

Larvicidal Activity of Some Medicinal Plants Naturally Growing in Turkey Against *Aedes albopictus* (Diptera: Culicidae)

Emine KILIÇKAYA SELVİ^{1*} Asu USTA¹ M. Mustafa AKINER²

¹Department of Chemistry, Faculty of Arts and Sciences, Recep Tayyip Erdogan University, 53100 Rize, TURKEY.

²Department of Biology, Faculty of Arts and Sciences, Recep Tayyip Erdogan University, 53100 Rize, TURKEY

*  <https://orcid.org/0000-0003-0291-5362> ¹  <https://orcid.org/0000-0002-1826-9599> ²  <https://orcid.org/0000-0002-7658-3236>

Received date: 21.02.2018

Accepted date: 11.04.2019

How to cite: Kılıçkaya Selvi, E., Usta, A. & Akiner, M.M. (2019). Larvicidal activity of some medicinal plants naturally growing in Turkey against *Aedes albopictus* (Diptera: Culicidae). *Anatolian Env. and Anim. Sciences*, 4(1), 53-59. Doi: <https://doi.org/10.35229/jaes.530139>
Atf yapmak için: Kılıçkaya Selvi, E., Usta, A. & Akiner, M.M. (2019). Türkiye’de doğal olarak yetişen bazı tıbbi bitkilerin *Aedes albopictus* (Diptera: Culicidae)’a karşı larvasidal aktiviteleri. *Anadolu Çev. ve Hay. Dergisi*, 4(1), 53-59. Doi: <https://doi.org/10.35229/jaes.530139>

Abstract: Massive use of chemical based insecticides led to detrimental effects on the public health and environment. Environment friendly and biodegradable natural products of plant origin alternatives to synthetic larvicids have received attention as agents for vector control. Thus, many researchers have studied on alternative methods for pest control instead of chemical ones.

In this study, some medicinal valuable plants naturally growing in Turkey were selected for this purpose and the larvicidal effect against Asian tiger mosquito *Aedes albopictus* larvae was investigated. Bioassays were carried out by applying late 3th to early 4th instar larvae of *Aedes albopictus* to two different dose of plant extracts. The larval mortality was counted after 24 and 48 h after experiments. Bioassay results revealed that *Leucanthemum vulgare* was the most effective for ethyl acetate extract in 500 ppm concentration, 100 % for 24 h, 100 % for 48 h. *Salvia verticillata* induced a second high mortality after 24 h treatment for two doses of both extracts. *Inula vulgaris* and *Matricaria chamomilla* produced high and moderate mortality for 500 ppm dose of both extracts, respectively.

Our results evidenced that the high larvicidal activity was detected products obtained from crude extracts with ethyl acetate and methyl alcohol of *Salvia verticillata*, *Leucanthemum vulgare*, *Inula vulgaris* and *Matricaria chamomilla*. But there is a need for further studies about possible candidate species for new and safer control products against mosquito control.

Keywords: *Aedes albopictus*, larvicidal activity, mosquito control, plant extracts.

Türkiye’de Doğal Olarak Yetişen Bazı Tıbbi Bitkilerin *Aedes albopictus* (Diptera: Culicidae)’a Karşı Larvasidal Aktiviteleri

Öz: Kimyasal insektisitlerin yoğun kullanımı halk sağlığı ve çevre üzerinde olumsuz etkilere sebep olmuştur. Bu durum, sentetik larvasitlere alternatif, bitki kökenli çevre dostu ve biyobozunur doğal ürünler vektör kontrolünde dikkat çekmiştir. Bu nedenle birçok araştırmacı, kimyasal olanlar yerine haşere kontrolü için alternatif yöntemler üzerinde çalışmıştır.

Bu çalışmada, Türkiye’de doğal olarak yetişen bazı tıbbi bitkiler, bu amaç için seçilmiş ve Asya kaplani sivrisinek *Aedes albopictus* larvalarına karşı larvasit etkileri araştırılmıştır. Biyoassaylar, *Aedes albopictus*’un geç 3. ile erken 4. evre larvalarının iki farklı bitki ekstraktına uygulayarak gerçekleştirilmiştir. Larva ölümleri, deneylerden 24 ve 48 saat sonra sayılmıştır. Biyoassay sonuçları, *Leucanthemum vulgare*’nin etil asetat özütünün 500 ppm’lik konsantrasyonda 24 saatte % 100, 48 saatte % 100 oranında en etkili olduğunu gösterdi. *Salvia verticillata*, her iki ekstraktın iki dozu için 24 saat uygulamadan sonra ikinci yüksek ölüm oranı yaratmıştır. *Inula vulgaris* ve *Matricaria chamomilla*, her iki ekstraktın 500 ppm’lik dozu için sırasıyla yüksek ve orta ölüm oranı göstermiştir.

Sonuçlarımız, yüksek larvasidal aktiviteye, *Salvia verticillata*, *Leucanthemum vulgare*, *Inula vulgaris* ve *Matricaria chamomilla*’nın etil asetat ve metil alkollü ham özütlerinden elde edilen ürünlerin sahip olduğunu göstermiştir. Ancak, sivrisinek kontrolüne karşı yeni ve daha güvenli kontrol ürünleri için olası aday türler hakkında daha fazla araştırma yapılması gerekir.

