

Beta-Sitronellol ve (-)-Menton Monoterpenlerinin İnsan Meme Kanseri (MCF-7) Hücre Hattında Sitotoksik Etkilerinin İncelenmesi

Investigation of Cytotoxic Effect of Monoterpenes Beta-Citronellol and (-)-Menthone in Human Breast Cancer (MCF-7) Cell Line

Sevcan MAMUR¹

¹Gazi Üniversitesi, Yaşam Bilimleri Uygulama ve Araştırma Merkezi

ÖZ

Amaç: Monoterpenler, ilaç, gıda veya kozmetik endüstrisinde kullanılan, bitki esans yağlarının ana bileşenleridir. Birçok terpenoid, antialerjik, antiviral, anti-inflamatuar ve antikanser gibi çeşitli farmakolojik aktiviteler sergiler. Bununla birlikte, bu bileşiklerin insan meme kanseri (MCF-7) hücreleri üzerine sitotoksik etkileri ile ilgili yeterli çalışma bulunmamaktadır. Bu çalışmanın amacı, Beta-Sitronellol (BS) and (-)-Menton (MNT) monoterpenlerinin potansiyel sitotoksik etkilerini MCF-7 hücre hattında 3-(4,5-dimethylthiazolyl-2)-2,5 diphenyl tetrazolium bromid (MTT) testi ile incelemektir.

Yöntem: Bu çalışmada BS ve MNT monoterpenlerinin sitotoksik etkileri, MCF-7 hücre hattında 24 ve 48 saat muamele sürelerinde MTT testi ile analiz edilmiştir. Bunun için hücrelere BS (2, 10, 50, 250, 1250 µg/mL) ve MNT (16, 80, 400, 2000, 10.000 µg/mL)'nin farklı konsantrasyonları uygulanmıştır.

Bulgular: BS ve MNT, 24 saatlik uygulama süresinde tüm konsantrasyonlarda, (%) hücre canlılığını önemli düzeyde azaltmıştır. Ayrıca 48 saatlik uygulamada, BS (250, 1250 µg/mL) ve MNT (16, 400, 2000, 10.000 µg/mL), (%) hücre canlılığını anlamlı oranda düşürmüştür.

Sonuç: Sonuç olarak; BS ve MNT, her iki muamele süresinde de MCF-7 hücreleri üzerinde sitotoksik etki göstermiştir. Ancak sitotoksitenin belirlenmesine yönelik farklı hücre hatlarında daha fazla çalışma yapılmalıdır.

Anahtar Kelimeler: Monoterpen, Beta-Sitronellol, (-)-Menton, MCF-7, MTT testi

ABSTRACT

Objective: Monoterpenes are the primary components of plant essential oils which can use in medicines, food or cosmetic industries. Many terpenoids exhibit a various pharmacological activities such as antiallergic, antiviral, anti-inflammatory and anticancer. However no enough data are found about cytotoxicity of these compounds on human breast cancer (MCF-7) cells. The aim of this study was to evaluate the potential cytotoxic effects of monoterpenes Beta-Citronellol (BC) and (-)-Menthone (MNT) using 3-(4,5-dimethylthiazolyl-2)-2,5 diphenyl tetrazolium bromide (MTT) assay in MCF-7 cell line.

Methods: In this study, the cytotoxic effects of monoterpenes BC and MNT were analyzed by MTT assay at 24 and 48 hour treatment times in the MCF-7 cell line. MCF-7 cells were treated with different concentrations of BC (2, 10, 50, 250, 1250 µg/mL) and MNT (16, 80, 400, 2000, 10.000 µg/mL).

Results: BC and MNT were significantly decreased cell viability (%) at all concentrations at 24-h treatment period. Furthermore, BC (250, 1250 µg/mL) and MNT (16, 400, 2000, 10.000 µg/mL) were significantly reduced the cell viability at 48-h treatment.

Conclusion: As a result, BC and MNT showed cytotoxic effect on MCF-7 cells in both treatment times. Further studies should be conducted to determination cytotoxicity using different cell lines.

