

Black Sea Journal of Agriculture Volume 7 | Special Issue

Special Issue of ICABGEH-23

VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry September 18 - 20, 2023 in Krakow, POLAND

ISSN: 2618 - 6578

BLACK SEA JOURNAL OF AGRICULTURE (BSJ AGRI)

Black Sea Journal of Agriculture (BSJ Agri) is a double-blind, peer-reviewed, open-access international journal published electronically since January 2018. The articles in this special issue were presented at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23), which was held in Krakow, POLAND, September 18 - 20, 2023.

ISSN: 2618 - 6578 Phone: +90 362 408 25 15 Fax: +90 362 408 25 15 Email: bsjagri@blackseapublishers.com Web site: http://dergipark.gov.tr/bsagriculture Publication date and place: October 15, 2024- Samsun, TÜRKİYE Publishing kind: Electronically

OWNER

Prof. Dr. Hasan ÖNDER

DIRECTOR IN CHARGE

Assoc. Prof. Uğur ŞEN

EDITOR BOARDS

EDITOR IN CHIEF

Prof. Dr. Hasan ÖNDER, Ondokuz Mayis University, TÜRKİYE Assoc. Prof. Uğur ŞEN, Ondokuz Mayis University, TÜRKİYE

SECTION EDITORS*

Prof. Dr. Berna KILIÇ, Section Editor of Aquaculture, Ege University, TÜRKİYE

Prof. Dr. Kürşat KORKMAZ, Section Editor of Soil Science and Plant Nutrition, Ordu University, TÜRKİYE

Prof. Dr. Mahmut KESKİN, Section Editor of Animal and Veterinary Sciences, Hatay Mustafa Kemal University, TÜRKİYE

Prof. Dr. Mehmet KURAN, Section Editor of Agricultural Biotechnology, Ondokuz Mayis University, TÜRKİYE

Prof. Dr. Muharrem ÖZCAN, Section Editor of Horticulture, Ondokuz Mayis University, TÜRKİYE Prof. Dr. Şerife Evrim ARICI, Section Editor of Plant Protection, Applied Sciences University of Isparta, TÜRKİYE

Assoc. Prof. Dr. Hasan Gökhan DOĞAN, Section Editor of Agricultural Economics, Kirsehir Ahi Evran University, TÜRKİYE

Prof. Dr. Kadir Ersin TEMİZEL, Section Editor of Agricultural Structures and Irrigation, Ondokuz Mayis University, TÜRKİYE

Assoc. Prof. Dr. Kadyrbay CHEKİROV, Section Editor of Biological Researches, Kyrgyz Turkish Manas University, KYRGYZSTAN

Assoc. Prof. Dr. Kemal Çağatay SELVİ, Section Editor of Agricultural Machinery and Technologies Engineering, Ondokuz Mayis University, TÜRKİYE

Assoc. Prof. Dr. Ömer SÖZEN, Section Editor of Field Crops, Kirsehir Ahi Evran University, TÜRKİYE

Assoc. Prof. Dr. Şerife Evrim ARICI, Section Editor of Süleyman Demirel University, TÜRKİYE

Assoc. Prof. Dr. Ümran ÇİÇEK, Section Editor of Nutrition and Food Sciences, Tokat Gaziosmanpasa University, TÜRKİYE

* The ranking is arranged alphabetically within the academic title

EDITORIAL - ADVISORY BOARD*

Prof. Dr. Ahmet BALKAYA, Ondokuz Mayis University, TÜRKİYE Prof. Dr. Alireza TERINEJAD, Azarbaijan Shahid Madani University, IRAN Prof. Dr. Zeynel CEBECİ, Çukurova University, TÜRKİYE Prof. Dr. Gheorghe VOICU, Politehnica University of Bucharest, ROMANIA Prof. Dr. Savaş ATASEVER, Ondokuz Mayis University, TÜRKİYE Prof. Dr. Sherein Saeid ABDELGAYED, Cairo University, EGYPT Prof. Dr. Habil Sorin STEFAN BIRIS, Politehnica University of Bucharest, ROMANIA Prof. Dr. Mustafa BOĞA, Nigde Ömer Halisdemir University, TÜRKİYE Dr. Jiban SHRESTHA, Nepal Agricultural Research Council, NEPAL Dr. Mikó JÓZSEFNÉ JÓNÁS, University of Szeged, HUNGARY Dr. Valentin VLADUT, National Research and Development Institute, ROMANIA

* The ranking is arranged alphabetically within the academic title

SCIENTIFIC COMMITTEE OF ICABGEH-23*

Prof. Dr. Adem KAMALAK, Sutcu Imam University, TÜRKİYE Prof. Dr. Cathy DWYER, SRUC, SCOTLAND Prof. Dr. Cigdem TAKMA, Ege University, TÜRKİYE Prof. Dr. Dorian J. GARRICK, Massey University, NEW ZEALAND Prof. Dr. Edit Mikó JONAS, University of Szeged, HUNGARY Prof. Dr. Hao CHENG, Iowa State University, USA Prof. Dr. Karl SCHELLANDER, University of Bonn, GERMANY Prof. Dr. Kaspar BINENEFELD, Humboldt University, GERMANY Prof. Dr. Khalid JAVED, University of Veterinary and Animal Science, PAKISTAN Prof. Dr. Kustantinah ADIWIMARTA, Gadjah M. University, INDONESIA Prof. Dr. Mahmut KESKIN, Mustafa Kemal University, TÜRKİYE Prof. Dr. Mahmut YILMAZ, Kırsehir Ahi Evran University, TÜRKİYE Prof. Dr. Maria DATTENA, AGRIS, ITALY Prof. Dr. Masroor A.BAJWA, University of Balochistan, PAKISTAN Prof. Dr. Mohammad M. TARIQ, University of Agriculture, PAKISTAN Prof. Dr. Muhammad KHALID, University of London, UK Prof. Dr. Rhan L. FERNANDO, Iowa State University, USA Prof. Dr. Savas ATASEVER, Ondokuz Mayis University, TÜRKİYE Prof. Dr. Sherein Saeid ABDELGAYED, Cairo University, EGYPT Assoc. Prof. Dr. Abdulmojeed YAKUBU, Nasarawa State University, NIGERIA

Assoc. Prof. Dr. Aleksandra DUNISLAWSKA, Bydgoszcz University of Science and Technology, POLAND Assoc. Prof. Dr. Farhat IQBAL, University of Balochistan, PAKISTAN Assoc. Prof. Dr. Petr KUNC, University of Prague, CZECHIA Assoc. Prof. Dr. Saltanat ISSABEKOVA, Saken Seifullin Kazakh Agrotechnical University, KAZAKHSTAN Assist. Prof. Dr. Farhood GOLMOHAMMADI, Islamic Azad University, IRAN Assist. Prof. Dr. Ivona DIMITROVA, University of Forestry, BULGARIA Assist. Prof. Dr. Lilija DEGOLA, Latvian University, LATVIA Assist. Prof. Dr. Pramote PAENGKOUM, Suranaree University, THAILAND Dr. Fadjeria YAACOUB, University of Biskra, ALGERIA Dr. Gracinda Andre MATAVEIA, E Mondlane University, MOZAMBIQUE Dr. Thobela Louis TYASI, University of Limpopo, SOUTH AFRICA

* The ranking is arranged alphabetically within the academic title

STATISTIC EDITOR

Prof. Dr. Taner TUNÇ, Ondokuz Mayis University, TÜRKİYE

ENGLISH EDITOR

Asst. Prof. Dr. Betül ÖZCAN DOST, Ondokuz Mayis University, TÜRKİYE

REVIEWERS OF THE ISSUE*

Prof. Dr. Belgin Coşge ŞENKAL, Yozgat Bozok University, Department of Field Crops, TÜRKİYE

Prof. Dr. İsmail ÜLGER, Erciyes University, Department of Animal Science, TÜRKİYE

Prof. Dr. Mustafa Boğa, Niğde Ömer Halisdemir University, Department of Animal Science, TÜRKİYE

Assoc. Prof. Dr. Aycan Mutlu YAĞANOĞLU, Atatürk University, Department of Animal Science, TÜRKİYE

Assoc. Prof. Dr. Bülent BÜLBÜL, Dokuz Eylül University, Department of Reproduction and Artificial Insemination, TÜRKİYE

Assoc. Prof. Dr. Çağrı Özgür ÖZKAN, Kahramanmaraş Sütçü İmam University, Department of Animal Science, TÜRKİYE

Assoc. Prof. Dr. Canan KOP BOZBAY, Eskişehir Osmangazi University, Department of Animal Science, TÜRKİYE

Assoc. Prof. Dr. İsa COŞKUN, Kırşehir Ahi Evran University, Department of Animal Science, TÜRKİYE

Asist. Prof. Dr. Çiğdem ŞEREMET, Ege University, Department of Animal Science, TÜRKİYE

Asist. Prof. Dr. Fatma AKBAY, Malatya Turgut Özal University, Department of Field Crops, TÜRKİYE

Asist. Prof. Dr. Muhammed Ali KARA, Siirt University, Department of Animal Science, TÜRKİYE

Asist. Prof. Dr. Zeynep YARDIM, Aydın Adnan Menderes University, Department of Animal Science, TÜRKİYE

Dr. Ebru ERGÜNEŞ BERKIN, Islahiye District Directorate of Agriculture and Forestry,. Department of Animal Science, TÜRKİYE

Dr. Kalbiye KONANÇ, Ordu University, Department of Animal Science, TÜRKİYE

* The ranking is arranged alphabetically within the academic title

Volume 7, Special Issue 4 (ICABGEH-23)

Table of Contents

Research Articles

Merve GÜNDÜZ, Arda SÖZCÜ, Üzeyir ÇAĞIRICI..……………………………………………..………464-476

doi: 10.47115/bsagriculture.1419351

Open Access Journal e-ISSN: 2618 – 6578

Research Article

Volume 7 - Special Issue (ICABGEH-23): 429-433/ August 2024

EFFECT OF VARIETY ON THE POTENTIAL NUTRITIVE VALUE OF OAT HAYS

Bilal SELÇUK1*, Adem KAMALAK2, Mustafa YILDIRIM³

¹Ondokuz Mayıs University, Ladik Vocational School, Department of Plant and Animal production, 55760, Ladik, Samsun, Türkiye ²Kahramanmaraş Sütçü İmam University, Faculty of Agriculture, Department of Animal Science, 46100, Kahramanmaraş, Turkiye ³Kahramanmaraş Sütçü İmam University, Faculty of Agriculture, Department of Field Crops, 46100, Kahramanmaraş, Turkiye

Abstract: This study was conducted to determine the differences in the chemical composition, *in vitro* gas production (GP), methane production (CH4), metabolizable energy (ME), and *in vitro* organic matter digestion (IVOMD) of oat hay varieties commonly used in ruminant feeding (Küçükyayla, Kahraman, Kırklar, ST-4, Yeniceri, Sebat, and Arslanbey). The oat hay varieties in the study were harvested during the flowering period in the 2019-2020 season in Kahramanmaras province. The *in vitro* findings of this study revealed significant differences among oat hay varieties in terms of their chemical composition, *in vitro* gas production, methane production, ME and IVOMD (P<0.001). The crude protein (CP) content of oat hays ranged from 7.61% to 9.57%, neutral detergent fiber (NDF) ranged from 64.46% to 72.96%, acid detergent fiber (ADF) ranged from 36.74% to 41.70%, crude ash (CA) ranged from 6.56% to 7.91%, metabolizable energy ranged from 6.96 to 7.98 MJ kg⁻¹ DM, IVOMD ranged from 67.30% to 74.90%, and methane production rate ranged from 15.42% to 16.35%. The Yeniceri variety stood out with a NDF content of 64.46%, an ADF content of 36.74%, a ME of 7.98 MJ kg-1 DM, and an IVOMD of 74.90%. ST-4 had the highest *in vitro* gas production with 49.46 ml, while Sebat had the highest methane production rate with 15.42%. In conclusion, considering the chemical composition and fermentation parameters, the Yeniceri variety can be considered a potential source of forage, but further *in vivo* studies are needed to assess their effects on feed intake and animal production.

Keywords: Oat hay varieties, in vitro, Methane production, Metabolic energy

1. Introduction

Oats are preferred as a feed ingredient in animal nutrition due to their high nutritional value, antioxidant content, and facilitative effect on digestion (Peterson et al., 2005; Naneli and Sakin, 2017). Oat grain is known as a functional food due to its distinct chemical composition, nutritional value, and beneficial effects on health compared to other cereals (Biel et al., 2014; Sterna et al., 2016). Consequently, significant efforts have been made in recent years in agronomic research, leading to the emergence of a substantial number of new oat varieties with different agronomic characteristics (Buerstmayr et al., 2007; Redaelli, 2008; Martínez et al., 2010).

The nutrient content of forages included in the rations of ruminant animals and their microbial digestion in the rumen, as well as the extent to which they are converted into metabolic energy, are of vital importance for ruminants (Ørskov and McDonald, 1979). Ruminants, due to their digestive systems, undergo ruminal fermentation of the consumed feed, resulting in the production of methane (CH4), which is released into the environment. This emission leads to an energy loss of approximately 12-16% from the feed (Johnson and Johnson, 1995; IPCC, 2001). Therefore, low methaneproducing feeds should be preferred in ruminant rations (López et al., 2010). Additionally, factors such as the timing of forage harvesting, maturity stage, and processing methods applied to these feeds, such as grinding, drying, wilting, etc., greatly affect their potential nutritional values and fermentation parameters (Doane et al., 1997; Sanderson et al., 1997; Filya et al., 2002).

The aim of the current experiment was to evaluate effect of variety on chemical composition; *in vitro* gas production, methane production, ME and IVOMD.

2. Materials and Methods

The trial was conducted at the experimental field of the Faculty of Agriculture, Kahramanmaras Sutcu Imam University, during the 2019–2020 season. Seven oat genotypes (Kucukyayla, Kahraman, Kırklar, ST-4,

Yeniceri, Sebat and Arslanbey) were used in the study. The experiment was set up in a randomized complete block design with three replications. The plot size was 7 m long and 6 rows wide, with a spacing of 20 m between rows. The seeding rate was maintained at 500 seeds/m2. Fertilization was applied at a rate of 12 kg/ha of pure nitrogen, half of which was applied as diammonium phosphate (DAP) fertilizer as top dressing, while the other half was applied as urea during the tillering stage. The oat varieties were harvested during the flowering period, and the remaining oat hay was brought to the laboratory for further analysis.

Dry matter, crude ash, crude protein and ether extract of oat hay samples were analyzed using the method of AOAC (1990). NDF and ADF contents of oat hay samples were analyzed with the method suggested by Van Soest (1991). All chemical analyses were carried out in triplicate.

The *in vitro* gas production technique was utilized to determine the gas production and methane production of oat hay. Rumen fluid was obtained from three sheep from a private abattoir in Kahramanmaras province and transferred to the laboratory in a thermos. The rumen fluid was then filtered through four layers of cheesecloth while being agitated with CO₂. Approximately 200 mg of oat hay samples were weighed into 100 ml glass syringes in triplicate. Subsequently, 30 ml of buffered rumen fluid (1:2 V/V) was added to the glass syringes containing oat hay samples and transferred to a water bath set at 39 °C for 24 hours of incubation. To obtain blanks, the same amount of buffered rumen fluid without substrate was added to four glass syringes (Menke et al., 1979). Gas and CH⁴ production of the oat hay samples were measured after 24 hours of incubation. After the 24 hour incubation, the total gas production and the percentage of CH⁴ in oat hay samples were determined using an infrared methane analyzer (Sensor Europe GmbH, Erkrath, Germany) (Goel et al., 2008).

The methane productions of oat hay samples as ml were calculated as follows in Equations 1-3:

CH4 production (ml)

$$
= Total gas production (ml)
$$

\n
$$
* Percentage of CH4 (%)
$$
 (1)

The ME and IVOMD of oat hay samples were estimated with equations suggested by Menke and Steingass (1988).

$$
ME\left(\frac{MJ}{kg}DM\right) = 1.06 + (0.1570 * GP) + (0.084
$$

 * CP) + (0.220 * EE) - (0.081
 * CA) (2)

$$
IVOMD (\%) = 28.49 + (0.7967 * GP) + (0.325
$$

* *CP*) (3)

here, GP= gas production of 200 mg sample at 24 h incubation (ml), CP= crude protein (%), EE= ether extract $(\%)$, and CA= crude ash $(\%)$.

2.1. Statistical Analyses

One-way analysis of variance (ANOVA) was used to determine the effect of variety on chemical composition, *in vitro* gas production, methane production, ME and OMD of oat hay samples. Differences (P<0.05) among the mean of oat hay varieties were determined with Tukey's multiple range tests (Genç and Soysal, 2018).

3. Results

The effect of variety on the chemical composition of oat hay is presented in Table 1. Variety had a significant impact on the chemical composition of oat hay (P<0.05). The dry matter content of oat hay varieties ranged from 23.25% to 26.26%. The crude ash content of oat varieties ranged from 6.56% to 7.91%. The crude protein content of oat hay varieties ranged from 7.61% to 9.57%, with the highest values found in Kucukyayla, and Arslanbey variety, the lowest in Kırklar. Previous studies have reported crude protein contents of oat hay ranging from 4.2% to 9.2% (Sehu et al., 1998; Gursoy, 2023). The NDF (neutral detergent fiber) and ADF (acid detergent fiber) contents of oat varieties ranged from 64.46% to 72.96% and 36.74% to 41.70%, respectively.

The effect of variety on gas production, methane production, metabolizable energy (ME) and *in vitro* organic matter digestibility (IVOMD) of oat hay is shown in Table 2. Variety had a significant impact on methane production, ME, and IVOMD of oat hay. Gas production and methane production ranged from 50.17 to 58.73 ml and 7.98 to 9.37 ml, respectively. The percentage of CH⁴ in oat hay ranged from 15.42% to 16.35%. ME and IVOMD of oat hay from different varieties ranged from 6.96 to 8.42 MJ (kg/DM) and 67.30% to 74.87%, respectively, with the highest values observed in Yeniceri and the lowest in ST-4.

