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## Drilling and core data from the Gulf of Gemlik (SE Sea of Marmara): Holocene fauna and flora assemblages

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Research Article

### Keywords:

Foraminifer, Ostracod, Mollusc, Nannoplankton, Diatome, Gemlik Gulf

### ABSTRACT

This study was conducted to determine fauna and flora assemblages of Holocene sequences from Gemlik Gulf (SE Marmara Sea) and to obtain their similarities and differences between the assemblages of Gemlik and İzmit Bays. Total of 201 dark gray colored, fine to medium grained sandy clay samples were studied. In the drilled samples, 22 genera and 38 species were identified from the foraminifera characterizing the infralittoral zone. 40 genera and 58 species of foraminifera characterizing the upper circalittoral zone were identified. In addition, Black Sea originated *Ammonia parasovica* was found for the first time in cores taken from Gemlik Gulf. Nannofossil species characterizing the open-shallow marine environment *Emiliania huxleyi*, *Reticulofenestra parvula*, *Coronosphaera* spp., *Syracosphaera* spp., *Helicosphaera* spp. beside *Gephyrocapsa oceanica*, small *Gephyrocapsa* spp., *Scyphosphaera porosa* were obtained. A total of 27 genera and 37 species from ostracods were defined in the samples from gulf. Moreover, the quite abundant mollusc assemblage characterizing the shallow marine environment and fewer genera and species of diatoms were identified. The studied sequence is represented by NN21 *Emiliania huxleyi* biozone at the Holocene. Foraminifers and ostracods of the Gemlik Gulf are observed as Mediterranean-Aegean Sea originated assemblages. The mollusc shows the shallow marine community.

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## 1. Introduction

The study area is the Gulf of Gemlik located in southeast of Marmara Region (Figure 1). In order to reveal information about the Black Sea-Marmara-Mediterranean connection, many studies have been carried out both with marine and non-marine data obtained from the drilling and bottom sediments in the Sea of Marmara and the Gulf of Gemlik (Meriç, (1995); Görür et al. (1997); İslamoğlu and Chepalyga,

(1998); Çağatay et al. (2000, 2003, 2009); Aksu et al. (1999, 2002); Yalıtırak and Alpar, (2002); Sperling et al. (2003); Kerey et al. (2004); Mudie et al. (2004); Meriç et al. (2005, 2009, 2018); Chepalyga (2007); Hiscott et al. (2007); Yanko-Hombach et al. (2007); McHugh et al. (2008); Marret et al. (2009); İslamoğlu, (2002, 2009); Brückner et al. (2010); Vidal et al. (2010); Gasperini et al. (2011); Nazik et al. (2011); Mertens et al. (2012); Taviani et al. (2014); Vardar et al. (2014); Filikçi et al. (2017).

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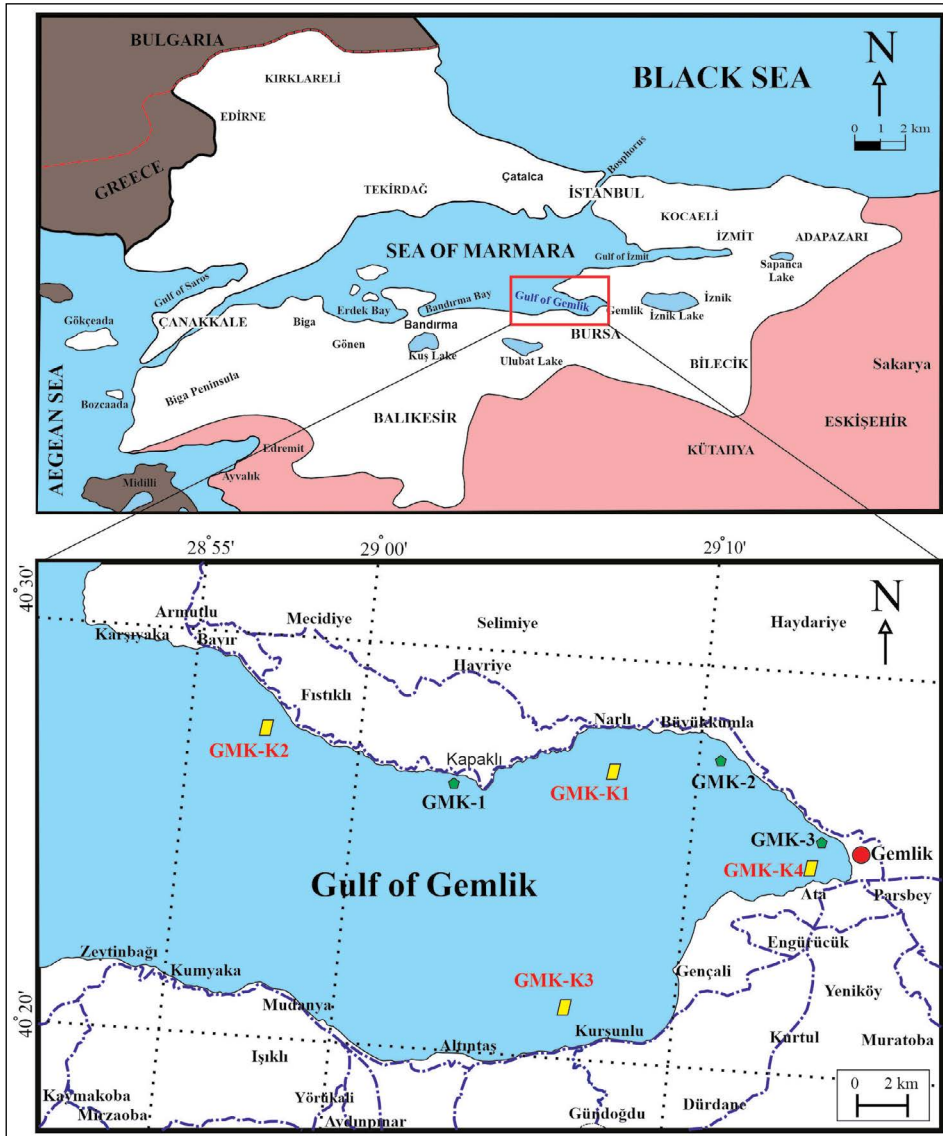


Figure 1- Location map of drilling and cores taken from the Gulf of Gemlik (GMK drilling, GMK-K core locations).

In this study, the foraminifer, ostracod, mollusc, nannoplankton and diatom assemblages in the deposits identified in three drillings holes (GMK-1, GMK-2, GMK-3) from the northeast and east of the Gulf of Gemlik and 4 cores taken from its north, northeast and southeast (GMK-K1, GMK-K2, GMK-K3, GMK-K4) were studied and compared with the Mediterranean-Marmara-Black Sea assemblages in order to reveal similarities and differences.

When previous studies carried out in the region are compared, the foraminifer, ostracod and mollusc faunas of the Sea of Marmara, and also the nannoplankton and diatom assemblages from the deep sediments of the Gulf of Gemlik, as indicated by the

present study, are considered to be richer compared to those reported by Meriç et al. (2005).

## 2. Material and Methods

Sediment samples used in this study were taken from 3 drillings carried out in north, northeast and southeast of the Gulf of Gemlik (GMK-1, GMK-2, GMK-3) (Figures 1-3). Samples come from 4 cores ranging from between 0,70-1,00 m in length. These were collected from the north, northeast and southeast of the bay (GMK-K1, GMK-K2, GMK-K3, GMK-K4) during summer 2015 (Figures 1-4, 5 and table 1). The drilling samples mostly consist of blackish gray, clayey and sandy gravel, medium-grained sandy and

