Biosynthesis of Silver Nanoparticles Using Rosa canina Extract and Its Anti-cancer and Anti-metastatic Activity on Human Colon Adenocarcinoma Cell Line HT29

Rosa canina Özütlü Kullanılarak Gümüş Nanopartiküllerin Biyosentez ve İnsan Kolon Adenokarsinoma Hücre Hattı HT29 Üzerindeki Anti-kanser ve Anti-metastatik Aktiviteleri

Çiğdem AYDIN ACAR1, Suray PEHLİVANOĞLU2

1 Burdur Mehmet Akif Ersoy University, Bucaş School of Health, Burdur, Turkey
2 Necmettin Erbakan University, Department of Molecular Biology and Genetics, Konya, Turkey

Abstract: Despite progress in conventional treatment methods for colon cancer, it remains the fourth leading cause of cancer-related deaths in the world. Therefore, more effective new treatment strategies for colon cancer are needed. Silver nanoparticles (AgNPs) synthesized using plant extracts have shown therapeutic applications and make it to be a good anti-cancer candidates. The aim of this study was to evaluate the anti-metastatic and anti-cancer activity of biosynthesized silver nanoparticles from Rosa canina extract on the human colon adenocarcinoma cell line HT29. The biosynthesis of AgNPs was carried out using Rosa canina extract. R-AgNPs were characterized by techniques such as UV-vis spectrophotometry and scanning electron microscopy (SEM). HT29 cells were incubated with different concentrations of AgNPs (0-20 µg/mL) for 48 h. The cytotoxic activity of the synthesized R-AgNPs against human colon adenocarcinoma cells HT29 was investigated by MTT assay and the IC50 value was found to be 7.89 µg/mL at 48 h incubation. Anti-metastatic potential of R-AgNPs were determined on HT29 cells using scratch assay. R-AgNPs induced a significant decrease of cell motility in dose-dependent manner. In conclusion, these findings suggest that the biosynthesized AgNPs may be promising new therapeutic agents for the treatment of human colon cancer.

Keywords: Silver nanoparticles, Colon cancer, Metastasis, AgNPs.

Öz: Kolon kanseri, geleneksel tedavi yöntemlerinde kaydedilen ilerlemelere rağmen, dünyada kansere bağlı ölüm nedenleri arasında dördüncü sıradadır. Bu nedenle, kolon kanserinin tedavisi için daha etkili yeni tedavi stratejilerine ihtiyaç duyulmaktadır. Bitki özütlü kılanan gümüş nanopartiküller (AgNP'ler), terapötik uygulamalar göstermiş ve bu durum onları bir kanser önleyici ajan olarak daライン getirmiştir. Çalışmamızda, kuşburnu bitisinin sulu özü kullanılarak gerçekleştirildi. R-AgNP'ler, UV-görüntü spektroskopmetre ve taramalı elektron mikroskobu (SEM) gibi çeşitli analitik tekniklerle karakterize edildi. HT29 hücreleri 48 saat süresince çeşitli konsantrasyonlarda R-AgNPler (0-20 µg/mL) muamele edildi. Sentezlenen R-AgNP'lerin insan kolon kanserinin hücrelerine karşı sitotoksik aktivitesi MTT assay ile değerlendirildi ve IC50 değeri 7.89 µg/mL olduğu belirlendi. R-AgNP'lerin anti-metastatik potansiyeli, HT29 hücrelerinde scratch assay kullanılarak çözüldü. R-AgNP'lerin doza bağımlı olarak hücre hareketliliğini önemli bir azalmaya neden olduğu belirlendi. Sonuç olarak, bu bulgular biosentezlenen gümüş nanopartiküllerin insan kolon kanseri tedavisi için umut vertici yeni terapötik ajanlar olabileceğini göstermektedir.

Anahtar Kelimeler: Gümüş nanopartiküller, Kolon kanseri, Metastaz, AgNP.

