

# Larvicidal Activities of Essential Oils Extracted from Five Algerian Medicinal Plants against *Culiseta longiareolata* Macquart. Larvae (Diptera: Culicidae).

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#### ABSTRACT

**Objective:** The use of essential oils in mosquito control is considered as a potential alternative of synthetic insecticides. The current study aimed to assess the larvicidal activity of the essential oils extracted from five medicinal plants collected from northeastern Algeria against the *Culiseta longiareolata* larvae, a vector of the *Plasmodium* species in birds and one of the most abundant mosquito species in Algeria.

**Materials and Methods:** The essential oils extracted from: *Thymus vulgaris, Artemisia herba-alba, Juniperus phoenicea, Rosmarinus officinalis,* and *Eucalyptus globulus* were tested against the 3<sup>rd</sup> and 4<sup>th</sup> instar *Culiseta longiareolata* larvae. The larvae were exposed to a series of concentrations of the tested essential oils for 24h. The concentrations that caused between 10% and 90% mortality were replicated four times, and the entire test was repeated three times. The collected data were used to determine the LC<sub>50</sub> and LC<sub>90</sub> values,

**Results:** The tested oils revealed an efficient larvicidal activity. *T. vulgaris* showed 100% mortality at 80ppm final concentration, while the other tested oils showed 100% mortality at 200ppm. Furthermore, the lethal concentrations that caused 50% and 90% mortality (LC<sub>50</sub> and LC<sub>90</sub>) were varying. *T. vulgaris* was the most efficient essential oil (LC<sub>50</sub>=25.64ppm, LC<sub>90</sub>=50.53ppm), followed by *J. Phoenicea* (LC<sub>50</sub>=59.83ppm, LC<sub>90</sub>=137.68ppm), *R. officinalis* (LC<sub>50</sub>= 64.18ppm, LC<sub>90</sub>= 96.55ppm), *A. herba-alba* (LC<sub>50</sub>=86.67ppm, LC<sub>90</sub>=139.55ppm), then *E. globules* (LC<sub>50</sub>=95.83ppm, LC<sub>90</sub>= 168.25ppm).

**Conclusion:** The use of essential oils or their principal active components as  $\alpha$ -pinene, 1,8-cineole and Camphor may serve as an eco-friendly method to control mosquito larvae. Nevertheless, the field application of essential oils and their principal components remains a fundamental step to evaluate the field efficacy of these botanic extracts and to note their possible secondary effects on non-targeted organisms.

Keywords: Aromatic medicinal plants, Culiseta longiareolata, Essential oil, Larvicidal activity, Mosquitoes

#### INTRODUCTION

Culicidae, or mosquitoes as commonly known, is a family of Diptera insects that reproduce quickly and abundantly. Simultaneously, this family includes major vectors for many deadly and dangerous diseases. Therefore, the importance of the mosquito family in terms of public health makes mosquito control an important initiative to minimize the negative effects of mosquito-born-diseases. Mosquito control may depend on various strategies; the most common in the past decades was the use of synthetic insecticides as inexpensive and available products. However, the use of synthetic insecticides has over time created environment pollution and resistance problems (1, 2).

Recently, eco-friendly methods were developed to control mosquitoes. For instance, the enhancement



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of behavior-based control tools and the development of repellent and toxic products based on botanic components can target different mosquito life stages (3, 4). Essential oils (EOs) extracted from different parts of plants were frequently tested for their mosquitocidal activity (5). These primary botanic materials present various biological activities. They can act as insecticides where they can affect the oviposition, survival, larval duration, pupation and insect emergence (6, 7). However, the larvae stage appears to be more appropriate to control mosquito populations because of the high reproduction rates and larvae food mechanisms that allow a high number of mosquito individuals to be targeted simultaneously. Therefore, the assessment of the larvicidal efficacy of various plant derivatives was the main objective of many research papers (8-11).