4. Discussion

Gas production in feeds occurs as a result of the reaction between fermentable carbohydrates and buffer solutions, leading to the production of volatile fatty acids (Wolin, 1960). It has been reported that an increase in ruminal gas production may be associated with an increase in fermentable carbohydrate content (Sampath et al., 1995). The *in vitro* gas production values in the study indicate that the Yeniceri and Kucukyayla varieties fermented well compared to other varieties. In a study conducted by Lopez et al. (2010), feed ingredients were classified based on their anti-methanogenic properties. Feed ingredients with percentages ranging from 11% to 14% were classified as low, 6% to 11% as moderate, and 0% to 6% as high anti-methanogenic character. The findings of the current study revealed that oat hay varieties did not exhibit any anti-methanogenic effect according to the classification by Lopez et al. (2010).

It has been reported that if the crude protein (CP) content of feed ingredients is below 8%, the enzymatic activities of microorganisms in the rumen may be limited, resulting in an inadequate supply of ammonia in the rumen (Norton, 2012; Cappellozza et al., 2013).

ab: Column means with common superscripts do not differ (P<0.05), SEM= standard error mean, DM= dry matter (%), CA= crude ash (%), CP= crude protein (%), EE= ether extract (%), ADF= acid detergent fiber (%), NDF= neutral detergent fiber (%), P<0.05.

ab: column means with common superscripts do not differ (P<0.05), SEM= standard error mean, GP= gas production (ml), CH₄= methane production (ml), CH4= methane production (%), ME= metabolic energy (MJ/kg DM), IVOMD= *in vitro* organic matter digestibility (%), P<0.05.

With the exception of the Kırklar variety, the CP levels of oat hay varieties in the study can be considered sufficient for the proper functioning of microbial activity in the rumen. The difference in CP contents of oat varieties between the two experiments is possibly associated with differences in climatic conditions, fertilization, and soil type of the growing site (Sehu et al., 1998; Gursoy, 2023). In rations, it is desired to have low acid detergent fiber (ADF) levels as it is difficult to digest in the rumen (Van Soest, 2018). An increase in ADF levels in the ration has been reported to result in a feeling of fullness in ruminants, leading to a decrease in feed intake and consequently a decrease in the utilization of energy and protein from the feed (Yavuz, 2005). Therefore, it is recommended to have ADF levels in ruminant rations between 21% and 30% (Balthrop et al., 2011). The ADF contents of oat hay varieties in the study were found to be higher than the desired optimal level.

The metabolizable energy (ME) values of oat hays obtained in the current experiment are consistent with the values indicated by NRC (2007). Furthermore, an increase in the cell wall content in feeds has been reported to result in a decrease in ME and organic matter digestibility (OMD) values (Sagocak, 2011).

5. Conclusion

Variety had a significant effect on chemical compositions, gas production, CH⁴ production, ME and IVOMD of hay. There is considerable amount of variation among oat varieties in terms of chemical compositions, gas production, CH⁴ production, ME and IVOMD of oat hay. The oat hays from different varieties had provided new raw materials with a range of nutritional characteristics and will provide not only energy and protein but also fiber for ruminant animals. Based on the chemical composition and fermentation parameters, variety Yeniceri can be recommended for hay production since it has a high CP, ME and IVOMD. However, *in vivo* studies are needed to determine the effects of oat hay varieties on feed intake and growth performance on ruminant animals.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C=Concept, D= design, S= supervision, DCP= data collection and/or processing, DAI= data analysis and/or interpretation, L= literature search, W= writing, CR= critical review, SR= submission and revision, PM= project management, FA= funding acquisition.

Conflict of Interest

The authors declared that there is no conflict of interest.

Ethical Consideration

Ethics committee approval was not required for this study because of there was no study on animals or humans.

Acknowledgments

This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- AOAC. 1990. Official method of analysis, 15th ed. Association of Official Analytical Chemists, Washington, US, pp: 66-88.
- Balthrop J, Brand B, Cowie RA, Danier J, de Boever JL, de Jonge LH, Piotrowski C. 2011. Quality assurance for animal feed analysis laboratories (No. 14). FAO, pp: 152.
- Biel W, Jacyno E, Kawecka M. 2014. Chemical composition of hulled, dehulled and naked oat grains. S Afr J Anim Sci, 44(2): 189-197.
- Buerstmayr H, Krenn N, Stephan U, Grausgruber H, Zechner E. 2007. Agronomic performance and quality of oat (Avena sativa L.) genotypes of worldwide origin produced under Central European growing conditions. Field Crops Res, 101: 341-351.8.
- Cappellozza BI, Bohnert DW, Schauer CS, Falck SJ, Vanzant ES, Harmon DL, Cooke RF. 2013. Daily and alternate day supplementation of urea or soybean meal to ruminants consuming low-quality cool-season forage: II. Effects on ruminal fermentation. Livestock Sci, 155(2-3): 214-222.
- Doane PH, Schofield P, Pell AN. 1997. Neutral detergent fiber disappearance and gas and volatile fatty acid production during the in vitro fermentation of six forages. J Anim Sci, 75(12): 3342-3352.

Filya I, Karabulut A, Canbolat O, Degirmencioglu T, Kalkan H.

2002. Bursa bolgesinde yetistirilen yem hammaddelerinin besleme degeri ve hayvansal organizmada optimum degerlendirme kosullarının in vivo ve in vitro yontemlerle saptanması uzerinde arastırmalar. UU Ziraat Fak Bil Aras Inc Ser, 25: 1-16.

- Genç S, Soysal Mİ. 2018. Parametric and nonparametric post hoc tests. BSJ Eng Sci, 1(1): 18-27.
- Goel G, Makkar HP, Becker K. 2008. Effects of Sesbania sesban and Carduus pycnocephalus leaves and Fenugreek (Trigonella foenum-graecum L.) seeds and their extracts on partitioning of nutrients from roughage-and concentratebased feeds to methane. Anim Feed Sci Technol, 147(1-3): 72-89.
- Gursoy E. 2023. Samanların besin degeri ve sindirilebilirligini artırma yontemleri. Kadirli Uyg Bil Fak Derg, 3(1): 160-169.
- IPCC. 2001. Climate Change 2001: The Scientific Basis. Contribution of Working Group Ito the Third Assessment Report of the Intergovernmental Panel on Climate Change.
- Johnson KA, Johnson DE. 1995. Methane emissions from cattle. J Anim Sci, 73(8): 2483-2492.
- López S, Makkar HP, Soliva CR. 2010. Screening Plants and Plant Products for Methane Inhibitors. In: Vercoe, P., Makkar, H., Schlink, A. (eds) In vitro screening of plant resources for extra-nutritional attributes in ruminants: nuclear and related methodologies. Springer, Dordrecht, the Netherlands, pp: 191-231. https://doi.org/10.1007/978-90-481-3297-3_10
- Martínez MF, Arelovich HM, Wehrhahne LN. 2010. Grain yield, nutrient content and lipid profile of oat genotypes grown in a semiarid environment. Field Crops Res, 116(1-2): 92-100.
- Menke KH, Raab L, Salewski A, Steingass H, Fritz D, Schneider W. 1979. The estimation of the digestibility and metabolizable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor in vitro. J Agri Sci (Camb), 93: 217-222.
- Menke KH, Steingass H. 1988. Estimation of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. Anim Res Dev, 28: 7-55.
- Naneli I, Sakin MA. 2017. Bazı yulaf çeşitlerinin (Avena Sativa L.) farklı lokasyonlarda verim ve kalite parametrelerinin belirlenmesi. Tarla Bitkileri Merkez Aras Enst Derg, 26: 37- 45. https://doi.org/10.21566/tarbitderg.359057.
- Norton BW. 2012. The nutritive value of tree legumes. URL: http://www.fao.org/ag/AGP/AGPC/doc/Pubicat/Guttshel/ x5556e0j.htm (Accesed date, November 07, 2022).
- NRC. 2007. National Research Council: Nutrient requirements of small ruminants.
- Ørskov ER, McDonald I. 1979. The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. J Agri Sci, 92(2): 499- 503.
- Peterson DM, Wesenberg DM, Burrup DE, Erickson CA. 2005. Relationships among agronomic traits and grain composition in oat genotypes grown in different environments. Crop Sci, 45: 1249-1255.
- Redaelli R, Lagana P, Rizza F, Nicosia OLD, Cattivelli L. 2008. Genetic progress of oats in Italy. Euphytica, 164: 679-687.
- Sagocak AT. 2011. Determination of potential nutritive value of leaves Arbutus adrachne. MSc Thesis, Kahramanmaras Sutcu Imam University, Institute of Science, Kahramanmaraş, Türkiye, pp: 63.
- Sampath KT, Wood CD, Prasad CS. 1995. Effect of urea and by products on the in vitro fermentation of untreated and urea treated finger millet (Eleusine coracana) straw. J Sci Food Agri, 67(3): 323-328.

Sanderson R, Lister, SJ, Sargeant, A, Dhanoa MS. 1997. Effect of

particle size on in vitro fermentation of silages differing in dry matter content. In Proceedings of the British Society of Animal Science Vol. 1997, Cambridge University Press, Cambridge, UK, pp: 197-197.

- Sehu A, Yalçin S, Önol AG. Koçak D. 1998. Prediction of dry matter intake and live weight gain in lambs by some characteristics of roughages. Turkish J Vet Anim Sci, 22(6): 475-484.
- Sterna V, Zute S, Brunava L. 2016. Oat grain composition and its nutrition benefice. Agri Agricultural Sci Proc, 8: 252-256.

Van Soest PJ. 2018. Nutritional ecology of the ruminant. Cornell

University Press, New York, US, pp: 488.

- Van Soest PV, Robertson JB, Lewis, BA. 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. J Dairy Sci, 74(10): 3583-3597.
- Wolin MJ. 1960. A theoretical rumen fermentation balance. J Dairy Sci, 43(10): 1452-1459.
- Yavuz M. 2005. Bazı ruminant yemlerinin nispi yem degeri ve in vitro sindirim degerlerinin belirlenmesi. J Agri Fac Gaziosmanpasa Univ, 22(1): 97-101.

doi: 10.47115/bsagriculture. 1480587

Open Access Journal e-ISSN: 2618 – 6578

Research Article

Volume 7 - Special Issue (ICABGEH-23): 434-438/ August 2024

EFFECTS OF SPIRULINA (*S. PLATENSIS***) ADDITION TO DIETS OF BREEDER QUAILS**

Batuhan DİLBER1, Rabia GÖÇMEN2*

¹Atatürk University, Faculty of Agriculture, Department of Agricultural Biotechnology, 25240, Erzurum, Türkiye ²Selçuk University, Faculty of Agriculture, Department of Animal Science, 42130, Konya, Türkiye

Abstract: The experiment aimed to investigate how different levels of Spirulina (spirulina platensis) supplementation in the diet of breeder Japanese quail (Coturnix coturnix japonica) affect their laying performance, egg quality, and hatchability parameters. The study was carried out with 96 breeding quails (72 female, 24 male). Quails were randomly distributed to 4 treatment groups with 24 subgroups (1 male 3 female). The feeding trial lasted for 8 weeks, during which time four diets were prepared with powdered Spirulina supplementation at 0% (control), 0.75%, 1.5%, and 2.25% to the basal diet. At the end of the experiment, feed consumption was higher in the control group than in the other groups (P<0.05). The control group had the highest feed conversion ratio (FCR) while the group fed with 2.25% SP had the lowest FCR (P<0.05). No considerable differences were observed between the groups in terms of egg production, body weight, egg weight, and egg mass. Similarly, eggshell thickness, eggshell weight, eggshell ratio, egg shape, albumen, and yolk index parameters were not significantly affected by the treatment. L* value, which is one of the egg yolk color parameters, decreased linearly with increasing spirulina levels in the diet, while a* and b^* values increased (P<0.01). Differences between treatment groups in terms of incubation parameters were not significant. In conclusion; the addition of 0.75%, 1.5%, and 2.25% SP to the diets of breeder Japanese quail had positive effects on feed intake and feed conversion ratio, and egg yolk color, but had no negative effects on egg quality and hatching parameters. It was concluded that spirulina can be used at a 2.25% level in breeder Japanese quail diets.

Keywords: Breeder Japanese quail, Spirulina (*spirulina platensis*), Incubation parameters, Performance, Egg yolk color, Egg quality

1. Introduction

Spirulina, an edible cyanobacterium, has a spiral-shaped and filamentous structure. This bacterium is commonly found in alkaline lakes in nature. Previously, it was classified as a type of blue-green algae (Becker, 2007; Gupta et al., 2008). This is the reason why it attracts attention as animal feed, and human food due to its high crude protein, essential amino acid, fat, vitamin, and mineral content in its dry matter (Mišurcová et al., 2014; Salmeán et al., 2015). Spirulina platensis has antioxidant, anti-inflammatory, immune-modulating, and hepatoprotective functions (Jeyaprakash and Chinnaswamy, 2005; Abdel-Daim et al., 2015; Ibrahim and Abdel-Daim, 2015; Jamil et al., 2015). It can be grown in bioreactors or open ponds without using agricultural land (Chen et al., 2016). It is a sustainable resource that can be obtained without using agricultural land. Microorganisms, which can synthesize the nutrients (carbohydrates, protein, lipids, minerals, and vitamins) that form the basis of nutrition from simple organic and inorganic substances, gain great importance in solving the problem of food insufficiency in the world. Cellulose

is not found in the cell wall of Spirulina Platensis. This can be absorbed by improving mucosal digestion and intestinal function. While the population of beneficial microorganisms (such as Bifidobacter, and Lactobacillus) increases, harmful microorganisms (such as Candida) are suppressed. The increase in the Lactobacillus-type microorganism population positively affects food digestion and absorption (Seyidoglu et al., 2017). There are some desirable properties in a source that can be used as an alternative in animal nutrition. Ideally, the features expected from this new source are high nutritional value and efficiency of conversion to animal products, optimizing quality of product, and using water and land efficiently (Poppi and McLennan, 2010). Spirulina is a strong candidate with the potential to meet all of these features. This trial was carried out to find out the effects of powdered spirulina added to breeder quail diets in the amounts of 0, 0.75, 1.5, and 2.25% on egg quality, performance, and hatching parameters. The efficacy of spirulina as a viable feed additive was evaluated in the investigation.

2. Materials and Methods

A total of 96 breeder quails (*Coturnix coturnix japonica*), 72 females and 24 males, aged 10 weeks, were used in the study. Four treatment groups were created to distribute quails, with each group having 24 subgroups. In each subgroup, there were 3 female quails and 1 male quail. For eight weeks, a specialized feed was prepared to meet the nutritional requirements of breeding quail. The

feed was formulated to contain 2900 Kcal/kg metabolizable energy and 20% crude protein (Council, 1994). Spirulina (Spirulina platensis) powder form was used in the experiment. A set of rations for a trial was prepared, including a basal ration (0%) mainly made from corn and soybean meal. The preparation involved the use of Spirulina (*Spirulina platensis*) at different percentages of 0.75%, 1.5%, and 2.25% (Table 1).

The experiment was carried out in multi-storey breeder cages with 3 cells (45x30x25 cm) on each floor in the quail unit located at the Faculty of Agriculture Department of Animal Science Farm of Selçuk University. In the experiment, quails were given trial rations for 8 weeks. During the trial, a 16-hour light/8-hour dark lighting program was applied. Feed and water were given ad libitum. At the beginning and end of the experiment, the animals in each cage were weighed as a group. In the experiment, two periods of measurement were made, each lasting 4 weeks. The given results are the average of the measurements taken in two periods. The amount of feed given was recorded daily. At the end of each period, feed intake (FI) was calculated from these records as the daily average FI per quail. For each period, the feed conversion ratio was calculated by dividing the daily average feed intake (in grams) per quail by the egg mass (in grams) during that period. Egg production (EP) was recorded daily and determined from these records at the end of each period. The weight of eggs (EW) was calculated by taking an average of the egg weights collected during the final two days of each period. To determine egg mass, the formula EM= (egg production x egg weight)/100 was used. Eggs were gathered during the final two days of each period for the experiment, where various factors were measured including shell thickness, weight, shape index, yolk and albumen index, and Haugh unit. The colorimeter (Konica Minolta CR410) was used to measure L*, a*, and b* values of egg yolk. End of the experiment, to determine the hatching properties, the eggs were collected for 7 days, and those with hatchability properties were numbered according to groups and placed in the incubator. The number of chicks hatched after the seventeenth day and the number of fertile and non-fertile eggs was determined by breaking the eggs that did not hatch after the twentieth day. The data obtained was used to calculate the hatching efficiency, fertility rate, and hatchability (Erensayın, 2000). One-way analysis of variance (ANOVA) was used for statistical analysis of the experiment's data (Minitab, 2000) and to determine the differences between the means, Duncan's multiple comparisons test was utilized.

3. Results

The effects of supplementing breeder Japanese quail diets with SP at varying levels (0%, 0.75%, 1.5%, and 2.25%) on parameters such as LWC, FI, FCR, EP, EW, and EM are presented in Table 2. The live weight change, egg weight, egg mass, and egg production of breeder quails were not affected by the addition of spirulina in their diets. However, the inclusion of spirulina resulted in a decrease in feed intake and feed conversion ratio. The lowest feed consumption was in the 2.25% SP group (P<0.05). The best feed conversion ratio was in the 2.25% SP group (P<0.05). Table 3 presents the effects of adding spirulina to the diets of breeder Japanese quails on egg quality parameters, including yolk L^* , a^* , and b^* values. The effect of spirulina supplements on breeder Japanese quail diets on eggshell thickness, eggshell weight, eggshell ratio, yolk index, and Haugh Unit were not statistically significant. Table 4 presents the outcomes of incorporating spirulina into the diets of breeder quails, and its effect on the incubation parameters. There were no significant differences observed in the incubation parameters, including chick's weight, hatchability, and fertility, between the treatment groups (P>0.05), indicating that the treatment did not have a significant impact on these parameters.

a, bThe differences indicated by different letters on the same row are statistically significant, P<0.05

Table 3. The effect of dietary different levels of SP on the egg quality, yolk L*, a* and b* values of breeder Japanese quails

A, BThe differences indicated by different letters on the same row are statistically significant, P<0.01

4. Discussion

Numerous studies have shown that spirulina has an impact on poultry performance. It has been reported that diets containing Spirulina positively affect the production performance of laying poultry (Takashi, 2003; Nikodémusz et al., 2010). Similarly, adding spirulina has been reported to improve the FCR of laying hens (Kharde et al., 2012; Shanmugapriya et al., 2015). According to Mariey et al. (2012), adding 0.2 % spirulina to the laying hen feed improved FCR, EW, EM, EP, and FCR (P<0.05). It is similar to the results of the current study, but studies are reporting that adding spirulina to poultry diets does not affect performance data. Reportedly, the inclusion of spirulina in quail diets does not have any impact on their feed intake (Hajati and Zaghari, 2019; Omri et al., 2019). Zahroojian et al. (2013) stated that SP addition to laying hen diets had no statistically significant effect on EP, FI, FCR, and EW (P>0.05). Additionally, Hajati and Zaghari (2019) stated that SP did not affect FCR, EW, and EP. The reason for the differences in the results may be the amount of SP used the housing conditions of the animals, the animal species, the compositions of feed raw

materials and production systems, etc.