Key words: Monoterpen, Beta-Citronellol, (-)-Menthone, MCF-7, MTT assay

Sorumlu Yazar: Sevcan MAMUR

Gazi Üniversitesi, Yaşam Bilimleri Uygulama ve Araştırma Merkezi, Ankara, TÜRKİYE
smamur@gazi.edu.tr

Geliş Tarihi: 10.10.2018 – Kabul Tarihi: 21.05.2019

*Bu çalışma 26 Ekim - 28 Ekim 2018 tarihlerinde Aydın ili, Adnan Menderes Üniversitesi'nde düzenlenen Uluslararası Tarım, Çevre ve Sağlık Kongresi'nde sözel bildiri olarak sunulmuştur

1. INTRODUCTION

Essential oils are synthesized by aromatic plants as secondary metabolites (1, 2). The most compounds in the essential oils are monoterpenes (3). Monoterpenes are widely used in medicines, preservation, food flavoring and fragrances or cosmetic industries (4, 5). They exert a wide range of pharmacological activities such as antiallergic, antiviral, anti-inflammatory and anticancer (6, 7). Some researchers revealed that terpenes are important cancer chemopreventive and chemotherapeutic agents (8-10).

Beta-Citronellol is a monoterpene naturally found in the essential oil of various plant worldwide (11). This monoterpene is a widely used as a fragrant constituents in perfumes (12) and a flavoring agent in food and beverages (12, 13). It has an antihypertensive effects (11) and anti-inflammatory effects in rodent (14).

(-)-Menthone along with Menthol and other compounds are a cyclic monoterpene which is generally members of the *Mentha* genus (15). (-)-Menthone is synthesized in the 8-step pathway of menthol biosynthesis described in great detail by Croteau et al. (16). It is a major essential oil constituent of a very limited number of aromatic plants, known to shown various biological activity such as antimicrobial, antifungal, anticancer and anti-inflammatory effects (15). (-)-Menthone, isomenthone and other compounds impart the cooling minty taste and smell to plants (15, 17). Therefore they are major flavouring additives besides vanilla and citrus. Also it is used in a variety of consumer products ranging from oral-care products such as toothpaste to confections such as chocolate and chewing gum as well as in over-the counter medicinal products for its cooling and biological effects (15).

Many terpenes have cytotoxic effect and they can inhibit the survival of cancer cells (18-21). Breast cancer is one of the most common female malignant tumors which primary leads cancer deaths in worldwide (22, 23). However no data are found about the cytotoxicity of these compounds in human breast cancer (MCF-7) cell line. The aim of this study was to evaluate the cytotoxic effects of monoterpenes Beta-citronellol (BC), and (-)-Menthone (MNT) using 3-(4,5-dimethylthiazolyl-2)-2,5 diphenyl tetrazolium bromide (MTT) assay in MCF-7 cell line.

2. METHODS

Chemicals

The test substances Beta-citronellol (3,7-dimethyl-6-octen-1-ol, Cas No: C83201-5G, %95) and (-)-Menthone [(1R,4S)-p-Menthan-3-one, (2S,5R)-2-Isopropyl-5-methylcyclohexanone, Cas No: 218235-25G, %90] were obtained from Sigma. Dulbecco's Modified Eagle Medium with phenol red (DMEM, Cas. No: F0445), Dulbecco's Modified Eagle Medium without phenol red (DMEM, Cas. No: F0475), PBS (L1825), foetal bovine serum (FBS, Cas. No: S0613), penicillin/streptomycin (Cas. No: A2213), L-glutamine (Cas. No: K0283) and trypsin (Cas. No: L2163) were obtained from Biochrome. In vitro toxicology assay kit (MTT based, Tox-1), DMSO Hybrimax (Cas. No: D2650) were obtained from Sigma.

The molecular structure of Beta-citronellol and (-)-Menthone were respectively shown in Figure 1.

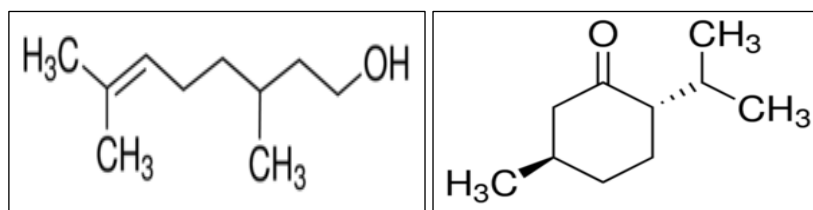


Figure 1. Molecular Structure of Monoterpenes a) Beta-citronellol (24) b) (-)-Menthone (25)

Cell Culture

Human breast cancer (MCF-7) cells were cultured at 37 °C and 5% CO₂ in Dulbecco's Modified Eagle Serum (DMEM) medium supplemented with 10% fetal bovine serum (FBS), 1 (%) penicillin/streptomycin and 2 mM L-glutamine. The cells were seeded in T75 flasks to grown. After the cells were reached a sufficient number, the culture medium was transferred to 96 multiwell plates and incubated 24 h for the adhesion of cells.