Culiseta longiareolata (Macquart 1838) constitutes with the Culex pipiens (Linnaeus 1758) complex the most abundant species in Algeria. It usually breeds near human habitations, however, the females prefer to feed on bird blood (12). Cs longiareolata has uniquely adaptive and survivor features. Kiflawi et al. (13) have confirmed that the females of this species showed an adaptive response against the risk of predation and negative density effects where they avoid laying their eggs in predator pools. Further, Cs longiareolata is considered as a primary vector of Plasmodium (Giovannolaia) circumflexum (Kikuth 1931), Plasmodium relictum (modified from Garnham 1966) and Plasmodium polare (Manwell 1934) in birds, and its capacity to transmit P. relictum in Algeria was proven experimentally (14, 15). In this context, we have assessed the larvicidal activity of EOs extracted from five aromatic medicinal plants, harvested from Northeastern Algeria, against Cs longiareolata larvae. The efficacy of the tested EOs will be evaluated by calculating the  $LC_{_{50}}$  and  $LC_{_{90}}$  values and by comparing them with the  $LC_{_{50}}$  and LC<sub>on</sub> values of the same EOs tested previously against other targeted mosquito species.

#### MATERIALS AND METHODS

#### **Mosquito Collection**

*Culiseta longiareolata* larvae were collected regularly from three clean fixed and controlled pools in Algeria, where the mosquitoes were not exposed to any insecticides. Larvae of the third and fourth instar were used directly in the test; eggs, first and second instar larvae were reared in room temperature  $(27^{\circ}C\pm 2^{\circ}C)$ , in a 12 h light: 12 h dark photoperiod, until the fourth instar was reached.

#### **Essential Oils Extraction**

The aerial parts of the tested plants were collected from different regions in the Mediterranean and semi-arid climate northeastern Algeria: *Thymus vulgaris* L. from Guelma, *Artemisia herba-alba* Asso from M'Sila, *Juniperus phoenicea* L. from Jijel, *Rosmarinus officinalis* Linn from Bouira and *Eucalyptus globules* L. from Batna. The plants' collection started at the beginning of the summer (June) in 2018. The samples were air-dried at room temperature. The dried plants were

submitted to classical steam distillation for 3-6 h. The samples were exposed to the water vapor produced in the flask crosses, the vapor was charged with the EO, and then was condensed in the condenser. The EO floated on the water surface was then recuperated. The yield of the EOs was between 0.8 and 1.5%.

#### **Larvicidal Bioassay**

According to WHO guidelines for laboratory and field testing of mosquito larvicides (16), we tested the larvicidal activity of EOs extracted from the leaves of five aromatic medicinal plants T. vulgaris, A. herba-alba, J. phoenicea, R. officinalis ,E. globulus against Culiseta longiareolata larvae under laboratory conditions. The EOs were extracted by steam distillation, they were next serially diluted in ethanol to obtain 10%, 1%, 0.1% and 0.01% of stock solution, and 0.1-1ml of the previous dilutions were added to 100ml of water to obtain the final concentrations. A series of concentrations and controls were applied on 25 mosquito larvae distributed in five cups containing 100ml of water. A total of 8925 larvae were tested. We started the test with the lowest concentrations. The concentrations that showed less than 10% mortality were excluded. Concentrations that showed 10% mortality or more were replicated 4 times, and each test was run three times. After 24h of exposure, moribund and dead larvae were counted. We have chosen four concentrations which caused between 10% and 90% mortality to determine the LC<sub>50</sub> and LC<sub>oo</sub> values. The data obtained from the four replicates in the three tests were pooled for analysis.

#### Statistical Analyses

Data were subjected to probit analysis using SPSS software V25 (Using probit model because of the normal distribution of data); and final concentrations were transformed to log10. Lethal concentration  $LC_{50}$  and  $LC_{90}$  with a 95% confidence limit (CL) suspected of killing 50% and 90% of the population respectively, were calculated and presented with the regression equations (Y= a +b\*x) and regression coefficients (R<sup>2</sup>).

