# MORPHOLOGICAL CHARACTERIZATION OF STREPTOCOCCUS THERMOPHILUS AND LACTOBACILLUS DELBRUECKII SUBSP. BULGARICUS VIRULENT PHAGES

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#### Abstract

In this study, 25 phages of *S. thermophilus* and 25 phages of *L. bulgaricus* were inspected and identified morphologically by electron microscopy. In *S. thermophilus* phages the diameters of hexagonal heads were found to range between 47-74 nm, while non-contractile tails were 182 to 290 nm long and 7-14 nm wide. In these phages tail plaque, collar and fiber like structures were not found. Identified *S. thermophilus* phages were placed in the *Siphoviridae* family according to Ackermann's and/or Group B in Bradley's classification. Also for *L. bulgaricus* phages isometric hexagonal capsid and non-contractile tails were between 117-162 nm long and 7 to 13 nm wide. These phages were also placed in *Siphoviridae* family in Ackermann's classification and/or group B in Bradley's. Importantly, it was also determined that in some of these phages collar and tail plaque structures were present. It is thought that minor modifications in the preparation of the *S. thermophilus* and *L. bulgaricus* phages, and the type of electron microscope used were significant factors that affected the visibility of the tail structures.

Keywords: S. thermophilus, L. bulgaricus, phage, morphological characterization, electron microscope

## STREPTOCOCCUS THERMOPHİLUS VE LACTOBACİLLUS DELBRUECKİİ SUBSP. BULGARİCUS VİRÜLENT FAJLARININ MORFOLOJİK KARAKTERİZASYONU

#### Özet

Bu çalışmada 25 adet *S. thermophilus* ve 25 adet *L. bulgaricus* fajının elektron mikroskobik incelemesi yapılarak morfolojik karakterizasyonu gerçekleştirilmiştir. *S. thermophilus* fajlarında izometrik, hegzagonal baş çapının 53-74 nm, kontraktil olmayan kuyruk uzunluğunun 182-290 nm ve kuyruk genişliğinin de 7-14 nm arasında değiştiği görülmüştür. Bu fajlarda yaka, kuyruk plağı ve fibril benzeri yapıya rastlanmamıştır. İncelenen tüm fajlar, elde edilen verilere dayanılarak diğer *S. thermophilus* fajları gibi Ackermann sınıflaması *Siphoviridae* familyasına ve/veya Bradley sınıflaması B grubuna dâhil edilmiştir. *S. thermophilus* fajlarında olduğu gibi *Lb. bulgaricus* fajlarında da izometrik, hegzagonal kapsit ve kontraktil olmayan kuyruk yapısı belirlenmiştir. Kapsit çapları 47-73 nm arasında değişirken, kontraktil olmayan kuyruk uzunlukları 117-162 nm ve kuyruk enleri 7-13 nm arasında bulunmuştur. Ackermann sınıflaması *Siphoviridae* familyasına ve/veya Bradley sınıflaması B grubuna dâhil edilen bu fajlarda yaka, kuyruk tablası ve fibril yapısının varlığı dikkat çekmiştir. *S. thermophilus* ve *L. bulgaricus* faj örneklerinin hazırlanmasındaki farklılıkların ve kullanılan elektron mikroskop tiplerinin kuyruk yapılarının görünebilirliğini etkilediği düşünülmüştür.

Anahtar kelimeler: S. thermophilus, L. bulgaricus, faj, morfolojik karakterizasyon, elektron mikroskop

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## INTRODUCTION

Knowledge of thermophilic phages is quite restricted when compared to that of lactococcal phages (1), owing to the high incidence of phage infections in the cheese production industry (2). Thus lactococcal phages have been under investigation since first being determined by Whitehead and Cox in 1935 (3). However the first phage specific to Streptococcus thermophilus (S. thermophilus) was determined in 1952 (4) and a Lactobacillus delbrueckii subsp. bulgaricus (L. bulgaricus) phage was isolated first by Reddy and Reinbold in 1974 with further investigations being done in 1982 (5). In addition, in recent years researches have focused more on thermophilic phages because of the frequent infections in cheese and yogurt plants which they cause (6).