Anahtar sözcükler: *Aedes albopictus*, larvasidal aktivite, sivrisinek kontrolü, bitki ekstraktı.

INTRODUCTION

Mosquitoes are responsible for transmitting several infectious diseases like malaria, yellow fever, dengue, West Nile encephalitis, filariasis, Zika, Chikungunya (Nauen, 2007; Kovendan et al., 2012; Govindarajan et al., 2013). For this reason, public health is becoming a challenging problem and mosquitoes and their transmitting diseases have a serious social and economic impact for many countries in tropical and subtropical areas (Bossche & Coetzer, 2008). The distribution of mosquitoes is dynamic in space and time and their life cycles are influenced by environmental conditions (Crans, 2004). Climate change seems to be a driving force for establishing new invasion areas for invasive *Aedes* mosquitoes (Caminade et al., 2012, Campbell et al., 2015). Mosquito-borne diseases are endemic in more than 100 countries all over the world, and new ones are added every year (Ogden 2018). They are causing serious public health problems and mortality of two million people every year. More than two billion people are at risk around the world related to mosquito-borne diseases (Klempner et al., 2007).

In Turkey, various genera of mosquitoes, such as *Aedes*, *Anopheles*, *Culex* are important for mosquito-borne diseases. Recently *Aedes albopictus* and *A. aegypti* introduced and established populations in Eastern Black Sea part and raises concerns about a possible resurgence of the pathogens that can be transmitted by this vector species (Akner et al., 2016). Increase of the human population and movement are raising the transmission risk of the pathogens especially yellow fever, dengue, Chikungunya, West Nile fever and Zika. Limited funds of mosquito control, lack of awareness among the residents and highly competent behaviour of the invasive vector species resulted in an increase in mosquito-borne diseases (Gubler, 1998). Mosquito control efforts are the best strategy for protection against mosquito-borne diseases. Usage of the synthetic insecticides is the most important method for controlling mosquito and other pests for human and animal health. But, extensive usage of synthetic insecticides causes environmental problems and development resistance. Furthermore, it causes toxic and detrimental/lethal effects on non-target organisms (Roberts & Andre, 1994; Milam et al., 2000; Dinesh et al. 2015, Sakthivadivel et al., 2015). Phytochemicals obtained from plants are mainly biodegradable, non-phytotoxicity and more importantly, they are renewable (Çalmaşur et al., 2006; Boulogne et al., 2012). In this respect, plant and plant-derived substances regain interest to control of insect pests.

Plant secondary products as terpenoids, flavonoids and phenolic compounds which serve as storage compounds in plants are important for plant defence mechanisms against insects and used as an excellent alternative to synthetic or chemical insecticide in many parts of the World (Luthria et al., 1993; Ghosh et al. 2012; Ali et al., 2015; Hikal et al., 2017). Especially, plants with high terpenoid, phenolic compound and alkaloid content are known to be used for this purpose. Therefore, the insecticidal, larvicidal, ovicidal,

antifeedant and repellent activity of many plants belongs to Asteraceae and Lamiaceae families due to the rich terpenoid content in the crude extract or their components have been described in several studies (Çalmaşur et al., 2006; Pavela 2008, Boussaada et al., 2008; Çetin et al., 2009; Govindarajan & Karuppanan, 2011, Boulogne et al., 2012; Cheah et al., 2013).

Salvia verticillata L., *Phlomis lychitis* L., *Salvia tomentosa* Mill., are belonging to the Lamiaceae family. Many species of the genera *Salvia* and *Phlomis* over the Mediterranean region native to Turkey, North Africa, Europe and Asia are used as herbal tea for many decades in folk medicine as sore throats, colds, gastrointestinal disorder among others (Orhan et al., 2007; Tepe et al., 2007; Amor et al., 2009; Lopez et al., 2010; Aşkun et al., 2010). The other five plants (*Matricaria chamomilla* L., *Leucanthemum vulgare* (Vall.) Lam., *Triploarasperrum caucasicum* (Wild.) Haya, *Erigeron annuus* (L.) Pers, *Inula vulgaris* (Lam.) Trevisan) belonging to the Asteraceae family are also subject of chemical screening and bioactivity studies and used for folk medicine (Singh et al., 2011; Kumar et al., 2014; Magharri et al., 2015; Assi et al., 2017). There are a number of reports on the isolation of various secondary metabolites and phytochemical analysis from all of these plants, e.g., monoterpenoids, sesquiterpenoids, diterpenoids, alkaloids, flavonoids and other phenolic compounds (Sefidkon et al., 1999; Chalchat et al., 2001; Pitarokili et al., 2006; Amor et al., 2009; Nazaruk and Kalemba, 2009; Aşkun et al., 2010; Ayoughi et al., 2011; Singh et al., 2011; Kumar et al., 2014; Magharri et al., 2015; Hatipoğlu et al., 2016; Assi et al., 2017; Kim et al., 2018). *Salvia verticillata* L., *Salvia tomentosa*, *Matricaria chamomilla*, *Leucanthemum vulgare* (Vall.) Lam., *Erigeron annuus* (L.) Pers plant extracts have been determined by some researchers in recent years with phytochemical analyses by using classic or spectral methods. According to these studies, the major chemical compositions of Turkish *S. verticillata* L. and *S. tomentosa* plant extracts are monoterpenes such as β -pinene, α -pinene, borneol, camphor and sesquiterpenes such as caryophyllene oxide, α - / β -caryophyllene, γ -muurulone (Chalchat et al., 2001; Pitarokili et al., 2006; Aşkun et al., 2010). The essential oils of *M. chamomilla*, *L. vulgare* (Vall.) Lam. and *E. annuus* (L.) Pers extracts contain α -bisabolol, α - / β -farnesene, guaiazulene, β -caryophyllene, caryophyllene oxide as major components (Ayoughi et al., 2011; Kumar et al., 2014; Magharri et al., 2015; Kim et al., 2018). These components are found in these plants but with different amounts. Differences among chemical compositions of the crude oils or essential oils widely depend on conditions such as climate, variety, origin, time and soil factors.