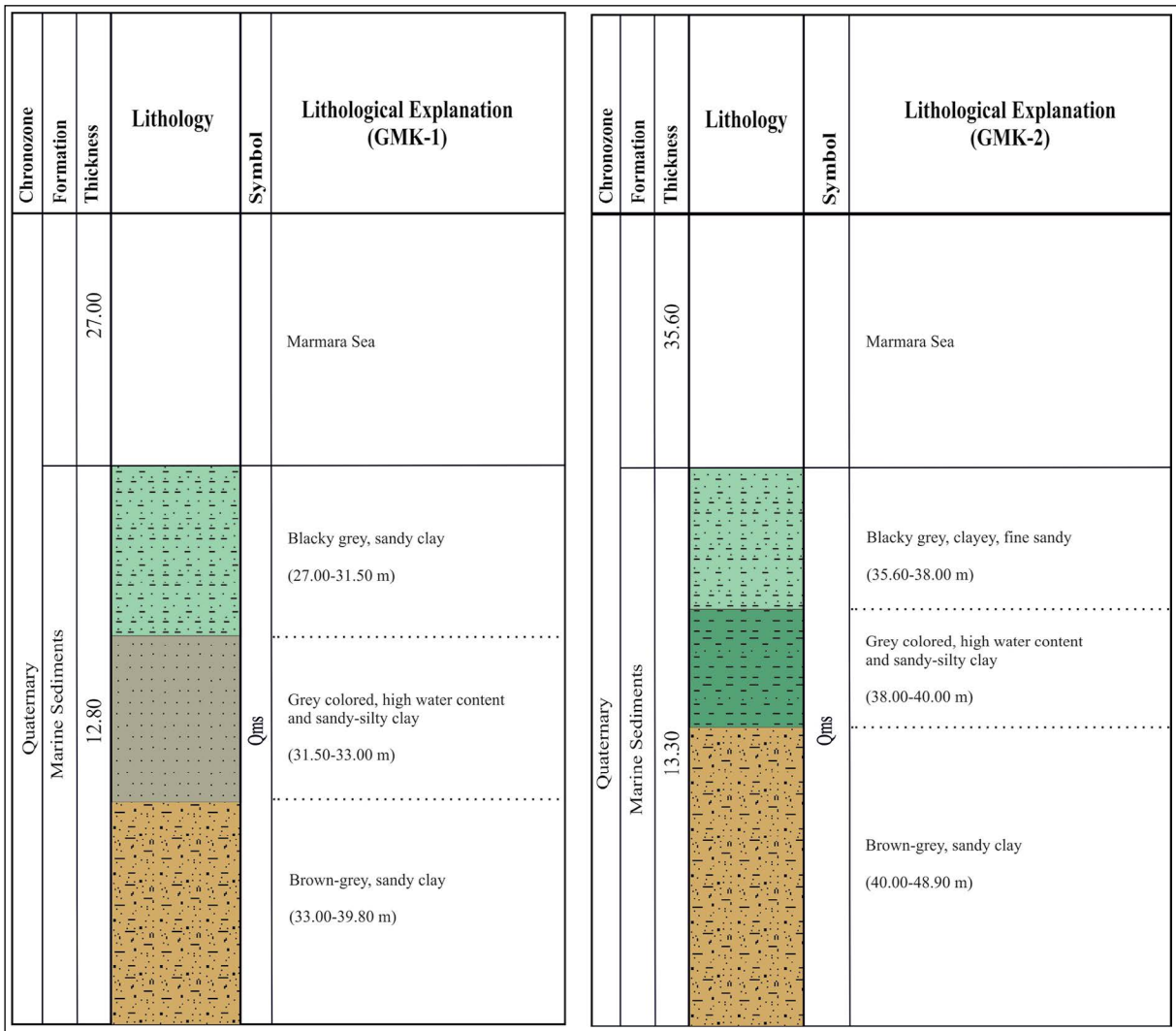


Figure 2- The columnar section of the locations in which drilling samples were taken in the Gulf of Gemlik (GMK-1, water depth 27,00 m and GMK-2, water depth 35,60 m) (not to scale).

silty clay with mollusc shells. The core samples are dark gray, fine sand (with high water content), silty and sandy clay units.

A total of 201 samples were studied as follows: 150 samples (with 10 cm intervals) from the cores of GMK-1 and GMK-2, 14 samples from GMK-3 drilling (with 20 cm intervals) and 37 samples taken from 4 cores (with 10 cm intervals).

To identify foraminifer, ostracod and molluscs, 10% hydrogen peroxide was added to wet sediment samples weighing 5 g, and then left for 24 hours. Afterwards, the samples were washed through a 0.063 mm sieve with pressurized water, allowed to dry in a 50 °C oven and further sieved through 2.00, 1.00,

0.500, 0.250, 0.125 mm size sieves and then examined under a binocular microscope.

The nannofossil smear-slides were prepared by the traditional method, as summarized by Perch-Nielsen (1985a, b). Preparations were examined under the light microscope with the help of 100x magnification oil-immersion objective lens and 10x ocular magnification (using the traditional method) and displayed by using approximately 40x magnification with an intermediate lens+digital camera. Imaging under polarized (PL), normal (NL), contrast (CL) light was as used by Perch-Nielsen (1985a, b) for microscope studies. In addition, the imaging methods under PL using a gypsum wedge (GL) was employed as suggested by Reinhardt (1972) and Romein (1979). Also, imaging

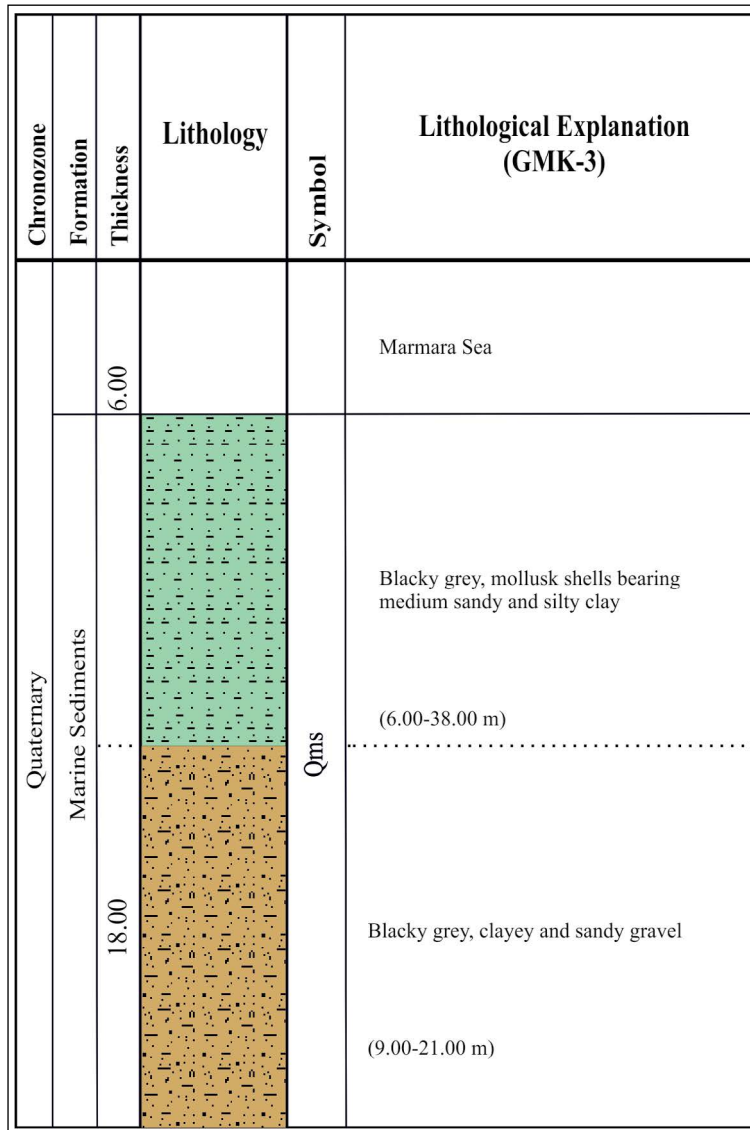


Figure 3- The columnar section of locations in which drilling samples in the Gulf of Gemlik were taken (GMK-3, water depth: 6 m) (not to scale).

under PL was carried out using a quartz wedge (QL), as recommended by Sagular (2009). Selected images of the nannofossil species identified are shown in plate 1.

Samples taken from the drilling holes for diatom analysis were first cleaned by using 10% HCl at the Department of Geological Engineering of the Faculty of Engineering in Aksaray University; this material was then prepared for paleontological study by mounting on a slide using entellan and a coverglass. The preparations were examined under 1600x magnification using a polarizing microscope and an oil-immersion objective with a Leica DM 2500 P. The

fossil diatom genera and species were then identified and photographed (Plate 2)

### 3. Micro and Macro Faunal Findings

#### 3.1. Foraminifera

In the samples from GMK-1, GMK-2 and GMK-3, a total of 22 genera and 39 species were identified as follows: *Textularia bocki* Höglund, *Adelosina cliarensis* (Heron-Allen and Earland), *A. mediterraneensis* (Le Calvez and Le Calvez), *Spiroloculina dilatata* d’Orbigny, *S. excavata* d’Orbigny, *S. ornata* d’Orbigny, *Siphonaperta*

Chronozone Formation Thickness		Lithology	Symbol	Lithological Explanation (GMK-K1)	Chronozone Formation Thickness		Lithology	Symbol	Lithological Explanation (GMK-K2)
Quaternary Marine Sediments	?	> 70 cm	Qms	Current Sediments: Grey colored, high water content, fine sandy, silty clay	?	> 100 cm	Qms	Current Sediments: Dark grey colored, high water content, fine silt, sandy clay	
		81.00 m		Marmara Sea				75.00 m	Marmara Sea
				Unsampled section				Unsampled section	

Figure 4- The columnar section of locations in which core samples in the Gulf of Gemlik were taken (GMK-K1, water depth 81,00 m and GMK-K2, water depth 75,00 m) (not to scale).

Chronozone Formation Thickness		Lithology	Symbol	Lithological Explanation (GMK-K3)	Chronozone Formation Thickness		Lithology	Symbol	Lithological Explanation (GMK-K4)
Quaternary Marine Sediments	?	> 100 cm	Qms	Current Sediments: Dark grey colored, high water content, fine sandy, silty clay	?	> 100 cm	Qms	Current Sediments: Dark grey colored, mollusc shelly, fine sand interbedded, silty clay	
		98.00 m		Marmara Sea				20.00 m	Marmara Sea
				Unsampled section				Unsampled section	

Figure 5- The columnar section of locations in which core samples in the Gulf of Gemlik were taken (GMK-K3, water depth 98,00 m and GMK-K4, water depth 20,00 m) (not to scale).



Table 1- Coordinates and other specifications of drilling and core locations.