Introduction

Colorectal cancer, also known as colon cancer, is a type of cancer that begins in the colon. Colon cancer is the third leading cause of cancer-related deaths worldwide. In 2018, it was diagnosed in...
over one million people and caused death to 880,792 persons worldwide (Bray et al., 2018). Colon cancer risk factors include a person's lifestyle, diet, age, ethnicity and family history (Slattery, 2000). Although some types of cancer are preventable, this is not the case for colon cancer because colon cancer is often detected in advanced stages and is usually not diagnosed until symptoms appear. There are a variety of traditional treatment options such as surgery, radiotherapy and chemotherapy for treating this cancer, but the success rates are very low in advanced stage colon cancer. Because of all these reasons new treatment models in the treatment of colon cancer patients are needed.

Nanotechnology is a rapidly growing area with the advances in science and technology that, aims to produce new materials on the nanoscale (Albrecht et al., 2006). In past decades, metal nanoparticles have been used frequently as new anti-cancer agents owing to their features like cytotoxicity, functionality and compatibility. Especially, silver nanoparticles (AgNPs) have attracted the attention of researchers due to their various industrial applications (Yesilot and Aydin, 2019). AgNPs can be synthesized using physical, chemical and biological methods. The chemical synthesis is a commonly used approach for the synthesis of AgNPs that use toxic chemicals, leading to non-eco-friendly biological products. In the recent years, green synthesis is preferred due to it is cost effective, environment friendly and easily synthesize large quantities properties compared to other methods (Thakkar et al., 2010). For the green synthesis of nanoparticles, plants extracts, bacteria, fungi, algae, yeast and viruses, which are naturally available resources can be used (Ahmed et al., 2016; Raveendran et al., 2003; Daphne et al., 2018). Plant-mediated synthesis is seen as the most suitable method among green synthesis methods because it allows the formation of more stable nanoparticles in a short time (Iravani, 2011).

Rosehip (Rosa canina) is a species of plant belonging to the Rosa genus in the Rosaceae family. Rose hip are widely distributed in several areas, including Europe, Africa, Middle and West Asia and Anatolia. In recent years, rosehip has been consumed in many cultures as a food and has been used frequently for the treatment of some diseases and has attracted more attention due to its documented therapeutical properties (Chrubasik et al., 2007). Rose hip contains high concentrations of ascorbic acid, phenolic compounds and healthy fatty acids (Larsen et al., 2003). It is a medicinal plant commonly used in traditional medicine for the treatment of colds, asthma, hemorrhoids, infections, chronic pains, arthritis, and inflammatory diseases. Also, rose hip is known to have many biological properties such as diuretic, anti-oxidant, anti-ulcerogenic, anti-obesity, anti-diabetic, anti-carcinogenic, neuroprotective, and anti-microbial effects (Mármol et al., 2017; Cheng et al., 2016; Turan et al., 2017). Thus, in this study, we investigated the synthesis of AgNPs using an aqueous extract of the rose hip and evaluated their anti-cancer and anti-metastatic activities on human colon cancer cells.

Materials and Methods

Preparation of aqueous extract of Rose hip fruit (Rosa canina)

The dried fruit of Rosehip was finely powdered. For preparation of extract, 20 g of rosehip was boiled in 50 mL deionized water for 2 min. The aqueous extract was subsequently filtered through whatman no. 1 filter paper and then was stored at 4°C until further use.

Biosynthesis of silver nanoparticles

For the synthesis of silver nanoparticles (R-AgNPs), 5mL of rosehip extract was mixed with 95 mL of AgNO₃ (5mM). The reaction mixture was incubated at microwave oven (1200W, 50Hz) for 1 min. The progress of the reaction was routinely monitored by observing colour change from yellow to brown, which indicated the formation of R-AgNPs.
Characterization of silver nanoparticles

A combination of analytical techniques was utilized to characterize the synthesized R-AgNPs including UV-vis Spectroscopy, Scanning Electron Microscopy (SEM) and Energy Dispersive X-ray Spectroscopy (EDX). UV–Vis spectrophotometer (T60, PG Instruments) was used to confirm the reduction of metal ions and was scanned in the range of 300–700 nm.