Selim et al. (2018) conclusively demonstrated that the supplementation of spirulina (0, 0.1, 0.2, and 0.3%) to laying hen rations did not affect the shell percentage, albumen index, yolk index, shape index, and Haugh Unit. Similarly, studies are reporting that adding different amounts of spirulina to laying hen diets does not affect egg quality (Zahroojian et al., 2013; Curabay et al., 2021). The spirulina supplement to breeder quail diets decreased the L^* value and increased the a^* and b^* values. A decrease in L* results in decreased brightness, while an increase in a* and b* leads to increased redness and yellowness. Egg yolks in laying hens supplemented with spirulina were reported to be redder and less yellow than the control group (P< 0.01) (Omri et al., 2019). The inclusion of 2.5% SP in the diet of quails resulted in an increase in the a* and b* values of the breast meat (Göçmen, 2022). Studies are reporting that adding SP to the diet increases the egg yolk score (Mariey et al., 2012; Selim et al., 2018). Spirulina's high carotenoids, including β-carotene, zeaxanthin, and xanthophylls, accumulate in the yolk, resulting in its characteristic color (Anderson et al., 1991; Takashi, 2003; Kotrbáček et al., 2013). Spirulina is a good source of xanthophyll for yolk (Ross and Dominy, 1990). According to Abouelezz (2017) report, adding 1% spirulina to the feed and 0.25% to the drinking water increased fertility in Japanese quails. The addition of up to 12% spirulina to Japanese quail diets positively affected fertility and hatchability (Ross and Dominy, 1990).

5. Conclusion

As a result, spirulina addition to breeder quail diets at 0 %, 0.75 %, 1.5 %, and 2.25 % levels did not affect EP, EW, and EM, but improved FI and FCR. There were no significant findings in regards to eggshell thickness, shell weight, shell ratio, yolk index, and Haugh Unit. None of these factors showed any significant variation or change. Egg yolk L^* value decreased, and a^* and b^* values increased. Decreased feed intake as a result of the supplementation of spirulina to quail feeds in the experiment and improvement of FCR without affecting egg production are the desired changes in performance. The color of egg yolk is a parameter that affects consumer preference. The color of egg yolk can be altered by Spirulina, which is a natural feed additive. Further scientific research is required to establish the appropriate practices for incorporating spirulina into poultry nutrition.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C=Concept, D= design, S= supervision, DCP= data collection and/or processing, DAI= data analysis and/or interpretation, L= literature search, W= writing, CR= critical review, SR= submission and revision, PM= project management, FA= funding acquisition.

Conflict of Interest

The authors declared that there is no conflict of interest.

Ethical Consideration

In the year 2023, a research study was carried out at the Livestock Facilities of Selçuk University's Faculty of Agriculture. The authors declare that the study was carried out by the animal welfare rules specified in Article 9 of the Republic of Turkey Law No. 5996.

Acknowledgments

This study was based on Batuhan Dilber's master's thesis. This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Abdel-Daim MM, Farouk SM, Madkour FF, Azab SS. 2015. Antiinflammatory and immunomodulatory effects of Spirulina platensis in comparison to Dunaliella salina in acetic acidinduced rat experimental colitis. Immunopharmacol Immunotoxicol, 37(2): 126-139.
- Abouelezz F. 2017. Evaluation of spirulina algae (Spirulina platensis) as a feed supplement for japanese quail: nutiritional effects on growth performance, egg production, egg quality, blood metabolites, sperm-egg penetration and fertility. Egyptian Poult Sci J, 37(3): 707-719.
- Anderson DW, Tang CS, Ross E. 1991. The xanthophylls of Spirulina and their effect on egg yolk pigmentation. Poult Sci, 70(1): 115-119.
- Becker EW. 2007. Micro-algae as a source of protein. Biotechnol Adv, 25(2): 207-210.
- Chen J, Wang Y, Benemann JR, Zhang X, Hu H, Qin S. 2016. Microalgal industry in China: challenges and prospects. J

Applied Phycol, 28: 715-725.

- Council NR. 1994. Nutrient requirements of poultry: 1994. National Academies Press, New York, USA, pp: 28.
- Curabay B, Sevim B, Cufadar Y, Ayasan T. 2021. Effects of adding spirulina platensis to laying hen rations on performance, egg quality, and some blood parameters. J Hellenic Vet Med Soc, 72(2): 2945-2952.
- Erensayın C. 2000. Bilimsel-teknik pratik tavukçuluk. Nobel Yayın Dağıtım, İstanbul, Türkiye, ss: 54.
- Göçmen R. 2022. Use of Spirulina Platensis in Japanese quail diets in fattening period and responses of performance, meat quality and immunity. J Hellenic Vet Med Soc, 73(3): 4511- 4516.
- Gupta R, Bhadauriya P, Chauhan VS, Bisen PS. 2008. Impact of UV-B radiation on thylakoid membrane and fatty acid profile of Spirulina platensis. Current Microbiol, 56: 156-161.
- Hajati H, Zaghari M. 2019. Spirulina Platensis in poultry nutrition. Cambridge Scholars Publishing, London, UK, pp: 145.
- Ibrahim AE, Abdel-Daim MM. 2015. Modulating effects of Spirulina platensis against tilmicosin-induced cardiotoxicity in mice. Cell J (Yakhteh): 17(1): 137.
- Jamil AR, Akanda MR, Rahman MM, Hossain MA, Islam MS. 2015. Prebiotic competence of spirulina on the production performance of broiler chickens. J Adv Vet Anim Res, 2(3): 304-309.
- Jeyaprakash K, Chinnaswamy P. 2005. Effect of Spirulina and Liv-52 on cadmium induced toxicity in albino rats. Inter J Experiment Biol, 43(9): 773-781.
- Kharde S, Shirbhate R, Bahiram K, Nipane S. 2012. Effect of Spirulina supplementation on growth performance of broilers. Indian J Vet Res, 21(1): 66-69.
- Kotrbáček V, Skřivan M, Kopecký J, Pěnkava O, Hudečková P, Uhríková I, Doubek J. 2013. Retention of carotenoids in egg yolks of laying hens supplemented with heterotrophic Chlorella. Czech J Anim Sci, 58(5): 193-200.
- Mariey Y, Samak H, Ibrahem M. 2012. Effect of using Spirulina platensis algae as a feed additive for poultry diets: 1 productive and reproductive performances of local laying

hens. Egyptian Poult Sci J, 32(1): 201-215.

- Minitab I. 2000. Minitab: release 13 for Windows. Minitab Incorporated.
- Mišurcová L, Buňka F, Ambrožová JV, Machů L, Samek D, Kráčmar S. 2014. Amino acid composition of algal products and its contribution to RDI. Food Chem, 151: 120-125.
- Nikodémusz E, Páskai P, Tóth L, Kozák J. 2010. Effect of dietary Spirulina supplementation on the reproductive performance of farmed pheasants. Technic Artic Poult Indust, 2010: 1-2.
- Omri B, Amraoui M, Tarek A, Lucarini M, Durazzo A, Cicero N, Santini A, Kamoun M. 2019. Arthrospira platensis (Spirulina) supplementation on laying hens' performance: Eggs physical, chemical, and sensorial qualities. Foods, 8(9): 386.
- Poppi D, McLennan S. 2010. Nutritional research to meet future challenges. Animal Production Sci, 50(6): 329-338.
- Ross E, Dominy W. 1990. The nutritional value of dehydrated, blue-green algae (Spirulina plantensis) for poultry. Poult Sci, 69(5): 794-800.
- Salmeán GG, Castillo LHF, Chamorro-Cevallos G. 2015. Nutritional and toxicological aspects of Spirulina (Arthrospira). Nutri, 32(1): 34-40.
- Selim S, Hussein E, Abou-Elkhair R. 2018. Effect of Spirulina platensis as a feed additive on laying performance, egg quality and hepatoprotective activity of laying hens. European Poult Sci, 82, 1-13.
- Seyidoglu N, Inan S, Aydin C. 2017. A prominent superfood: Spirulina platensis. Superfood and functional food the development of superfoods and their roles as medicine. Intech, 22: 1-27.
- Shanmugapriya B, Babu S, Hariharan T, Sivaneswaran S, Anusha M, Raja PU. 2015. Synergistic effect of Spirulina platensis on performance and gut microbial load of broiler chicks. Indo-Asian J Multidiscip Res, 1(2): 149-155.
- Takashi S. 2003. Effect of administration of Spirulina on egg quality and egg components. Anim Husband, 57(1): 191-195.
- Zahroojian N, Moravej H, Shivazad M. 2013. Effects of dietary marine algae (Spirulina platensis) on egg quality and production performance of laying hens. J Agri Sci Technol, 15(7): 1353-1360.

doi: 10.47115/bsagriculture.1488437

Open Access Journal e-ISSN: 2618 – 6578

Research Article

Volume 7 - Issue (ICABGEH-23): 439-441 / August 2024

INVESTIGATING HEREDITARY DISORDERS OF SIMMENTAL FROZEN BULL SEMEN IMPORTED TO TÜRKİYE DURING 10- YEAR PERIOD

Mustafa ÇAM1*, Şeref İNAL¹

¹Selcuk University, Faculty of Veterinary Medicine, Department of Animal Science, 42003, Konya, Türkiye

Abstract: This study was aimed to determine hereditary defects of frozen Simmental bull semen imported to Turkiye between 2013 and 2022. As the source of this study, the websites of various companies that produce semen, bull catalogues, and databases of bull semen companies have been used. A total of 13 websites including Simmental semen databases and bull catalogues were investigated. The result of the study showed that 438 of 1301 Simmental bull's frozen semen carry at least one genetic defect during 10-year period of import. The most common hereditary defects are Bovine Male subfertility (BMS) for 164 frozen bull semen, Trombopathia (TP) for 155 bull semen and Fleckvieh Haplotype 4 (FH4) for 127 bull semen respectively. Significant effect was observed between different years and the rate of carriers was significantly decreased after 2019. As for origin of bull semen, the bull semen imported from Czech Republic showed significantly the least hereditary defect rate compared with those imported from Austria, Germany and Italy.

Keywords: Simmental bull sperm, Hereditary disorders, Genetic defects

***Corresponding author:** Selcuk University, Faculty of Veterinary Medicine, Department of Animal Science, 42003, Konya, Türkiye **E mail:** mustafa.cam@selcuk.edu.tr (M. ÇAM)

Mustafa ÇAM https://orcid.org/0000-0002-1821-191X **Received:** May 22, 2024 Şeref İNAL https://orcid.org/0000-0003-4746-8930

Accepted: July 22, 2024 **Published:** August 15, 2024

Cite as: Çam M, İnal Ş. 2024. Investigating hereditary disorders of Simmental frozen bull semen imported to Türkiye during 10-year period. BSJ Agri, 7(Special Issue; ICABGEH-23): 439-441.

1. Introduction

The practice of artificial insemination (AI) is so prevalent in Türkiye. Nearly all the cows and heifers in the registered farms get pregnant by AI. A great portion of frozen semen used in Türkiye is imported. The procedures and principles regarding the import of semen, ovum and embryo were issued by General Directorate of Livestock of Ministry of Agriculture and Forestry. There are some important hereditary diseases which should be taken into consideration. However, there is no regulation or prohibition against importing Simmental bull frozen semen carried hereditary disorders. Hereditary defects are important for genetic pollution of livestock herds in the worldwide. Therefore, the raised bulls or the frozen semen should be tested for hereditary diseases and the results of analyses should be specified in pedigrees or information cards. İnal and Çam (2016) reported just the prevalence of hereditary disorders of imported semens in 2015. Since there is lacking of hereditary disorder information for frozen bull semen imported to Türkiye, a comprehensive study is needed to investigate occurrence of Simmental bull semen using the all revealed data via General Directorate of Livestock of Ministry of Agriculture and Forestry. So this study was aimed to determine hereditary defects of frozen Simmental bull semen imported to Türkiye between 2013 and 2022.

2. Materials and Methods

The name and ID of the bull, the importing company and the number of straws are the list of imported bulls issued by General Directorate of Livestock of Ministry of Agriculture and Forestry. In this study, hereditary status of imported Simmental bulls between 2013 and 2023 were examined and a category was created according to years and origins.

Table 1. Websites of various companies used in the study

BSI Agri / Mustafa CAM and Seref İNAL 439

Bulls were combined for general examination, reducing the number to 1302 bulls (13,113,327 straws) (Table 1). Some bulls with beef origin were excluded from the study due to the fact that their pedigrees don't include hereditary disorders.

Table 1 showed websites of various companies that produce semen, bull catalogues and database of bull semen in Simmentals.

Effect of number of carriers on different year and origin was analyzed using Chi-Square analysis in SPSS (ver. 25.0). %5 confidence interval was accepted for the significance level of the tests

3. Results and Discussion

Simmental breeding in Türkiye is getting popular day by day. The primary purpose of Simmental cattle breeding imported to Türkiye is for dairy production. So the majority of the Simmental bulls imported between 2013 and 2022 were for dairy production. Totally, 438 of 1301 bulls carried at least one genetic defect as a result of the research. The type of each hereditary defects of Holstein bulls were summarized in Table 2.

Table 2. Hereditary defects of the Simmental frozen semen imported between 2013 and 2022

¹Hereditary Defect, ²Arachnomelia, ³Dwarfism, ⁴Flekvieh Haplotype 2, 5Flekvieh Haplotype 4, 6Flekvieh Haplotype 5, ⁷Braunvieh Haplotype 2, ⁸Bovine Male Subfertility, ⁹Zinc Deficiency, ¹⁰Trombopathia.

Table 3. Effect of year on occurrence of hereditary defects of the Simmental bull frozen semen

Year	Bull (n)	$CR1$ (n)	SNC ²	TSN ³
2013	92	39.1ab (36)	176599	533881
2014	146	44.5 ^a (65)	414876	1008256
2015	168	45.8 ^a (77)	596778	1252819
2016	233	42.1a (98)	682012	1705780
2017	159	39.0ab (62)	524924	1472013
2018	229	31.4abc (72)	764619	2171580
2019	183	24.6cd(45)	344425	1454485
2020	244	30.3bcd (74)	467988	1343418
2021	214	$26.2cd$ (56)	333572	1258967
2022	174	$21.8d$ (38)	186432	912128

a, b, c, d; different superscripts in the same column show significant differences (P<0,001). 1carriers rate, 2straw number of carriers ³total straw number.

The significant decrease in number of carriers imported in different years became prominent after 2018 (Table 3). Among 1301 Simmental bulls, It was determined that 4 bulls carried Dwarfism, 8 bulls carried Zinc Deficiency. Flekvieh Haplotype 2 (FH2) gene causing deficiency of growth in Simmentals can be another form of dwarfism (Burgstaller et al., 2016). It was determined that 55 Simmental bulls carried FH2. FH4 gene for Simmentals decreases the pregnancy rate like Holstein haplotype genes (Ling et al., 2018). Calves with FH4 genes can't survive because the homozygous FH4 gene causes zygote death. It was determined that 127 Simmental bulls of frozen semen carried FH4. FH5 is a new haplotype for Simmental causing death with symptoms of heart and liver failure in 48 hours after birth (Emmerling, 2015). 9 Simmental bulls carried FH5. 42 Simmental bulls carried Braunvieh Haplotype 2 (BH2) gene which is normally found in Brown Swiss Haplotype. Calves carrying BH2 genes as homozygous are either born dead or die after birth. 37 Simmental bulls were determined to be carrier of Trombopathia characterized by impaired blood coagulation (Ling et al., 2018). 32 bulls were found to have carrier of Bovine Male Subfertility gene causing sterility of bulls (Ling et al., 2018) and not problem for Türkiye (Table 4). It would be a problem in case Türkiye will start to have AI bulls for breeding. Given that 4705983 doses of frozen semen belonging to these bulls were imported and used in Türkiye, which can be concluded that Simmental breeding faces a grave serious genetic pollution.

Table 4. Effect of bull origin on occurrence of hereditary defects of the Simmental frozen semen

Origin	n	CR ¹	PNC ²	TPN ³
AT ⁴	227	$30,8b$ (70)	830146	2073004
CZ ₅	103	17.,5c(18)	237278	877278
DF ₆	861	$38.2a$ (329)	2391557	6558026
IT ⁷	48	35.4^{ab} (17)	73601	441074

 a, b ; different superscripts in the same column show significant differences (P<0.001) ¹carriers rate, ²straw number of carriers ³total straw number ⁴Austria ⁵Czech Republic ⁶Germany ⁷Italy

5. Conclusion

This study highlights the importance of genetic pollution in imported bull semen. Even though there was a limited study investigating hereditary defects of imported semen to Türkiye; this research was the first to examine all hereditary disorders throughout ten-year period. The result of this study encourages authorities to make more precautions during importing frozen bull semen against genetic pollution.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C=Concept, D= design, S= supervision, DCP= data collection and/or processing, DAI= data analysis and/or interpretation, L= literature search, W= writing, CR= critical review, SR= submission and revision, PM= project management, FA= funding acquisition.

Conflict of Interest

The authors declared that there is no conflict of interest.

Ethical Consideration

Ethics committee approval was not required for this study because of there was no study on animals or humans.