Borehole and Cores	GMK-1: GS-84 6 Degree Coordinates		Water Depth	Sample Length
	Longitude	Latitude	(m)	(m)
GMK-1	666899.00 d E	4480487.00 m N	27,00	12,80
GMK-2	677320.00 d E	4480755.00 m N	35,60	13,30
GMK-3	682561.00 d E	4477768.00 m N	6,00	3,00
GMK-K1	672101.00 d E	4481326.00 m N	81,00	0,70
GMK-K2	658957.00 d E	4481138.00 m N	75,00	1,00
GMK-K3	669332.26 d E	4471289.16 m N	98,00	1,00
GMK-K4	681950.40 d E	4477270.34 m N	20,00	1,00

*aspera* (d'Orbigny), *Cycloforina contorta* (d'Orbigny), *C. rugosa* (d'Orbigny), *Massilina secans* (d'Orbigny), *Quinqueloculina bidentata* d'Orbigny, *Q. disparilis* d'Orbigny, *Q. jugosa* Cushman, *Q. lamarckiana* d'Orbigny, *Q. seminula* (Linné), *Miliolinella elongata* Kruit, *M. subrotunda* (Montagu), *Pseudotriloculina laevigata* (d'Orbigny), *P. oblonga* (Montagu), *P. rotunda* (d'Orbigny) *Triloculina marioni* Schlumberger, *Lagena laevis* (Montagu), *Polymorphina* sp., *Globobulimina affinis* (d'Orbigny), *Valvulineria bradyana* (Fornasini), *Rosalina bradyi* Cushman, *Lobatula lobatula* (Walker and Jacob), *Planorbulina mediterraneensis* d'Orbigny, *Asterigerinata mamilla* (Williamson), *Nonion depressulum* (Walker and Jacob), *Ammonia compacta* Hofker, *A. parkinsoniana* (d'Orbigny), *Porosonion subgranosum* (Egger), *Elphidium advenum* (Cushman), *E. complanatum* (d'Orbigny), *E. crispum* (Linné), *E. depressulum* Cushman, *E. jenseni* (Cushman), *E. macellum* (Fichtel and Moll).

In contrast, in the samples from GMK-K1, GMK-K2, GMK-K3 and GMK-K4, a total of 40 genera and 58 species were found, namely: *Ammodiscus planorbis* Höglund, *Eggerelloides scabrus* (Williamson), *Bigenerina nodosaria* d'Orbigny, *Textularia bocki* Höglund, *T. cf. pala* Czjcek, *Adelosina cliarensis* (Heron-Allen and Earland), *Spiroloculina excavata* d'Orbigny, *S. tenuiseptata* Brady, *Siphonaperta aspera* (d'Orbigny), *Cycloforina contorta* (d'Orbigny), *C. tenuicollis* (Wiesner), *Quinqueloculina seminula* (Linné), *Biloculinella depressa* (Wiesner), *B. wiesneri* (Le Calvez and Le Calvez), *Miliolinella subrotunda* (Montagu), *Pseudotriloculina laevigata* (d'Orbigny), *P. rotunda* (d'Orbigny), *Pyrgo elongata* (d'Orbigny), *P. inornata* (d'Orbigny), *Triloculina tricarinata* d'Orbigny, *Sigmoilinita costata* (Schlumberger), *S. edwardsi* (Schlumberger), *S. tenuis* (Czjcek),

*Sigmoilopsis schlumbergeri* (Silvestri), *Dentalina inornata* d'Orbigny, *Neolenticulina peregrina* (Schwager), *Amphicoryna scalaris* (Batsch), *Lagena strumosa* Reuss, *Brizalina alata* (Seguenza), *B. spathulata* (Williamson), *Cassidulina carinata* Silvestri, *Rectuvigerina phlegeri* Le Calvez, *Bulimina aculeata* d'Orbigny, *B. costata* d'Orbigny, *B. elongata* d'Orbigny, *B. marginata* d'Orbigny, *Globobulimina affinis* (d'Orbigny), *G. pseudospinescens* (Emiliani), *Reussella spinulosa* (Reuss), *Fursenkoina acuta* (d'Orbigny), *Valvulineria bradyana* (Fornasini), *Discorbinella bertheloti* (d'Orbigny), *Hyalinea balthica* (Schröter), *Planorbulina mediterraneensis* d'Orbigny, *Asterigerinata mamilla* (Williamson), *Nonion depressulum* (Walker and Jacob), *Nonionella turgida* (Williamson), *Chilostomella mediterraneensis* Cushman and Todd, *Gyrodinoides lamarckiana* (d'Orbigny), *Aubignyna perlucida* (Heron-Allen and Earland), *Ammonia compacta* Hofker, *A. parasovica* Stshedrina and Mayer, *A. parkinsoniana* (d'Orbigny), *Porosonion subgranosum* (Egger), *Elphidium complanatum* (d'Orbigny), *E. crispum* (Linné), *E. cf. incertum* (Williamson), *E. macellum* (Fichtel and Moll) (Table 2). These genera and species were identified by using Meriç et al. (1995, 2005); Sakıncı, (2008); Meriç et al. (2014). In total, 77 species were observed in the drillings and cores.

### 3.2. Ostracods

Ostracods were found in 82 samples of GMK-1, GMK-2, GMK-3 and 37 samples of GMK-K1, GMK-K2, GMK-K3, GMK-K4, in which a total of 27 genera and 37 species were identified (Table 3), according to Van Morkhoven (1963); Breman (1975); Bonaduce et al. (1975); Yassini (1979); Athersuch et al. (1989); Guillaume et al. (1985); Tunoğlu (1999, 2002); Şafak (1999); Guernet et al. (2003).

Table 2- The foraminifer contents of boreholes and cores.

FORAMINIFERA	BOREHOLES			CORES			
	GMK-1	GMK-2	GMK-3	GMK-K1	GMK-2	GMK-3	GMK-4
<i>Ammodiscus planorbis</i> Höglund				*		*	
<i>Eggerelloides scabrus</i> (Williamson)							*
<i>Bigenerina nodosaria</i> d'Orbigny				*	*		
<i>Textularia bocki</i> Höglund		*		*		*	
<i>Textularia cf. pala</i> Czjcek				*	*	*	
<i>Adelosina cliarensis</i> (Heron-Allen and Earland)		*			*		
<i>Adelosina mediterraneensis</i> (Le Calvez and Le Calvez)		*					
<i>Spiroloculina dilatata</i> d'Orbigny	*						
<i>Spiroloculina excavata</i> d'Orbigny	*	*		*	*	*	
<i>Spiroloculina ornata</i> d'Orbigny		*					
<i>Spiroloculina tenuiseptata</i> Brady				*	*	*	
<i>Siphonaperta aspera</i> d'Orbigny	*	*				*	
<i>Cycloforina contorta</i> (d'Orbigny)	*	*	*				*
<i>Cycloforina rugosa</i> (d'Orbigny)	*						
<i>Cycloforina tenuicollis</i> (Wiesner)				*			
<i>Massilina secans</i> (d'Orbigny)	*	*	*				
<i>Quinqueloculina bidentata</i> d'Orbigny	*						
<i>Quinqueloculina disparilis</i> d'Orbigny	*		*				
<i>Quinqueloculina jugosa</i> Cushman	*						
<i>Quinqueloculina lamarckiana</i> d'Orbigny		*					
<i>Quinqueloculina seminula</i> (Linné)	*	*	*	*	*	*	*
<i>Biloculinella depressa</i> (Wiesner)						*	
<i>Biloculinella wiesneri</i> (Le Calvez and Le Calvez)				*	*	*	
<i>Miliolinella elongata</i> Kruit		*					
<i>Miliolinella subrotunda</i> (Montagu)	*				*		
<i>Pseudotriloculina laevigata</i> (d'Orbigny)	*	*				*	
<i>Pseudotriloculina oblonga</i> (Montagu)	*		*				
<i>Pseudotriloculina rotunda</i> (d'Orbigny)	*						*
<i>Pyrgo elongata</i> (d'Orbigny)				*			
<i>Pyrgo inornata</i> (d'Orbigny)				*			
<i>Triloculina marioni</i> Schlumberger	*						
<i>Triloculina tricarinata</i> d'Orbigny				*			
<i>Sigmoilinita costata</i> (Schlumberger)					*		
<i>Sigmoilinita edwardsi</i> (Schlumberger)					*		
<i>Sigmoilinita tenuis</i> (Czjcek)					*	*	
<i>Sigmoilopsis schlumbergeri</i> (Silvestri)						*	
<i>Dentalina inornata</i> d'Orbigny						*	
<i>Neolenticulina peregrina</i> (Schwager)				*			
<i>Amphicoryna scalaris</i> (Batsch)				*	*		
<i>Lagena laevis</i> (Montagu)	*						
<i>Lagena strumosa</i> Reuss				*	*		
<i>Polymorphina</i> sp.	*						
<i>Brizalina alata</i> (Seguenza)				*	*	*	
<i>Brizalina spathulata</i> (Williamson)				*	*	*	*
<i>Cassidulina carinata</i> Silvestri				*	*	*	
<i>Rectuvigerina phlegeri</i> Le Calvez				*	*	*	
<i>Bulimina aculeata</i> d'Orbigny				*	*	*	
<i>Bulimina costata</i> d'Orbigny						*	

Table 2- continued.