Many *Salvia* species have been reported to have insecticidal activities. According to these records, methanolic extract of *Salvia verticillata* L. and hexane extract of *Salvia tomentosa* Mill. have moderate larvicidal activity against *Culex tritaeniorhynchus* and *Culex pipiens* (Diptera:

Culicidae), respectively (Pavela, 2008; Gün et al., 2011). Also, essential oils of *Salvia tomentosa* Mill. have insecticidal activity against *Lipaphis pseudobrassicae* (Aphididae: Homoptera), *Spodoptera exigua* (Lepidoptera) (Sampson et al., 2005; Polatoğlu et al., 2017). Khodadad et al. (2007) reported that acaricidal potential against female *Rhipicephalus annulatus* (Ixodidae) of *Matricaria chamomile* ethanolic extract. In another report, ethanolic extract of *Matricaria chamomile* has showed oviposition deterrent, larvicidal and repellent activity against *Culex quinquefasciatus* (Al-Mehmadi & Al-Khalaf 2008; Gad & El-DaKheel 2009). Literature survey has shown that there are no such studies for our other studied plants.

The present study aims to evaluate potential larvicidal activity of eight different plants (Lamiaceae and Asteraceae), are grown naturally in Turkey against *Aedes albopictus* larvae.

MATERIALS and METHODS

Plant material: Fresh aerial parts of *Salvia verticillata* L. and *Salvia tomentosa* Mill. were collected from Antalya in southwest part of Turkey in April 2018 and May 2018, respectively (Table 1). Dried aerial parts of *Phlomis lychitis* L. were purchased from a market in Konya, southwest part of Turkey in May 2018. *Matricaria chamomilla* L., *Leucanthemum vulgare* (vall.) Lam., *Triploaraspium caucasicum* (Wild.) Haya, *Erigeron annuus* (L.) Pers, *Inula vulgaris* (Lam.) Trevisan, were collected from Rize in northeast part of Turkey in May-August 2018 (Table 1) and transported to the laboratory. The identification of the plants was performed by Prof. Vagif Atamov (Faculty of Science and Arts, Recep Tayyip Erdogan University in Rize, Turkey).

Plant Extracts: The solvents (*n*-hexane, CHCl₃, MeOH, AcOEt) used were either of analytical grade or bulk solvents distilled before use. Electric blender was used as a Waring Commercial (CT, USA). Ultrasonic bath was Bandelin Sonorex (Berlin, Germany).

Fresh plant materials were cleaned and washed with distilled water and ethyl alcohol mixture (1:1, v/v). Then, these materials were dried for one week at room temperature with reduced humidity (25 %) and whole plants were powdered using an electric blender. The resulting powder of each plant samples was divided into two flasks, 100g / flask. Extraction procedure is the method developed by Selvi et al., (2018). Each sample was defatted with 100 mL of chloroform at 25 °C for 30 minutes. Two different solvent extractions of each sample were performed with 2 x 400 mL of solvent at room temperature for 2 h in a shaker. The most preferred solvent for phenolic compounds extraction is methanol, because the phenolic composition at plant samples is mostly soluble and stable in this solvent. Ethyl acetate is mostly suitable for flavonoid extraction (Dmitrienko et al. 2012; Mokrani and Madani 2016; Sukeksi and Sarah 2016). Therefore, plant extractions were performed using two different polarities: methanol and ethyl acetate. The crude extracts were filtrated and dried under vacuum in a rotary evaporator at 40°C and then lyophilized. A stock solution of each crude extract was prepared in DMSO and stored below 4 °C until testing for bioassay. Crude oil yields obtained from extractions with different solvents of studied plant have been given in Table 1. In this study, larvicidal activities of 50 and 500 ppm doses of crude extracts obtained from eight plants against *Ae albopictus* (Diptera: Culicidae) have been tested.

Table 1. Plants used in this study, their parts used, origins and crude oil yields.