#### RESULTS

Five plant EOs were tested to evaluate their larvicidal activity, and the tested oils revealed various mortality percentages at different concentrations (Table 1). The majority of the tested oils showed 100% mortality at 200ppm final concentration, except for T. vulgaris that showed 100% mortality at 80ppm. Further, the oils started to affect the larvae life at different concentrations; the lowest concentration that caused equal or more than 10% mortality was 20ppm for T. vulgaris, 40ppm for J. phoenicea, 50ppm for A. herba-alba and R. officinalis and 70ppm for E. globules (Table1). The 24h LC<sub>50</sub> and LC<sub>90</sub> estimate, upper and lower values obtained from the larvicidal activity test of EOs extracted from the five plants in addition to the regression equations and regression coefficients are presented in Table 2. T. vulgaris was the most efficient with 25.64 (16.58-32.03) LC<sub>50</sub> and 50.53 (40.15-82.43) LC<sub>90</sub>, while A. herba-alba was the least efficient. Likewise, the influence degree of increasing one unit of EOs concentration on their larvicidal activity was different. Among the tested EOs, the augmentation of one unit of *R. officinalis* concentration showed the highest influence in increasing the  $LC_{s_0}$  and  $LC_{q_0}$  (b=7.16). The R<sup>2</sup> was close to 1 in all probit analysis, the minimal residuals obtained between the observed and expected values was shown by *E. globulus* EO ( $R^2$ =0.99) (Table 2; Figures 1-5).

**Table 1:** The mortality observed to the *Culiseta longiareolata* larvae, caused by the application of the tested essential oils at different concentrations, with the arithmetic mean (AM) and standard error (SE).

		Dead in a total of 300 larvae (AM±SE)								
IC (%)	Aliquot (ml)	FC (ppm)	Thymus vulgaris	Juniperus phoenicea	Artemisia herba-alba	Rosmarinus officinalis	Eucalyptus globules			
	0,2	20	93 (7.75±1.53)	-	-	-	-			
-	0,4	40	253 (21.08±1.97)	94 (7.83±1.23)	-	-	-			
-	0,5	50	257 (21.42±1.59)	103 (8.58±1.23)	24 (2±0.75)	75 (6.25±1.54)	-			
1	0,6	60	286 (23.83±0.42)	156 (13±1.58)	57 (4.75±1.52)	106 (8.83±1.56)	-			
1.	0,7	70	-	-	-	-	68 (5.67±1.1)			
	0,8	80	300 (7.75±1.53)	176 (14.67±1.91)	89 (8.17±1.36)	236 (19.67±0.85)	107 (8.92±1.02)			
	0,9	90	-	-	-	-	134 (11.17±1.6)			
-	1	100	300 (25±0.0)	255 (21.25±1.55)	216 (18±2.03)	274 (22.83±1.21)	159 (13.25±1.69)			
10	0,2	200	300 (25±0.0)	300 (25±0.0)	300 (25±0.0)	300 (25±0.0)	300 (25±0.0)			

IC(initial concentration), FC (final concentration)

**Table 2:** The LC<sub>50</sub> and LC<sub>90</sub> values of essential oils extracted from *T. vulgaris, A. herba-alba, J. phoenicea, R. officinalis* and *E. globules* against the 3<sup>rd</sup> and 4<sup>th</sup> instar larvae of the *Culiseta longiareolata*, after 24 hours exposure period; with regression equations and regression coefficients (R2).

	LC50 (ppm) 95% Cl			LC90 (ppm) 95% Cl					
Essentialoils	Estimate	Lower	Upper	Estimate	Lower	Upper	Sig (df)	Regression equation	R2
Thymus vulgaris	25.64	16.58	32.03	50.53	40,15	82.43	p>0.05 (2)	y=-6.15+4.36*x	0.97
Juniperus phoenicea	59.83	45.36	75.81	137.68	97.21	<250	p>0.05 (3)	y=-6.49+3.66*x	0.9
Artemisia herba-alba	86.67	66.59	<250	139.55	98.03	<250	p>0.05 (2)	y=-11.77+6.08*x	0.93
Rosmarinus officinalis	64.18	55.41	72.56	96.55	82.73	139.84	p>0.05 (2)	y=-12.93+7.16*x	0.98
Eucalyptus globules	95.83	92.27	101.09	168.25	146.59	201.87	p>0.05 (2)	y=-10.45+5.28*x	0.99
Sig (significance level), df (degrees of freedom)									