<i>Bulimina elongata</i> d'Orbigny				*	*	*	
<i>Bulimina marginata</i> d'Orbigny				*	*	*	
<i>Globobulimina affinis</i> (d'Orbigny)	*			*	*	*	
<i>Globobulimina pseudospinescens</i> (Emiliani)						*	
<i>Reussella spinulosa</i> (Reuss)						*	
<i>Fursenkoina acuta</i> (d'Orbigny)					*		
<i>Valvulineria bradyana</i> (Fornasini)		*		*	*	*	
<i>Rosalina bradyi</i> Cushman	*						
<i>Discorbinella bertheloti</i> (d'Orbigny)				*	*	*	
<i>Hyalinea balthica</i> (Schröter)				*	*	*	
<i>Lobatula lobatula</i> (Walker and Jacob)	*	*	*				
<i>Planorbulina mediterraneensis</i> d'Orbigny		*		*			
<i>Asterigerinata mamilla</i> (Williamson)	*					*	
<i>Nonion depressulum</i> (Walker and Jacob)	*						*
<i>Nonionella turgida</i> (Williamson)				*	*	*	
<i>Chilostomella mediterraneensis</i> Cushman and Todd				*	*	*	
<i>Gyrodinoides lamarckiana</i> (d'Orbigny)				*	*	*	
<i>Aubignyna perlucida</i> (Heron-Allen and Earland)							*
<i>Ammonia compacta</i> Hofker	*	*	*		*		*
<i>Ammonia parasovica</i> Stshedrina and Mayer							*
<i>Ammonia parkinsoniana</i> (d'Orbigny)	*	*	*				*
<i>Porosonion subgranosum</i> (Egger)			*				*
<i>Elphidium advenum</i> (Cushman)	*		*				
<i>Elphidium complanatum</i> (d'Orbigny)	*	*	*		*		*
<i>Elphidium crispum</i> (Linné)	*	*	*				*
<i>Elphidium depressulum</i> Cushman	*	*					
<i>Elphidium cf. incertum</i> (Williamson)					*		*
<i>Elphidium jenseni</i> (Cushman)	*						
<i>Elphidium macellum</i> (Fichtel and Moll)	*		*				*

When the boreholes and cores were compared to each other in terms of the Gulf of Gemlik ostracod community, it was found that the number of genera and species are richer in the drilling samples of GMK-1 and GMK-2 and the core samples of GMK-K2 and GMK-K4 (Table 3). When the drilling locations are considered, it is observed that the northern and eastern parts of the bay are rich in ostracod fauna, similar to the ostracod fauna found in the bottom sediments of the Gulf of Gemlik, as emphasized by Meriç et al. (2005).

The ostracod community found in this study was compared with the ostracod communities in the Aegean islands, the Adriatic Sea, Algeria, Sea of Marmara (Gökçeada-Bozcaada-Çanakkale), the southern Marmara shelf, the Western Black Sea, and the İzmir, Edremit and Gemlik gulfs as reported by Nazik (2001), Meriç et al. (2002, 2005,

2008), Parlak and Nazik (2016), and in the studies mentioned above. There are numerous similarities in genera and species, with *Carinocythereis carinata* (Roemer), *Hiltermannicythere turbida* (Mueller), *H. Rubra* (Mueller), *Costa edwardsii* (Roemer), *Pterygocythereis jonesii* (Baird), *Palmoconcha agilis* (Ruggieri), *Cytheridea acuminata* (Bosquet) species being widespread. In addition, *Loxocauda pellucida* (Mueller), which is known from the Mediterranean, Aegean and Marmara Seas (from Pliocene to the Recent), was found only in the core samples. *Ilyocypris bradyi* Sars, indicative of fresh water inflow, was found at depths of 6,00-6,20 m; i.e. in the topmost part of the GMK-3 drilling. *Heterocypris salina* (Brady) was found at higher levels of the core of GMK-K4 drilling at depths of 20,10-20,20 m. As a result, it was inferred that the ostracod community obtained from the Gulf of Gemlik drillings and cores can be equated with the ostracods of Mediterranean-Aegean seas.



Table 3- The ostracod content of boreholes and cores.

OSTRACODA	BOREHOLES			CORES			
	GMK-1	GMK-2	GMK-3	GMK-K1	GMK-K2	GMK-K3	GMK-K4
<i>Cytherella alvearium</i> Bonaduce, Ciampo and Masoli		*			*		
<i>Carinocythereis carinata</i> (Roemer)	*	*					*
<i>Carinocythereis rhombica</i> Stambolidis	*	*					*
<i>Aurila arborescens</i> (Brady)			*				
<i>Aurila convexa</i> (Baird)		*					
<i>Tyrrenocythere amnicola</i> (Sars)							*
<i>Hiltermannicythere rubra</i> (Mueller)	*						*
<i>Hiltermannicythere turbida</i> (Mueller)	*						*
<i>Pterygocythereis jonesii</i> (Baird)	*	*					*
<i>Cytheretta judaea</i> (Brady)	*				*		
<i>Costa batei</i> (Brady)					*		
<i>Costa edwardsii</i> (Roemer)	*	*		*	*	*	*
<i>Costa tricostata</i> (Reuss)		*					
<i>Callistocythere intricatoides</i> (Ruggieri)	*						*
<i>Callistocythere pallida</i> (Mueller)	*						
<i>Cyprideis torosa</i> (Jones)	*		*				
<i>Cytheridea acuminata</i> (Bosquet)	*	*		*			*
<i>Leptocythere</i> sp.	*	*					
<i>Leptocythere multipunctata</i> (Seguenza)							*
<i>Urocythereis crenulosa</i> (Terquem)	*		*		*		
<i>Buntonia</i> sp.					*		
<i>Semicytherura acuta</i> Mueller	*						
<i>Semicytherura incongruens</i> (Mueller)	*	*					
<i>Semicytherura inversa</i> (Seguenza)	*						*
<i>Paracytheridea depressa</i> Mueller	*				*		
<i>Cytheropteron</i> sp.				*			
<i>Palmoconcha agilis</i> (Ruggieri)	*	*		*	*		*
<i>Loxoconcha bairdi</i> Mueller	*						
<i>Loxoconcha stellifera</i> Mueller	*	*	*		*		
<i>Sagmatocythere versicolor</i> (Mueller)					*		
<i>Loxocauda pellucida</i> (Mueller)				*		*	
<i>Cushmanidea turbida</i> (Mueller)	*	*			*		*
<i>Pontocypris rara</i> Mueller	*	*		*			
<i>Argilloecia</i> sp.					*		
<i>Xestoleberis dispar</i> (Mueller)	*						*
<i>Heterocypris salina</i> (Brady)							*
<i>Ilyocypris bradyi</i> Sars			*				

### 3.3. Nannoplanktons

The studies were carried out on mud/unconsolidated mudstone samples from GMK-1, GMK-2, GMK-3 drillings and GMK-K1, GMK-K2, GMK-K3 and GMK-K4 cores. In general, the nannofossil assemblages including: *Gephyrocapsa oceanica*, small *Gephyrocapsa* spp., *Scyphosphaera porosa* are present, in addition to *Emiliana huxleyi*, *Reticulofenestra parvula*, *Coronosphaera* spp.,

*Bylosphaera* spp., *Helicosphaera* spp. characterize an open-shallow marine environment. Biostratigraphic evaluation of this group allowed NN21 *Emiliana huxleyi* biozone to be determined. According to these data, Holocene marine deposition occurred.

In addition, both synsedimentary and reworked nannofossil assemblages, and their representative biozones and environmental characteristics were determined in mud samples belonging to the boreholes

and cores of the Gulf of Gemlik. In these stratigraphic interpretations, the data related from other coeval fossils, such as dinoflagellate cysts, ascidian spicules, diatom and sponge spicules were also used (see Table 4 and Plate 1). Some synsedimentary (Holocene) nannofossil species were simply reported as “coccospheres”.

In mud samples of the GMK-1 drilling, 18 nannofossil species were observed, of which 12 are “synsedimentary”, 5 of them “reworked from Cenozoic rocks” and a single one is “reworked from Cretaceous rocks”. In these samples, 4 dinoflagellate cysts (*Thoracosphaera* spp.) were recognized, 2 of them are “synsedimentary” and the other 2 are “reworked” (Table 4). In mud samples from the GMK-2 drilling, 39 nannofossil species were recorded, of which 16 species are “synsedimentary”, 20 of them were “reworked” from Cenozoic units, which of them are recorded as

3 dinoflagellate species, 2 are “synsedimentary” and 1 “reworked” (*Thoracosphaera* spp.) (Table 4) is. Finally, in the mud samples of the GMK-3 drilling, 15 nannofossil species were observed, 2 of them were “synsedimentary”, 8 of them were reworked from the Cenozoic and 5 of them were reworked from the Cretaceous. Two “synsedimentary” dinoflagellate cysts (*Thoracosphaera* spp.) were also distinguished (Table 4). As shown in Table X, the nannofossil assemblages *Syracosphaera* spp. and *Coronosphaera* spp., in addition to *Emiliana huxleyi*, *Reticulofenestra parvula*, indicate the presence of the NN21 *Emiliana huxleyi* Zone. It is concluded that the sediments, excluding *Gephyrocapsa omega* individuals in the GMK-1 and GMK-2 drillings, represent shallow-open sea deposition during the Holocene period (Varol and Houghton, 1996; Young, 1998; Young et al., 2014). Although, there are few nannofossils in the samples from the GMK-3 drilling, the abundance

Table 4- The nannofossils assemblage and biostratigraphical distribution of boreholes and cores.