To this day, morphological studies have shown *S. thermophilus* and *L. bulgaricus* phages to have a single morphotype. All have a hexagonal capsid and a non-contractile tail, and all of them are placed in the *Siphoviridae* family according to Ackermann's classification and/or in group B in Bradley's classification (2, 5-9). A *L. bulgaricus* phage that shows a different morphotype from other thermophilic phages is the only phage having a contractile tail and is placed in the *Myoviridae* family according to Ackermann's classification (5).

During the last 6 years, a thermophilic phage problem has periodically occurred in modern yogurt factories in Turkey (10). In order to solve the phage problem, it is important that, in addition to conventional prevention measures, factories should rotate between commercial strains the sensitivity of which has been tested against native phage collections (11, 12). It is also important that factories should use natural (indigenous) phage resistant strains because it has been shown that natural strains are much more resistant to phages than commercial strains (6). For this purpose 23 S. thermophilus and 25 L. bulgaricus phages were provided from our phage collection. This research focuses on the morphological characterization of these 48 phages using a transmission electron microscope (TEM). Owing to the limited data on the isolation of S. thermophilus phages from raw milk samples (12) the incidence of such phages in raw milk was also investigated. Two S. thermophilus phages isolated from raw milk samples were characterized morphologically and identified taxonomically along with other 48 phages.

## MATERIALS AND METHODS

## Phage and strain cultures

Twenty three S. thermophilus phages and twenty five L. bulgaricus phages were provided from our collection. These phages had been isolated and purified from bulk, yogurt, whey and ayran-Turkish buttermilk collected from dairy plants in the Ankara region. Bacterial strains (S. thermophilus B3, 709, 231 and L. bulgaricus Y4, V1, V2, 231) from which the phages were isolated were mixed starter cultures used industrially. The two S. thermophilus phages ( $\Phi$ 1B3-A,  $\Phi$ 2B3-A) from raw milk samples from the Afyon region were also isolated using the B3 industrial strain as a host organism. The isolation of all phages was carried out single plaque isolation technique as described previous study (13). To study the growth of S. thermophilus strains and their phages, modified M17 Broth (thMl7 Broth), modified M17 Agar (th M17 Agar) and modified M17 Soft Agar (th M17 Soft Agar) (0.45% agar) were used (14). L. bulgaricus strains and their phages were grown in MRS Broth (Merck, Darmstadt, Germany), MRS Agar (1.5% agar) and MRS Soft Agar (0.6% agar). In order to achieve a better adsorption of L. bulgaricus phages, CaCl, (10 mmol/l) was added to all three types of culture medium (15). All cultures and phages were incubated for 18 h at 43°C.

## **Electron microscopy of phages**

S. thermophilus and L. bulgaricus phages were concentrated through the centrifugation of phage lysates containing 107-108 phage particles (pfu/ ml) for 2 h at 20000 rpm at 4°C. Some of S. thermophilus phage pellets were dissolved in 20 µl 0.3 M ammonium acetate and an equal volume of dye solution (10 µl amonium molibdate and 10 µl sodium phosphotungstate, 2 % - 3 %, w/v, pH 5.0  $\pm$ 0.02, Sigma Chem. Co., USA) was added (16). The phage-dye mixture was then dropped on to 400 mesh grids covered with carbon formvar (3.05 mm, Agar Scientific Ltd. UK). After waiting for 15 minutes, the excess dye was removed. Electron micrographs were taken with TEM, models JEOL JEM S 100 and JEOL JEM 100-C under 80 kV power. The pellets of the other *S. thermophilus* ( $\Phi$ 1B3-A, Ф2ВЗ-А, ФВЗ-Х12, ФВЗ-Х13, ФВЗ-Х18, Ф2З1-Х9,  $\Phi$ 231-X23) and all of the *L. bulgaricus* phages were suspended in 0.1 % (w/v) ammonium acetate first (17), then 20  $\mu$ l 3 % phosphotungstate was added and finally they were placed on the grids (18). Fifteen minutes later, the grids were washed with deionised water and dried on a filter paper (19). Micrographs of the phages were taken under 80 kV power by using a LEO 906 E electron microscope. Phage size calculations are based on the averages of 5 to 10 measurements (20).