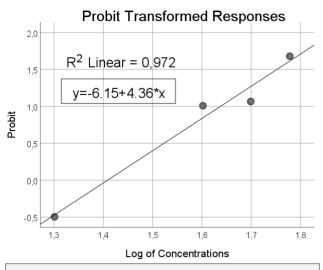


Figure 1. Probit transformed responses with equation regression and coefficient of determination  $R^2$  for *Thymus vulgaris* essential oil tested on  $3^{rd}$  and  $4^{th}$  instars larvae of *Culiseta longiareolata* for 24 h.

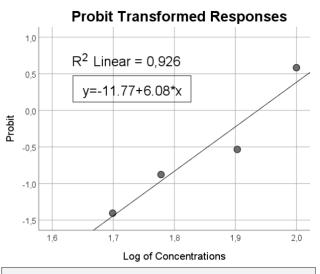


Figure 3. Probit transformed responses with equation regression and coefficient of determination  $R^2$ , for *Artemisia* herba-alba essential oil tested on  $3^{rd}$  and  $4^{th}$  instars larvae of *Culiseta longiareolata* for 24 h.

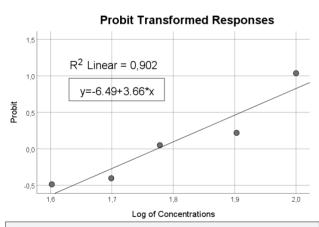


Figure 2. Probit transformed responses with equation regression and coefficient of determination  $R^2$  for *Juniperus Phoenicia* tested on  $3^{rd}$  and  $4^{th}$  instars larvae of *Culiseta longiareolata* for 24 h.

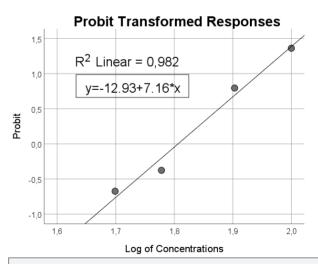


Figure 4. Probit transformed responses with equation regression and coefficient of determination  $R^2$  for *Rosmarinus officinalis* essential oil tested on  $3^{rd}$  and  $4^{th}$  instars larvae of *Culiseta longiareolata* for 24 h.

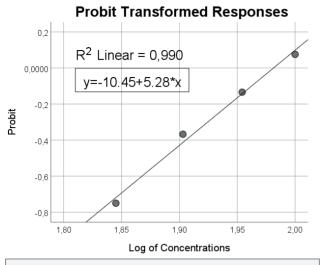


Figure 5. Probit transformed responses with equation regression and coefficient of determination  $R^2$  for and *Eucalyptus globulus* essential oil tested on  $3^{rd}$  and  $4^{th}$  instars larvae of *Culiseta longiareolata* for 24 h.

## DISCUSSION

The current study has confirmed that the EOs extracted from the aromatic medicinal plants *T. vulgaris*, *A. herba-alba*, *J. phoenicea*, *R. officinalis* and *E. globulus* present an efficient larvicidal activity against the *Culiseta longiareolata* larvae; however, the mortality responses obtained were varying.