Acknowledgments

The study was received financial support from Scientific Research Projects Coordination Unit, Selcuk University (Project Number: 23701205). This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Burgstaller J, Url A, Pausch H, Schwarzenbacher H, Egerbacher M, Wittek T. 2016. Clinical and biochemical signs in Fleckvieh cattle with genetically confirmed Fanconi-Bickel syndrome (cattle homozygous for Fleckvieh haplotype 2). Berl Munch Tierarztl Wochenschr, 129(3-4): 132-137.
- Emmerling R, 2015. Fleckvieh Haplotyp 5 (FH5). URL: https://www.lfl.bayern.de/itz/rind/122227/index.php (accessed date: May 28, 2024).
- İnal Ş, Çam M. 2016. Türkiye'ye 2015 yılında sperması ithal edilen boğalardaki kalıtsal kusurlar. Euroasian J Vet Sci, 32(4): 278-284.
- Ling A, Aggrey S, Rekaya R. 2018. Comparison of quantitative trait nucleotide assisted selection and gene editing for improvement of complex traits. J Anim Sci, 96: 125-126.

doi: 10.47115/bsagriculture.1457467

Open Access Journal

e-ISSN: 2618 – 6578

Review

Volume 7 - Special Issue (ICABGEH-23): 442-448 / August 2024

USE OF SOME INDUSTRIAL PLANTS IN POULTRY

Rabia Vildan ŞAHİN1*, Hasan Alp ŞAHİN²

¹Republic of Türkiye Ministry of Agriculture and Forestry, Black Sea Agricultural Research Institute, 55300, Samsun, Türkiye *²Ondokuz Mayıs University, Research Institute of Hemp, 55200, Samsun, Türkiye*

Abstract: The rapid increase in the human population has also increased the demand for raw materials in many fields. The increasing population has brought along the problem of opening agricultural areas to settlement. For this reason, versatile plants that bring high efficiency from the unit area without harming the existing natural resources and can contribute in many fields such as food, textile, biofuel and animal nutrition arouse excitement among scientists. Industrial plants also provide the diversification of agriculture, enable the simultaneous development of plant and animal production, and make significant contributions to local economies, especially in developing countries. Industrial crops play a crucial role in various aspects of human society, economy, and sustainability. Their importance extends beyond food production and consumption, as they contribute to a wide range of industries and offer several benefits. One of these sectors which affected by industrial plants is poultry nutrition. Industrial crops are utilized to meet the energy, protein, vitamin, fibre, Omega-3, antioxidant, phytochemicals, phosphorous and mineral requirements of poultries. Such as soybeans, sunflower, maize, cottonseed and flax are widely used in the feeding of poultries. They serve as high-energy and protein sources, promoting healthy growth and productivity. These crops are specifically cultivated to provide essential nutrients and meet the dietary needs of poultries. As a conclusion, industrial crops serve as essential ingredients of poultry feeding methods, providing the crucial nutrients, energy, and protein required for the appropriate growth, development, and productivity of poultries. Their inclusion in diets ensures the optimal health and well-being of poultries in various agricultural settings. This review article has examined some important industrial plants as poultry feed.

Keywords: Industrial crops, Poultry nutrition, Poultry

1. Introduction

Industrial crops play a significant role in the provision of nourishment for poultry, serving as both nutritious and cost-effective sources of feed for livestock. These plants are regularly assessed as potential feed content due to their nutritional importance and widespread availability. As a result of the studies, it was stated that white meat consumption will increase by 73% by 2050 (Percival, 2022). It is predicted that increasing world population and production costs will negatively affect the poultry industry (FAO, 2014; Cerisuelo and Calvet, 2020). Poultry production is sustainable with proper nutrition. The more efficiently feeding is done, the more resource demand will decrease and accordingly, costs will decrease (Greenhalgh et al., 2020). The suitability of a plant for poultry consumption is determined by a set of criteria that include the presence of various nutrient substances in different parts of the plant, as well as their digestibility, palatability, metabolizable, and the absence of harmful substances in different plant components (McDonald et al., 1998) and the agronomic properties, concerning cultivation, harvesting difficulties and financial issues. Regarding nutrient substances the essential nutrients in animal feeding items are

(McDonald et al., 1998) proteinic nitrogenous compounds, with special reference to their major amino acid ingredients, nonproteinic nitrogenous compounds, carbohydrates, mainly starch, fats as oils, and cellulosic compounds as fibres, vitamins, and minerals.

Current developments show that the search for alternative feed sources is inevitable in the poultry industry. This paper offers an overview of several crucial industrial plants and their impact on poultry nutrition.

2. Poultry Industry

The World Food and Agriculture Organization, alongside various scholarly investigations, concurs that the human population is projected to reach approximately 9.2 billion individuals by the year 2050, thereby necessitating an augmented global food demand ranging from 35% to 56%. In light of these projections, the task of meeting this heightened demand while maintaining the nutritional adequacy of the food supply becomes increasingly complex and arduous (FAO, 2014; van Dijk et al., 2021). The shorter growth biology of poultry compared to other farm animals, the short period between two generations, and the fact that most of them have an omnivorous diet make them preferable to other animal species. In recent

decades, the demand for poultry meat and, accordingly, per capita consumption rates have increased (OECD, 2022). Compared to other animal foods, the poultry industry has become preferable in terms of the highquality animal protein, amino acids, energy and microelements it contains (Bohrer, 2017). As a consequence of the escalating need for sustenance brought about by the burgeoning human populace, the endeavour of supplying nutrient-dense food with constrained resources poses a formidable challenge for the field of animal husbandry. Developed countries such as the USA, China and Brazil rank first in world chicken meat production (FAO, 2021). Although the amount of chicken meat consumed per capita in the world is 14.7 kg, this rate reaches 64 kg in Israel and 50.1 kg in America (FAO, 2021).

In the past few decades, there have been notable advancements in the field of boiler production, which have undoubtedly revolutionized the industry. These remarkable developments have brought about significant improvements in various aspects of broiler manufacturing, resulting in enhanced efficiency and performance. For instance, between the years 1985 and 2010, there has been a substantial increase in the live weight and feed conversion rates of 35-day-old chickens. Specifically, the live weight has surged from 1.4 kg to an impressive 2.4 kg, while the feed conversion rates have also witnessed a commendable rise from 1.5 to 2.3. These findings highlight the tremendous progress made in optimizing the growth and productivity of poultry, thereby contributing to the overall advancement of the agricultural sector (Siegel, 2014). These increases have been associated with improvements in breeding, nutrition, vaccination, biosecurity guidance, disease prevention and control, and poultry environmental management. Considering these risks, it is important that protein production continues in a sustainable manner.

The objective of this overview is to describe the importance of nutrition in the poultry industry. The topics include the relationship between different industrial crops and poultry diets.

2.1. Sunflower

Sunflower seeds, which are characterized by their distinctively large and flat shape, belong taxonomically to the genus *Helianthus annuus*, which is a member of the family *Asteraceae*, commonly known as the aster, daisy, or sunflower family (Soliman et al., 1996; Canibe et al., 1999; San Juan and Villamide, 2001). The sunflower plant, which has been extensively recorded and studied, holds a prominent position in the hierarchy of essential resources in the realm of nutrition and agriculture. It stands as the second most significant reservoir of feed and protein, offering a valuable source of sustenance for both humans and animals alike. Sunflower seeds contain 20% of protein, whereas protein contents of the oil press cakes and extraction residues range from 30 to 50% (Dorrell and Vick, 1997). Sunflower seeds are considered rich with their energy content of 3,691 to 5,004 kcal

(Daghir et al., 1980; Cheva-Isarakul and Tangtaweewipat, 1990). Furthermore, it also serves as the third most crucial source of oil, a highly sought-after commodity utilized extensively by both humans and animals for various purposes. In addition to its contributions in the realm of nutrition and oil production, the sunflower plant also holds the distinction of being the fourth largest supplier of oilseeds, further solidifying its position as a vital component of the global agricultural industry (Lusas, 1985; Aboul Ela et al., 2000; Garcés et al., 2009). In a research endeavour that took place in 1997, an investigation was carried out, wherein it was ascertained and subsequently documented that the inclusion of sunflower meal within the poultry diets could serve as a feasible alternative to the utilization of soybean meal as a feed ingredient (Soliman, 1997). Research findings have indicated that the inclusion of sunflower in the diets of chickens at a level as high as 200g/kg does not appear to have any detrimental impacts on their growth performance and feed conversion (Valdivie et al., 1982; El-Sherif et al., 1995). In a separate and distinct academic inquiry, it has been conclusively ascertained that the existence and inclusion of substantial quantities of sunflower within the dietary compositions of poultry have been empirically proven to not yield any deleterious repercussions on key factors, such as the efficacy of productivity performance as well as the overall quality criteria pertaining to eggs (Tsuzuki et al., 2003; Casartelli, et al., 2006; Rezaei and Hafezian, 2007). On the other hand, in another study, the inclusion of sunflowers up to 20% (El-Sherif et al., 1997; Tavernari et al., 2008) or at even higher ratios (Rama Rao et al., 2006; Mushtaq et al., 2009) did not have any adverse effects on live body weight or body weight gain. In another study, sunflower was replaced with soybean meal up to 30% in chicken rations and the results were observed. According to the results of the study, it was ascertained that the consumption of feed by the animals and the subsequent increase in their live weight exhibited a significant enhancement of 13.17% and 12.04%, respectively, thereby indicating a marked improvement in their overall growth performance, all of which occurred in the absence of any adverse consequences (Furlan et al., 2001). According to the findings of scientific researchers, it has been determined that the inclusion of sunflower in chicken diet can reach a significant threshold of up to 15% without encountering any detrimental consequences. Additional investigations conducted by these very same scientists further elucidated that not only did the yolk index and Haugh unit score remain unaffected, but they also failed to exhibit any adverse effects. However, it is worth noting that the thickness of the shell exhibited an increase of 15% as a result of the incorporation of sunflower (Mirza and Sial 1992).

2.2. Soybean

Soybean (*Glycine max* L.), which is one of the basic nutrients with its high nutritional values, belongs to the *Leguminosae* family and has a high nutritional content.

Soybean, a grain rich in yellow vegetable protein, has its origins in China and has since become a widely distributed and cost-effective source of vegetable protein, comprising approximately 48 to 50% of its composition, while also boasting low-fat content (Garcia et al., 1997). Feed cost is seen as the main challenge in poultry production (Ravindran, 2013). Due to the prohibition of animal proteins from inclusion in feed in some regions (Van Harn and Veldkamp, 2011), vegetable protein is becoming an essential nutrition of feed for poultry (Ravindran, 2013), and SBM (soybean meal) remains the most crucial and preferred source of high-quality protein for animal feed manufacturing (FAO, 2014). Soy-based feeds, regarded as a means of obtaining animal protein, are experiencing a surge in popularity owing to their superior quality (Banaszkiewicz, 2011).

In their seminal work published in 1995, Fan et al. assert with utmost confidence that soybean meal is an unparalleled and supreme reservoir of protein, thereby solidifying its status as an unrivalled source of this essential macronutrient. Furthermore, they astutely emphasize that soybean meal bestows upon its consumers a staggering quantity of approximately 480 grams per kilogram of dry matter protein that is of unparalleled and exceptional caliber, thus solidifying its position as the epitome of protein quality (Yamka et al., 2003). Soybean has been reported to have the highest lysine digestibility and lowest crude fibre content among all other oilseed meals (Willis, 2003). Soybeans purportedly account for a significant proportion, specifically two-thirds, of the protein content found in animal feed on a global scale. Furthermore, soybeans contribute to over a quarter of the fats and oils utilized in the same context. Notably, three-quarters of the overall international trade in high-protein meals is attributed to soybeans (Peisker, 2001). It has been stated that the level of addition of SBM to diets varies from 25% in chick diets to 30-40% in the diets of old broiler chickens and laying hens (McDonald et al., 2002; Willis, 2003). One research study conducted an observation that soybeans possess a high level of digestibility suitable for avian species, regardless of their breed or age (Newkirk, 2010). Soybean meal has been reported to provide high energy density and feed conversion efficiency in broiler and turkey diets (Erdaw et al., 2016).

2.3. Maize

Alongside the tangible proof of sustenance discovered amidst selected archaeological excavations conducted within the geographical confines of Mexico, a scholarly consensus has emerged, positing that the crop of maize (scientifically classified as *Zea mays* L.) assumed a pivotal role in the early agrarian practices of ancient farmers, commencing some 7,000 to 10,000 years ago (Smith, 2001). Maize which is especially preferred for feeding poultry, is the most important feed grain grown worldwide. (Stamen, 2010). Since maize has consistently possessed the utmost level of dietary energy value when compared to other grains over an extensive span of time,

it has undeniably emerged as the grain of choice in the realm of nutrition for domestic avian creatures in developing nations, including but not limited to Latin America, Africa, and Asia (Larbier and Leclercq, 1994). The primary consequence that arises from the process is commonly denoted as "draff" or "distillers dried grains" (DDG), and it possesses a significant protein content. In addition to DDG, it is also possible to incorporate another resulting substance known as "soluble," which encompasses the minutest remaining particles of both maize and yeast. Due to its elevated protein concentration, presence of trace elements and vitamins, and enhanced accessibility of phosphorus, DDGs have gained substantial popularity as feed ingredients for the purpose of poultry cultivation (Larbier and Leclercq, 1994). Corn grain contains nutrients in various amounts. They are approximately 4% fat, 9% protein, 73% starch, and 14% other components (mostly fibre). Oil is stored mainly in the seed, while starch and protein are mainly found in the endosperm, which makes up the majority of the seed (Tan and Morrison 1979). The digestible energy content of corn grain was indicated to be between 3.75– 4.17 kcal/g in a particular study (Fetuga et al., 1979). The metabolized energy value of corn was stated as 3.6 kcal/g according to a study conducted on poultry (Nelson et al., 1974; Fetuga et al., 1979). Moreover, the recorded value for the digestibility of gross energy in chickens amounted to 86% (Fetuga et al., 1979).

2.4. Flaxseed

Flaxseed (*Linum usitatissimum* L.), a prominent oilseed plant, holds immense significance for its applications in industrial, food, and feed sectors. The appellation "flaxseed" derives from its Latin name, *Linum usitatissimum* L., which aptly translates to "very useful." With an extensive history spanning over five millennia, this plant has been widely consumed as a dietary staple since ancient times. Studies have reported that flaxseed contains approximately 40% lipid, 30% dietary fibre and 20% protein (Daun et al., 2003; Oohma, 2003). The primary objective of incorporating flax in poultry diets is to alter the composition of fatty acids in animal tissue, with the specific aim of generating eggs that are abundant in omega-3 fatty acids, thereby rendering them suitable for consumption by humans. A study on feeding 10%, 20% and 30% flaxseed for 28-day periods was reported for laying hens in 1990. According to this study, omega-3 fatty acids in eggs were reported to increase significantly at each level of flaxseed supplementation (Caston and Leeson, 1990). Feeding poultry with diets that are enriched with omega-3 has been observed to result in an augmentation of the omega-3 levels found in both eggs and meat. Enriched poultry products, as a result of their nutritional composition, possess the capability to function as a feasible and practical alternative for individuals who are seeking to augment their daily intake of omega-3 fatty acids, thereby presenting consumers with an advantageous opportunity to fortify their regular diet with this essential nutrient (Leskanich and Noble, 1997). During the course of the study, the researchers introduced two different concentrations of linseed oil, namely 3% and 6%, into the diet of laying hens for a duration of 25 weeks. Throughout this period, the researchers meticulously observed and analysed the outcomes of this dietary manipulation. Notably, the findings from this investigation revealed that the addition of flaxseed oil had a remarkable and notable impact on the nutritional composition of the eggs, particularly with regards to the enrichment of α-linolenic acid and the overall content of omega-3-polyunsaturated fatty acids. Therefore, the results of this research strongly indicate the significant role of linseed oil in enhancing the nutritional quality of eggs, thereby highlighting its potential benefits in promoting human health. Furthermore, it was observed by the researchers that the level of cholesterol present in the yolk underwent a decline of approximately 5% in comparison to the control group. Additionally, when the hens were provided with the enriched diets, no significant alterations were identified in the quality parameters of the eggs, such as their texture, colour, or flavour. Moreover, the laying efficiency of the hens and the consumption of feed per egg remained consistent and unaffected by the dietary enrichment (Kozlowska et al., 2008).

2.5. Cottonseed

Cottonseed meal, used in certain amounts in poultry feeding, is a by-product obtained from the cotton plant. Cottonseed meal (CSM), which is derived from the process of extracting oil from cotton seeds, serves as a by-product of the oil industry. With protein levels ranging from 30% to 50%, CSM emerges as a notably abundant reservoir of this essential macronutrient. Furthermore, CSM is also found to be replete with a diverse array of amino acids, further enhancing its nutritional value and potential applications (He et al., 2015). One research investigation conducted an observation in which they found that chickens that are 7– 21 days old have the capability to consume a diet that contains up to 20% cottonseed meal, provided that the formulation is supplemented with either 1.5% or 3% Lysine (Azman and Yılmaz, 2005). Mishra et al., 2015 have shown that low gossypol cottonseed meal (0.001% of free gossypol) can be safely incorporated at 5% (1-14 days of age) and 10% (15-35 days) as a partial replacement of soybean meal in diets containing the same level of crude protein and essential amino acids as control diet, without any adverse effect on performance (Mishra et al., 2015). The present study investigated the impact of different levels of cottonseed meal inclusion (0%, 4%, 8%, and 12%) in the diets of 1 and 42-day-old chickens, with regards to their digestibility, performance, and microflora composition. The results of this study demonstrated that the inclusion of 4% and 8% cottonseed meal in the feeds positively influenced the aforementioned parameters. The findings indicated that these particular levels of cottonseed meal inclusion were

beneficial in terms of enhancing the digestibility and performance of the chickens, while also influencing the composition of the microflora in a favourable manner (Sun et al., 2013).