ORIGINAL CLASSIFICATION	CHRONOZONE	BIOZONE	Nannofossil species	BOREHOLES			CORES				
				GMK-1	GMK-2	GMK-3	GMK-K1	GMK-K2	GMK-K3	GMK-K4	
SYNSEDIMENTARY	HOLOCENE	NN21 <i>Emiliana huxleyi</i> Zone	<i>Alisphaera</i> sp. (Gran and Braarud)		*						
			<i>Anthosphaera</i> sp. "coccosphere"		*						
			<i>Braarudosphaera bigelowii</i> (Gran and Braarud)	*	*		*	*	*		
			<i>Cocolithus pelagicus</i> (Wallich)				*		*		
			<i>Coronosphaera binodata</i> (Kamptner)	*	*		*	*	*		
			<i>Coronosphaera mediterranea</i> Lohmann	*	*		*	*	*	*	
			<i>Dictyococcites antarcticus</i> Haq	*							
			<i>Dictyococcites productus</i> (Kamptner)	*	*		*	*	*	*	
			<i>Emiliana huxleyi</i> (Lohmann)	*	*	*	*	*	*	*	
			<i>Emiliana huxleyi</i> (Lohmann) "coccosphere"		*			*	*	*	
			küçük <i>Gephyrocapsa</i> sp.		*			*	*		
			<i>Gephyrocapsa oceanica</i> Kamptner					*		*	
			<i>Helicosphaera carteri</i> (Wallich)	*	*		*	*	*		
			<i>Helicosphaera hyalina</i> Gaarder	*	*			*	*		
			<i>Helicosphaera wallichii</i> (Lohmann)					*	*		
			<i>Papposphaera</i> sp. Tangen "coccosphere"					*			
			<i>Pontosphaera japonica</i> (Takayama)					*			
			<i>Pseudoemiliana lacunosa ovata</i> (Kamptner)					*			
			<i>Reticulofenestra parvula</i> (Okada and McIntyre)	*	*	*	*	*	*	*	*
			<i>Rhabdosphaera clavigera</i> Murray and Blackman	*	*			*		*	
<i>Syracosphaera histrica</i> Kamptner	*				*		*				
<i>Syracosphaera pulchra</i> Lohmann	*	*			*	*	*	*			
<i>Thoracosphaera grantifera</i> Fütterer	*	*	*	*	*	*	*	*			
<i>Thoracosphaera tuberosa</i> Kamptner	*	*	*	*	*	*					

Table 4- continued.

REWORKED	PALEOGENE-NEOGENE	?	<i>Arkhangelskiella specillata</i> Vekshina	*					
			<i>Blackites spinosus</i> (Deflandre and Fert)				*		
			<i>Chiasmolithus grandis</i> (Bramlette and Riedel)						*
			<i>Clausicoccus fenestratus</i> (Deflandre and Fert)	*					
			<i>Cocolithus miopelagicus</i> Bukry						*
			<i>Cocolithus pelagicus</i> (Wallich)	*	*	*			*
			<i>Cruciplacolithus tenuis</i> (Stradner)	*					
			<i>Cyclicargolithus floridanus</i> (Roth and Hay)	*	*		*		
			<i>Dictyococcities hesslandii</i> (Haq)		*	*	*	*	*
			<i>Dictyococcites scrippsae</i> Bukry and Percival	*					
			<i>Discoaster gemmifer</i> Stradner						*
			<i>Discoaster saipanensis</i> Bramlette and Riedel			*			
			<i>Ericsonia formosa</i> (Kamptner)	*	*		*		
			<i>Micrantolithus basquensis</i> Martini	*					
			<i>Neococcolithus dubius</i> (Deflandre)	*					
			<i>Pemma</i> sp. Klumpp	*					
			<i>Pontosphaera obliquipons</i> (Deflandre)					*	
			<i>Pontosphaera pectinata</i> (Bramlette and Sullivan)	*	*				
			<i>Prinsius martinii</i> (Perch-Nielsen)			*			
			<i>Reticulofenestra dictyoda</i> (Deflandre)		*				
			<i>Reticulofenestra gelida</i> (Geitzenauer)					*	
			<i>Reticulofenestra haqii</i> Backman	*		*	*		
			<i>Reticulofenestra hampdanensis</i> Edwards			*			
			<i>Reticulofenestra lockeri</i> Müller				*	*	
			<i>Reticulofenestra minuta</i> Roth	*					
			<i>Reticulofenestra minutula</i> (Gartner)	*					*
			<i>Reticulofenestra pseudumbilicus</i> (Gartner)	*	*		*	*	*
			<i>Scyphosphaera globulata</i> Bukry and Percival	*					
			<i>Sphenolithus abies</i> Deflandre				*		
			<i>Sphenolithus moriformis</i> (Bronnimann and Stradner)	*	*				*
			<i>Sphenolithus radians</i> Deflandre	*	*				*
			<i>Thoracosphaera heimii</i> Lohmann	*	*		*	*	
			<i>Toweius crassus</i> (Bramlette and Sullivan)						*
			UPPER CRETACEOUS	?	?	<i>Aspidolithus parvus parvus</i> (Stradner)			*
<i>Braarudosphaera bigelowii</i> (Gran and Braarud)						*			
<i>Braarudosphaera discula</i> Bramlette and Riedel								*	
<i>Cribrosphaerella ehrenbergii</i> (Arkhangelsky)							*		
<i>Cyclagelosphaera reinhardtii</i> (Perch-Nielsen)							*		
<i>Ellipsogelosphaera britannica</i> (Stradner)									*
<i>Ellipsogelosphaera ovata</i> (Bukry)									*
<i>Microrhabdulus attenuatus</i> Deflandre								*	
<i>Micula decussata</i> Vekshina									
<i>Micula staurophora</i> (Gardet)						*			
<i>Prediscosphaera cretacea</i> (Arkhangelsky)						*		*	*
<i>Quadrum gartneri</i> Prins and Perch-Nielsen									*
<i>Rhomboaster cuspis</i> Bramlette and Sullivan								*	
<i>Stradneria crenulata</i> Bramlette and Martini							*		
<i>Thoracosphaera saxea</i> Stradner	*						*		
<i>Watznaueria barnesae</i> Black	*	*				*	*	*	*

of dinoflagellate cysts such as *Thoracosphaera* spp. (*Th. granifera*, *Th. tuberosa*) that represent shallow-sea conditions indicates that the deposition, which generally represents the Holocene, records a steadily deepening marine environment. Although there are no nannofossils at some levels (as specified in the tables for the drilling samples), the presence of dinoflagellate cysts such as *Thoracosphaera* spp., which represent a shallow-marine setting (*Th. granifera*), suggests that there has been at least one short-period decrease in sea level during the Holocene.

In the mud samples of the GMK-K1 core, 18 nannofossil species were determined, of which 15 are “synsedimentary” and 3 are from Cenozoic and Cretaceous rocks, which of them are recorded as 3 dinoflagellate species (*Thoracosphaera* spp.), 2 of them are “synsedimentary” and 1 of them is “reworked” (Table 4). In the mud samples from the core GMK-K2, 23 nannofossil species are “reworked”, of which 12 are “synsedimentary”, 11 are from the Cenozoic and Cretaceous, which of them are recorded as 3 dinoflagellate species, 2 of them are “synsedimentary” and 1 of them is “reworked” (*Thoracosphaera* spp). Also, in mud samples from core GMK-K3, 20 nannofossil species are present, of which 14 are “synsedimentary”, and 6 were reworked from the Cenozoic and Cretaceous. In addition, 1 “synsedimentary” dinoflagellate species (*Thoracosphaera granifera*) was observed. In the mud samples of GMK-K4, 18 nannofossil species occur, of which 6 are “synsedimentary” and 12 were “reworked” from the Cenozoic and Cretaceous (Table 4).

In addition to an abundance of *Emiliania huxleyi* and *Reticulofenestra parvula* in the mud samples from GMK-K2 (75,00-76,00 m) and GMK-K3 (98,00 - 99,00 m), two deepest cores in Gemlik Bay, the existence of *Gephyrocapsa oceanica* and small *Gephyrocapsa* species (survivors after extinction of *Gephyrocapsa omega*) indicate that marine sedimentation began at the base of Holocene.

#### 3.4. Diatoms

The number of genera and species of diatoms in the drilling and core samples is very small. In the drilling of GMK-2, *Stephanodiscus lucens* Hustedt occurs at 39,00-39,10 m also, *Perissonoe cruciata*

(Janisch and Robenhorst) Andrews and Stocizel at 41,00-41,10 and 41,40-41,50 m. In the drilling of GMK-3, *Eunotia* sp. occurs at 7,20-7,40 m and *Campylodiscus echeneis* Ehrenberg at 8,20-8,40 m. (Plate 2). Between these, *Stephanodiscus lucens* is a planktonic freshwater form commonly found in the high-temperature waters. *Perissonoe cruciata* (Janisch and Robenhorst) is a warm-water form, which is common in marine, littoral environments. *Eunotia* sp. is common in waters with both low and high temperatures, and is also a benthic (epiphytic, epilithic) fresh water form; it is common in low-nutrient (oligosaprobic) waters, which have both low and high water temperatures (oligotrophic, eutropic) and pH>7 (alkaline). *Campylodiscus echeneis* Ehrenberg is generally a marine form (Krammer-Lange Bertalot, 1988, 1991; Soinien and Könönen, 2004; Round et al., 2007; Lerin and Cambra, 2007; Lange Bertalot et al., 2011; Krizmanici et al., 2015).

In terms of the diatom flora, *Stenopterobia* sp., which is a cosmopolitan form, occurs in the cores of GMK-K3 between 98,30-98,40 m and 98,60-98,70 m, and individuals belonging to the same genus rarely occur at 21,90-22,00 m in GMK-K4 (Krammer, Lange-Bertalot, 1988 and URL) (Plate 2).

*Stephanodiscus lucens* Hustedt and *Eunotia* sp., which are encountered in small numbers amongst the diatom assemblages in the drilling samples, are freshwater forms and are likely to have been transported to the environment by rivers. *Perissonoe cruciata* (Janisch and Robenhorst) and *Campylodiscus echeneis* Ehrenberg are marine forms. Especially, *Perissonoe cruciata* (Janisch and Robenhorst) represents both a marine environment and a littoral warm-water environment.

The diatom species and genera described in boreholes and cores are widespread throughout the Holocene.