*T. vulgaris* is a flowering herb that has a worldwide distribution (17). From the total of the tested oils, the *T. vulgaris* EO was the most efficient. This EO was previously assessed by Knio et al. (18) against the *Ochlerotatus caspius* (Pallas 1771) larvae; however,

its toxicity against Oc caspius (LC50=33.65ppm; LC90=50.85ppm) was less than that shown by our T. vulgaris EO. Likewise, the larvicidal activity of the EOs extracted from the Juniperus species was tested in previous studies: J. Phoenicea against Aede salbopictus (Skuse 1894) (LC<sub>50</sub>= 55.5ppm; LC90= 77ppm), and J. virginiana L. against Ae aegypti (Linnaeus 1762) and Cx pipiens (19, 20). Comparing our results, our J. phoenicea EO showed lower larvicidal activity against Cs longiareolata. Moreover, the larvicidal activity of R. officinalis EO was assessed against Ae albopictus (LC<sub>50</sub><250ppm), Cx tritaeniorhynchus (Giles 1901) (LC<sub>50</sub>= 115.38ppm; LC90= 211.53ppm) and Anopheles subpictus Grassi (LC<sub>50</sub>= 64.5ppm; LC90= 113.74ppm) (21, 22). The R. officinalis EOs tested against Ae albopictus, Cx tritaeniorhynchus and An subpictus in the previous researches showed lower values than the toxicity results that we obtained by testing the same EO against Cs longiareolata.

The other EOs E. globules and A. herba-alba were less efficient; however, their lethal concentrations were notable. E. grandis L. EO and its major components were assessed for their larvicidal activity against Aedes aegypti by Lucia et al. (23). The EO showed 32.4ppm LC<sub>50</sub> and the principal components  $\alpha$ -pinene (52.71%) and 1,8-cineole (18.38%) showed 15.4ppm and 57.2ppm LC 50 respectively. The principal leaf oil components of E. globules harvested from Algeria are  $\alpha$ -pinene and 1,8-cineole, according to Samir et al. (24). However, our E. globules EO tested against Cs longiareolata was less efficient (LC<sub>50</sub>= 95.83ppm). Furthermore, EOs extracted from Artemisia genus were assessed for their larvicidal activity against various mosquito species. Our A. herbaalba EO tested against Cs longiareolata larvae was more efficient (LC<sub>50</sub> = 86.67ppm) than A. vulgaris L. that was tested by Ilahi and Ullah (25) against Cx quinquefasciatus (LC<sub>50</sub>= 803.2ppm), but less efficient than A. absinthium L. tested by Govindarajan and Benelli (26) against An stephensi (Liston 1901), An subpictus, Ae aegypti, Ae albopictus, Cx quinquefasciatus (Say 1823), and Cx

**Table 3:** Principal component percentages of *T. vulgaris, A. herba-alba, J. Phoenicea, R. officinalis and E. globules* harvested from Algeria, according to previous works.

Principalcomponents	T. vulgaris (29)	J. phoenicea (30)	A. herba-alba (31)	R. officinalis (32)	E. globules (24)
Carvacrol	11.41	-	-	-	
Thymol	25.57	-	-	-	-
α-Pinene	12.1	34.5	Tr	5.4	8.8
α-Terpinylacetate	-	14.7	-	-	-
p-Cymene	26.36	-	-	-	-
Thymoquinone	10.5	-	-	-	-
β-Phellandrene	-	22.4	-	-	-
Camphor	-	-	19.4	14.6	-
1,8-Cineole	-	-	Tr	12.2	71.3
β–Caryophyllene	-	-	-	10.9	-
Borneol	-	-	-	10.6	-
γ-terpinene	-	-	23.8	-	-
β-thujone	-	-	15.0	-	-
chrysanthenone	-	-	15.8	-	-
trans-pinocarveol	-	-	16.9	-	-

*tritaeniorhynchus* ( $LC_{50}$ =41.85, 52.02, 46.33, 57.57, 50.57, and 62.16 ppm respectively). Various mosquito species were targeted in the previous researches to assess the larvicidal activity of EOs. However, *Cs longiareolata* was not previously targeted by EOs, but by the lichen metabolites evaluated by Cetin et al. (27), that showed high larvicidal activity against *Cs longiareolata*.