2.6. Hempseed

The Hemp plant, scientifically classified as *Cannabis sativa* L., which is a member of the *Cannabinaceae* family, has been widely recognized and acknowledged for its significant contributions throughout history in the realms of food production, medicinal applications, and the provision of various types of fibre (Russo and Reggiani, 2015). Hemp, a widely distributed and extensively cultivated plant, carries substantial industrial importance owing to its diverse applications as a source of whole seeds, dehulled seeds, seed flour, oil, and fibre (Callaway, 2004). In countries such as Iran, Pakistan and Türkiye, roasted salted hemp seeds are still sold as snacks in herbalists and are also used as bird feed (e.g., canary, pigeons) during mating period to increase stamina of male birds (Karimi and Hayatghaibi, 2006). Hemp seeds, which serve as an excellent protein source for poultry, have been documented to possess a noteworthy abundance of edestin and albumin, two types of storage proteins that exhibit exceptional quality and are characterized by their effortless digestibility and possession of indispensable amino acids (Callaway, 2004). Hemp seeds contain 20-25% high quality protein, 30-36% fat, 30-40% fiber and 6-7% moisture. In addition, while the energy content in the seed is 2200 kJ/100 g, the energy content in the seed meal is reported as 1700 kJ/100 g (Callaway, 2004). Khan et al. (2010) fed 160 day-olds broiler chicks with basal diet and basal diet+hemp seed (5, 10 and 20%). The findings of the study demonstrated that the inclusion of hemp seeds at a concentration of 20% in the diets of broiler chickens resulted in a significant improvement in body weight gain. In addition to this, the presence of hemp seeds in the diet led to a reduction in feed intake, indicating that the birds were able to achieve higher body weights with lower feed consumption. Moreover, the study also found a positive correlation between feed conversion and the inclusion of hemp seeds, suggesting that the birds were able to convert the feed more efficiently when hemp seeds were included in their diets. This is particularly noteworthy as hemp seeds are known to possess a rich nutritional profile, making them a potential alternative source of protein in organic poultry feeds. By incorporating hemp seeds into the diets of broiler chickens, it is possible to enhance their growth and performance while also reducing the reliance on traditional protein sources. Therefore, the inclusion of hemp seeds in organic poultry feeds holds promising potential in improving the overall sustainability and efficiency of broiler production systems. Thus, Eriksson and Wall (2012), added hemp seed cake in organic broiler diets (10% in starter and 20% in grower diets). It has been observed in many studies that it has a beneficial impact on egg weight (Gakhar et al., 2012; Halle and

Schöne, 2013; Skrivan et al., 2019) and eggshell thickness (Konca et al., 2019) in laying hens. Including hemp seeds in the diet instead of rapeseed oil has been suggested as a potential means of enhancing bone health in laying hens and cockerels, as per previous assertions (Skrivan et al., 2019; Skrivan et al., 2020).

3. Conclusion

Industrial plants are one of the major sectors that affect and are linked with many different sectors like food, textile, biofuel, and animal nutrition. They contain nutrients that are important for the poultry industry, which has an important place in world food production, in terms of the development of birds with high protein, vitamins, minerals, amino acids, and omega-3 content and the production of the products expected from them, and they constitute a source of raw materials for the poultry industry. The nutritional content, taste and digestibility of meat and eggs obtained from poultry are directly related to the environment in which the animals live and their diet, and has been addressed by scientists for many years as a subject that arouses interest and is worth researching. For years, the effects of many different plants on poultry nutrition have been investigated for this purpose. Although the plants discussed in this article have been studied for many years and are mostly used in the poultry nutrition industry, scientific studies on the subject are continuing and the effects of different industrial plant varieties on poultry nutrition can be evaluated.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C=Concept, D= design, S= supervision, L= literature search, W= writing, CR= critical review, SR= submission and revision.

Conflict of Interest

The authors declared that there is no conflict of interest.

Acknowledgments

This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Aboul Ela SS, Attia AI, El-Tawil M, Soliman MM. 2000. Sunflower meal as a substitute for soybean meal in broiler rations. Conf. of Social and Agricultural Development of Sinai, El-Arish, Egypt, pp: 31.
- Azman MA, Yılmaz M. 2005. The growth performance of broiler chicks fed with diets containing cottonseed meal supplemented with lysine. Revue de Medecine Vet, 156: 104- 106.
- Banaszkiewicz T. 2011. Nutritional value of soybean meal, in: EL-SHEMY, H.A. (Ed) Soybean and nutrition, InTech, Rijeka, Croatia, pp: 1-20.
- Bohrer BM. 2017. Review: Nutrition density and nutritional value of meat products and non-meat foods high in protein. Trends Food Sci Technol, 65: 103-112.
- Callaway JC. 2004. Hempseed as a nutritional resource: An overview. Euphytica, 140: 65-72.
- Canibe N, Pedrosa MM, Robredo LM, Knudsen KEB. 1999. Chemical composition, digestibility and protein quality of 12 sunflower (Helianthus annus L) cultivars. J Sci Food Agri, 79(13): 1755-1782.
- Casartelli EM, Filardi RS, Junqueira OM, Laurentiz AC Assuena V, Duarte KF. 2006. Sunflower meal in commercial layer diets formulated on total and digestible amino acids basis. Braz J Poult Sci, 8(3):167-171. http://dx.doi.org/10.1590/s1516- 635x2006000300005
- Caston G, Leeson S. 1990. Dietary flax and egg consumption. Poultry Sci, 69: 1617-1620.
- Cerisuelo A, Calvet S. 2020. Feeding in monogastric animals: A key element to reduce its environmental impact. ITEA Info Tecnica Econ Agraria, 116(5): 483-506. https://doi.org/10.12706/itea.2020.039
- Cheva-Isarakul B, Tangtaweewipat S. 1990. Effect of different levels of sunflower seed in broiler rations. Poultry Sci, 70: 2284-2294.
- Daghir NJ, Raz MA, Uwayjan M. 1980. Studies the utilization of full fat sunflower seed in broiler rations. Poultry Sci, 59: 2273-2278.
- Daun J, Barthet V, Chornick T, Duguid S. 2003. Structure, composition, and variety development of flaxseed. In: Thompson L, Cunanne S. edition. Flaxseed in Human Nutrition. 2nd edition Champaign, Illinois, US, pp: 1-40.
- Dorrell DG, Vick BA. 1997. Properties and processing of oilseed sunflower. In: Schneiter AA. edition, Sunflower technology and production American Society of Agronomy, Madison, Wisconsin, US, pp: 709-744.
- El-Sherif K, Gippert T, Gerendai D. 1995. Effect of different levels of expeller sunflower seed meal in broiler diets. Anim Breed Feed, 44(5): 427-435.
- El-Sherif K, Gippert T, Gerendai D. 1997. Effect of feeding sunflower meal supplemented with energy and amino acids on laying hen performance. Anim Breed Feed, 46(1): 87-94.
- Erdaw MM, Bhuiyan MM, Iji PA. 2016. Enhancing the nutritional value of soybeans for poultry through supplementation with new-generation feed enzymes. World's Poultry Sci J, 72(2): 307-322.
- Eriksson M, Wall H. 2012. Hemp seed cake in organic broiler diets. Animal Feed Sci Technol, 171: 205-213.
- Fan MZ, Sauer WC, De Lange CFM. 1995. Amino acid digestibility in soybean meal, extruded soybean and full-fat canola for early-weaned pigs. Animal Feed Sci Technol, 52(3- 4): 189-203.
- FAO. 2014. Protein sources for the animal feed industry. proceedings: FAO Expert consultation and workshop, April 29 -May 3, Bangkok, Thailand, pp: 54.
- FAO. 2021. Livestock Primary. Meat Chicken Production Quantity. URL: https://www.fao.org/faostat/en/#data/QL. (accessed date: June 12, 2023).
- Fetuga BL, Babatunde GM, Oyenuga VA. 1979. Comparison of the energy values of some feed ingredients for the chick, rat and pig. J Agri Sci, 17: 3-11.
- Furlan AC, Mantovani C, Murakami AE, Moreira I, Claudio S, Nunes ME. 2001. Utilização do farelo de girasol na alimentação de frangos de corte. R Bras Zootec, 30(1): 158- 164. http://dx.doi.org/10.1590/S1516- 35982001000100023.
- Gakhar N, Goldber E, Jing M Gibson R, House J. 2012. Effect of feding hemp seed and hemp seed oil on laying hens performance and egg yolk fatty acid content: evidence of their dafey and efficacy for lyinh hen diets. Poultry Sci, 91(3): 701-711.
- Garcés R, Martínez-Force E, Salas JJ, Venegas-Calerón M. 2009. Current advances in sunflower oil and its applications. Lipid Technol, 21(4): 79-82.
- Garcia MC, Torre M, Marina ML, Laborda F, Rodriquez AR. 1997. Composition and characterization of soyabean and related products. Critical Rev Food Sci Nutrit, 37(4): 361-391.
- Greenhalgh S, Chrystal PV, Selle PH, Liu Y. 2020. Reduced-crude Protein Diets in Chicken-meat Production: Justification for an Imperative. World's Poultry Sc J, 76(3): 537-548. https://doi.org/10.1080/00439339.2020.1789024.
- Halle I, Schöne F. 2013. Influence of rapeseed cake, linseed cake and hemp seed cake on laying performance of hens and fatty acid composition of egg yolk. J Für Verbrauch Und Lebensm, 8: 185-193.
- He Z, Zhang H, Olk DC. 2015. Chemical composition of defatted cottonseed and soy meal products. PloS One, 10: e0129933.
- Karimi I, Hayatghaibi H. 2006. Effects of Cannabis sativa L. seed (hempseed) on serum lipid and protein profiles of rat. Pakistan J Nutrit, 5(6): 585-588.
- Khan RU, Durrani FR, Chand N, Anwar H. 2010. Influence of feed supplementation with Cannabis sativa on quality of broilers carcass. Pakistan Vet J, 30(1): 34-38.
- Konca Y, Yuksel T, Yalcin H, Beyzi SB, Kaliber M. 2019. Effects of heat-treated hempseed supplementation on performance, egg quality, sensory evaluation and antioxidant activity of laying hens. British Poultry Sci, 60(1): 39-46. https:/doi.org/10.1080/00071668.2018.1547360.
- Kozlowska J, Munoz GA, Kolodziejczyk PP. 2008. Food and feed applications for flaxseed components. 2008 International Conference on Flax and other Bast Plants, July 21-23, Saskatoon, Canada, pp: 299-307.
- Larbier M, Leclercq B. 1994. Nutrition and Feeding of Poultry. J. Wiseman, Trans. & Ed. Nottingham University Press, Loughborough, UK, pp: 305.
- Leskanich CO, Noble R. 1997. Manipulation of the n-3 polyunsaturated fatty acid composition of avian eggs and meat. World's Poultry Sci J, 53: 155-183.
- Lusas EW. 1985. Sunflower seed protein in new protein foods. Academic Press Public Inc., New York. US, pp: 393. http://dx.doi.org/10.1016/B978-0-12-054805-7.50019-1
- Mc Donald P, Greenhalgh JFD, Edwards R, Morgan CA. 2002. Animal nutrition. 6th edit. Pearson Education Limited, Edinburgh, England, pp: 525-378.
- McDonald P, Edwards RA, Greenhalgh JFD, Morgan CA. 1998. Animal Nutrition 5th edition (3rd reprint). Scientific and Technical Publishers, Longman, US, pp: 72.
- Mirza MA, Sial MA. 1992. Sunflower meal as a major vegetable protein source in layers ration. Arch Anim Nutr, 42(3-4): 273- 277.
- Mishra A, Ray S, Sarkar SK, Haaldar S. 2015. Cotton seed meal as a partial replacement for soybean meal in Cobb 400 broiler rations. Indian J Anim Nutrit, 32: 69-74.
- Mushtaq T, Sarwar M, Ahmad G, Mirza MA, Ahmad T, Noreen U, Mushtaq MMH, Kamran Z. 2009. Influence of sunflower mealbased diets supplemented with exogenous enzyme and digestibile lysine on performance, digestibility and carcass response of broiler chickens. Anim Feed Sci Technol, 149(3- 4): 275-286.
- Nelson TS, May MA, Miles Jr. RD. 1974. Digestible and Metabolisable energy content of feed ingredient for rats. J Anim Sci, 38: 555-558.
- Newkirk R. 2010. Soybean. Feed industry guide, 1st edition Canadian International Grains Institute, Ottowa, Canada, pp: 48.
- OECD. 2022. Organisation for Economic Co-operation and Development: Agricultural Outlook 2022-2031. URL: https://policycommons.net/artifacts/2552558/oecd-faoagriculturaloutlook-2022-2031/3675435 (accessed date: January 10, 2023).
- Oohma B. 2003. Processing of flaxseed fiber, oil, protein, and lignan. In: Thompson, L., Cunnane, S. Editores. Flaxseed in Human Nutrition. 2nd. Edn. Champaing, Illinois, US, pp: 363- 386.
- Peisker M. 2001. Manufacturing of soy protein concentrate for animal nutrition. Cahiers Opt Mediter, 54: 103-107.
- Percival R. 2022. The meat paradox: eating, empathy, and the future of meat. London: Little, Brown Book Group Limited, London, UK, pp: 54.
- Rama Rao SV, Raju MVLN, Panda AK, Redely MR. 2006. Sunflower seed meal as a substitute for soybean meal in commercial broiler chicken diets. Br Poult Sci, 47(5): 592- 598. http://dx.doi.org/10.1080/00071660600963511
- Ravindran V. 2013. The role of poultry in human nutrition, in: FAO. Poultry Devel Rev, pp: 1-8. URL: URL: http://www.fao.org/3/a-i3531e.pdf (accessed date: January 10, 2023).
- Rezaei M, Hafezian H. 2007. Use of different levels of high fiber sunflower meal in commercial Leghorn type layer diets. Int J Poult Sci, 6(6): 431-433. http://dx.doi.org/10.3923/ijps.2007.431.433
- Russo R, Reggiani R. 2015. Evaluation of protein concentration, amino acid profile and antinutritional compounds in hempseed meal from dioecious and monoecious varieties. American J Plant Sci, 6: 14-22.
- San Juan LD, Villamide MJ. 2001. Nutritional evaluation of sunflower products for poultry as affected by the oil extraction process. Poult Sci, 80(4): 431-437.
- Siegel PB. 2014. Evolution of the modern broiler and feed efficiency. Annu Rev Anim Biosci, 2: 375-338.
- Skrivan M, Englmaierova M, Taubner T, Skrivanova E. 2020. Effects of dietary hempseed and flaxseed on growth performance, meet fatty acid compositions, liver tocopherol concentration and bone strenght of cockerels. Animals, 10(3): 458.
- Skrivan M, Englmaierova M, Vit T, Skrivanova E. 2019. Hempseed increases gamma-tocopherol in egg yolks and the breaking strength of tibias in laying hens. Plos One, 14(5): e0217509.
- Smith BD. 2001. Documenting plant domestication: the consilience of biological and archaeological approaches. Proc Natl Acad Sci, 98: 1324-1326.
- Soliman AA. 1996. Evaluation of the productivity and performance of broiler breeder hens fed practical or vegetable diets containing high levels of barley and sunflower

meal with multi-enzymes supplement during the pre-laying and laying periods. PhD Thesis, Alexandria University, Faculty of Agriculture, Alexandria, Egypt, pp: 142.

- Stamen B. 2010. Feed preference index on cereal grains for poultry. URL: http://hdl.handle.net/1811/45500 (accessed date: February 23, 2023).
- Sun H, Tang JW, Yao XH, Wu YF, Wang X, Feng J. 2013. Effects of dietary inclusion of fermented cottonseed meal on growth, cecal microbial population, small intestinal morphology, and digestive enzyme activity of broilers. Tropical Anim Health Prod, 45: 987-993.
- Tan SL, Morrison WR. 1979. Lipids in the germ, endosperm and pericarp of the developing maize kernel. J Am Oil Chem Soc, (56): 759-764.
- Tavernari FC, Albino LFT, Morata RL, Dutra Júnior WM, Rostagno HS, Viana MTS. 2008. Inclusion of sunflower meal, with or without enzyme supplementation, in broiler diets. Braz J Poult Sci, 10(4): 233-238. http://dx.doi.org/10.1590/s1516-635x2008000400007.
- Tsuzuki ET, de Garcia ERM, Galli JR, Murakami AE, Sakamoto MI. 2003. Utilization of sunflower seed in laying hen rations. Braz J Poult Sci, 5(3): 179-182.
- Valdivie M, Sardinas O, Garcia JA. 1982. The utilization of 20% sunflower seed meal in broiler diets. Cuban J Agric Sci, 16: 167-171.
- Van Dijk, M, Morley T, Rau ML, Saghai Y. 2021. A meta-analysis of projected global food demand and population at risk of hunger for the period 2010-2050. Nat Food, 2: 494-501.
- Van Harn J, Veldkamp T. 2011. Pig MBM in broiler feed did not meet expectations. All about Feed, 2: 24-26.
- Willis S. 2003. The use of soybean meal and full-fat soybean meal by the animal feed industry. In the 12th Australian soybean conference. Soy Australia, Bundaberg, Australia, pp: 47.
- Yamka RM, Jamikorn U, True AD, Harmon DL. 2003. Evaluation of soyabean meal as a protein source in canine foods. Anim Feed Sci Technol, 109(1-4): 121-132.

doi: 10.47115/bsagriculture.1467433

Open Access Journal e-ISSN: 2618 – 6578

Review

Volume 7 - Special Issue (ICABGEH-23): 449-457/ August 2024

GENERAL STATUS OF RUMINANT LIVESTOCK IN THE TURKISH REPUBLIC OF NORTHERN CYPRUS

Çağrı Halil KOCADAL1*, Mustafa BOĞA²

¹Niğde Ömer Halisdemir University, Faculty of Agriculture, Department of Animal Science, 512440, Niğde, Türkiye ²Niğde Ömer Halisdemir University, Bor Vocational School, 512440, Niğde, Türkiye

Abstract: Among other agricultural activities, animal production has the most important strategic importance for the country's economy. In the Turkish Republic of Northern Cyprus (TRNC), animal production constitutes 45% of the total agricultural production. In response to the increasing population, the need for animal food continues to increase day by day. State hatcheries and many private companies run animal production activities along with small-scale family enterprises. Almost all animal feed raw materials are imported. On the other hand, most of the food of animal origins that is marketed (e.g. meat, milk, egg etc.) is produced in TRNC and the rest is imported when it is necessary. Halloumi, which covers almost all animal-based imported goods, is imported into many countries especially Turkey and Kuwait and contributes to the country's economy. This study concentrates on the general state of animal production in TRNC and aims to present the current state of ruminant animals (66 thousand head of cattle, 197 thousand head of sheep and 78 thousand head of goat) in the country's livestock farming, their contributions to the economy and the type of their production.

Keywords: TRNC, Agriculture, Animal husbandry, Import-export, Halloumi

1. Introduction

Animal production, which is a branch of agriculture, has a strategic importance in terms of its contribution to other fields of agriculture and the country's economy (Görgülü, 2009). Grain crises (difficulties in finding grain and sudden increases in price) due to recent epidemics and conflicts between countries with strategic importance have disrupted production in the food sector. For this reason, food prices have doubled in countries that are foreign-dependent in food production, and it has become difficult for human beings to reach the nutrients they need (Gürlük and Turan, 2008, Doğruyol A. 2021.). Due to these factors, it has once again proven the importance of countries to carry out their own agricultural activities. Animal production outputs constitute the largest share of agricultural production in the economy. Animal products such as meat and meat products, milk and dairy products, leather and fleece, which are needed by the society, are met through bovine and ovine breeding (Paksoy and Özçelik, 2008; Semerci and Çelik, 2016).