Plant species	Plant family	Common name	Plant part	Origin and date	Crude oil yield (%)	
					MeOH	EtOAc
<i>Salvia verticillata</i> L.	Lamiaceae	lilac sage	aerial part	Antalya, April 2018	2.58	1.72
<i>Phlomis lychitis</i> L.	Lamiaceae	lampwick plant	aerial part	Konya, May 2018	3.77	2.46
<i>Salvia tomentosa</i> Mill.	Lamiaceae	balsamic sage	aerial part	Antalya, May 2018	2.07	4.73
<i>Matricaria chamomilla</i> L.,	Asteraceae	chamomile	aerial part	Rize, May 2018	2.62	1.97
<i>Leucanthemum vulgare</i> (vall.) Lam.	Asteraceae	ox-eye daisy	aerial part	Rize, July 2018	2.94	2.47
<i>Triploaraspium caucasicum</i> (Wild.) Haya	Asteraceae	vulture weed	aerial part	Rize, July 2018	4.62	4.03
<i>Erigeron annuus</i> (L.) Pers	Asteraceae	fleabane	aerial part	Rize, August 2018	4.55	2.47
<i>Inula vulgaris</i> (Lam.) Trevisan	Asteraceae	ploughman's spikenard	aerial part	Rize, August 2018	2.78	3.33

Test organisms: *Aedes albopictus* was collected from Rize province in July 2018 in around whole sales market (41,0416 Lat 40,5771 Lan) / small industrial area (41,0453 Lat 40,5784 Lan) and used for the larvicidal activity. Whole samples collected larval stages in inside used tires. Alive collected larvae transferred to the laboratory and colonised in the laboratory condition. Mosquitoes were held at 26 ± 2°C, 65 ± 10 % relative humidity and a photoperiod

regime of 14:10 h (L:D) in the insectarium. The larvae were fed on larval food. Adult mosquitoes were periodically fed with 10% glucose solution (Gerbert et al., 1994, Imam et al., 2014). F1 and F2 generation larvae were used for tests.

Larvicidal bioassay: The larvicidal activity against *Ae albopictus* was analysed with minor modifications adopted for the study in standard protocol recommended by the World Health Organization (WHO) (WHO, 2005). The

tests were conducted in 250 mL plastic container. Tested third or early fourth instar mosquito larvae were obtained from laboratory colonized mosquitoes of F1 and F2 generation. From the stock solution, different concentrations 50 and 500 ppm with distilled water were prepared. Twenty healthy larvae were released into each 250 mL plastic container containing 200 mL of water and two hours after test extracts (50-500 ppm) were added the plastic container. Controls made with DMSO solution were performed simultaneously. Larval mortality was observed for 24 and 48 hours. The larval percent mortality was calculated. All tests were done in two replications and control mortality was corrected using Abbott's formula (Abbott, 1925).

RESULTS AND DISCUSSION

The results of larvicidal affects of *S. verticillate* L., *S. tomentosa* (Mill.) extracts showed low level effects on *Ae albopictus* except *S. verticillate* L. methanol and ethyl acetate extracts 500 ppm dose. *S. verticillate* L. bioassay results showed high larval mortality (up to 85 % for 500 ppm dose) but *S. tomentosa* extracts showed low larval mortality (under 15 % for 50 and 500 ppm doses). Similarly, *P. lychnitis* L. extracts showed low level mortality for two doses (under 20 %) (Table 2). Gün et al. (2011) showed larvicidal effects of some *Salvia* species hexane extracts on *Culex pipiens* larvae. They used four different *Salvia* species (*S. tomentosa*, *S. sclerae*, *S. argentea*, *S. syriaca*) and tested against *Cx pipiens* 3th and 4th instar larvae. They indicated that *S. tomentosa* is the most effective *Salvia* species against *Cx pipiens* larvae. In contrast to these results, *S. verticillate* species showed higher larvicidal effect against *Ae albopictus* larvae and gived high mortality rate after 24 and 48 h in our study. *S. tomatosa* mortality rates were very low and also found 15 % after 48 h. Çetin et al. (2006) tested larvicidal activities some Lamiaceae species against *Cx pipiens* larvae. In that study, ethanol extract of the *Salvia sclerae* showed high larvicidal activity and 200 ppm ethanol extract gived 90 % mortality after 24h. Kaliopoulos et al. (2010) reported the larvicidal activity of *Salvia fruticosa* Mill., *Salvia pomifera* L. subsp. *calycina* (Sm.) Hayek and *Salvia pomifera* L. subsp. *pomifera* essential oils against *Cx pipiens* larvae. All these studies have shown different/similar larvicidal activity of many *Salvia* species related to the polar/apolar solvent extracts and mosquito species.