The results obtained confirm the previous studies; the use of EOs can serve as an eco-friendly method to control mosquito larvae. However, the noted variability in the efficacy level of the tested oils may be due to their chemical composition and the percentages of their principal components as a-Pinene, Camphor and 1,8-Cineole (Table 3); whereas, the direct use of the principal components of EOs may produce a higher efficacy in mosquito control. This hypothesis was proven in the study conducted by Lucia, Gonzalez-Audino (23), where the principal components of Turpentineand E. grandis EO showed lower  $LC_{so}$  than that obtained by the use of the entire *E. grandis* EO. Moreover, the repellency effect of the thyme EO compounds against Culex pipiens mosquito evaluated by Park et al. (28) showed higher repellent efficacy of α-Terpinene and Carvacrol than the commercial formulation diethyltoluamide (DEET), and an equal efficacy between the Thymol component and the DEET.

## CONCLUSION

The EOs extracted from the aromatic medicinal plants and their principal components may serve as safe products to control the *Culiseta longiareolata* larvae in Algeria; nevertheless, their practical application remains a fundamental step to evaluate their field efficacy and to note their possible secondary effects on non-targeted organisms.

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#### REFERENCES

- Mossa ATH, Mohafrash SM, Chandrasekaran N. Safety of Natural Insecticides: Toxic Effects on Experimental Animals. BioMed Research International. 2018; 2018: 1-17.
- 2. Hemingway J, Ranson H. Insecticide resistance in insect vectors of human disease. Annu Rev Entomol. 2000; 45(1): 371-91.
- 3. Benelli G. Research in mosquito control: current challenges for a brighter future. Parasitol Res. 2015; 114(8): 2801-5.
- 4. Benelli G, Jeffries C, Walker T. Biological control of mosquito vectors: past, present, and future. Insects. 2016; 7(4): 52.
- Pavela R. Essential oils for the development of eco-friendly mosquito larvicides: a review. Industrial crops and products. 2015; 76: 174-87.

- Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils–a review. Food Chem Toxicol. 2008; 46(2): 446-75.
- Bessah R, Benyoussef E-H. La filière des huiles essentielles Etat de l'art, impacts et enjeux socioéconomiques. Revue des Energies Renouvelables. 2015; 18(3): 513-28.
- Elimam AM, Elmalik KH, Ali FS. Efficacy of leaves extract of Calotropis procera Ait.(Asclepiadaceae) in controlling Anopheles arabiensis and Culex quinquefasciatus mosquitoes. Saudi J Biol Sci. 2009; 16(2): 95-100.
- Park IK, Lee SG, Shin SC, Park JD, Ahn YJ. Larvicidal activity of isobutylamides identified in Piper nigrum fruits against three mosquito species. J Agric Food Chem. 2002; 50(7): 1866-70.
- Ghosh A, Chowdhury N, Chandra G. Plant extracts as potential mosquito larvicides. The Indian journal of medical research. 2012; 135(5): 581.
- Markouk M, Bekkouche K, Larhsini M, Bousaid M, Lazrek H, Jana M. Evaluation of some Moroccan medicinal plant extracts for larvicidal activity. J Ethnopharmacol. 2000; 73(1-2): 293-7.
- 12. Al-Jaran TK, Katbeh-Bader AM. Laboratory studies on the biology of Culiseta longiareolata (Macquart)(Diptera: Culicidae). Aquat Insects. 2001; 23(1): 11-22.
- Kiflawi M, Blaustein L, Mangel M. Predation-dependent oviposition habitat selection by the mosquito Culiseta longiareolata: a test of competing hypotheses. Ecol Lett. 2003; 6(1): 35-40.
- 14. Valkiunas G. Avian malaria parasites and other haemosporidia: CRC press; 2004.
- Santiago-Alarcon D, Palinauskas V, Schaefer HM. Diptera vectors of avian Haemosporidian parasites: untangling parasite life cycles and their taxonomy. Biological Reviews. 2012; 87(4): 928-64.
- Organization WH. Guidelines for laboratory and field testing of mosquito larvicides. Geneva: World Health Organization. 2005.
- 17. Hosseinzadeh S, Kukhdan AJ, Hosseini A, Armand R. The application of Thymus vulgaris in traditional and modern medicine: a review. Global J Pharmacol. 2015; 9: 260-6.
- Knio K, Usta J, Dagher S, Zournajian H, Kreydiyyeh S. Larvicidal activity of essential oils extracted from commonly used herbs in Lebanon against the seaside mosquito, Ochlerotatus caspius. Bioresource technology. 2008; 99(4): 763-8.
- Lee H-S. Mosquito larvicidal activity of aromatic medicinal plant oils against Aedes aegypti and Culex pipiens pallens. J Am Mosq Control Assoc. 2006; 22(2): 292-6.
- Giatropoulos A, Pitarokili D, Papaioannou F, Papachristos DP, Koliopoulos G, Emmanouel N, et al. Essential oil composition, adult repellency and larvicidal activity of eight Cupressaceae species from Greece against Aedes albopictus (Diptera: Culicidae). Parasitol Res. 2013; 112(3): 1113-23.
- Conti B, Canale A, Bertoli A, Gozzini F, Pistelli L. Essential oil composition and larvicidal activity of six Mediterranean aromatic plants against the mosquito Aedes albopictus (Diptera: Culicidae). Parasitol Res. 2010; 107(6): 1455-61.
- Govindarajan M. Larvicidal and repellent properties of some essential oils against Culex tritaeniorhynchus Giles and Anopheles subpictus Grassi (Diptera: Culicidae). Asian Pac J Trop Med. 2011; 4(2): 106-11.
- Lucia A, Gonzalez-Audino P, Seccacini E, Licastro S, Zerba E, Masuh H. Larvicidal effect of Eucalyptus Grandis essential oil and turpentine and their major components on Aedes Aegypti larvae. J Am Mosq Control Assoc. 2007; 23(3): 299-303.
- Samir B, Benayache F, Benyahia S, Chalchat J-C, Garry R-P. Leaf Oils of some Eucalyptus Species Growing in Algeria. Journal of Essential Oil Research. 2001; 13(3): 210-3.