MTT Cell Viability Assay

The 3-(4,5-dimethylthiazolyl-2)-2,5 diphenyltetrazolium bromide (MTT) assay was applied according to the methods of Mossman et al. (26) with some modifications Mamur et al. (27). Firstly, the cells were plated on 96-multiwell plates, including to per well 5x10³ cells. And then MCF-7 cells were treated with different concentrations of BC (2, 10, 50, 250, 1250 µg/mL) and MNT (16, 80, 400, 2000, 10.000 µg/mL) for 24 and 48 hours. Afterwards the processes was finished, the cell viability (%) and the half of inhibitory (IC₅₀) values were calculated.

Statistical Analysis

MTT cell viability assay was analyzed by One Way ANOVA, followed by Dunnet's multiple comparison test and P value less than 0.05 was considered as the statistically significance level.

3. RESULTS

Beta-citronellol (BC)

The cytotoxicity result of Beta-citronellol (BC) shows in Table 1. BC was significantly decreased the cell viability (%) at all concentrations at 24-h treatment period. Furthermore, BC was significantly reduced the cell viability at 250, 1250 µg/mL for 48-h treatment on human breast cancer (MCF-7) cell line. The half of inhibitory value (IC₅₀) evidenced by BC was 1250 µg/mL for 24-h and at 250 µg/mL concentration for 48-h (Figure 2).

Table 1. Cytotoxic effect of Beta-citronellol on MCF-7 cell line

Test substance	24 hour		48 hour
	Concentrations (µg/mL)	N	Mean±SD
Control	0.00	3	1.2690±0.1443
Beta-citronellol	2	3	0.8767±0.1460*
	10	3	0.8560±0.0812*

Table 1. Cytotoxic effect of Beta-citronellol on MCF-7 cell line (continued)

50	3	0.6840±0.1298 *	0.7070±0.4171
250	3	0.6720±0.0752 *	0.5443±0.0155 *
1250	3	0.6420±0.1857 *	0.3297±0.1715 *

MCF-7: human breast cancer cell line, SD: standart deviation

* Significantly different from the control P< 0.05 (One way ANOVA-Dunnet Test)

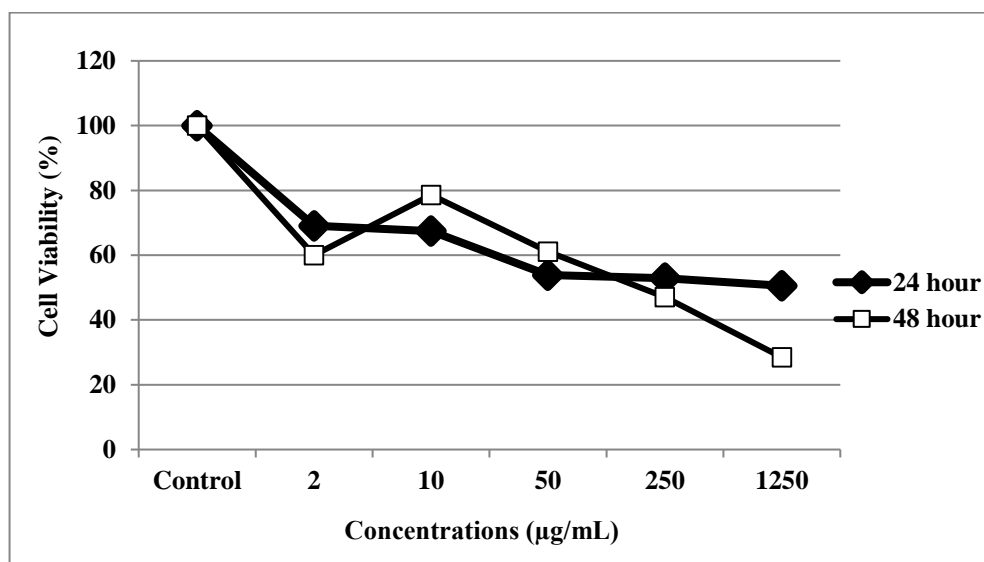


Figure 2. Cell viability (%) results of Beta-citronellol on MCF-7 cell line

(-)-Menthone (MNT)