Thanks to the Mediterranean climate of the island of Cyprus, it has a suitable structure for plant and animal production (Anonymous, 2022a). It is known that food resources should be used more efficiently with the increase in consumption due to the increasing population in the Northern Cyprus (Anonymous, 2022b).

This article is designed as a review article supported by an according to the data published by the Ministry of Agriculture and Natural Resources of Northern Cyprus in 2021, TRNC statistical institution and literature review data in various scientific databases.

2. Agricultural Sector in North Cyprus 2.1. Geographical and Economic Structure

The island of Cyprus is the third largest island in the Mediterranean after Sicily and Sardinia. The island, with a total surface area of 9251 km2, consists of three natural geographical structures: Beşparmak Mountains, Troodos Mountains and Mesarya plain. The climate that is effective in Cyprus is the Mediterranean climate. The characteristic feature of the climate is hot and dry summers, mild and less rainy winters (Anonymous, 2022c, Anonymous. 2022g).

Due to the internal turmoil in the 1960s, the island of Cyprus was divided into two regions, Turks in the north of the island and Greeks in the south, in 1974. The Turkish Republic of Northern Cyprus was established on November 15, 1983, and its surface area is 3242 km² (Koday, 1995; Anonymous, 2022c). Its population in 2011 was 286,257. The majority of the population is also located in non-rural areas, mainly in the cities of Nicosia and Famagusta (Anonymous, 2022d). In addition,

800,000 people come from Turkey and other countries for touristic purposes, especially in the summer months. Therefore, with the increasing population, the North Cyprus' need for animal and vegetable products will increase. According to the statistical data of 2021, approximately 8.38% of the Gross National Product (GNP) originates from the agriculture sector and approximately 45% of this is the livestock sector (Anonymous, 2022d).

2.2. General Condition of Livestock

The agricultural sector in Northern Cyprus consists of plant and animal production (bovine and ovine animals and fisheries). According to the 2021 data of the Ministry of Agriculture and Natural Resources (TDKB), animal production in Northern Cyprus is a large, medium, and small-scale business model.

There are 890 cattle breeding, 3550 sheep and 2510 goat breeding enterprises in the northern part of Cyprus. There are very few large-scale companies in the poultry sector. In 2021, 8,124,764 dozen eggs were produced throughout the country, and 15 thousand tons of chicken

meat was produced. Chicken meat has the highest share in meat production in 2021. Poultry production and the processing and distribution of the products obtained from these animals within the country and abroad are also carried out by private companies. Chicken feet, which are slaughterhouse waste in poultry, are exported to the People's Republic of China. Chicken and piece chicken products (such as hips, breasts) are exported to Middle East countries in the country. In addition, 1 turkey production enterprise operates. In recent years, advertising activities to promote turkey meat have made turkey meat in demand in the country and paved the way for production by private enterprises.

Although North Cyprus is an island country, the contribution of fisheries to national income is limited and below the expected level due to the low fish stocks on the coasts outside the Karpaz region and the inability to engage in open sea fishing. According to 2018 data, fish production is reported to be approximately 650 tons (Anonymous, 2022e) (Table 1, Figure 1).

Table 1. The condition of the agricultural sector in the TRNC between the years 2016-2021 (Anonymous, 2022e, Anonymous. 2022f.)

Years	2016	2017	2018	2019	2020	2021
Agricultural s-Sector	5,6	4,5	6,2	5,5	6,0	8,3
Herbal Production	2,7	1,9	2,3	2,6	2,8	4,2
Animal Production	2,5	2,1	3,5	2,5	2,7	3,8
Fisheries	0,3	0,5	0.4	0.4	0.4	0,3

Figure 1. The situation of the agricultural sector in the TRNC between the years 2016-2021 (Anonymous, 2022e).

2.3. Cattle and Sheep Breeding

In today's livestock, there are products of animal origin mostly obtained from cattle breeding. Therefore, cattle

are a factory producing edible and usable animal products such as meat, milk and leather. For this reason, cattle breeding are widely carried out in almost all parts

of the world, especially in the developed and intensive regions of plant production (Canbolat, 2015; Ergün and Bayram, 2021). According to the statistical data of the Ministry of Agriculture and Natural Resources of Northern Cyprus in 2021, there is a total of 65701 cattle. Most of the milk needed by the TRNC is met by the Holstein breed, which is widely used in production. Depending on the rearing conditions, 6500 kg of milk per cow and a total of 159894 tons (2021) of milk are produced annually. 86.87% of milk production in the northern part of Cyprus is provided by dairy cows. Males of the Holstein breed, which are good in terms of milk yield, are used for fattening. In recent years, artificial insemination of Simmental breeds is carried out with Holstein breeds fourth lactation and above cows, which are combined productive, in order to increase cattle fattening performance and to obtain better quality carcasses among producers. Thus, the fattening performance is tried to be increased (Anonymous, 2022a).

Ovine breeding, which is one of the other branches of the livestock sector, has a wide area of use by obtaining products such as meat, milk, wool, mohair and leather (Semerci and Çelik, 2016). According to the statistical data of the Ministry of Agriculture and Natural Resources in 2021, the number of small-scale enterprises engaged in sheep and goat breeding is 4500, the number of medium-sized enterprises is 1480 and the number of large enterprises is 80. There are a total of 196,890 sheep and 77,852 goats in these enterprises (Anonymous, 2022i). There are two state breeding farms engaged in small cattle breeding. Genetic and breeding studies are carried out in these enterprises and they contribute to the country's livestock. (Anonymous, 2022e)

While a decrease was observed in the number of sheep and goats in 2017-2021, the Ministry of Agriculture and Natural Resources and the European Union regarding the registration of Halloumi/Halloumi as a Protected Product of Origin and Protected Geographical Indications (PDO-Protected Designation of Origin) in the European Union (EU). Grant supports have increased in order to develop small cattle breeding and increase production (Anonymous, 2022a). In Table 2, when 2017 and 2021 years are compared, it is given that while the number of cattle producers decreased, the number of cattle increased.

Chios, Awassi, Asaf and their hybrid breeds are used in sheep breeding in North Cyprus. In goat breeding, production is made with Hair, Aleppo (Damascus), Saanen, Alpine, and hybrids. 79% of the 24 tons of ovine milk produced in 2021 is sheep milk and 21% is goat milk (Anonymous, 2022h).

2.4. Production and Marketing of Bovine and Ovine Products

Population growth in the Turkish Republic of Northern Cyprus is ensured every year by the number of tourists and the number of students coming to the island for higher education (Anonymous, 2022d). Thus, it is estimated that the demand for food in Northern Cyprus will increase exponentially. Northern Cyprus is under an embargo (North Cyprus is a country recognized only by Turkey in the world, and is an independent state founded on November 15, 1983. Since its establishment, it has been under embargoes in economic, social and cultural aspects by the whole world, especially the Greek Cypriot part, foreign expansion has been prevented, the fact that the domestic industry is very weak, and that it is dependent on foreign sources for many products emphasizes the indispensable importance of agricultural production (Doğan, 2009).

Milk production is a universal agricultural production, people milk their dairy animals in almost all countries around the world and almost one billion people live on dairy farms (Anonymous, 2022i). For this reason, while milk production plays a major role for the world food sector, it is also very important for the development of rural areas. When the share of products constituting agricultural exports in terms of value for 2015 is analyzed, it is seen that dairy products have a significant share with 58%. Until the 1960s, families in the villages of Cyprus extensively raised sheep and goats, and from the milk of these animals, they obtained dairy products, mainly halloumi, nor, talar cheese and yoghurt. Industrial type production in the dairy sector started in 1957 in the TRNC, for the first time, with the collection of milk from the producers, and the production of halloumi and perforated cheese. TRNC started to export halloumi for the first time to England, and with the increase in milk production, exports increased with the introduction of private factories. Halloumi was the second leading export product until 1994 (Anonymous, 2022k; Anonymous, 2022b). Until the Abat Decisions, a significant amount of halloumi was exported to other European countries and Arab countries, especially England. As a result of the embargoes applied to the Turkish Cypriots after the Abat Decisions, exports to England and other European countries were prevented. This isolation still continues (Aran, 2006).

Halloumi, which is widely produced in the country among dairy products and is unique to the island; it is exported to many countries, especially Turkey, Kuwait, Saudi Arabia and the United Arab Emirates. The contribution of the state to this export increases the production of halloumi day by day. In 2021, the European Union Commission decided to register the halloumi product as the name of origin (PDO) and included Cyprus as the base country. It has been decided that the milk to be used for the production of traditional halloumi within the scope of PDO should be prepared with 51% ovine (sheep-goat) and 49% bovine (cattle) milk (Anonymous, 2022a). TRNC Ministry of Economy and Energy Chamber of Commerce 2021 halloumi export data are in Table 3 below (Anonymous, 2022c).

Species	2017		2021		
	Number of Producers	Number of Animals	Number of Producers	Number of Animals	
Cattle	998	63.270	890	65,701	
Sheep	3,619	203.419	3,550	196,890	
Goat	2,560	83.467	2,510	77,852	

Table 2. Number of animals and producers between 2017-2021 (Anonymous, 2022i)

Table 3. 2021 Halloumi export data (Anonymous, 2022c)

Countries	Quantitiy	Turkish Lira	Dollars
USA	82,577,00	£4,186,192,77	\$509,043,50
Australia	10,004,00	£244,763,78	\$26,774,71
Bahrain	96.042.00	£4,208,040,91	\$445,043,10
United Arab Emirates	1,088,192,00	£41,854,831,20	\$4,628,825,92
Palestine	3,000,00	£109,691,21	\$13,300,00
Iraq	60,202,00	£ 2,683,887,69	\$281,805,09
Qatar	83,103,00	£4,416,724,88	\$507,479,81
Kuwait	2,733,342,00	£89,807,766,75	\$10,270,363,59
North Korea	27,700,00	£964,577,13	\$107,180,00
Saudi Arabia	1,278,500,00	£47,437,442,80	\$5,348,001,00
Thailand	210,00	E 7,515,58	\$903,00
Turkey	3,366,094,00	£107,656,159,78	\$12,427,984,50
0 _{man}	24,214,00	$\pm 1,066,664,12$	\$131,054,40
Jordan	562,550,00	£19,585,067,82	\$2,332,815,00
Total	9,415,730,00	£324,229,326,42	\$37,030,573,62

In addition, bottled pasteurized milk production was started in 1963. The KOOP Milk Factory, which is still operating today, was established in 1969 (Anonymous, December 2022k; Anonymous, 2022b). (Anonymous, 2022i) (Table 4). Icebox: It is the event that small-scale producers carry and pour their milk, which they milk due to the lack of electricity in animal shelters, into the freezers in a hygienic environment in their homes by means of cans or jugs.

Foods of animal origin play an important role in nutrition in terms of the proteins, minerals and vitamins they contain. Especially meat and meat products contain essential amino acids needed in human nutrition in ideal proportions. However, due to the availability of iron in the body and being a very good source of vitamin B12, it shows that it is a food that plant-based foods cannot replace meat and meat products (Derinöz et al., 2021). In a healthy diet, 40-50% of the body's protein needs are composed of foods of animal origin. The access of the individuals who make up the society to animal protein sources is affected by the agricultural policies of the countries and the implementations of these policies.

In terms of the belief, tradition and custom of the society we live in, cattle, sheep and goat species are preferred in the production of meat and meat products (Gürer, 2021). According to the Northern Cyprus' 2021 statistical data, chicken has the highest share in meat production, while cattle take the second place. These values are given in Table 5.

According to the data of 2021, approximately 49% of the red meat production in our country is beef, followed by mutton with a share of 40% and goat meat with a share of 11%. In Table 6, it is observed that the amount of beef has not changed in the last three years, and the amount of sheep and goat meat has decreased. When the data of 2021 is examined, it is noteworthy that the production of sheep and beef is similar (Anonymous, 2022a). Red meat production amounts for the years 2019-2021 are given in Table 6.

Leather, which has a lot of usefulness with its perfection and many aspects, and has been used by human beings since ancient times, is a product that is rarely mentioned among animal products, but has very valuable features. The biggest share in the rapid development in the leather industry belongs to sheep skins, especially sheep skins. The skin is an organic material and it is an armour that has the task of protecting the animal in general, with the hair or wool cover on it, the connective tissue fibres called collagen that make it up (Yakalı, 1979). A total of 271,679 leathers were produced in the North Cyprus in 2021. Approximately 66% of the total leather production is sheepskin and 26% is goatskin (Anonymous, 2022a). The amount of leather produced according to the species is given in Table 7.

Table 4. Milk production by years (Anonymous, 2022i)

Table 5. Meat production by species in 2021 (Anonymous, 2022i)

Table 6. Red meat production (Tonnes) for the last three years (Anonymous, 2022i)

Sheared dirty wool from sheep is called "fleece". In a broad sense, fleece means all the hairs that are removed from the animals as shirts during shearing and that can be twisted and made into yarn (Kaymakçı and Sönmez, 1992). Wool is the sheared, washed and cleaned form of the fleece shirt covering the sheep (Tüfekçi and Olfaz, 2015). In fabric production, the washed and cleaned form of fleece is used (Kaymakçı and Sönmez, 1992).

Although animals have not been exported as living material in the Northern Cyprus, leather and fleece are exported to Turkey at certain rates. Turkish Republic of Northern Cyprus Trade Office, leather and wool export values for the years 2017-2021 are given in Table 8 (Anonymous, 2022h).

Bovine and ovine manures are a valuable nutrient provider for the soil and a regulator of soil conditions. When properly matured and applied to the soil, it is a better and more economical nutrient provider than commercial fertilizers. Animal manure is a fertilizer rich in organic matter content. Therefore, it increases the water holding capacity of the soil and provides the soil with nutrients such as nitrogen (N), phosphorus (P), potassium (K) and sulfur (S) (Kacar and Katkat, 2009; Konca and Uzun, 2012). Due to the exorbitant increases in chemical fertilizer prices in recent years, the value of ruminant fertilizers, whose value has not been understood for years and has been launched as if they have no economic value, has started to gain importance today (Teoman and Yaşar, 2016). In parallel with the increase in the number of animals in the North Cyprus in 2017 and 2021, there was an increase in the amount of manure. In Table 9, the amount of bovine and ovine manure produced in the North Cyprus is given in tons (Anonymous, 2022i).

Table 8. Leather and wool exports in TRNC for the years 2017-2021 (Anonymous, 2022h)

Table 9. Amount of bovine and ovine manure produced in North Cyprus (Anonymous, 2022i)

2.5. Feed Raw Materials and Production

Animals need nutrients such as water, carbohydrates, proteins, fats, vitamins and minerals to survive and produce various products. Animals meet their needs with this nutrient through feed and water consumption. Feeds can be of vegetable and animal origin, as well as their processing residues or by-products (Gürsoy and Macit, 2020).

Feeds are generally examined in three classes. These include roughage, dense (concentrated) feeds and feed additives. Forages are plant-based feeds with high cellulose content, low protein and energy content used as animal feed in fresh, dried or silage form. Dense (concentrated) feeds are those that are rich in energy and protein content and have a high digestibility of nutrients. Feed additives, on the other hand, are substances used to increase feed efficiency, improve the quantity and quality of animal products, reduce costs, and protect animal health (Kutlu and Çelik, 2016). It is stated that more than 60% of livestock operating expenses in our country, as in the world, are feed expenses (Görgülü, 2009).

Producers mostly meet their roughage needs from the lands they cultivate or by renting lands belonging to the state. The negativities seen due to climate change affect the roughage needs of the producers. The state allows the import of roughage in order to eliminate the disruptions in production depending on the climatic conditions, and provides certain amounts of support to animal producers in order to eliminate the problems experienced and for sustainable production. In addition, the grant feed contribution is made by the state. Quality roughage is produced in some of the cultivated lands, while grain group (barley, wheat and oat) is produced in the remaining parts. According to the data of the North Cyprus Ministry of Agriculture and Natural Resources, the amount of roughage produced by region for 2021 is given in Table 10 (Anonymous, 2022i).

The producers separate and preserve some of the grain produced as seeds for the next year. On the other hand, producers who make both plant production and animal production together use the excess grain to meet the concentrated feed needs of the animals. On the other hand, producers who only produce plants sell the remaining part of the seed product to the TRNC Soil Products Authority (TÜK) according to the price determined by the TRNC Ministry of Agriculture and Natural Resources. The distribution of cereals produced according to the statistical data of 2021 is shown in Table 11 (Anonymous, 2022i).

In the TRNC, the most barley is produced, followed by wheat. Producers who make their own feed, registered with the Ministry of Agriculture and Natural Resources Livestock Department of the TRNC, benefit from the discounted barley purchase given by the ministry and reduce the cost of barley ration purchased below the current price.

In our country, almost all of the raw materials used in feed production are imported, as in other areas. According to the statistical data of the TRNC Ministry of Agriculture and Natural Resources, the raw materials imported in 2021 and their quantities are given in Table 12 (Anonymous, 2021a, b).

There are 6 feed factories in the Northern Cyprus. While only 1 of these factories is state supported, the others are privately owned. However, there is one feed additive manufacturing enterprise in North Cyprus.

Table 10. Amount of roughage produced by regions in 2021 (Anonymous, 2022i)

3. Discussion

The island of Cyprus has two separate political entities: the Turkish Republic of Northern Cyprus (TRNC) and the Greek Cypriot Administration of Southern Cyprus (GCASC). The livestock sector is significant agriculturally and economically in both regions. However, economic and political disparities have led to divergent development paths in these two regions' livestock sectors.