## RESULTS

#### Morphology of S. thermophilus phages

Phages isolated by using *S. thermophilus* 709, 231 and B3 industrial hosts had isometric, hexagonal and assumptively icosahedral capsids, 47-74 nm in diameter, and non-contractile tails 182-290 nm long and 7-14 nm wide (Table 1). None of the phages displayed collar, tail plaque or fiber structures. The presence of a plaque-like structure was suspected only in phages  $\Phi$ 231-X9 and  $\Phi$ B3-X18 (Figure 1a,b). The morphological characterization of 25 *S. thermophilus* phages showed that the phages belong to the *Siphoviridae* family in Ackermann's and/or in Group B in Bradley's classification.

Table 1 Morphological properties of S. thermophilus phages



Figure 1. S. thermophilus phages. (a) Φ231-X9, 60000x; (b) ΦB3-X18, 100000x; (c) Φ709-X4, 45 000x

#### Morphology of L. bulgaricus phages

The four industrial strains of *L. bulgaricus* were 231, Y4, V1 and V2, and were used as host organisms for these 25 phages. The capsids were 47 to 73 nm in diameter and isometric hexagonally shaped. It was determined that all of the phages carried non-contractile tails without a cover structure and the tails were 117 to 162 nm long and 7 to 13 nm wide (Table 2). While in the phages  $\Phi$ HV1 (Figure 2a),  $\Phi$ GV2,  $\Phi$ G1V2 and  $\Phi$ G4V2 the presence of the structures like collar pieces and tail plaques were determined, in the phages  $\Phi$ H2Y4,  $\Phi$ JY4,  $\Phi$ FY4

Phage Code	Capsid diameter (nm)	Tail length (nm)	Tail width (nm)	Collar (nm)	Tail plaque (nm)	Fiber
Φ709-X1	64	235	12	_1	-	-
Φ709-X2	61	245	12	-	-	-
Ф709-ХЗ	63	190	8	-	-	-
Φ709-X4	74	290	10	-	-	-
Φ709-X5	67	220	12	-	-	-
Ф231-Х6	62	182	12	-	-	-
Ф231-Х7	69	290	12	-	-	-
Ф231-Х9	60	230	12	-	-	-
Ф231-Х10	68	221	12	-	-	-
ΦB3-X11	53	193	9	-	-	-
ΦB3-X12	53	214	9	-	-	-
ФВЗ-Х1З	60	220	12	-	-	-
ΦB3-X14	55	235	13	-	-	-
ΦB3-X15	73	230	14	-	-	-
ФВЗ-Х16	54	234	11	-	-	-
ΦB3-X17	57	210	10	-	-	-
ΦB3-X18	54	220	10	-	-	-
ФВЗ-Х19	63	217	12	-	-	-
ФВЗ-Х20	47	224	10	-	-	-
Ф231-Х21	62	272	7	-	-	-
Ф231-Х22	67	230	10	-	-	-
Ф231-Х23	73	220	10	-	-	-
Ф1В3-А	57	244	10	-	-	-
Ф2В3-А	57	244	10	-	-	-

Phage Code	Capsid diameter (nm)	Tail length (nm)	Tail width (nm)	Collar (nm)	Tail plaque (nm)	Fiber
H2Y4Φ(X1)	54	134	11	_1	30x15	-
MY4Φ(X2)	55	151	11	-	-	-
JY4Φ(X3)	55	138	13	-	32x15	-
FY4Φ(X4)	55	141	13	-	24x13	-
НЗҮ4Ф(Х5)	58	131	13	-	22x13	-
H1Y4Φ(X6)	49	128	11	-	22x14	-
IY4Φ(X7)	56	156	10	-	+2	-
SİYY4Φ(X8)	58	142	9	-	+	-
709BY4Ф(X9)	50	138	9	-	15x9	-
SİBY4Φ(X10)	55	138	11	-	22x12	-
GY4Φ(X11)	73	127	10	-	+	-
PY4Φ(X12)	56	151	10	-	+	-
F231Φ(X17)	53	147	9	-	+	-
HV1Φ(X19)	55	143	11	30x7	19x11	-
FV1Φ(X20)	52	144	11	+	22x13	-
GV2Φ(X21)	57	137	11	34x15	28x19	-
G1V2Φ(X22)	54	136	11	38x7	24x13	-
G3V2Φ(X23)	47	133	7	-	-	-
G4V2Φ(X24)	56	143	9	30x9	24x13	-
G5V2Ф(X25)	58	162	8	-	+	-
GKV2Φ(X26)	51	133	9	-	-	-
KV2Φ(X27)	54	145	10	-	-	-
LV2Ф(X28)	53	117	10	-	34x13	-
AV2Ф(X29)	58	138	9	-	-	-
SİYV2Φ(X30)	51	149	10	-	+	-