Table 2 summarizes the results of cytotoxic effect of a treatment with (-)-Menthone (MNT) on MCF-7 cell line. MNT was significantly reduced the cell viability (%) at all concentrations at 24-h treatment period. In addition, MNT was significantly decreased the cell viability at 48-h treatment (16, 400, 2000, 10.000 µg/mL). Moreover, IC50 value for MNT was determined as 80 µg/mL concentration for 24-h and at 2000 µg/mL concentration for 48-h treatment (Figure 3).

Table 2. Cytotoxic effect of (-)-Menthone on MCF-7 cell line

Test substance			24 hour	48 hour
	Concentrations (µg/mL)	N	Mean±SD	Mean±SD
Control	0.00	3	1.4210±0.5214	1.037±0.030
(-)-Menthone	16	3	0.7957±0.0965 *	0.7723±0.1057 *
	80	3	0.7243±0.1668 *	0.8520±0.2119
	400	3	0.6010±0.0898 *	0.6640±0.0034 *
	2000	3	0.5483±0.0319 *	0.6270±0.1490 *
	10.000	3	0.2097±0.0560 *	0.7280±0.0511 *

MCF-7: human breast cancer cell line, SD: standart deviation

* Significantly different from the control P< 0.05 (One way ANOVA-Dunnet Test)

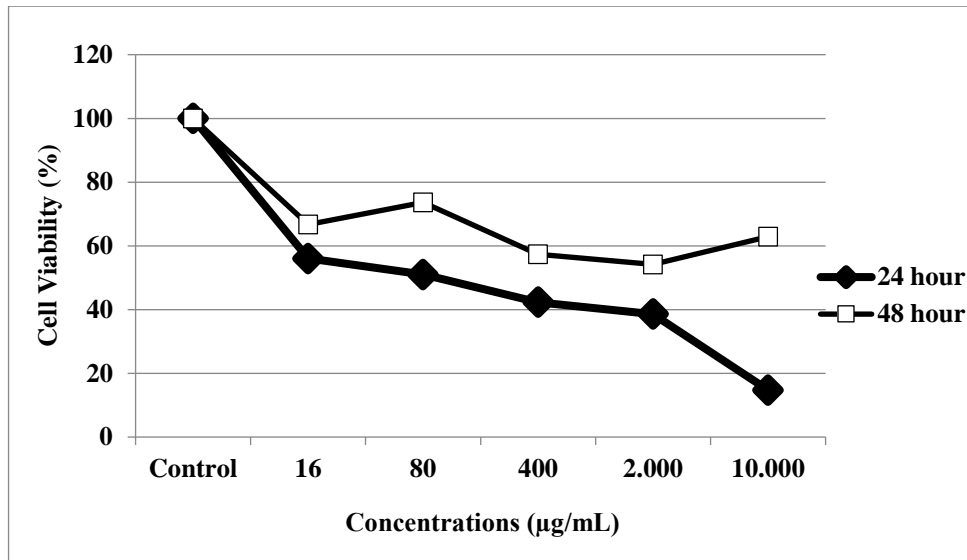


Figure 3. Cell viability (%) results of (-)-Menthone on MCF-7 cell line

4. DISCUSSION

Monoterpenes are an essential oils of many aromatic plants. They are provided active components exhibiting antibacterial, antiviral, antifungal, antioxidant and anticancer effects (3, 28). Recently, monoterpenes have become of clinical relevance due to their ability to suppress cancer development (21). Various studies have indicated that many terpenes possible use in primary prevention and therapy of many types of cancer (18, 20, 21, 29) and their ability to suppress cancer development, e.g., in the breast (30), lung (31, 32), pancreas (33), prostate (34), colon (19). Monoterpenes are widely used in cosmetic, pharmaceutical and medical applications, as well as in the food industry (4). For this reason, it is necessary to determine their safety of terpenes for human health.

Cytotoxicity is an necessary parameter for evaluating chemical substances for toxicity and health risks (35). MTT assay is mostly used colorimetric analysis and it used to detect cytotoxicity following an exposure to toxic substances measured mitochondrial activity (36). According to MTT protocol, the living cells to reduce MTT into a blue formazon product (37).