Cell culture

The human colon adenocarcinoma cell Line HT29 obtained from Animal Cell Culture Collection (HUKUK, Sap Institute, Ankara, Turkey) were cultured DMEM (Dulbecco’s Modified Eagles Medium), supplemented with 10% heat inactivated Fetal Bovine serum (FBS), L-glutamine, 100μg/mL Penicillin and 100μg/mL Streptomycin and were incubated at 37°C with 5% CO2 in humidified atmosphere.

Cell viability assay

The cytotoxicity effect of the synthesized R-AgNPs on HT29 cancer cells was evaluated by MTT assay. The cells were grown in DMEM medium containing 10% FBS. For experiments, the cells (1x10^4 cells/well) were plated in 96-well plates with the medium containing 10% FBS and incubated for 24 h with 5% CO2 at 37 °C in humidified atmosphere. Later, the medium was replaced with DMEM containing 1% FBS and the R-AgNPs (20, 10, 5, 2.5, 1.25, 0.625, 0.312, 0.156 and 0.075 μg/mL). Cells without nanoparticles were used as a control. After treatment, the plates were incubated for 48 h and 10 μL of MTT prepared at a concentration of 5 mg/mL was added to each well and incubated for 4 h. Purple colour formazan crystals formed were then dissolved in 100 μL of dimethylsulphoxide (DMSO). Viability of the cells was evaluated by measuring the absorbance at 570 nm in a multiwell ELISA plate reader (Multiscan GO, Thermo Fisher Sci.). The absorbance at 570 nm was also measured for wells without any sample as blank. The % cell viability was evaluated using the following equation:

\[
\text{Cell viability} \% = \left( \frac{\text{the OD of treated well}}{\text{the OD of control well}} \right) \times 100
\]

IC50 value was calculated from the dose-response curve obtained with this assay.

Scratch Assay

Cell migration was determined by using an *in vitro* scratch assay. HT29 cells were seeded in 24 well culture plates and grown to 100% confluence. A scratch was made in the monolayer of cells with a sterile pipette tip. Measures were taken to maintain the same scratch angle in the test and control wells. After a scratch was made, the cells were washed with PBS to remove unattached cells. At 10X magnification of an inverted microscope equipped with a camera, images of control and test wells (0.1μg/mL and 1μg/mL R-AgNPs) were captured at 0 h, 12 h and 24 h. Further the captured images were analyzed by Image J software.

Colony formation assay

The colony formation assay is capable of evaluating *in vitro* cell death based on the ability of a single cell to transform into a colony. Briefly, cells were seeded as single cells (1000 cells/well) in 6-well plates and allowed to adhere for 24 h. Cells were treated with two different concentrations (0.1 and 1 μg/mL) of AgNPs at 37°C for 14 days. At the end of the incubation period, the cells were washed and fixed with methanol for 5 min, and then stained with 0.1% crystal violet for 15 min and colony formation in each dish was photographed. The clonogenic index was then determined by counting the cell colonies in each well.

Results

Biosynthesis of R-AgNPs and Characterization

In this study, rose hip extract was used to synthesis of R-AgNPs. Figure 1A shows silver nitrate, rosehip extract and R-AgNPs. It was observed that
the extract had a yellowish-red color before reaction with the silver ions. After a few minutes of mixing the rosehip extract and the AgNO3 solution, a brown color change was observed indicating the formation of AgNPs. Further, the R-AgNPs were characterized using UV-vis spectroscopy. The UV-visible absorption spectra of R-AgNPs were measured in the range of the wavelength range 300-750 nm. A surface plasmon peak was observed at 430 nm for the R-AgNPs (Figure 1B). The surface nature of the R-AgNPs were determined by SEM. (Figure 1C). It was seen that according to EDX, there were peaks of Ag metal in the elemental composition of R-AgNPs (Figure 1D).

**AgNPs-induced cytotoxicity in HT29 human colon adenocarcinoma cells**

The cytotoxicity potential of the biosynthesized R-AgNPs on human colon cancer cells has been examined using the HT29 cell line. The cells were exposed to various concentrations of AgNPs for 48 h, then the cytotoxicity effects of R-AgNPs assessed using the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide] assay. The MTT results showed that the AgNPs decreased cell viability in terms of dose and time compared to control group (Figure 2). The inhibitory concentration (IC50 value) was calculated to be 7.98 μg/mL after 48 h of cell treatment.