Different daisy extracts results showed moderate and high mortality especially in methanol and ethyl acetate high dose experiments. *M. chamomilla* bioassay revealed that the low level mortality of methanol extracts low dose for two test duration. But, the high dose of this extract gived 75 % mortality for 48 h duration. Ethyl acetate extracts bioassay results showed moderate (ethyl acetate 50 ppm 48 h) and high mortality for two test duration. A similar study of Gad & El-DaKheel (2009) showed that the larvicidal activity of *M. chamomilla* against *Cx quinquefastiatus* larvae and

determined 0.310 mg/L LC₅₀ value for 4th instar larvae. Al-Mehmadi (2011) also reported that the larvacidal activity of *M. chamomilla* extracts against *Cx quinquefastiatus* and indicated that the duration is important for the LC₅₀. They showed 0.505 mg/L after 28 h LC₅₀ value and 0.301 mg/L after 48 h from treatment. Another study of Al-Mehmadi & Al-Khalaf (2008) reported that ethanolic extracts of *M. chamomilla* had oviposition deterrent and skin repellent against *Cx quinquefastiatus*. Mahyoub et al. (2014) reported that larvicidal effects of *M. chamomilla* extract against *Ae aegypti* larvae. They indicated that the concentration is important for the increasing larval mortality against 4th instar *Ae aegypti* larvae. Candido et al. (2013) reported that the high larval and pupal mortality with *R. communis* and *C. phyllacanthus* oils against *Ae aegypti* larvae. They also indicated that the lethality was enhanced with increasing time of exposure to plant products. Our results also indicated that mortality increasing with time.

L. vulgare (vall.) Lam. extracts also showed similar results as *M. chamomilla*, but methanol 500 ppm and ethyl acetate 50 ppm results showed moderate mortality for two test duration. The highest mortality result for this plant was seen at 500 ppm of ethyl acetate extract (100 %, 24 h and 48 h).

T. caucasicum (Wild.) Haya extracts showed low level mortality for whole extracts and duration except ethyl acetate 500 ppm 48h duration (75 % mortality). Almost similar results were found in *E. annuus* (L.) Pers and ethyl acetate 500 ppm 48h duration showed 67.5 % mortality.

I. vulgaris (Lam.) Trevisan methanol and ethyl acetate low doses showed low level mortality for two test duration. In contrast to these results, high doses of methanol extract gived moderate mortality for two test duration and high doses of ethyl acetate extracts gived high mortality for two test duration. In a similar study, He et al. (2014) described larvicidal activity of another *Inula* species (*I. racemosa*) root ethanol extract against *Ae albopictus* larvae. They showed that 25.23 µg/mL LC₅₀ value of ethanol extract.

There are numerous studies on plant extracts against mosquito larvae. Insects and especially mosquitoes control becomes more difficult day by day related to the insecticide resistance. Literature survey has shown that there is no report on larvicidal activity of *P. lychnitis*, *L. vulgare*, *T. caucasicum*, *E. annuus* unlike other tested species. Botanical originated insecticides may have a chance for safe control option beyond the harmful effects of synthetic insecticides and also may help to solve insecticide resistance obstacle for mosquito control. This study indicates that the methanol and ethyl acetate extract of *Salvia* and some daisy species naturally found in our country have a potential to be used for the control of *Ae albopictus*. Therefore, more detailed studies are needed for exact and effective candidate of invasive *Aedes* species recently distributed in Turkey.

Table 2. Mortality rate of *Aedes albopictus* by different concentrations of some plant extracts belonging to Asteraceae and Lamiaceae families.

Concentration (ppm)	% mortality			
	Ethyl acetate		Methyl alcohol	
	24 hour	48 hour	24 hour	48 hour
<i>Salvia verticillata</i> L.				
50	0	7.5	2.5	5
500	90	100	85	90
<i>Salvia tomentosa</i> Mill.				
50	0	0	0	2.5
500	2.5	15	7.5	15
<i>Phlomis lychnitis</i> L.				
50	0	0	2.5	5
500	10	17.5	2.5	2.5
<i>Matricaria chamomilla</i> L.				
50	12.5	57.5	0	25
500	90	95	30	75
<i>Leucanthemum vulgare</i> (vall.) Lam.				
50	50	62.5	7.5	10
500	100	100	45	50
<i>T. caucasicum</i> (Wild.) Haya				
50	17.5	20	2.5	20
500	30	75	10	10
<i>Erigeron annuus</i> (L.) Pers				
50	10	30	15	17.5
500	35	67.5	17.5	22.5
<i>Inula vulgaris</i> (Lam.) Trevisan				
50	10	30	5	27.5
500	75	95	57.5	62.5