In this study results indicated that Beta-citronellol (BC) (250, 1250 µg/mL) and (-)-Menthone (MNT) (all treatment except at 80 µg/mL) were significantly decreased the cell viability (%) for 48-h treatment on human breast cancer (MCF-7) cells line. Moreover, both monoterpenes had cytotoxic effects in all concentrations for 24-h treatment period on MCF-7 cells. The half of inhibitory value (IC₅₀) evidenced by BC was 1250 µg/mL for 24-h and at 250 µg/mL for 48-h. Furthermore, the IC₅₀ value of MNT was determined as 80 µg/mL for 24-h and at 2000 µg/mL for 48-h. According to our results observed that BC is more cytotoxic effect than MNT on MCF-7 cells.

To the best of our knowledge, no studies have been conducted on cytotoxicity of Beta-citronellol and (-)-Menthone on MCF-7 cell line. However, the cytotoxic effects of these terpenes and their derivatives in several cell line have been previously reported. Citronellol exhibited a weak cytotoxic effects against HL60 tumor cells (38). Some studies have demonstrated citronellal affect hepatocellular processes in rodents and humans (21). Maßberg et al. (21) investigated that physiological effect upon long-term stimulation with (-)-citronellal,

a proliferation assay was performed. (-)-Citronellal (500 µM) treated with Huh7 cells for 24 h, 48 h and 5 day. According to their results, stimulation over a period of 5 day resulted in a significant inhibition of cell proliferation (52%) compared to the control. (-)-Citronellal does not affect the cell viability of Huh7 cells with propidium iodide staining. Another monoterpenes is a (-)-Menthone, which is a derivatives from Menthol. Menthol was a cytotoxic effect on murine leukemia WEHI-3 cells in a concentration-dependent manner (39). Lin et al. (40) showed menthol induced cytotoxicity by inhibiting the expression of topoisomerases I, II alpha and II beta and promoting the expression of NF-κB in SNU-5 cells. In addition this terpene induced the antiproliferative activity of 1α,25-dihydroxyvitamin D3 in LNCaP cells (41). Wang et al. (42) found that menthol was inhibited the proliferation and motility of prostate cancer DU145 cells.

Researches with other terpenes have shown that many monoterpenes were determined to have cytotoxic effect in different cell line. Slamenova et al. (4) observed that eugenol had a cytotoxic effect in VH10 human fibroblasts, in Caco-2 human colonic cells, but not in HepG2 human hepatoma cells. Thymol exhibited cytotoxic effect against P815 mastocytoma cells (43), human colonic cells (Caco-2), human hepatoma cells (HepG2), and hamster lung cells (V79) (44). Similarly, Jaafari et al. (45) determined that carvacrol, thymol, carveol, carvone, eugenol and isopulegol were cytotoxic activity on murine mastocytoma (P-815), human chronic myelogenous leukemia (K-562), acute T lymphoblastoid leukemia (CEM), human breast adenocarcinoma (MCF-7) cells. Besides, authors indicated that the carvacrol is the most cytotoxic monoterpene. Jin et al. (46) indicated geraniol inhibited the proliferation in BXPc-3 human pancreatic cancer cell. In 2017, geraniol was performed a significant reduction in cell viability of human hepatoma (HepG2) cell line and in peripheral blood mononuclear cells (2).

The mechanism of cytotoxicity of monoterpenes is currently unknown. However, some studies reported that many cancer cells are characterized by severe changes in energy metabolism, mitochondrial overproduction and permanent oxidative stress (3, 47). Because of essential oils capacity to interfere with mitochondrial functions, may add prooxidant effects and thus become genuine anticancer agents (3).

5. CONCLUSION

In the present study, the cytotoxic potential of monoterpenes Beta-Citronellol and (-)-Menthone were evaluated using MTT assay in MCF-7 cell line. In literature, this is the first study to determine the cytotoxic effects of Beta-Citronellol and (-)-Menthone on MCF-7 cells. Based on our findings, both monoterpenes exhibit cytotoxic effects on MCF-7 cells. Therefore, further investigations are needed in order to assess the cytotoxicity on human and ensure its safety.