#### 3.5. Molluscs

The drilled samples contain very rich gastropod and bivalve assemblages. In drilled samples of GMK-1, GMK-2, GMK-3, there is a rich community of gastropod, namely: *Gibbula albida* (Gmelin), *G. rarilineata* (Michaud), *Phorcus mutabilis* (Philippi), *Bittium latreillii* (Payraudeau), *B. reticulatum* (da Costa), *B. submamillatum* (de Rayneval and

Ponzi), *Turritella communis* Risso, *Similiphora similior* (Bouchet and Guillemot), *Epitonium clathrus* (Linné), *Melarhappe neritoides* (Linné), *Rissoa auriformis* Pallary, *R. splendida* Eichwald, *Pussilina inconspicua* (Alder), *P. lineolata* (Michaud), *P. marginata* (Michaud), *Alvania geryonia* (Nardo), *Obtusella intersecta* (S. Wood), *O. macilentata* (Monterosato), *Hydrobia acuta* (Draparnaud), *Hyalia vitrea* (Montagu), *Euspira intricata* (Donovan), *Tritia pygmaea* (Lamarck), *Clathromangelia strigilata* Pallary, *Sorgenfreispira brachystoma* (Philippi), *Mangelia scabrida* Monterosato, *Mangelia* sp., *Raphitoma* sp., *Eulimella acicula* (Philippi), *Parthenina juliae* (de Folin), *Megastomia conoidea* (Brocchi), *Odostomia megerlei* (Locard), *Ondina crystallina* Locard, *Turbonilla acutissima* Monterosato, *Retusa leptoneilema* (Brusina), *R.*

*minutissima* (Monterosato), *R. obtusa* (Montagu), *Cylichna cylindracea* (Pennant) (Table 5a), and bivalves which are; *Nucula hanleyi* Winckworth, *N. sulcata* Bronn, *Lembulus pella* (Linné), *Mytilus galloprovincialis* Lamarck, *Musculus subpictus* (Cantraine), *Modiolula phaseolina* (Philippi), *Ostrea edulis* Linné, *Ctena decussata* (O. G. Costa), *Lucinella divaricata* (Linné), *Myrtea spinifera* (Montagu), *Kurtiella bidentata* (Montagu), *Acanthocardia paucicostata* (G. B. Sowerby II), *Parvicardium scriptum* (Bucquoy, Dautzenberg and Dollfus), *Papillicardium papillosum* (Poli), *Spisula solida* (Linné), *S. subtruncata* (da Costa), *Abra prismatica* (Montagu), *Atlantella pulchella* (Lamarck), *Timoclea ovata* (Pennant), *Gouldia minima* (Montagu), *Pitar mediterraneus* (Aradas and Benoit), *P. rudis* (Poli), *Ruditapes decussatus* (Linné), *Corbula gibba* (Olivi),

Table 5a- The mollusc content of the drillings.

MOLLUSCA		BOREHOLES		
		GMK-1	GMK-2	GMK-3
GASTROPODA	<i>Gibbula albida</i> (Gmelin)	*		*
	<i>Gibbula rarilineata</i> (Michaud)			*
	<i>Phorcus mutabilis</i> (Philippi)		*	
	<i>Bittium latreillii</i> (Payraudeau)	*	*	*
	<i>Bittium reticulatum</i> (da Costa)	*		*
	<i>Bittium submammillatum</i> (de Rayneval and Ponzi)	*	*	*
	<i>Turritella communis</i> Risso	*	*	
	<i>Similiphora similior</i> (Bouchet and Guillemo)	*		
	<i>Epitonium clathrus</i> (Linnaeus)	*		
	<i>Melarhappe neritoides</i> (Linnaeus)	*		
	<i>Rissoa auriformis</i> Pallary		*	*
	<i>Rissoa splendida</i> Eichwald	*	*	*
	<i>Pussilina inconspicua</i> (Alder)	*	*	
	<i>Pusillina lineolata</i> (Michaud)	*	*	*
	<i>Pusillina marginata</i> (Michaud)	*		
	<i>Alvania geryonia</i> (Nardo)	*		
	<i>Obtusella intersecta</i> (S. Wood)	*	*	
	<i>Obtusella macilentata</i> (Monterosato)	*		
	<i>Hyalia vitrea</i> (Montagu)		*	
	<i>Hydrobia acuta</i> (Draparnaud)	*		
	<i>Euspira intricata</i> (Donovan)	*	*	*
	<i>Tritia pygmaea</i> (Lamarck)	*	*	
	<i>Tritia reticulata</i> (Linnaeus)			*
	<i>Sorgenfreispira brachystoma</i> (Philippi)	*		
	<i>Clathromangelia strigilata</i> Pallary		*	
	<i>Mangelia scabrida</i> Monterosato	*	*	
<i>Raphitoma</i> sp.	*	*		
<i>Eulimella acicula</i> (Philippi)	*	*		
<i>Parthenina juliae</i> (de Folin)	*			

Table 5a- continued.

GASTROPODA	<i>Megastomia conoidea</i> (Brocchi)		*	*
	<i>Odostomia megerlei</i> (Locard)	*	*	
	<i>Turbonilla acutissima</i> Monterosato	*		
	<i>Retusa leptoneilema</i> (Brusina)	*		
	<i>Retusa obtusa</i> (Montagu)	*	*	
	<i>Cylichna cylindracea</i> (Pennant)	*	*	
BIVALVIA	<i>Nucula hanleyi</i> Winckworth		*	
	<i>Nucula sulcata</i> Bronn	*		
	<i>Lembulus pella</i> (Linnaeus)	*	*	*
	<i>Musculus subpictus</i> (Cantraine)	*		
	<i>Modiolula phaseolina</i> (Philippi)	*	*	*
	<i>Ostrea edulis</i> Linnaeus			*
	<i>Ctena decussata</i> (O. G. Costa)	*		
	<i>Lucinella divaricata</i> (Linnaeus)	*	*	*
	<i>Myrtea spinifera</i> (Montagu)	*	*	
	<i>Kurtiella bidentata</i> (Montagu)	*	*	
	<i>Acanthocardia paucicostata</i> (G. B. Sowerby II)	*		
	<i>Parvicardium scriptum</i> (Bucquoy, Dautzenberg and Dollfus)	*	*	*
	<i>Papillicardium papillosum</i> (Poli)	*		
	<i>Spisula solida</i> (Linnaeus)	*	*	
	<i>Spisula subtruncata</i> (da Costa)	*		
	<i>Abra prismatica</i> (Montagu)		*	
	<i>Atlantella pulchella</i> (Lamarck)	*	*	
	<i>Timoclea ovata</i> (Pennant)	*	*	
	<i>Gouldia minima</i> (Montagu)	*	*	*
	<i>Pitar mediterraneus</i> (Aradas and Benoit)	*		
<i>Pitar rudis</i> (Poli)	*			
<i>Ruditapes decussatus</i> (Linnaeus)	*			
<i>Corbula gibba</i> (Olivi)	*			
<i>Hiatella arctica</i> (Linnaeus)	*			
<i>Hiatella rugosa</i> (Linnaeus)			*	

*Hiatella arctica* (Linné), *H. rugosa* (Linné) were found (Table 5a) (Cossignani et al., 2011; Scaperrotta et al., 2009-2015) (Plates 3 and 4).

When the cores of GMK-K1, GMK-K2, GMK-K3 and GMK-K4 are considered in terms of mollusc assemblages; i.e. the following bivalves were identified: *Kelliella miliaris* (Philippi), *Spisula subtruncata* (da Costa), *Parvicardium exiguum* (Gmelin), *Loripes dentatus* (Defrance), *Timoclea ovata* (Pennant), *Corbula gibba* (Olivi), *Abra* sp., from gastropods; *Turritella communis* Risso, *Alvania cimicoides* (Forbes), *Panthenina intersincta* (J. Adams), *Bittium reticulatum* (de Costa), *Ecrobia* cf. *maritima*, *Rissoa* sp., *Turbonilla* sp.; the scaphopod, *Dentalium* sp. was also observed (Table 5b). The above assemblage is Holocene and characterizes a shallow-marine environment. Owing to the partial

fossilization of the shells, they are considered to be of Early Holocene age (Neveeskaja, 1965, 1974; Perna, 2003; Nielsen et al., 2006; Taviani et al., 2014; Çağatay et al., 2015; Büyükmeriç, 2016).

#### 4. Discussion and Conclusion

As a result of the studies, it is understood that the foraminifer assemblage of the Gulf of Gemlik was under the influence of the Mediterranean-Aegean Sea communities. It is remarkable that there is a great difference between the samples from the drilling and the cores in terms of the number of genera and species. Although, a total of 22 genera and 38 species, characterizing the infra-littoral zone in drilling samples were identified, 40 genera and 58 species were observed, which characterize the circa-littoral zone in the cores. In addition, to observing large

Table 5b- The mollusc content of core samples.

MOLLUSCA	CORES			
	GMK-K1	GMK-K2	GMK-K3	GMK-K4
<i>Kelliella miliaris</i> (Philippi)	*	*	*	*
<i>Spisula subtruncata</i> (da Costa)		*	*	*
<i>Parvicardium exiguum</i> (Gmelin)		*		*
<i>Loripes dentatus</i> J.L.M. Defrance				*
<i>Timoclea ovata</i> (Pennant)				*
<i>Corbula gibba</i> Olivi				*
<i>Abra</i> sp.				*
<i>Turritella communis</i> Risso		*		
<i>Alvania cimicoides</i> (Forbes)		*		
<i>Parthenina interstincta</i> (J. Adams)				*
<i>Bittium reticulatum</i> (da Costa)				
<i>Ecrobia</i> cf. <i>maritima</i> (Milaschewitsch)	*			
<i>Rissoa</i> sp.		*		
<i>Turbonilla</i> sp.				*
<i>Dentalium</i> sp.	*			

number of species of Black Sea origin, *Ammonia parasovica* in samples from GMK-K4, taken from the eastern part of the gulf, is another notable feature for the study area, and is the first record of this species in the Gulf of Gemlik. The same genus and species were encountered at different levels of the drillings of KS-2, S-5 and S-3, carried out in the Gulf of İzmit (Meriç et al., 1995). It is thought that this species of Black Sea origin continued its life in the Gulf of Gemlik after the Gulf of İzmit in Holocene. Both the boreholes and the core samples have a very rich fauna in terms of the ostracod assemblage. The ostracod assemblage of the boreholes and the core samples resembles each other. *Loxocauda pellucida*, which is known in the Mediterranean, Aegean and Marmara Seas, was encountered only in core samples. The ostracod genera and species identified belong to typical Mediterranean-Aegean Sea fauna.