In cytotoxicity studies, colony forming assay is one of the standard method for verifying clonogenic survival ability of cells exposed to potential therapeutic agent. After treatment with R-AgNPs, a colony formation assay was performed to measure the proliferation of HT29 cancer cells. HT29 cancer cells were seeded at appropriate dilutions and treated with two different concentrations of R-AgNPs (0.1 and 1 μg/mL). Cultures were kept under normal culture conditions for 14 days and the colony formations were analyzed. The results showed that R-AgNPs treatment inhibited significantly the colony forming ability of HT29 cells in a dose dependent manner (Figure 3).
Metastasis is a hallmark of cancer progression and the leading cause of mortality among cancer patients (Hanahan and Weinberg, 2011). The migration of cancer cells is the first step in metastasis, and these processes are called as tumor invasion (Clark and Vignjevic, 2015). The in vitro scratch assay is a simple and commonly used method to measure cell migration (Liang et al., 2007). Thus, we have used in vitro scratch assay to evaluate the effect of R-AgNPs on cell migration. Our results showed that the R-AgNPs treatment reduced migration ability of colon cancer cells HT29 at period of 12h and 24h. R-AgNPs at 1 μg/mL concentration showed more effective inhibitory cell migration determined through scratch assay (Figure 4).

**Discussion**

Metal nanoparticles are sophisticated agents with a broad range of applications in various fields. In particular, silver nanoparticles have great potential as antimicrobial agents, diagnostic tools, imaging, biomedical device coating tools, drug and gene delivery carriers in nanomedicine (Panacek., 2006). Different methods are available for the synthesis of silver nanoparticles. However, the use of highly toxic and harmful to the environment materials is a necessity in many methods used in the production of silver nanoparticles. Furthermore, nanoparticle production with these methods is limited, expensive and harmful to the environment due to high-energy dissipation (Mathur., 2017). The biological method used in the production of nanoparticles uses biological resources such as plants, algae, fungi, yeasts and bacteria.

In this study, we tried to define the synthesis of R-AgNPs with rosehip extract and then further use of synthesized nanoparticles for clinical purposes.

![Figure 4. Effects of R-AgNPs on HT29 colon cancer cell migration, assessed by scratch assay. HT29 cells were imaged with an inverted microscope (original magnification, ×10) at 0, 12 and 24h post-scratching. The images were analyzed by Image J software.](Image)

**Figure 4.** Effects of R-AgNPs on HT29 colon cancer cell migration, assessed by scratch assay. HT29 cells were imaged with an inverted microscope (original magnification, ×10) at 0, 12 and 24h post-scratching. The images were analyzed by Image J software.

In this study, we tried to define the synthesis of R-AgNPs with rosehip extract and then further use of synthesized nanoparticles for clinical purposes. The synthesized R-AgNPs were characterized via UV-vis spectrophotometry and SEM. It is known that the characteristic absorption peak of AgNPs is centered at 400-450 nm depending on the size and distribution of the nanoparticles (Anandalakshmi et al., 2016). In our study, an absorption peak at 430 nm was observed, suggesting that the synthesized R-AgNPs were pure. It is well known that a strong surface plasmon peak is observed in the presence of various metal nanoparticles with a width of 2-100 nm (Sastry et al., 1997; Sastry et al., 1998).

Recently, many studies have suggested that the biosynthesized silver nanoparticles from plant extracts have an encouraging cytotoxicity effect against tumor cells (Venugopal et al., 2016; Das et al., 2013; Behboodi et al., 2018; Alavi and Karimi, 2017). In the literature, the cytotoxic and anticancer effects of the extracts obtained from Rosa canina have been reported against several cancer cells (Kilinc et al., 2019; Turan et al., 2018; Jiménez et al., 2016). The plant extracts used in these studies were able to inhibit the growth of cancer cells at much higher concentrations than nanoparticles. There is only one study showing the use of Rosa canina in silver nanoparticle synthesis and a different method has been used. However,
the effects of nanoparticles on cancer cells were not evaluated in this study (Pulit et al., 2014).