KAYNAKLAR

1. Gherlardini, C., Galeotti, N., Mazzanti, G. (2001). Local anaesthetic activity of monoterpenes and phenylpropanes of essential oils. *Planta Medica*, 67, 564–566.
2. Queiroz, T.B., Santos, G.F., Ventura, S.C., Hiruma-Lima, C.A., Gaivão, I.O.M., Maistro, E.L. (2017). Cytotoxic and genotoxic potential of geraniol in peripheral blood

- mononuclear cells and human hepatoma cell line (HepG2). *Genetics and Molecular Research*, 16 (3), gmr16039777, DOI <http://dx.doi.org/10.4238/gmr16039777>.
3. Bakkali, F., Averbeck, S., Averbeck, D., Idaomar, M. (2008). Biological effects of essential oils-a review. *Food and Chemical Toxicology*, 46, 446-475 <https://doi.org/10.1016/j.fct.2007.09.106>.
 4. Slamenova, D., Horvathova, E., Wsolova, L., Sramkova, M., Navarova, J. (2009). Investigation of anti-oxidative, cytotoxic, DNA-damaging and DNA-protective effects of plant volatiles eugenol and borneol in human-derived HepG2, Caco-2 and VH10 cell lines. *Mutation Research*, 677, 46–52.
 5. Unlu, M., Ergene, E., Unlu, G.V., Zeytinoglu, H.S., Vural, N. (2010). Composition, antimicrobial activity and *in vitro* cytotoxicity of essential oil from *Cinnamomum zeylanicum* Blume (Lauraceae). *Food and Chemical Toxicology*, 48, 3274–3280.
 6. Balasundram, N., Sundram, K., Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99(1), 191-203.
 7. Alu'datt, M.H., Rababah, T., Alhamad, M.N., Al-Mahasneh, M.A., Almajwal, A., Gammoh, S., Ereifej, K., Johargy, A., Alli, I. (2017). A review of phenolic compounds in oil-bearing plants: Distribution, identification and occurrence of phenolic compounds. *Food Chemistry*, 218, 99–106.
 8. Crowell, P.L. (1999). Prevention and therapy of cancer by diethary monoterpenes. *The Journal of Nutrition*, 129, 775S–778S.
 9. Lampe, J.W. (2003). Spicing up a vegetarian diet: chemopreventive effects of phytochemicals. *American Society for Clinical Nutrition*, 78, 579S–583S.
 10. Nikolic, B., Mitic-Culafic, D., Vukovic-Gacic, B., Knezevic-Vukcevic, J. (2011). Modulation of genotoxicity and DNA repair by plant monoterpenes camphor, eucalyptol and thujone in *Escherichia coli* and mammalian cells. *Food and Chemical Toxicology*, 49, 2035-2045.
 11. Ribeiro-Filho, H.V., Silva de, C.M.S., Siqueira de, R.B., Lahlou, S., Santos, A.A., Magalhães, P.J.C. (2016). Biphasic cardiovascular and respiratory effects induced by β -citronellol. *European Journal of Pharmacology*. 775, 96–105.
 12. Elsharif, S.A., Buettner, A. (2017). Influence of the chemical structure on the odor characters of β -citronellol and its oxygenated derivatives. *Food Chemistry*, 232, 704–711.
 13. Murakami, A., Furukawa, J., Kawasaki, Y., Ota, R. (2013). Flavoring agent with natural fruit-like hop aroma and manufacturing method of beverages. In: *Kirin Brewery Co., Ltd. Japan*. pp. 19.
 14. Brito, R.G., Guimarães, A.G., Quintans, J.S.S., Santos, M.R.V., Sousa, D.P., Badaue-Passos, D., Quintans, L.J. (2012). Citronellol, a monoterpene alcohol, reduces nociceptive and inflammatory activities in rodents. *Journal of Natural Medicines*, 66 (4), 637–644.
 15. Kamatou, G.P.P., Vermaak, I., Viljoen, A.M., Lawrence, B.M. (2013). Menthol: A simple monoterpene with remarkable biological properties. *Phytochemistry*, 96, 15–25.
 16. Croteau, R.B., Davis, E.M., Ringer, K.L., Wildung, M.R. (2005). (-)-Menthol biosynthesis and molecular genetics. *Naturwissenschaften*, 92, 562–577.
 17. Lawrence, B.M. (2013). The story of India's mint oils and menthol. *Perfumer and Flavorist*, 38 (1), 26–35.
 18. Thoppil, R.J., Bishayee, A. (2011). Terpenoids as potential chemopreventive and therapeutic agents in liver cancer. *World journal of hepatology*, 3, 228-249.