REFERENCES

- Abbott, W.S. (1925).** A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology*, **18**, 265-267.
- Akiner, M.M., Demirci, B., Babuadze, G., Robert, V. & Schaffner F. (2016).** Spread of the invasive mosquitoes *Aedes aegypti* and *Aedes albopictus* in the Black Sea Region increases risk of chikungunya, dengue, and zika outbreaks in Europe. *PLOS Neglected Tropical Diseases*, **10**, e0004664.
- Ali, A., Tabanca, N., Demirci, B., Blythe, E.K., Ali, Z., Baser, K.H.C. & Khan, I.A. (2015).** Chemical composition and biological activity of four *Salvia* essential oils and individual compounds against two species of mosquitoes. *Journal of Agriculture and Food Chemistry*, **63**, 447-456.
- Al-Mehmadi, R.M & Al- Khalaf. A. A. (2008).** Oviposition deterrent and skin repellent activities of *Artemisia herba alba*, *Matricaria chamomilla* and *Mellia azedarach* against *C. quinquefasciatus*. *Saudi Journal of Biological Sciences*, **15**, 97-104.
- Al-Mehmadi, R.M. (2011).** Larvicidal, histopathological and ultra – structure studies of *Matricaria chamomell* extract rift valley fever mosquito *Culex quinquefasciatus* (Diptera: Culicidae). *Journal of Entomology*, **8**, 63-72.
- Amor, I.L., Boubaker, J., Sgaier, M.B., Bhourri, I.S.W, Neffati, A., Kilani, S., Bouhlel, I., Ghedira, K. & Chekir-Ghedira, L. (2009).** Phytochemistry and biological activities of *Phlomis* species, *Journal of Ethnopharmacology*, **125**, 183-202.
- Assi, M., Usta, J., Mounimne, Y., Aboul-Ela, M. & El-Lakany, A. (2017).** Phytochemical study and the antiproliferative activity of *Inula vulgaris* species grown in Lebanon. *International Journal of Pharmacy and Pharmaceutical Sciences*, **9**, 75-83.
- Aşkun, T., Başer, K.H.C., Tümen, G. & Kürkçüoğlu, M. (2010).** Characterization of essential oils of some *Salvia* species and their antimycobacterial activities. *Turkish Journal of Biology*, **34**, 89-95.
- Ayoughi, F., Barzegar, M., Sahari, M.A. & Naghdibadi, H. (2011).** Chemical compositions of essential oils of *Artemisia dracuncululus* L. and endemic *Matricaria chamomilla* L. and an evaluation of their antioxidative effects. *Journal of Agriculture Science and Technology*, **13**, 79-88.
- Bossche, V. & Coetzer, J.A. (2008).** Climate change and animal health in Africa. *Revue scientifique et technique*, **27**, 551-562.
- Boulogne, I., Petit, P., Ozier-Lafontaine, H., Desfontaines, L. & Loranger-Merciris, G. (2012).** Insecticidal and antifungal chemicals produced by plants: a review. *Environmental Chemistry Letter*, **10**, 325-347.
- Boussaada, O., Kamel, M.B.H., Ammar, S., Haouas, D., Mighri, Z. & Helal, A.N. (2008).** Insecticidal activity of some Asteraceae plant extracts against *Tribolium confusum*. *Bulletin of Insectology*, **61**, 283-289.
- Caminade, C., Medlock, J.M., Ducheyne, E., McIntyre, K.M., Leach, S., Baylis, M. & Morse, A.P. (2012).** Suitability of European climate for the Asian tiger mosquito *Aedes albopictus*: recent trends and future scenarios. *J. R. Soc. Interface* **9**, 2708-2717.
- Campbell, L.P., Luther, C., Moo-Llanes, D., Ramsey, J.M., Danis-Lozano, R. & Peterson, AT. (2015)** Climate change influences on global distributions of dengue and chikungunya virus vectors. *Phil. Trans. R. Soc. B*, **370**, 20140135.
- Candido, L.P., Cavaltini M.J., Beserra E.B. (2013).** Bioactivity of plant extracts on the larval and pupal stages of *Aedes aegypti* (Diptera, Culicidae). *Revista da Sociedade Brasileira de Medicina Tropical* **46**(4):420-425.
- Chalchat, J.C., Gorunovic, M.S., Petrovic, S.D. & Maksimovic, Z.A., (2001).** Chemical Compositions of Two Wild Species of the Genus *Salvia* L. from Yugoslavia: *Salvia aethiopsis* and *Salvia verficillafa*. *Essential Oil Research*, **13**, 416-418.
- Cheah, S.X., Tay, J.W., Chan, L.K. & Jaal, Z. (2013).** Larvicidal, oviposition, and ovicidal effects of *Artemisia annua* (Asterales: Asteraceae) against *Aedes aegypti*, *Anopheles sinensis*, and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitology Research*, **112**, 3275-3282.