According to the 2021 statistics of TRNC, approximately 45% of the Gross Domestic Product (GDP) is constituted by the livestock sector. Large livestock production, which holds a significant share in this sector, includes

approximately 65,000 heads of cattle with a production of 158,646 tons of cow milk. 97% of the produced milk in the country is cow milk. Sheep and goat farming is carried out through enterprises of various scales. Of the produced 4,619 tons of small ruminant milk, approximately 52% is sheep milk, and the remaining 48% is goat milk. As for red meat production in TRNC, beef holds a 49% share, while sheep and goat meat holds 40% and 11% shares, respectively (Anonymous, 2022i). The Greek Cypriot Administration of Southern Cyprus (GCASC) accounts for approximately 45% of its GDP in the livestock sector, according to statistical data from 2017. The same data indicates the existence of 66,000

cattle with a production of 215,370 tons of milk. Production includes 321,000 sheep and lambs, 257,000 goats and kids, and 350,000 pigs. 52% of the produced small ruminant milk is sheep milk, and 48% is goat milk. In red meat production, pork constitutes a significant share of 80%, followed by beef with 9%, sheep meat with 6%, and goat meat with 5% (Anonymous, 2022i and Papachristoforou and Markou, 2006).

GCASC's membership in the EU enables it to benefit from EU support and funding in the agricultural sector, facilitating the adoption of modern technology and management practices (Papachristoforou and Markou, 2006). On the other hand, TRNC must rely on its internal resources, as it faces international trade embargoes, which restrict access to external markets. While GCASC has alternative sources of red meat production like pork, enhancing product diversity, TRNC's livestock sector needs more access to international markets due to trade embargoes.

Statistics emphasize the importance of the livestock sector in both regions. However, the more diverse economic structure of GCASC provides broader opportunities for external trade in the livestock sector, making it more competitive internationally. In conclusion, although the livestock sectors in both TRNC and GCASC share similar characteristics, different economic and political factors influence their development. While the livestock sector in TRNC contributes significantly to the national economy, its potential is hindered by international embargoes. To enhance its contributions further, the sector requires better internal support and lifting international embargoes.

4. Conclusion

The Turkish Republic of Northern Cyprus is a country rich in agriculture due to its climatic characteristics. Especially animal production, which is one of the branches of agriculture, is increasing day by day with the use of resources and the formation of international market network and population growth. State-supported export promotion activities continue for the development and expansion of animal production. Despite the development of the cattle and sheep livestock sector, it is necessary to continue the activities in the development of the sector due to the problems and limitations such as the standardization of animal products, the lack of an effective marketing system, and the lack of information in small family businesses.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C= Concept, D= design, S= supervision, L= literature search, W= writing, CR= critical review, SR= submission and revision.

Conflict of Interest

The authors declared that there is no conflict of interest.

Acknowledgments

This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Anonymous. 2021a. Tarımsal istatistik yıllığı. https://istatistik.gov.ct.tr/TEMEL-
- %C4%B0STAT%C4%B0ST%C4%B0KLER/N%C3%9CFUS-SAYIMLARI (accessed date: December 12, 2022).
- Anonymous. 2021b. SÜTEK, Kuzey Kıbrıs Türk Cumhuriyeti, https://www.ktsutek.com/kurumsal.html (accessed date: December 14, 2022).
- Anonymous. 2022a. Avrupa bilgi merkezi. https://www.abbilgi.eu/tr/avrupakomisyonu,xalloymi/halloumi/hellimi-%E2%80%98mense-
- ismi-ko.html (accessed date: December 12, 2022). Anonymous. 2022b. International dairy federation. The economic importance of dairying. Brussels: IDF.https://shop.fil-idf.org/products/economic-importanceof-dairying?_pos=1&_sid=a38541166&_ss=r, (accessed date: December 14, 2022).
- Anonymous. 2022c. Kıbrıs Türk süt endüstrisi kurumu, https://www.ktsutek.com/kurumsal/hakkimizda.html (accessed date: December 12, 2022).
- Anonymous. 2022d. KKTC ekonomi ve enerji bakanlığı, https://ekonomi.gov.ct.tr/Portals/16/ekonomik%20indicato r.pdf. (accessed date: December 12, 2022).
- Anonymous. 2022e. KKTC istatistik kurumu, gayri safi milli hasıla, https://stat.gov.ct.tr/TEMEL- %C4%B0STAT%C4%B0ST%C4%B0KLER/GAYR%C4%B0-
- SAF%C4%B0-M%C4%B0LL%C4%B0-
- HASILA/GAYR%C4%B0-SAF%C4%B0-
- M%C4%B0LL%C4%B0-HASILA-2016-2021 (accessed date: December 12, 2022).
- Anonymous. 2022f. KKTC istatistik kurumu, https://istatistik.gov.ct.tr/TEMEL-
	- %C4%B0STAT%C4%B0ST%C4%B0KLER/N%C3%9CFUS-

SAYIMLARI (accessed date: December 12, 2022).

Anonymous. 2022g. KKTC talim terbiye, http://talimterbiye.mebnet.net/Kitaplar/2015YeniKitaplar/ Orta-Lise/Tarih8Tumu-k.pdf (accessed date: December 12, 2022).

- Anonymous. 2022h. KOOP süt fabrikası, https://www.koopsut.com/hikayemiz-vemisyonumuz/index.html (accessed date: December 12,
- 2022).
- Anonymous. 2022i. CYPRUS Agricultural 2022, https://www.data.gov.cy/sites/default/files/E_AGRICULTUR E-EN-060319.xls (accessed date: December 12, 2022).
- Anonymous. 2022k. Kuzey Kıbrıs Türk Cumhuriyeti ticaret dairesi,

https://ticaret.gov.ct.tr/RAPORLAR/%C4%B0STAT%C4%B0 ST%C4%B0K-%C5%9EUBES%C4%B0-RAPORLARI/01-

D%C4%B1%C5%9F-Ticaret-%C4%B0thalat-ve-

%C4%B0hracat-Raporlar%C4%B1 (accessed date: December 12, 2022).

- Aran L. 2006. Avrupa topluluğu adalet divanı kararları ışığında kıbrıs sorunu. TEPAV-The Economic Policy Research Foundation of, İstanbul, Türkiye, pp: 25.
- Canbolat Ö. 2015. Süt sığırlarının beslenmesi ve rasyon hazırlama yöntemleri. Medyay Kitapevi, İstanbul, Türkiye, ss: 1-458.
- Derinöz AN, Çufaoğlu G, Ayaz ND. 2021.Et türü tayininde kullanılan yöntemler. Akademik Et Süt Kurum Derg, 1: 8-18.
- Doğan A. 2009. Ekonomik gelişme sürecine tarımın katkısı: Türkiye örneği. Sosyal Ekon Araş Derg, 9(17): 365-392.
- Doğruyol A. 2021. Tarım devrimi ve zaman ölçümü. Sakarya İktisat Derg, 10(1): 103-114.
- Ergün OF, Bayram B. 2021. Türkiye'de hayvancılık sektöründe yaşanan değişimler. Bahri Dağdaş Hayvan Araşa Derg, 10(2): 158-175.
- Görgülü M. 2009. Büyük ve küçükbaş hayvan besleme. Çukurova Üniversitesi Ziraat Fakültesi Zootekni Bölümü, Ders Kitabı, Genel Yayın, ss: 244.
- Gürer B. 2021.Türkiye'de nüfusun yeterli ve dengeli beslenmesi açısından hayvansal gıda arz ve talebinin değerlendirilmesi. Gıda, 46(6): 1450-1466.
- Gürlük S, Turan Ö. 2008. Dünya gıda krizi: nedenleri ve etkileri.

Uludağ Üniv Ziraat Fak Derg, 22(1): 63-74.

- Gürsoy E, Macit M. 2020. Hasat zamanının kaba yemin kimyasal kompozisyonu ve kalitesi üzerine etkisi. Euroasia J Math Engin Nat Med Sci, 7(9): 168-177.
- Kacar B, Katkat A. 2009. Plant nutrition. Nobel publication Science and Biology Publication Series, Ankara, Türkiye, ss: 424-533.
- Kaymakçı M, Sönmez R. 1992. Koyun yetiştiriciliği. Hasat Yayıncılık Hayvancılık, Serisi:3, İstanbul, Türkiye, ss: 405.
- Koday Z. 1995. Kuzey Kıbrıs Türk Cumhuriyeti Devleti'nin cografi özellikleri. Atatürk Üniv Türkiyat Araşı Enstit Derg, 2: 17-45.
- Konca Y, Uzun O. 2012. Effect of animal waste on soil and environment. 4th Congress of Soil Scientists of Azerbaijan, 23-25 Mayıs Azerbaijan, Bakü, ss: 25-28.
- Kutlu HR, Çelik L. 2016. Yemler bilgisi ve yem teknolojisi kitabı. Çukurova Üniversitesi Ziraat Fakültesi Zootekni Bölümü, Ders Kitabı, Genel Yayın, ss: 25.
- Paksoy M, Özçelik A. 2008. Kahramanmaraş ilinde süt üretimine yönelik keçi yetiştiriciliğine yer veren tarım işletmelerinin ekonomik analizi. Tarım Bilim Derg, 14(4): 420-427.
- Papachristoforou C, Markou M. 2006. Overview of the economic and social importance of the livestock sector in Cyprus with particular reference to sheep and goats. Small Rumi Res, 62(3): 193-199.
- Semerci A, Çelik AD. 2016. Türkiye'de küçükbaş hayvan yetiştiriciliğinin genel durumu. Mustafa Kemal Üniv Zir Fak Derg, 21: 182-196.
- Teoman Ö, Yaşar P. 2016. Türkiye'de 2003 sonrası buğday ve gübre fiyatları ilişkisinin piyasa yapıları bakımından değerlendirilmesi. Afyon Kocatepe Üniv İktisadi İdari Bilim Fak Derg, 18(1): 55-72.
- Tüfekçi H, Olfaz M. 2015. Yapağının alternatif kullanım alanları. Bahri Dağdaş Hayvan Araşa Derg, 1(1-2): 18-28.
- Yakalı T. 1979. Deri verimleri ve önemli sorunları. Hayvansal Üretim 11(1): 38-40 1979.

doi: 10.47115/bsagriculture.1457465

Open Access Journal e-ISSN: 2618 – 6578

Review

Volume 7 - Issue (ICABGEH-23): 458-463/ August 2024

USABILITY OF HEMP PLANT IN POULTRY NUTRITION

Hasan Alp ŞAHİN1*, Rabia Vildan ŞAHİN²

¹Ondokuz Mayıs University, Hemp Research Institute, 55200, Samsun, Türkiye

²Republic of Türkiye Ministry of Agriculture and Forestry, Black Sea Agricultural Research Institute, 55300, Samsun, Türkiye

Abstract: Hemp (*Cannabis sativa* L.) is an annual plant with 2n=20 chromosomes, one-year, C3 group, cultivated for its long and strong fibers and seeds. It is a plant that should be cultivated in a controlled manner due to the presence of THC (tetrahydrocannabinol) in its natural structure. Producers who avoid control and monitoring have turned to alternative products instead of hemp farming. Countries that cultivate cannabis in large areas; France, China, and Canada. Some countries are updating their cannabis-related laws and making efforts to increase hemp acreage. It is known that hemp farming has been practiced in Anatolia since 1500 BC. Various products such as hemp seeds, hemp pulp, hemp oil have been used in various studies in poultry feeding studies, while no negative results have been encountered, but various positive results have been suggested and their use is recommended. It is of great importance to investigate the effects of cannabidiols, which have strong antimicrobial, antioxidant and immunostimulant effects, as they are expected to support the immune system in poultry, especially broilers. Some of the studies in which hemp products are used in poultry nutrition are as follows; Addition of hemp meal to laying hen diets has been reported to make no significant difference in egg production, feed consumption, feed utilization, body weight gain or egg quality, but results in lower palmitic acid concentrations and higher Linoleic acid (LA) and alpha-linolenic acid (ALA) concentrations, which are healthier for human consumption. Hemp seeds and hemp oil used in chicken rations caused an increase in the omega-3 polyunsaturated fatty acid content and color intensity of egg yolks, but no adverse effects were observed on the sensory profiles of cooked eggs. Addition of cannabis meal to fast growing broiler diets did not affect performance or mortality, and no effect on the number of *Clostridium perfringens* in the cecum was reported. Hemp seeds have a remarkable effect on the growth of broiler chicks and can help reduce feed expenditures for broiler rearing. In this paper, the use and usability of the hemp plant, whose production areas are expanding rapidly, in poultry will be examined.

Keywords: Poultry nutrition, Hemp, Antioxidant, Immune system, Feeding performance

1. Introduction

For the assessment and analysis of different feed sources to ensure the long-term viability and sustainability of the livestock and animal products sectors, it is crucial to conduct a thorough investigation into the presence and accessibility of lesser-known and underutilized feed sources (Salami et al., 2019).

Cultivated hemp, belonging to the annual plant species within the *Cannabinaceae* family, is characterized by its unique features. This plant, which has been the subject of foreign fertilization, possesses a chromosome count of 2n=20, showcasing its genetic makeup. As described by Small and Cronquist in 1976, hemp exhibits both dioic and monoic types, presenting a diverse reproductive system. In the dioic variety, male and female flowers are segregated on distinct plants, highlighting a clear distinction in their reproductive structures. Conversely, in monoic types, the male and female flowers coexist on the same plant, albeit in different sections of the inflorescence, demonstrating a fascinating botanical phenomenon. Renowned for its multifaceted utility, hemp serves as a pivotal industrial plant that caters to

various sectors, utilizing almost all parts of its anatomy. Its significance within the industrial realm stems from its widespread applications across different industries, positioning it as a versatile botanical resource. Within the realm of industrial plants, hemp is classified into various types, including *Cannabis sativa, Cannabis indica,* and *Cannabis ruderalis*, each possessing distinct characteristics and properties (Schultes et al., 1975). Over the years, the fibers, seeds, leaves, and flowers derived from the stems of hemp have emerged as the most valuable and extensively utilized components, underscoring their economic importance. The chemical composition of hemp is subject to multiple factors such as varietal differences, processing techniques during pressing, and seed processing methods, as expounded by House et al. (2010), shedding light on the intricate chemical profile of this industrially significant plant. The aim of this study is to evaluate the impact of hemp

plant utilization in poultry farming on the nutrition, health, and production performance of chickens. Our hypothesis suggests that hemp-based feed may enhance the growth rate of chickens, increase egg production, and strengthen the immune system of poultry. Additionally,

we anticipate that the utilization of hemp as poultry feed could offer environmental and economic sustainability advantages. This study aims to provide a comprehensive assessment to understand the potential role of hemp in the poultry industry.

2. Hemp Parts Used in Poultry Feeding

2.1. Hemp Seeds

Most varieties of hemp are cultivated with the primary goal of seed production, with the most valuable output being hemp oil (Callaway, 2004). The hemp seed typically contains 30-35% seed oil, which is extracted through the cold pressing method, ensuring the preservation of the oil's physical and chemical properties (Hazekamp et al., 2010). The remaining solid byproduct after oil extraction is known as hemp pulp. Depending on the processing technology used, a portion of the oil (0.4-10%) may still be retained in the hemp pulp (House et al., 2010). Hemp seeds are predominantly utilized in animal nutrition, primarily for ornamental birds that are not intended for food production, accounting for approximately 95% of their usage. The remaining 5% is allocated to the food sector. The chemical contents of hemp seeds are given in Table 1.

Table 1. Nutritional value of hemp seeds in terms of vitamins and minerals (Göre and Kurt, 2021)

Hemp seeds are known for their rich protein content and as a source of omega-3 fatty acids. Recent findings suggest that incorporating omega-3 and omega-6 fatty acids into poultry feed can have beneficial effects on immune responses, egg quality, meat nutrition, and growth. Performance within hemp seeds, essential fatty acids such as linoleic acid (omega-6) and α-linolenic acid (omega-3) can be found (Smith, 2000). The chemical contents of hemp fatty acids are given in Table 2.

Gakhar et al. (2012) demonstrated that adding 20% hemp seed to egg-chicken rations increased egg weight and omega-3 fatty acid content without impacting egg yield or specific gravity.

Khan et al. (2009) observed that including up to 20% dried and crushed hemp seeds in broiler diets led to increased breast and thigh weights. This positive outcome was attributed to the high-quality protein and lipid combination, along with the beneficial properties like the absence of trypsin inhibitors and the antioxidant activity of cannabidiol. Another study by Khan et al. (2010) showed that feeding broiler chickens with 20% hemp seed improved feed conversion rate, live weight gain, slaughter age, and mortality rate. Kalmendal (2008) explored the use of hemp seed cake in organic broilers and found that its nutritional value was comparable to rapeseed pulp. Mahmoudi et al. (2012) discovered that including hemp seed in broiler diets up to 7.5% did not negatively impact performance and even reduced serum cholesterol levels. Their subsequent study revealed that incorporating hemp seed meal in the diets of fastgrowing organic broilers had no adverse effects on production performance or mortality rates. These studies collectively suggest that hemp seeds can be safely incorporated into poultry diets up to 30% without compromising their intended purpose.

2.2 Hemp Flowers and Cannabinoid Extract from Flower

Cannabis sativa L. has been extensively studied, leading to the identification and isolation of over 500 compounds, as reported in various studies (Pellati et al., 2018; Al Ubeed et al., 2022). Among these compounds, a total of 566 have been pinpointed and grouped into more than 18 categories of secondary metabolites, showcasing the remarkable chemical diversity present in this plant species.

Source	Component	Flour oil	Cold pressed oil	Seed oil
1	Palmitic, C16:0	7.35	7.15	7.03
2	Stearic, C18:0	2.62	2.73	2.78
3	Oleik, $C18:1 \omega - 9c$	12.79	12.75	12.74
$\overline{4}$	Linoleic, $C18:2 \omega$ -6c	56.42	56.08	56.16
5	Arachidic, C20:0	0.74	0.89	0.81
6	γ -Linolenik, C18:3 ω -6	3.00	3.03	2.94
7	cis-II-Eicosinoic C20:l	0.45	0.26	0.37
8	α -Linolenik, c18:3 ω-3	14.55	14.89	15.02
9	cis-ll, 14-Eikosinoic C20:2	0.82	1.03	0.99
10	Behenik, C22:0	0.29	0.20	0.27

Table 2. The chemical contents of hemp fatty acids (Siano et al., 2019)

Notably, the flowers and leaves of *Cannabis sativa* L. are particularly rich in these bioactive compounds, as highlighted in recent research (Kopustinskiene et al., 2022). Specifically, the identified compounds include 125 cannabinoids, 198 non-cannabinoids, and 120 terpenes. Recent investigations in 2021 have further expanded this list to reveal the presence of 2 alkaloids, 34 flavonoids, 42 phenols, and 3 sterols within the plant matrix (Al Ubeed et al., 2022). This comprehensive characterization underscores the complexity and pharmacological potential of *Cannabis sativa* L., offering a wealth of chemical entities for further exploration and exploitation in various fields of science and medicine. Moreover, the distinct aromatic profile of Female Hemp plants can be attributed to terpenes such as pinene, limonene, terpineol, and borneol, which are synthesised within the plant tissues (McPartland and Russo, 2001). These terpenes not only contribute to the sensory experience associated with Hemp consumption but also play a crucial ecological role by repelling insects and inhibiting the growth of competing vegetation. A key defensive mechanism employed by *Cannabis sativa* L. involves the production of resin by glandular trichomes, which possess antibiotic and antifungal properties, serving as a natural shield against pests and pathogens. Within these trichomes, a myriad of secondary metabolites, including phytocannabinoids and terpenoids, interact synergistically to safeguard the plant from external threats while also imparting its distinctive odor profile (Andre et al., 2016).