19. Murthy, K.N.C., Jayaprakasha, G.K., Patil, B.S. (2012). D-limonene rich volatile oil from blood oranges inhibits angiogenesis, metastasis and cell death in human colon cancer cells. *Life Sciences*, 91, 429- 439.
20. Steward, W.P., Brown, K. (2013). Cancer chemoprevention: a rapidly evolving field. *British Journal of Cancer*, 109, 1-7.
21. Maßberg, D., Simon, A., Häussinger, D., Keitel, V., Gisselmann, G., Conrad, H., Hatt, H. (2015). Monoterpene (-)-citronellal affects hepatocarcinoma cell signaling via an olfactory receptor. *Archives of Biochemistry and Biophysics*, 566, 100–109.
22. Meng, C., He, Y., Wei, Z., Lu, Y., Du, F., Ou, G., Wang, N., Luo, X.G., Ma, W. (2018). Zhang TC. He H. MRTF-A mediates the activation of COL1A1 expression stimulated by multiple signaling pathways in human breast cancer cells. *Biomedicine & Pharmacotherapy*, 104, 718-728.
23. Song, Y., Lu, M., Qiu, H., Yin, J., Luo, K., Zhang, Z., Jia, X., Zheng, G., Liu, H., He, Z. (2018). Activation of FOXO3a reverses 5-Fluorouracil resistance in human breast cancer cells. *Experimental and Molecular Pathology*, 105, 57–62.
24. beta-Citronellol, β -Citronellol, C83201, Sigma-Aldrich (Erişim tarihi: 18.10.2018). <https://www.sigmaaldrich.com/catalog/product/aldrich/c83201?lang=en®ion=TR>
25. (-)-Menthone 90%, Sigma-Aldrich (Erişim tarihi: 18.10.2018). <https://www.sigmaaldrich.com/catalog/product/aldrich/218235?lang=en®ion=TR>
26. Mossman, T. (1983). Rapid colometric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65, 55-63.
27. Mamur, S., Yuzbasioglu, D., Yılmaz, S., Erikel, E., Unal, F. (2018). Assessment of Cytotoxic and Genotoxic Effects of Enniatin-A *In Vitro*. *Food Additives & Contaminants: Part A*, 35 (8), 1633-1644, <https://doi.org/10.1080/19440049.2018.1486513>.
28. Liu, S.J., Liao, Z.X., Liu, C., Sun, J.Y. (2015). Chemical constituents, anticancer, antimicrobial and antioxidant activities of essential oil from *Anaphalis lacteal* grown in Qinghai-Tibet Plateau. *Natural Product Research*, 29, 1369-1371.
29. Rabi, T., Bishayee, A. (2009). Terpenoids and breast cancer chemoprevention. *Breast Cancer Research and Treatment*, 115: 223-239.
30. Yuri, T., Danbara, N., Tsujita-Kyutoku, M., Kiyozuka, Y., Senzaki, H., Shikata, N., Kanzaki, H., Tsubura, A. (2004). Perillyl alcohol inhibits human breast cancer cell growth *in vitro* and *in vivo*. *Breast Cancer Research and Treatment*, 84, 251–260.
31. Xu, M., Floyd, H.S., Greth, S.M., Chang, W.C.L., Lohman, K., Stoyanova, R., Kucera, G.L., Kute, T.E., Willingham, M.C., Miller, M.S. (2004). Perillyl alcohol-mediated inhibition of lung cancer cell line proliferation: potential mechanisms for its chemotherapeutic effects. *Toxicology and Applied Pharmacology*, 195, 232–246.
32. Yeruva, L., Pierre, K.J., Elegbede, A., Wang, R.C., Carper, SW. (2007). Perillyl alcohol and perillic acid induced cell cycle arrest and apoptosis in non small cell lung cancer cells. *Cancer Letters*. 257, 216–226.
33. Wiseman, D.A., Werner. S.R., Crowell, P.L. (2007). Cell cycle arrest by the isoprenoids perillyl alcohol, geraniol, and farnesol is mediated by p21(Cip1) and p27(Kip1) in human pancreatic adenocarcinoma cells. *The Journal of pharmacology and experimental therapeutics*, 320, 1163-1170.
34. Jayaprakasha, G.K., Murthy, K.N.C., Demarais, R., Patil, B.S. (2012). Inhibition of prostate cancer (LNCaP) cell proliferation by volatile components from Nagami kumquats. *Planta Medica*, 78, 974-980.