The nannofossil assemblage in which *Emiliania huxleyi*, *Reticulofenestra parvula*, *Gephyrocapsa oceanica*, *Scyphosphaera porosa* species are present in the boreholes and cores, characterize an open shallow-marine environment, typical of the Holocene period, including the NN21 *Emiliania huxleyi* biozone in the samples studied. The nannoplankton community is richer than in terms of the number of genera and species in the Gulf of İzmit.

Although the drilling boreholes and cores represent a sparse community in terms of the diatom flora, its actual presence in the eastern Gulf of Gemlik is first emphasized by this study. When compared, it is observed that there is a great difference between the diatom assemblage of the Gulf of Gemlik and that of the İzmit Lake. The presence of fresh-water diatoms and ostracods in some boreholes and drilling samples also suggests that there was fresh water outflow to the marine Gulf of Gemlik at certain times.

The mollusc assemblage on the other hand includes characteristic Mediterranean genera and species. According to the data obtained, the Gulf of Gemlik remained under Mediterranean influence during the Holocene period. A remarkable feature is the great abundance of *Turritella communis* in a mud-rich environments between 27,10 to 40,00 m in GMK-1 and also between 35,10-40,80 m in the GMK-2 drilling. This shows that the life conditions were suitable in the Gulf of Gemlik for this species. This is the only species that adapted itself to the ambient low-oxygen conditions in the gulf. When all of the microfossil and the macrofossil findings were evaluated, it was determined that the Gulf of Gemlik assemblage is typical of the Mediterranean-Aegean Sea community during the Holocene period.



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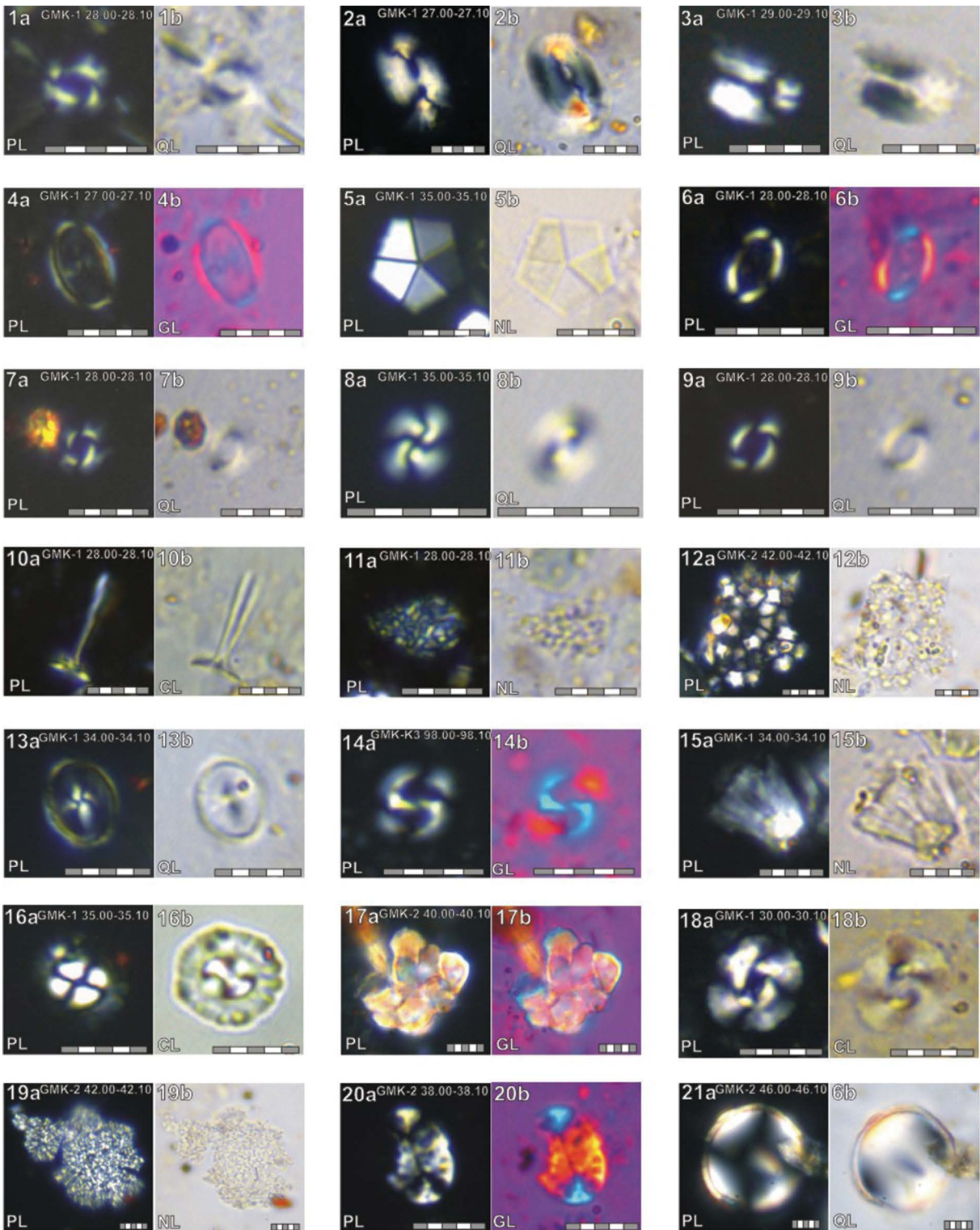
## **PLATES**

**PLATE 1**

Holocene nannofossil species found in drill and core samples of the Gemlik Bay (PL: polarized light, NL: normal light, CL: contrast light, GL: with gypsum wedge, QL: with quartz wedge; letter and number order on the upper left corner: drilling number and rock sampling depth, linear scale: 5 µm):

1. *Emiliana huxleyi*,
2. *Helicosphaera carteri*,
3. *Helicosphaera hyalina*,
4. *Syracosphaera histrica*,
5. *Braarudosphaera bigelowii*,
6. *Coronosphaera mediterranea*,
7. *Emiliana huxleyi*,
8. *Dictyococcites productus*,
9. *Reticulofenestra parvula*,
10. *Rhabdosphaera clavigera*,
11. *Thoracosphaera granifera*,
12. *Anthosphaera* sp.,
13. *Syracosphaera pulchra*,
14. *Gephyrocapsa oceanica*,
15. *Scyphosphaera porosa*,
16. *Coccolithus pelagicus*,
17. *Bonetia acuta* “Ascidian spicule”,
18. *Reticulofenestra dictyoda* “reworked”,
19. *Thoracosphaera tuberosa*,
20. *Pontosphaera pectinata* “reworked”,
21. *Scyphosphaera globulata*.

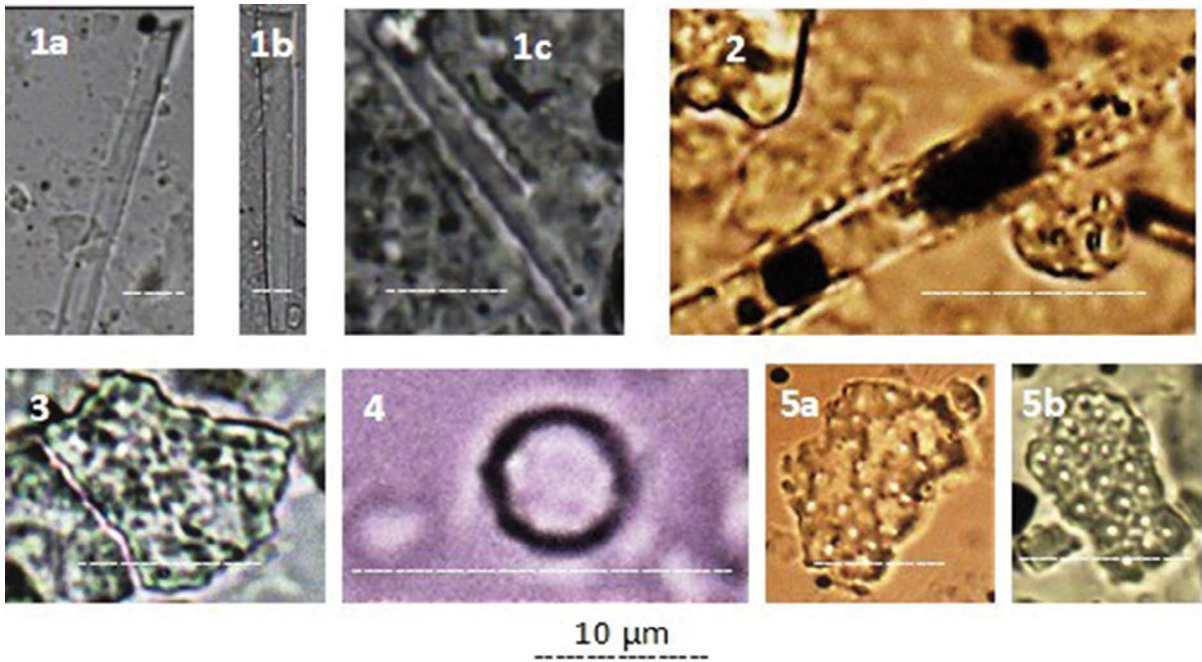




## PLATE 2

Diatom species found in drill and core samples from the Gulf of Gemlik.