- Crans, W.J. (2004).** A classification system for mosquito life cycles: life cycle types for mosquitoes of the northeastern United States. *Journal of Vector Ecology*, *1*, 1-10.
- Çalmaşur, Ö., Aslan, İ. & Şahin., F. (2006).** Insecticidal and acaricidal effect of three Lamiaceae plant essential oils against *Tetranychus urticae* Koch and *Bemisia tabaci* Genn. *Industrial Crops and Products*, *23*, 140-146.
- Çetin, H., Cinbilgel, I., Yanikoglu, A. & Gokceoglu, M. (2006)** Larvicidal Activity of some Labiatae (Lamiaceae) Plant Extracts from Turkey. *Phytotherapy Research*, *20*, 1088-1090.
- Çetin, H., Çilek, J.E., Aydın, L. & Yanıkoğlu, A. (2009).** Acaricidal effects of the essential oil of *Origanum minutiflorum* (Lamiaceae) against *Rhipicephalus turanicus* (Acari: Ixodidae). *Veterinary Parasitology*, *160*, 359-361.
- Dinesh, D.S., Kumari, S., Pandit, V., Kumar, J., Kumari, N., Kumar, P., Hassan, F., Kumar, V. & Das, P. (2015).** Insecticidal effect of plant extracts on *Phlebotomus argentipes* (Diptera: Psychodidae) in Bihar, India. *Indian Journal of Medicinal Research*, *142*, 95-100.
- Dmitrienko, S.G., Kudrinskaya, V.A. & Apyari, V.V. (2012).** Methods of extraction, preconcentration, and determination of quercetin. *Journal of Analytical Chemistry* *67*, 299-311.
- Gad, A.A. & El-DaKheel, A.A. (2009).** Larvicidal activities of *Cinnamomum osmophloeum* and *Matricaria chamomella* extracts against the filarial mosquito *Culex quinquefasciatus* (Diptera: Culicidae) and their effects on its haemogram. *The Egyptian Science Magazine*, *6*(1/2), 8-15.
- Gerberg, E.J., Barnard, D.R. & Ward, R.A. (1994).** Manual for mosquito rearing and experimental techniques. American Mosquito Control Association.
- Ghosh, A., Chowdhury, N. & Chandra, G. (2012).** Plant extracts as potential larvicides. *Indian Journal of Medical Research*, *135*, 581-598.
- Govindarajan, M., Sivakumar, R., Rajeswary, M. & Yogalakshmi, K. (2013).** Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae). *Experimental Parasitology*, *134*, 7-11.
- Gubler, D.J. (1998).** Resurgent vector-borne diseases as a global health problem. *Emerging Infectious Diseases*, *4*, 1-9.
- Gün, S.S., Cinbilgel, I., Öz, E. & Çetin H. (2011).** Larvicidal activity of some *Salvia* L. (Labiatae) plant extracts against the mosquito *Culex pipiens* L. (Diptera: Culicidae). *Kafkas Üniversitesi Veteriner Fakültesi Dergisi*, *17*(Suppl A), 61-65.
- Hatipoğlu, S.D., Zorlu, N., Dirmenci, T., Gören, A.C., Öztürk, T. & Topçu, G. (2016).** Determination of volatile organic compounds in forty five *Salvia* Species by thermal desorption-GC-MS technique. *Records of Natural Products*, *10*, 659-700.
- He, Q., Liu, X.C., Sun, R.Q., Deng, Z.W., Du, S.S. & Liu, Z.L. (2014).** Mosquito Larvicidal Constituents from the Ethanol Extract of *Inula racemosa* Hook. f. Roots against *Aedes albopictus*. *Journal of Chemistry*, *2014*(Article ID 738796), 1-6.
- Hikal, W.M., Baeshen, R.S. & Said-Al Ahl, H.A.H. (2017).** Botanical insecticide as simple extractives for pest control. *Cogent Biology*, *3*, 1404274.
- Imam, H., Zarnigar, S.G. & Seikh, A. (2014).** The basic rules and methods of mosquito rearing (*Aedes aegypti*). *Trop Parasitol*, *4*(1), 53-55.
- Khodadad, P.K. & Mehdi, R.A. (2007).** Biological activities of chamomile (*Matricaria chamomile*) flowers' extract against the survival and egg laying of the cattle fever tick (Acari Ixodidae). *Journal of Zhejiang University: Science B*, *8*, 693-696.
- Kim, D.Y., Won, K.J., Hwang, D.I., Park, S.M., Kim, H.B. & Lee, H.M. (2018).** Chemical Composition of Essential Oil from *Erigeron annuus* (L.) Pers. Flower and its Effect on Migration and Proliferation in Keratinocyte. *Journal of Essential Oil Bearing Plants*, *21*, 1146-1154.
- Klempner, M.S., Unnasch, T.R. & Hu, L.T. (2007).** Taking a bite out of vector-transmitted infectious diseases. *The New England Journal of Medicine*, *356*, 2567-2569.
- Koliopoulos, G., Pitarokili, D., Kioulos, E., Michaelakis, A. & Tzakou, O. (2010).** Chemical composition and larvicidal evaluation of *Mentha*, *Salvia*, and *Melissa* essential oils against the West Nile virus mosquito *Culex pipiens*. *Parasitology Research*, *107*, 327-335.
- Kovendan, K., Murugan, K., Kumar, A.N., Vincent, S. & Hwang, J.S. (2012).** Bioefficacy of larvicidal and pupicidal properties of *Carica papaya* (Caricaceae) leaf extract and bacterial insecticide, spinosad, against chikungunya vector, *Aedes aegypti* (Diptera: Culicidae). *Parasitology Research*, *110*, 669-678.
- Kumar, V., Mathela, C.S., Tewari, G., Singh, D., Tewari, A.K. & Bisht, K.S. (2014).** Chemical composition and antifungal activity of essential oils from three Himalayan *Erigeron* species. *LWT-Food Science and Technology*, *56*, 278-283.
- López, V., Jäger, A.K., Akerreta, S., Cavero, R.Y. & Calvo, M.I. (2010).** Antioxidant Activity and Phenylpropanoids of *Phlomis lychnitis* L.: A