35. Goyary, D., Chattopadhyay, P., Giri, S., Aher, V., Upadhyay, A., Veer, V. (2014). Ochratoxin A induces cytotoxicity, DNA damage and apoptosis in rat hepatocyte primary cell culture at nanomolar concentration. *World Mycotoxin Journal*, 7, 379–386.
36. Riss, T.L., Moravec, R.A., Niles, A.L., Duellman, S., Benink, H.A., Worzella, T.J., Minor, L. (2016). Cell Viability Assays. *Assay Guidance Manual*, 1-31.
37. Ahmad, J., Alhadlaq, H.A., Alshamsan, A., [Siddiqui, M.A.](#), [Saquib, Q.](#), [Khan, S.T.](#), [Wahab, R.](#), [Al-Khedhairi, A.A.](#), [Musarrat, J.](#), [Akhtar, M.J.](#), [Ahamed, M.](#) (2016). Differential cytotoxicity of copper ferrite nanoparticles in different human cells. *Journal of Applied Toxicology*, 36(10), 1284-1293, doi: 10.1002/jat.3299.
38. Wattenberg, L.W. (1991). Inhibition of azoxymethane-induced neoplasia of the large bowel by 3-hydroxy-3,7,11-trimethyl-1,6,10-dodecatriene (nerolidol). *Carcinogenesis*, 12,151-152.
39. Lu, H.F., Liu, J.Y., Hsueh, S.C., [Yang, Y.Y.](#), [Yang, J.S.](#), [Tan, T.W.](#), [Kok, L.F.](#), [Lu, C.C.](#), [Lan, S.H.](#), [Wu, S.Y.](#), [Liao, S.S.](#), [Ip, S.W.](#), [Chung, J.G.](#) (2007). (-)-Menthol inhibits WEHI-3 leukemia cells *in vitro* and *in vivo*,” *In Vivo*, 21 (2), 285–289.
40. Lin, J.P., Lu, H.F., Lee, J.H., [Lin, J.G.](#), [Hsia, T.C.](#), [Wu, L.T.](#), [Chung, J.G.](#) (2005). Menthol inhibits DNA topoisomerases I, II alpha and beta and promotes NF-kappaB expression in human gastric cancer SNU-5 cells. *Anticancer Research*, 25, 2069–2074.
41. Park, E.J., Kim, S.H., Kim, B.J., Kim, S.Y., So, I., Jeon, J.H. (2009). Menthol enhances an antiproliferative activity of 1 α ,25-dihydroxyvitamin D3 in LNCaP cells. *Journal of Clinical Biochemistry and Nutrition*, 44 (2), 125–130.
42. Wang, Y., Wang, X., Yang, Z., Zhu, G., Chen, D., Meng, Z. (2005). Menthol inhibits the proliferation and motility of prostate cancer DU145 cells. *Pathology Oncology Research*. 18 (4), 903-910.
43. Jaafari, A., Mouse, H.A., Rakib, E.M., M’barek, L.A., Tilaoui, M., Benbakhta, C., Boulli, A., Abbad, A., Ziad, A. (2007). Chemical composition and antitumor activity of different wild varieties of Moroccan thyme. *Revista Brasileira de Farmacognosia*, 17, 477-491.
44. Slamenova, D., Horvathova, E., Sramkova, M., Marsalkova, L. (2007). DNA-protective effects of two components of essential plant oils carvacrol and thymol on mammalian cells cultured *in vitro*. *Neoplasma*. 54, 108–112.
45. Jaafari, A., Tilaoui, M., Mouse, H.A., M’bark, L.A., Aboufatima, R., Chait, A., Lepoivre, M., Ziad, A. (2012). Comparative study of the antitumor effect of natural monoterpenes: relationship to cell cycle analysis. *Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy*, 22(3), 534-540.
46. Jin, X., Sun, J., Miao, X., Liu, G., Zhong, D. (2013). Inhibitory effect of geraniol in combination with gemcitabine on proliferation of BXPC-3 human pancreatic cancer cells. *Journal of International Medical Research*. 41(4) 993–1001.
47. Czarnecka, A.M., Golik, P., Bartnik, E. (2006). Mitochondrial DNA mutations in human neoplasia. *Journal of Applied Genetics*, 47, 67–78.