Table 2 Morphological properties of L. bulgaricus phages

<sup>1</sup> not exist <sup>2</sup> exist but can not be measured

(Figure 2b),  $\Phi$ H3Y4,  $\Phi$ H1Y4,  $\Phi$ 709BY4,  $\Phi$ SIBY4,  $\Phi$ FV1,  $\Phi$ LV2 only the presence of tail plaque was observed. Also, the presence of these structures was suspected in the phages  $\Phi$ IY4,  $\Phi$ SIYY4,  $\Phi$ GY4 (Figure 2c),  $\Phi$ PY4,  $\Phi$ F231,  $\Phi$ G5V2,  $\Phi$ SIYV2. There was no evidence of contractile tail covers in any of the phages. In the light of the characterization studies these 25 native *L. bulgaricus* phages were placed in the *Siphoviridae* family in Ackermann's classification and/or Group B in Bradley's.



Figure 2. L. bulgaricus phages (a)  $\Phi$ HV1, 60000x; (b)  $\Phi$ FY4, 60000x; (c)  $\Phi$ GY4, 60000x.

### DISCUSSION

It was observed that all of S. thermophilus phages had isometric hexagonal heads. Although it is possible to determine the taxonomical characteristics of the phages through the use of electron micrographs, it is difficult to identify whether the capsid structures were icosahedral, octahedral or dodecahedral (15). In addition, regarding the characteristics mentioned by Ackermann and DuBow (17), in electron micrographs of S. thermophilus phages 0709-X5, 0231-X7, 0231-X21, Ф231-X22, Ф231-X23, ФВ3-X11, ФВ3-X18 (Figure 1b) and  $\Phi$ B3-X19, hexagonal and pentagonal profiles were observed together while, importantly, hexagonal and spherical profiles were present together in the micrographs of  $\Phi$ 709-X4 (Figure 1c) and  $\Phi$ B3-X20. It can thus be said that the phages have an icosahedral head structure. In electron micrographs of other S. thermophilus phages, only hexagonal capsid profiles were determined. Furthermore, the tails penetrated the capsids, which is

a sign of icosahedral head structure, in the cases of phages Ф231-Х9 (Figure 1a), Ф231-Х22, ФВ3-Х12,  $\Phi$ B3-X13 and  $\Phi$ B3-X14. On the other hand, the determination of the pentagonal capsid only in tailed phages (17) increases the probability of them being placed in the Siphoviridae family with an isometric hexagonal profile having icosahedral capsid members. In this study, depending on the increasing number of the phages examined, the values of capsid diameter, tail length and tail width generally differed within wider limits. However, the 5 nm tail width determined by Kivi et al. (16) in one of 9 S. thermophilus phages and the 42 nm capsid diameter determined by Benbadis et al. (20) in one of 7 phages, differed from our phages in their extremely low values.