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- Ilahi I, Ullah F. Larvicidal activities of different parts of Artemisia vulgaris Linn. against Culex quinquefasciatus Say.(Diptera: Culicidae). International Journal of Innovation and Applied Studies. 2013; 2(2): 189-95.
- Govindarajan M, Benelli G. Artemisia absinthium-borne compounds as novel larvicides: effectiveness against six mosquito vectors and acute toxicity on non-target aquatic organisms. Parasitol Res. 2016; 115(12): 4649-61.
- Cetin H, Tufan-Cetin O, Turk A, Tay T, Candan M, Yanikoglu A, et al. Larvicidal activity of some secondary lichen metabolites against the mosquito Culiseta longiareolata Macquart (Diptera: Culicidae). Natural product research. 2012; 26(4): 350-5.
- Park B-S, Choi W-S, Kim J-H, Kim K-H, Lee S-E. Monoterpenes from thyme (Thymus vulgaris) as potential mosquito repellents. J Am Mosq Control Assoc. 2005; 21(1): 80-3.

- 29. Giordani R, Hadef Y, Kaloustian J. Compositions and antifungal activities of essential oils of some Algerian aromatic plants. Fitoterapia. 2008; 79(3): 199-203.
- Mazari K, Bendimerad N, Bekhechi C. Chemical composition and antimicrobial activity of essential oils isolated from Algerian Juniperus phoenicea L. and Cupressus sempervirens L. Journal of Medicinal Plants Research. 2010; 4(10): 959-64.
- Dob T, Benabdelkader T. Chemical composition of the essential oil of Artemisia herba-alba Asso grown in Algeria. Journal of Essential Oil Research. 2006; 18(6): 685-90.
- Djeddi DS, Bouchenah N, Settar I, Skaltsa H. Composition and antimicrobial activity of the essential oil of Rosmarinus officinalis from Algeria. Chemistry of Natural Compounds. 2007; 43(4): 487-90.