In our study, silver nanoparticle synthesis was performed for the first time by using Rosa canina extract and its anti-cancer and anti-metastatic effects were evaluated on HT29 colon cancer cells. The in vitro cytotoxic effect of R-AgNPs and cell inhibition (%) was evaluated by MTT assay compare to the control group. Untreated cells constituted the control group. The results of MTT assay showed that the increasing concentrations of biosynthesized R-AgNPs enhanced the inhibitory effect on the cell proliferation of HT29 cells after 48 h exposure. The IC50 value of biosynthesized R-AgNPs was determined to be 7.98μg/mL. The approximately 90% cell inhibition of colon cancer cells was observed at a maximum concentration of 20 μg/mL.

Grunathan et al. (2018) reported that the nariginen-mediated synthesis of AgNPs were very effective at low concentrations against HCT116 colorectal cancer cells (IC 50: 5 μg/mL) Anthemis atrapatana extract-mediated synthesis of AgNPs showed dose dependent cytotoxic effects against colon cancer cell lines (HT29) (Dehghanizade et al., 2018). Similarly, Abutilon indicum-mediated AgNPs exhibited dose-dependent antiproliferative effects on COLO 205 (human colon cancer) (IC50: 3 μg/mL) and MDCK (normal) cells (IC50: 75 μg/mL) (Mata et al., 2015). Likewise, Durai et al. (2014) reported that the para-hydroxybenzoate tetrahydrate (SPHT)-mediated synthesis of AgNPs showed dose- and time-dependent inhibition of cell viability on HCT15 and HT-29 colon cancer cell lines (IC50: 8 μg/mL). In this study, the potential cytotoxic effect of R-AgNPs on HT29 cells after short (48 hours) and long term (two weeks) exposure were demonstrated. In addition to MTT assay (48 hours), colony formation assay was performed to evaluate cell proliferation. Colony formation assay, used to evaluate the cytotoxic effects induced after long term exposure (two weeks). R-AgNPs was added at different concentrations (0.1 and 1μg/mL) and colonies were counted and analyzed after two weeks. The results showed that R-AgNPs inhibited HT29 colony formation as compared to the control cells (Figure 3). This result shows that R-AgNPs reduce the colony-forming potential of cancer cells and therefore can be used as an anti-cancer drug. The small number of colonies formed by cells exposed to R-AgNPs (1 μg/mL) confirmed the MTT assay cytotoxicity.

In the literature, cloning assay data were not found in any study that tested plant-mediated AgNPs in colon cancer cells.

In this study, the R-AgNPs inhibited lateral motility of HT29 cells in dose dependent manner. According to our results, the dose of 1µg/mL inhibited lateral motility by 97.4% for 24 h (Figure 4). The R-AgNPs showed the remarkable effect on cancer cell proliferation and migration. Also, the ability of R-AgNPs to show activity even at low concentrations may make them useful for in vivo experiments. There are few studies investigating the anti-metastatic effects of silver nanoparticles. The results of our study are consistent with previous studies reporting that silver nanoparticles inhibit the migration of cancer cells (Hussain et al., 2019; Shruti et al., 2015; Kavaz et al., 2018; Buranasukhon et al., 2017).

In conclusion, the silver nanoparticles were successfully synthesized from Rosa canina extract. The biosynthesized R-AgNPs exhibited anticancer and anti-metastatic effects against HT29 human colon cancer cells. Based on our findings, we suggest that biosynthesized AgNPs may be promising new therapeutic agents for the treatment of human colon cancer. Further studies are required to support our findings of the anti-cancer and anti-metastatic potential of biosynthesized AgNPs in vivo.

Conflicts of Interest

The authors declare no conflict of interest.

* This study was presented as an oral presentation at the 2nd International Conference on Technology and Science, 2019 on November 14- 16, 2019, Burdur-Turkey.
References