In S. thermophilus phages, the diameter of the capsids were found to be between 45 to 65 nm, the tails were 210-270 nm long and 7-13 nm wide (1, 14, 16, 19-25). Among the phages that we examined, the shortest (182 nm) and the longest tail (290 nm) was about 30 nm longer or shorter than the values found by other researchers. A phage determined by Reinbold et al. (5) also had a 290 nm long tail. In addition to that Suárez et al. (6) and Quiberoni et al. (2) finally isolated a phage having a 330 nm long tail. In another study electron microscopic analysis showed that phage 2972 had a 55-nm-diameter isometric capsid and a 260-nm-long noncontractile tail (25). Among 25 native S. thermophilus phages that were inspected morphologically, the presence of tail plaque was suspected in only two of the phages ( $\Phi$ 231-X9 and  $\Phi$ B3-X18) (Figure 1a,b) but in none of the phages were fiber or collar structures determined. Moreover Krusch et al. (14), Kivi et al. (16) and Prevots et al. (21) did not determine any collar structures out of 59, 7 and 9 phages they examined respectively. Krusch et al. (14) determined tail plaque structures in all of the phages they examined. It is known that fiber structures are rarely found (14, 22), and it is speculated that this is because these fragile structures are easily detached by mechanical effects or that perhaps mutants without fibers frequently occur. (26). However, in some S. thermophilus phages tail plaque and fibers were shown clearly (19, 21).

In this study while collar and tail plaques were observed in many of the *L. bulgaricus* phages, these structures could not be seen in *S. thermophilus* phages although the micrographs were taken at the same magnification. The reason for this may be either the modifications in the dying procedure or the use of three different electron microscopes. *L. bulgaricus* phages had also isometric hexagonal heads and noncontractile tails. In one exceptional case Reinbold et al., isolated and micrographed a *L. bulgaricus* phage having a contractile cover. This phage was categorised in *Myoviridae* in Ackermann's classification (5). The temperate phages mV1 and mV4 were studied by Cluzel et al. (27), and the morphology of the ch2 virulent phage was determined by Chow et al. (28). The lb 539 temperate phages was examined by Auad et al. (15). These four phages were also placed in Group B in Bradley's classification.

The capsid diameters (50 nm) of the mV1 and mV4 phages (27) were the same as in the case of the ch2 virulent phage (28). Their tail lengths were also close to each other, being 180 nm and 170 nm respectively. However in the lb 539 temperate phage the diameter of the capsid was 47 nm and the tail was 159 nm long (15). The new virulent phage phiLdb which was isolated by Wang et al. has an icosahedral capsid of  $47.7\pm0.9$  nm in diameter and a long noncontractible tail of  $129.8\pm2$  nm and several fibres (9). 25 native *L. bulgaricus* phages examined in this study were found to have a wider range of capsid diameter (47-73 nm), tail length (117-162 nm) and width (7-13 nm).

In this study, the capsid diameters and tail width determined in L. bulgaricus phages were similar to the other L. bulgaricus phages. However when the tail lengths were compared, it was found that they had smaller tails. Only the lb 539 (15) and phiLdb (9) phages with 159 and 129 nm tail length lay in between the values that we measured, respectively. One other difference was clearly apparent in collar and tail plaque structures. According to the measurements of Chow et al. (28), both collar and tail plaque structures showed considerably higher values (Figure 2a). However it is thought that the differences in the values are not because of the structural differences of the native phages originating in Turkey, but because tail structures are not clearly identified since the micrographs in this study were taken at a lower magnification.

# The frequency of thermophilic phage isolation from raw milk

In our former studies using native and industrial strains, although the lactococcal phages were isolated from raw milk at a high frequency (29, 30), a lack of success in the isolation of *S. thermophilus* and *L. bulgaricus* phages with native and industrial hosts, led us to reconsider raw milk as a highly suitable isolation source for these phages (31). Since there is only a limited literature on this subject (12) a minor trial study was conducted; first, 18 raw milk samples were compared with 3 S. thermophilus (B3, 709, 231) strains, 2 phages (Ф1В3-А and  $\Phi$ 2B3-A), which were effective only with the B3 host, were isolated from 2 raw milk samples and their morphological characterization was undertaken with the other phages in the collection. Then 9 industrial S. thermophilus (231, 632, 709, V1, V2, Y1, Y4, CH-1, B3) strains were used as a host. However, out of 3 raw milk samples, no phage could be isolated. Additionally, the same 3 samples were tried with a total of 6 L. bulgaricus strains, five of which were industrial (Y1, Y4, V1, V2, 231) and one being a reference strain (Lactobacillus bulgaricus ATTC 11842), from the 2 raw milk samples 7 of the phages which were effective with V1, V2, Y4 and L. bulgaricus ATTC 11842 strains could be isolated. In the light of the data, phages specific to L. bulgaricus are more frequently isolated than the phages specific to S. thermophilus from raw milk. However it is much more difficult and less common to isolate thermophilic phages from raw milk samples.