- Traditional Herbal Tea. *Plant Foods for Human Nutrition*, **65**, 179-185.
- Luthria, D.L., Ramakrishnan, V. & Banerji, A. (1993).** Insect antifeedant activity of furochromones: Structure-activity relationships. *Journal of Natural Products*, **56**, 671-675.
- Magharri, E., Razavi, S.M., Ghorbani, E., Nahar, L. & Sarker, S.D. (2015).** Chemical composition, some allelopathic aspects, free-radical-scavenging property and antifungal activity of the volatile oil of the flowering tops of *Leucanthemum vulgare* Lam. *Records of Natural Products*, **9**, 538-545.
- Milam, C.D., Farris, J.L. & Wilhide, J.D. (2000).** Evaluating mosquito control pesticides for effect on target and nontarget organisms. *Archives of Environmental Contamination and Toxicology*, **39**, 324-328.
- Mokrani, A. & Madani, K. (2016).** Effect of solvent, time and temperature on the extraction of phenolic compounds and antioxidant capacity of peach (*Prunus persica* L.) fruit. *Separation and Purification Technology*, **162**, 68-76.
- Nauen, R. (2007).** Insecticide resistance in disease vectors of public health importance. *Pest Management Science*, **63**, 628-633.
- Nazaruk, J. & Kalemba, D. (2009).** Chemical composition of the essential oils from the roots of *Erigeron acris* L. and *Erigeron annuus* (L.) Pers. *Molecules*, **14**, 2458-2465.
- Ogden, L. E. (2018).** Climate change, pathogens, and people. *BioScience*, **68**(10), 733-739.
- Orhan, I., Kartal, M., Naz, Q., Ejaz, A., Yilmaz, G., Kan, Y., Konuklugil, B., Sener, B. & Choudhary, M.I. (2007).** Antioxidant and anticholinesterase evaluation of selected Turkish *Salvia* species. *Food Chemistry*, **103**, 1247-1254.
- Pavela, R. (2008).** Larvicidal effects of various Euro-Asiatic plants against *Culex quinquefasciatus* Say larvae (Diptera: Culicidae). *Parasitology Research*, **102**, 555-559.
- Pitarokili, D., Tzakou, O. & Loukis, A. (2006).** Essential oil composition of *Salvia verticillata*, *S. verbenaca*, *S. glutinosa* and *S. Candidissima* growing wild in Greece. *Flavour and Fragrance Journal*, **21**, 670-673.
- Polatoğlu, K., Karakoç, E.C., Yücel, Y.Y., Gücel, S., Demirci, B., Demirci, F. & Başer, K.H.C. (2017).** Insecticidal activity of *Salvia veneris* Hedge. Essential oil against coleopteran stored product insects and Spodoptera exigua (Lepidoptera). *Industrial Crops and Products*, **97**, 93-100.
- Roberts, D.R. & Andre, R.G. (1994).** Insecticide resistance issues in vector borne disease control. *The American Journal of Tropical Medicine and Hygiene*, **50**, 21-34.
- Sakthivadivel, M., Gunasekaran, P., Sivakumar, M., Arivoli, S., Raveen, R. & Tennyson, S. (2015).** Mosquito larvicidal activity of *Hyptis suaveolens* (L.) Poit (Lamiaceae) aerial extracts against the filarial vector *Culex quinquefasciatus* Say (Diptera: Culicidae). *Journal of Medicinal Plants Studies*, **3**, 1-5.
- Sampson, B.J., Tabanca, N., Kirimer, N., Demirci, B., Baser, K.H.C., Khan, I.A., Spiers, J.M. & Wedge, D.E. (2005).** Insecticidal activity of 23 essential oils and their major compounds against adult *Lipaphis pseudobrassicae* (Davis) (Aphididae: Homoptera). *Pest Management Science*, **61**, 1122-1128.
- Sefidkon, F. & Khajavi, M.S. (1999).** Chemical composition of the essential oils of two *Salvia* species from Iran: *Salvia verticillata* L. and *Salvia santolinifolia* Boiss. *Flavour Fragrance Journal*, **14**, 77-78.
- Selvi, E.K., Turumtay, H., Demir, A. & Turumtay, E.A. (2018).** Phytochemical profiling and evaluation of the hepatoprotective effect of *Cuscuta campestris* by high-performance liquid chromatography with diode array detection. *Analytical Letters*, **51**, 3-15.
- Singh, O., Khanam, Z., Misra, N. & Srivastava, M. (2011).** Chamomile (*Matricaria chamomilla* L.): An overview. *Pharmacognosy Review*, **5**, 82-95.
- Sukeksi, L. & Sarah, M. (2016).** Characterizations and extraction of polyphenols from residual pulp of pink guava as source of antioxidants. *Journal of Engineering and Applied Sciences*, **11**, 5209-5216.
- Tepe, B., Eminağaoğlu, O., Akpulat, H.A. & Aydin, E. (2007).** Antioxidant potentials and rosmarinic acid levels of the methanolic extracts of *Salvia verticillata* (L.) subsp. *verticillata* and *S. verticillata* (L.) subsp. *amasiaca* (Freyn & Bornm.) Bornm. *Food Chemistry*, **100**, 985-989.
- WHO. (2005).** *Guidelines for laboratory and field testing of mosquito larvicides*, WHO/CDS/WHOPES/GCDPP/2005.13, Geneva, Switzerland.

*Corresponding author's:

Emine KILIÇKAYA SELVİ

Department of Chemistry, Faculty of Arts and Sciences, Recep Tayyip Erdogan University, 53100 Rize, TURKEY

E-mail: emine.selvi@erdogan.edu.tr

ORCID: <https://orcid.org/0000-0003-0291-5362>