1. *Stenopterobia* sp., a) GMK-K3, 98,30-98,40 m, b). GMK-K3, 98,60-98,70 m, c) GMK-K4, 21,90-22,00 m.
2. *Eunotia* sp., GMK-3, 07,20-07,40 m.
3. *Campylodiscus echeneis*, GMK-3, 08,20-08,40 m.
4. *Stephanodiscus lucens*, GMK-2, 39,00-39,10 m.
5. *Perissonoe cruciata*, a. GMK-2, 41,00-41,10 m, b. GMK-2, 41,40-41,50 m.

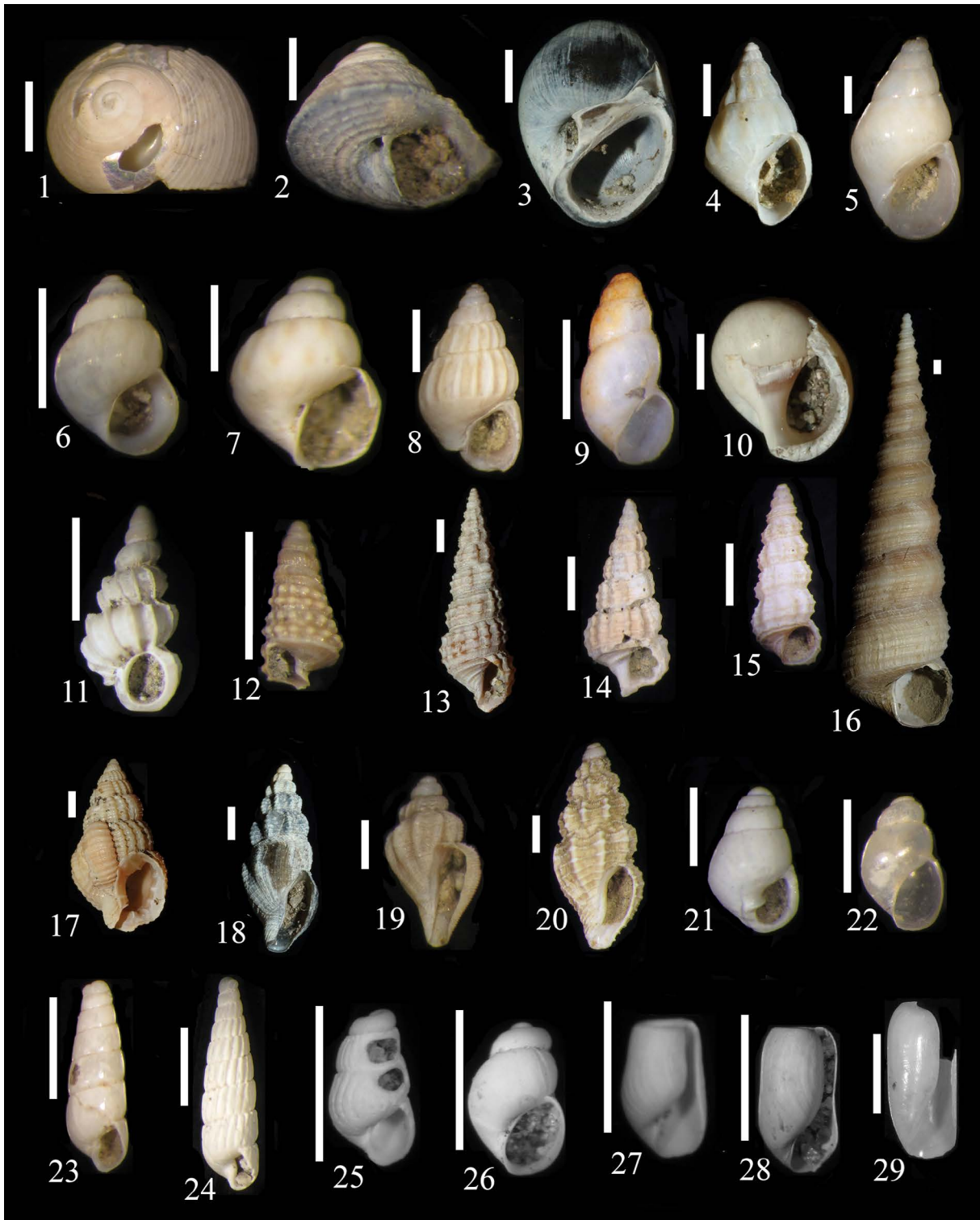


**PLATE 3**

Gastropod species found in drill samples from the Gulf of Gemlik.

1. *Phorcus mutabilis*, Gemlik-2, 40,20-40,30 m.
2. *Gibbula albida* Gemlik-1, 31,40-31,50 m.
3. *Melarhaphe neritoides*, Gemlik-1, 34,30-34,50 m.
4. *Rissoa splendida*, Gemlik-2, 38,00-38,10 m.
5. *Rissoa auriformis*, Gemlik-3, 07,20-07,40 m.
6. *Pussilina inconspicua*, Gemlik-2, 40,50-40,60 m.
7. *Pusillina lineolata*, Gemlik-1, 29,90-30,00 m.
8. *Pusillina marginata*, Gemlik-1, 30,50-30,60 m.
9. *Hyala vitrea*, Gemlik-2, 41,10-41,20 m.
10. *Euspira intricata*, Gemlik-1, 29,90-30,00 m.
11. *Epitonium clathrus*, Gemlik-1, 30,20-30,30 m.
12. *Similiphora similior*, Gemlik-1, 33,70-33,80 m.
13. *Bittium latreillii*, Gemlik-1, 33,50-33,60 m.
14. *Bittium reticulatum*, Gemlik-3, 07,20-07,40 m.
15. *Bittium submammillatum*, Gemlik-3, 06,40-06,60 m.
16. *Turritella communis*, Gemlik-2, 35,50-35,60 m.
17. *Tritia pygmaea*, Gemlik-1, 34,50-34,60 m.
18. *Sorgenfreispira brachystoma* ), Gemlik-1, 35,10-35,20 m.
19. *Mangelia scabrida*, Gemlik-1, 34,00-34,10 m.
20. *Clathromangelia strigilata*, Gemlik-2, 35,50-35,60 m.
21. *Megastomia conoidea*, Gemlik-3, 06,20-06,40 m.
22. *Ondina crystallina*, Gemlik-2, 46,40-46,50 m.
23. *Eulimella acicula*, Gemlik-2, 41,80-41,90 m.
24. *Turbonilla acutissima*, Gemlik-1, 33,40-33,50 m.
25. *Parthenina juliae*, Gemlik-1, 37,60-37,70 m.
26. *Odostomia megerlei*, Gemlik-1, 34,60-34,70 m.
27. *Retusa leptoneilema*, Gemlik-1, 34,70-34,80 m.
28. *Retusa obtusa*, Gemlik-1, 29,90-30,00 m.
29. *Cylichna cylindracea*, Gemlik-1, 30,30-30,40 m.

(Scale = 1 mm)





**PLATE 4**

Bivalve species found in drill samples from the Gulf of Gemlik.

1. *Nucula hanleyi*, Gemlik-2, 42,00-42,10 m.
2. *Nucula sulcata*, Gemlik-1, 35,00-35,10 m.
3. *Lembulus pella*, Gemlik-2, 39,20-39,30 m.
4. *Mytilus galloprovincialis*, Gemlik-3, 08,80-09,00 m.
5. *Ostrea edulis*, Gemlik-3, 06,20-06,40 m.
6. *Musculus subpictus*, Gemlik-1, 29,00-29,10 m.
7. *Modiolula phaseolina*, Gemlik-2, 44,60-44,70 m.
8. *Ctena decussata*, Gemlik-1, 34,00-34,10 m.
9. *Lucinella divaricata*, Gemlik-1, 28,10-28,20 m.
10. *Myrtea spinifera*, Gemlik-1, 27,80-27,90 m.
11. *Kurtiella bidentata*, Gemlik-1, 28,80-28,90 m.
12. *Acanthocardia paucicostata*, Gemlik-1, 27,90-28,00 m.
13. *Parvicardium scriptum*, Gemlik-1, 27,70-27,80 m.
14. *Papillicardium papillosum*, Gemlik-1, 3,30-33,40 m.
15. *Spisula solida*, Gemlik-2, 36,80-36,90 m.
16. *Spisula subtruncata*, Gemlik-1, 34,10-34,20 m.
17. *Abra prismatica*, Gemlik-2, 37,70-37,80 m.
18. *Atlantella pulchella*, Gemlik-1, 32,80-32,90 m.
19. *Timoclea ovata*, Gemlik-1, 27,80-27,90 m.
20. *Gouldia minima*, Gemlik-1, 33,60-33,70 m.
21. *Pitar mediterraneus*, Gemlik-1, 34,20-34,30 m.
22. *Pitar rudis*, Gemlik-1, 34,20-34,30 m.
23. *Ruditapes decussatus*, Gemlik-1, 34,20-34,30 m.
24. *Corbula gibba*, Gemlik-2, 42,10-42,20 m.
25. *Hiatella arctica*, Gemlik-1, 33,40-33,50 m.
26. *Hiatella rugosa*, Gemlik-3, 6,40-6,60 m.

(Scale = 1 mm)