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#### REFERENCES

1. Larbi D, Colmin C, Rousselle L, Decaris B, Simonet JM. 1990. Genetic and biological characterization of nine *Streptococcus salivarius* subsp. *thermophilus* bacteriophages. *Lait*, 70, 107-116.

2. Quiberoni A, Auad L, Binetti AG, Suárez VB, Reinheimer JA, Raya RR. 2003. Comparative analysis af *Streptococcus thermophilus* bacteriophages isolated from a yogurt industrial plant. *Food Microbiol*, 20, 461-469.

3. Moineau S, Borkaev M, Holler BJ, Walker SA, Kondo JK, Vedamuthu ER, Vanderbergh PA. 1996. Isolation and characterization of Lactococcal bacteriophages from cultured buttermilk plants in the United States. *J Dairy Sci*, 70, 2104-2111.

4. Reinbold GW. 1974. Bacteriophage For Italian Cheese Cultures. 11<sup>th</sup> Annual Marschall Invitational Italian Cheese Seminer, Wisconsin. 5. Reinbold GW, Reddy MS, Hammond EG. 1982. Ultrastructures of bacteriophages active against *Streptococcus thermophilus*, *Lactobacillus bulgaricus*, *Lactobacillus lactis* and *Lactobacillus helveticus*. *J Food Protect*, 45 (2), 119-124.

6. Suárez VB, Quiberoni A, Binetti AG, Reinheimer JA. 2002. Thermophilic lactic acid bacteria phages isolated from Argentinian dairy industries. *J Food Protect*, 65 (10), 1597-1604.

7. Le Marrec C, Sinderen D, Walsh L, Stanley E, Vlegels E, Moineau S, Heinze P, Fitzgerald G, Fayard B. 1997. Two groups of bacteriophages *Streptococcus thermophilus* can be distinguished on the basis mode packaing and genetic determinants for major structural proteins. *Appl Environ Microbiol*, 63 (8), 3246-3253.

8. Neve H, Freudenberg W, Diestel-Feddersen F, Ehlert R, Heller KJ. 2003. Biology of the temperate *Streptococcus thermophilus* bacteriophage TP-J34 and physical characterization of the phage genome. *Virology*, 315, 184–194.

9. Wang S, Kong J, Gao C, Guo T, Liu X. 2010. Isolation and characterization of a novel virulent phage (phiLdb) of *Lactobacillus delbrueckii*. *Int J Food Microbiol*, 137, 22–27.

10. Durlu-Özkaya F, İç N, Tunail N. 1999. The thermophilic starter cultures resistant to their phages and rotation. Special Issue for the 11<sup>th</sup> Congress of KÜKEM, 23 (2), 7-8.

11. Olsen JV. 1990. The Dairy Cultures of Chr. Hansen's. The latest development of dairy cultures. Dairy Seminar, Ankara Üniversity, Agricultural Faculty.

12. Bruttin A, Desiere F, d'Amico N, Guerin JP, Sidoti J, Huni B, Lucchini S, Brüssow H. 1997. Moleculer ecology of *Streptococcus thermophilus* bacteriophage infections in a cheese factory. *Appl Environ Microbiol*, 63, 3144-3150.

13. Kaleli D, Tunail N, Acar E. 2004. Virulent bacteriophages of *Streptococcus thermophilus* and lysogeny. *Milchwissenschaft*, 59 (9/10), 487-491.

14. Krusch U, Neve H, Luschei B, Teuber M. 1987. Characterization of virulent bacteriophages of *Streptococcus salivarius* subsp. *thermophilus* by host specifity and electron microscopy. *Kieler Milch. Forschungsberichte*, 39 (3), 155-167.

