage site leading to failure of APC to recognize it. APC is added to patient plasma, and a clotting assay is performed (PTT), with results expressed as a ratio of patient and normal:

$$\frac{\text{Patient aPTT} + \text{APC}}{\text{Patient aPTT} - \text{APC}}$$

APC-R can also be done by ELISA.

APCR may vary due to variation in reagents/laboratories it is therefore advocated that it be expressed as normalized APC ratio (nAPCR), wherein APC ratio of normal is performed in pooled plasma from at least 20 normals. This improves its accuracy and precision. nAPCR= APC ratio of patient/APC ratio of normal

APC resistance is said to be present when nAPCR <0.76; if found positive, molecular studies may be done for FV Leiden mutation.

APCR may be due to increased FVIII/V levels, inhibitors to APC or mutation at any of the 3 cleavage sites of FVa. In order to ensure specificity of APCR for FV mutation defects, it is advisable to use FV deficient plasma in a 'Modified Dahlback's test' wherein FV deficient plasma increase the specificity of the test for FV mutations. In cases with APCR, diagnosed by modified Dahlback's test, Polymerase chain reaction for FV Leiden may be performed.

Prothrombin gene mutation

- Recently, a *genetic defect in prothrombin gene* leading to prothrombin 20210 G →A mutation has been found in 1-2% of normals. As a result of this mutation, these patients have FII levels > 115iu/dl which increases the relative risk of VTE 2-5 times, further increases with pregnancy (15fold), OCP use (16fold)
- PG20210 A can be detected by molecular PCR technique, which can be performed despite concomitant anticoagulant therapy.

Hyperhomocysteinemia

- *Hyperhomocysteinemia* This may be due to mutations in MTHFR gene or due to deficiency of vitamin B12, B6, or folic acid.

Inherited severe hyperhomocysteinemia result from homozygous MTHFR and CBS deficiency or inherited errors of cobalamin deficiency. *Inherited mild to moderate homocysteinemia* results from the thermolabile variant of MTHFR (t1MTHFR) that is encoded by the C677T gene polymorphism.

Acquired hyperhomocysteinemia result from folic acid deficiency, vitamin B₆ and vitamin B₁₂ deficiency, renal insufficiency, hypothyroidism, inflammatory bowel disease, advanced age, carcinoma of the breast, ovaries, pancreas, ALL, drugs like methotrexate, theophylline, phenytoin.

Hyperhomocysteinemia (plasma level >18.5 micromol/L) has been associated with 2-4-fold increased VTE risk. For evaluation fasting total plasma homocysteine is measured first (normal level 5-15 μm/L) followed by testing the same 2-8 hours after an oral methionine load (100mg/kg); the later increases the sensitivity of occult B6 deficiency and obligate heterozygotes for CBS deficiency. Vitamin B₁₂ and folic acid do not

affect post-methionine homocysteine levels. With raised homocysteine level vitamin B₁₂ level should be measured before starting treatment.

Elevated F VIII levels (>150 IU/dl) confer a 6-fold increased risk of DVT. Increased thrombosis in patients with elevated FVIII levels is often seen in patients with O Blood group and vWF levels >150 IU/dl.

Conclusion

Investigation for idiopathic thrombophilia is essential for appropriate prophylactic and therapeutic anticoagulation, though only 30-40% have an underlying inherited cause 60% are idiopathic.

References

1. Egeberg O. Inherited antithrombin deficiency causing thrombophilia. *Thromb Diath Haemorrh* 1965; 13: 516-530.
2. Schafer AI: hypercoagulable states: molecular genetics to clinical practice. *Lancet* 1994; 344: 1739.
3. Steven R. Deitcher and George M. Rodgers, *Thrombosis and antithrombotic therapy*, Wintrobe's Clinical Hematology 2004; 1713-1758.
4. Andrew I Schafer et al. *Thrombotic disorders: Diagnosis and treatment: 2003 American Society of Hematology Education Programme Book*.
5. Jose A. Lopez *Deep Vein Thrombosis*. ASH et al. education program book, 2004.
6. Steven R Lentz. *Thrombophilia*. Carver College of Medicine, University of Iowa, May 2003
7. Steven R. Deitcher et al , *Hypercoagulable states*, The Cleveland Clinic, 2003
8. College of American Pathologists' Consensus Panel on Thrombophilia. *Arch Pathol Lab Med* 2001; 126: 1277-1433.
9. Gupta PK, Ahmed R, Kannan M, et al *Pro C^R Global: An Effective Screening Test for Thrombophilia*. *Am J Hematol* Issue 2003; 74: 208-210.
10. Dati F, Hafner G, Erbes H, Prellwitz W, Kraus M, Niemann F, Naoh M, Wagner C. *Pro C^R Global : the first functional screening assay for the complete protein C pathway*. *Clin Chem* 1997; 43: 1719-23.
11. Kenneth A. Bauer, *Hypercoagulable states; Hematology Basic Principles and Practice*, 3rd edition 2001: 2009-2039.
12. Martinelli I, Mannucci PM, De Stefano V, et al. Different risks of thrombosis in four coagulation defects associated with inherited thrombophilia: a study of 150 families. *Blood* 1998; 92: 2353-2358.
13. Sie P, Boneu B, Bierme R, et al. Arterial thrombosis and protein S deficiency of protein S. *Blood* 1995; 85: 3518-3523.

14. Rosendaal FR, Koster T, Vandembroucke JP et al. Reitsma PH. High risk of thrombosis in patients homozygous for factor V Leiden (activated protein C resistance). *Blood* 1995; 85:1504-1508.
15. Gerhardt A, Scharf RE, Beckmann MW, et al. Prothrombin and factor V mutations in women with history of thrombosis during pregnancy and puerperium. *N Engl J Med* 2000; 342: 374-380.
16. Rosendaal FR, Siscovick DS, Schwartz SM, et al. Factor V Leiden (resistance to activated protein C) increases the risk of myocardial infarction in young women. *Blood* 1997; 89: 2817-2821.
17. Kandice Kottke- Merchant, Alexander Duncan. Antithrombin deficiency: Issues in Laboratory Diagnosis. *Arch Pathol Lab Med* 2002; 126: 13261-336.