15. Auad L, Holgado R, Forsman P, Alatossava T, Raya RR. 1997. Isolation and characterization of a new *Lactobacillus delbrueckii* subsp. *bulgaricus* temperate bacteriophage. *J Dairy Science*, 80, 2706-2712.

16. Kivi S, Peltomäki T, Luomala K, Sarimo SS. 1987. Some properties of *Streptococcus thermophilus* bacteriophages. *Fotra Microbiol*, 32, 101-106.

17. Ackermann HW, DuBow MS. 1987. Viruses of Prokaryotes, Vol. 1: General Properties of Bacteriophages. CRC Press, Boca Raton, Florida.

18. Zhang X, Kong J, Qu Y. 2006. Isolation and characterization of a *Lactobacillus fermentum* temperate bacteriophage from Chinese yogurt. *J Appl Microbiol*, 101, 857–863. 19. Brüssow H, Frémont M, Bruttin A, Sidoti J, Constabla A, Fryder V. 1994. Detection and classification of *Streptococcus thermophilus* bacteriophages isolated from industrial milk fermentation. *Appl Environ Microbiol*, 60 (12), 4537-4543.

20. Benbadis L, Faelen M, Slos P, Fazel A, Mercenier A. 1990. Characterization and comparison of virulent bacteriophages of *Streptococcus thermophilus* isolated from yogurt. *Biochimie*, 72, 855-862.

21. Prevots F, Relano P, Mata M, Ritzenthaler P. 1989. Close relationship of virulent bacteriophages of *Strepto-coccus salivarius* subsp. *thermophilus* at both the protein and the DNA level. *J General Microbiol*, 135, 3337-3344.

22. Fayard B, Haefliger M, Accolas JP. 1993. Interaction of temperate bacteriophages of *Streptococcus salivarius* subsp. *thermophilus* with lysogenic affect phage DNA restriction pattern and host ranges. *J Dairy Res*, 60, 385-399.

23. Stanley E, Fitzgerald GF, Le Marrec C, Fayard B, Sinderen D. 1997. Sequence analysis and characterization of Φ01205, a temperate bacteriophage infecting *Streptococcus thermophilus* CNRZ1205. *Microbiology*, 143, 3417-3429.

24. Tremblay DM, Moineau S. 1999. Complete genomic sequence of the lytic bacteriophage DT1 of *Streptococcus thermophilus*. *Virology*, 255, 63-76.

25. Le 'vesque C, Duplessis M, Labonte J, Labrie S, Fremaux C, Tremblay D, Moineau S. 2005. Genomic Organization and Molecular Analysis of Virulent Bacteriophage 2972 Infecting an Exopolysaccharide-Producing *Streptococcus thermophilus* Strain. *Appl Environ Microbiol*, 71 (7), 4057–4068.

26. Neve H, Krusch U, Teuber M. 1989. Classification of virulent bacteriophages of *Streptococcus salivarius* subsp. *thermophilus* isolated from yogurt and Swiss-type cheese. *Appl Microbiol Biotechnol*, 30, 624-629.

27. Cluzel PJ, Veaux M, Rousseau M, Accolas JP. 1987. Evidence for temperate bacteriophages in two strains of *Lactobacillus bulgaricus*. J Dairy Res, 54, 397-405.

28. Chow J, Batt CA, Sinskey AJ. 1988. Characterization of *Lactobacillus bulgaricus* bacteriophage ch2. *Appl Environ Microbiol*, 54 (5), 1138-1142.

29. Aydar LY, Tunail N. 1995. Isolation and electron microscopic investigation of lactic phages which isolated from Turkey. *Milchwissenschaft*, 50 (6), 312-316.

30. Durlu F, Tunail N. 1991. Resistance of commercial cheese starter cultures to domestic bacteriophages in Turkey. The Third International Congres on Food Industry, 4-8 November, Kuşadası, İzmir, Turkey, 125-139.

31. Rio B, Binetti AG, Martı'na MC, Ferna'ndeza M, Magada'na AH, Alvarez MA. 2007. Multiplex PCR for the detection and identification of dairy bacteriophages in milk. *Food Microbiol*, 24, 75–81.