2.2.1. Some cannabinoids and their mechanism of action

The most well-known and studied cannabinoids in the Hemp plant are THC (tetrahydrocannabinol) and CBD (cannabidiol)The therapeutic potential of CBD, a prominent cannabinoid in *Cannabis sativa* L., has garnered significant attention due to its reputed health benefits across diverse medical conditions. CBD has been linked to an array of therapeutic effects, including relaxation, anxiety relief, and pain management, with documented efficacy in alleviating symptoms associated with cancer chemotherapy-induced vomiting, glaucoma, anorexia, neuropathic pain, spasticity, and other ailments. Approved for medical use in numerous countries, particularly in North America, CBD-based products are widely available in various formulations such as oils, capsules, topicals, and dietary supplements, catering to different consumer preferences. While CBD shares a molecular weight with THC, the psychoactive component of Hemp, it stands out for its analgesic, antiepileptic, antibacterial, anti-inflammatory, and antidepressant properties, making it a valuable therapeutic agent for both acute and chronic pain management (Beşir et al., 2022).

The multifaceted immunomodulatory and antioxidant actions of CBD are underpinned by intricate molecular mechanisms that involve interactions with various immune cells and inflammatory pathways. CBD exerts

modulatory effects on activated B cells and interferonβ/signal transducer and activator of transcription pathways, influencing cellular inflammation cascades such as the nuclear factor kappa-light chain enhancer pathway. By targeting adenosine receptor A2A and modulating adenosine reuptake, CBD can finely tune the activities of immune cells like neutrophils, macrophages, and T cells within the inflammatory milieu. Additionally, CBD impacts cytokines including interferon-c, interferonγ (Lee ve Erdelyi, 2016), tumor necrosis factor α (Magen et al., 2009; Rajesh et al., 2010; Khaksar ve Bigdeli, 2017; Wang et al., 2017), interleukin (IL)-1β (IL-1β) (Pazos et al., 2013; Wang et al., 2017), IL-6 (Lee ve Erdelyi, 2016) as well as adhesion molecules like ICAM1 and VCAM1, thereby regulating inflammatory responses at multiple levels (Rajesh et al., 2010). Furthermore, CBD exhibits anti-apoptotic effects by reducing caspase 9 activation (Castillo et al., 2010) and inhibiting caspase 3 (Rajesh et al., 2010; Da Silva et al., 2014; Santos et al., 2015), key components involved in apoptotic pathways. Importantly, CBD boosts the production of the antiinflammatory cytokine IL-10, further enhancing its immunomodulatory properties and underscoring its therapeutic potential in managing inflammatory disorders (Kozela et al., 2017).

The endocannabinoid system is a complex network within the human body that includes cannabinoid receptors, endocannabinoids, and enzymes responsible for the synthesis and breakdown of these compounds. It has been extensively studied in recent research as a potential target for treating various pathological conditions. This intricate system plays a crucial role in numerous physiological processes such as regulating energy balance, stimulating appetite, controlling blood pressure, providing analgesia, managing nausea and vomiting, facilitating memory and learning, and modulating immune system responses, as highlighted in studies by Balpınar and Aytaç (2021). Anandamide and 2-arachidonylglycerol are well-known endocannabinoids that act as signaling molecules within the brain, allowing for the adjustment of input signals from parent cells. Moreover, the endocannabinoid system has its own hemp molecules, which operate in a retrograde manner from dendrites to axon terminals, aiding in the fine-tuning of neuronal communication.

The endocannabinoid system has garnered significant attention due to its potential therapeutic implications in various health conditions. Notably, it interacts with a nuclear receptor called PPAR-γ, which is involved in regulating metabolic and inflammatory processes, as pointed out by O'Sullivan and Kendall (2010). Cannabinoids found in hemp, other than THC, interact with the endocannabinoid system receptors at a lower rate and with less affinity, as reported in studies by Pertwee (2008) and Elsohly et al. (2017). Among these compounds, cannabidiol (CBD) has emerged as a prominent phytocannabinoid with non-psychoactive properties, as emphasized by Aydoğan et al. (2020). CBD

has been recognized for its anxiolytic and antipsychotic effects, as highlighted in research by Zuardi et al. (1995) and Crippa et al. (2009). Additionally, CBD is known to counteract the psychoactive effects induced by THC, showcasing its potential therapeutic value in mitigating adverse reactions associated with THC consumption, as mentioned by Mechoulam et al. (2006).

3. Hemp Products and Poultry

Hemp seeds are reported to be abundant in various vitamins, minerals, essential amino acids, and essential fatty acids, making them a popular choice in poultry nutrition, particularly as bird feed (Silversides and LeFrancois, 2005; Goldberg et al., 2012; Eriksson and Wall, 2012; Konca et al. 2014). The utilization of hempderived CBD in research is quite limited, with a focus on clinical rather than nutritional studies. Clinical trials involving CBD are predominantly conducted pre-human use, with rats being the primary subjects, alongside occasional use of dogs and monkeys. Poultry nutrition commonly incorporates products derived from hemp seeds and seed derivatives, although it is essential to note that hemp seeds do not inherently contain CBD. Konieczka et al. (2023) explored the impact of CBD and CBD+Se (selenium) additives in chicken diets, revealing a protective effect against DNA damage in the intestinal mucosa, leading to improved interaction and enhanced fattening performance. Langer and Scilcher (1999) conducted a study on muscle pain in rats, administering 100mg/kg of 99% pure CBD intramuscularly after a series of intense workouts, noting a mild effect on muscle. However, CBD displayed anti-inflammatory properties on skeletal muscle, suggesting its potential use as an analgesic and anti-inflammatory agent. Moreover, the removal of cannabidiol (CBD) from the prohibited substances list allows athletes access to this nonpsychoactive hemp component, aligning with the World Anti-Doping Rules International Standard Prohibited List 2023.

Gustafsson and Jacobsson (2019) explored CBD's impact on embryo development by administering in-ovo CBD injections to chicken chicks on days 1, 4, and 7, leading to a 20% decrease in chick incubation efficiency due to developmental interference. Recent years have seen intensified research on in-ovo injection in poultry feed, focusing on amino acid development, growth performance, immune response, and digestive organ enhancement (Bhanja and Mandal, 2005). Studies have suggested that in-ovo feeding can bolster the population of bacteria resistant to enteric pathogens (Ferket 2006, Smirnov et al. 2006) and enhance intestinal resistance to *Salmonella enterica* in young chickens through in-ovo CpG-ODN administration. Furthermore, the in-ovo injection of oligosaccharides has been shown to impact digestive and storage capacities, with reports indicating that chick weight and growth are accelerated with the injection of amino acids, trace elements, fatty acids, and vitamins (Bakyaraj et al., 2012).

4. Conclusion

In conclusion, various hemp products like seeds, pulp, and oil have been studied in poultry feeding, yielding positive outcomes without encountering adverse effects, thereby recommending their utilization. The antimicrobial, antioxidant, and immunostimulant properties of Cannabidiols are anticipated to benefit the immune system of poultry, particularly broiler chickens, underscoring the importance of further investigations into their effects. Despite extensive research on hemp seeds and their derivatives in poultry nutrition, studies on the impact of CBD and other cannabinoids remain scarce. Only a single study on CBD nutrition and embryonic development was identified in our literature review. While one study facilitated embryo development (Gustafsson and Jacobsson, 2019), another utilized hemp extract (Konieczka et al., 2023).

The cost of 99.56% pure CBD was recorded at \$6000 per kilogram in the year 2016, but has since plummeted to the range of \$450 to \$375 per kilogram by 2024. This substantial reduction in price, amounting to approximately 15 times less in less than a decade, can be attributed to various factors such as the issuance of permits for hemp cultivation dedicated to CBD production within our nation, the steady expansion of hemp production areas, and the continuous advancements in technology related to CBD extraction processes. These developments have collectively contributed to the potential consideration of CBD as a viable additive for enhancing animal nutrition. The ongoing expansion of CBD production sites coupled with the continuous technological enhancements in extraction methods are driving the rapid decline in CBD prices, making it increasingly feasible for utilization across various industries provided it demonstrates positive outcomes.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

C=Concept, D= design, S= supervision, L= literature search, W= writing, CR= critical review, SR= submission and revision.

Conflict of Interest

The authors declared that there is no conflict of interest.

Acknowledgments

This article was presented as an oral presentation at the

VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Al Ubeed HMS, Bhuyan DJ, Alsherbiny MA, Basu A, Vuong QV. 2022. A comprehensive review on the techniques for extraction of bioactive compounds from medicinal cannabis. Molecules. 27: 604. https://doi.org/10.3390/molecules27030604.
- Andre CM, Hausman JF, Guerriero G. 2016. Cannabis sativa: the plant of the thousand and one molecules. Front Plant Sci, 7: 19. https://doi.org/10.3389/fpls.2016. 00019.
- Aydog an M, Terzı̇ YE, Gizlenci Ş, Acar M, Esen A, Meral H. 2020. Türkiye'de kenevir yetiştiriciliğinin ekonomik olarak yapılabilirliği: Samsun ili Vezirköprü ilçesi örneği, Anadolu J Agri Sci, 35: 35-50.
- Bakyaraj S, Bhanja SK, Majumdar S, Dash B. 2012. Modulation of post-hatch growth and immunity through in ovo supplemented nutrients in broiler chickens. J Sci Food Agri, 92(2): 313-320.
- Balpınar Ö, Aytaç S. 2021. Tibbi kenevir ve sağlik: Farmakolojik bir derleme. Ankara Ecz Fak Derg, 45(3): 631-651. https://doi.org/10.33483/jfpau.859372.
- Beşir A, Yazıcı Bektaş N, Mortaş M, Yazıcı F. 2022. Kenevirde THC ve CBD faktörlerinin değerlendirilmesi. Osmaniye Korkut Ata Üniv Fen Bil Enst Derg, 5(2): 1092-1104.
- Bhanja SK, Mandal AB. 2005. Effect of in ovo injection of critical amino acids on pre- and post-hatch growth, immunocompetence and development of digestive organs in broiler chickens. Asian-Aust J Anim Sci, 18(4): 524-531.
- Callaway JC. 2004. Hempseed as a nutritional resource: an overview. Euphytica, 140: 65-72. https://doi.org/10.1007/s10681-004-4811-6.
- Castillo A, Tolón MR, Fernández-Ruiz J, Romero J, MartinezOrgado J. 2010. The neuroprotective effect of cannabidiol in an in vitro model of newborn hypoxicischemic brain damage in mice is mediated by CB2 and adenosine receptors. Neurobiol Dis, 37(2): 434-440. https://doi.org/10.1016/j.nbd.2009.10.023.
- Crippa JA, Zuardi A W, Martín-Santos R, Bhattacharyya S, Atakan Z, McGuire P, Fusar-Poli P. 2009. Cannabis and anxiety: a critical review of the evidence. Hum Psychopharmacol, 24(7): 515-523.
- Da Silva VK, De Freitas BS, Da Silva Dornelles A, Nery LR, Falavigna L, Ferreira RDP. 2014. Cannabidiol normalizes caspase 3, synaptophysin, and mitochondrial fission protein DNM1L expression levels in rats with brain iron overload: implications for neuroprotection. Mol Neurobiol, 49(1): 222- 233. https://doi.org/10.1007/s12035-013-8514-7
- ElSohly MA, Radwan MM, Gul W, Chandra S, Galal A. 2017. Phytochemistry of Cannabis sativa L. Prog Chem Org Nat Prod, 103: 1-36.
- Eriksson M, Wall H. 2012. Hemp seed cake in organic broiler diets. Anim Feed Sci Technol, 171: 205-213.
- Ferket PR. 2006. Incubation and in ovo Nutrition affects Neonatal Development, 33rd Annual Carolina Poultry Nutrition Conference. September 26, RTP, NC, US, pp: 18-28.
- Gakhar N, Goldberg E, Jing M, Gibson R, House JD. 2012. Effect of feeding hemp seed and hemp seed oil on laying hen performance and egg yolk fatty acid content: evidence of their safety and efficacy for laying hen diets. Poult Sci, 91(3): 701-711.
- Goldberg EM, Gakhar N, Ryland D, Aliani M, Gibson RA, House

JD. 2012. Fatty acid profile and sensory characteristics of table eggs from laying hens fed hempseed and hempseed oil. J Food Sci, 74: 153-160.

- Göre M, Kurt O. 2021. Bitkisel üretimde yeni bir trend: Kenevir. Int J Life Sci Biotechnol, 4(1): 138-157. https://doi.org/10.38001/ijlsb.789970.
- Gustafsson SB, Jacobsson SOP. 2019. Effects of cannabinoids on the development of chick embryos in ovo. Sci Rep, 9: 13486. https://doi.org/10.1038/s41598-019-50004-7.
- Hazekamp A, Fischedick JT, Llano MD, Lubbe A, Ruhaak RL. 2010. Chemistry of cannabis. In: Liu HW (Ben), Mander L, editors. Comprehensive Natural Products II: Chemistry and Biology. Elsevier, London, UK, pp: 1033-1084.
- House JD, Neufeld J, Leson G. 2010. Evaluating the quality of protein from hemp seed (Cannabis sativa L.) products through the use of the protein digestibility-corrected amino acid score method. J Agri Food Chem, 58(22): 11801-11807. https://doi.org/10.1021/jf102636b.
- Kalmendal R. 2008. Hemp seed cake fed to broilers. Dissertation, Swedish University of Agricultural Sciences, Department of Animal Nutrition and Management, Uppsala, Sweden, pp: 43.
- Khaksar S, Bigdeli MR. 2017. Intra-cerebral cannabidiol infusion-induced neuroprotection is partly associated with the TNF-α/TNFR1/NF-кB pathway in transient focal cerebral ischaemia. Brain Inj. 31(13-14): 1932-1943. https://doi.org/10.1080/02699052.2017.1358397.
- Khan RU, Durrani FR, Chand N, Anwar H, Naz S, Farooqi FA, Manzoor MN. 2009. Effect of Cannabis sativa fortified feed on muscle growth and visceral organs in broiler chicks. Int J Biol Biotech, 6(3): 179-186.
- Khan RU, Durrani FR, Chand N, Anwar H. 2010. Influence of feed supplementation with Cannabis sativa on quality of broilers carcass. Pakistan Vet J, 30: 34-38.
- Konca Y, Yalcin H, Karabacak M, Kaliber M, Durmuscelebi FZ. 2014. Effect of hempseed (Cannabis sativa L.) on performance, egg traits and blood biochemical parameters and antioxidant activity in laying Japanese quail (Coturnix coturnix japonica). British Poult Sci, 55(6): 785-794. http://dx.doi.org/10.1080/00071668.2014.978264.
- Konieczka P, Szkopek D, Kinsner M, Kowalczyk P, Michalczuk M, Bie'n D, Banach J, Matusevi^xcius P, Bogucka J. 2023. Cannabidiol and nano-selenium increase microvascularization and reduce degenerative changes in superficial breast muscle in c. perfringens-infected chickens. Int J Mol Sci, 24: 237. https://doi.org/10.3390/ijms24010237.
- Kopustinskiene DM, Masteikova R, Lazauskas R, Bernatoniene J. 2022. Cannabis sativa L. Bioactive compounds and their protective role in oxidative stress and inflammation. Antioxidants, 11: 660.

https://doi.org/10.3390/antiox11040660.

- Kozela E, Juknat A, Vogel Z. 2017. Modulation of astrocyte activity by cannabidiol, a nonpsychoactive cannabinoid. Int J Mol Sci, 18(8): 1669. https://doi.org/10.3390/ijms18081669
- Langer E, Scilcher H. 1999. Propolis-Qualitat und Wirkungen von Proolis bzw. Propolis-zubereitugen. Dtsch Apoth Ztg, 37: 51-63.
- Lee WS, Erdelyi K. 2016. Cannabidiol limits T cell-mediated chronic autoimmune myocarditis: implications to autoimmune disorders and organ transplantation. Mol Med, 22(1): 1. https://doi.org/10.2119/molmed.2016.00007.
- Magen I, Avraham Y, Ackerman Z, Vorobiev L, Mechoulam R, Berry EM. 2009. Cannabidiol ameliorates cognitive and motor impairments in mice with bile duct ligation. J Hepatol, 51(3):

528-534. https://doi.org/10.1016/j. jhep.2009.04.021.

- Mahmoudi M, Farhoomand P, Azarfar A. 2012. Effects of graded levels of hemp seed (Cannabis sativa L.) on performance, organ weight and serum cholesterol levels on broilers. J Medic Plants, 2(42): 121-129.
- McPartland JM, Russo EB. 2001. Cannabis and Cannabis extracts: greater than the sum of their parts? J Cannabis Therapeut. 1: 103-132.

https://doi.org/10.1300/J175v01n03_08.

- Mechoulam R, Berry EM, Avraham Y, Marzo VD, Fride E. 2006. The endocannabinoid system, mechanism of action and Function. Endocannabinoids, feeding and suckling-from our perspective. Int J Obesity, 30: 24-28.
- O'Sullivan SE, Kendall DA. 2010. Cannabinoid activation of peroxisome proliferator-activated receptors: potential for modulation of inflammatory disease. Immunobiology, 215 (8): 611-616.
- Pazos MR, Mohammed N, Lafuente H, Santos M, Martínez-Pinilla E, Moreno E. 2013. Mechanisms of cannabidiol neuroprotection in hypoxic-ischemic newborn pigs: role of 5HT(1A) and CB2 receptors. Neuropharmacology, 71: 282- 291.
- Pellati F, Borgonetti V, Brighenti V, Biagi M, Benvenuti S, Corsi L. 2018. Cannabis sativa L. and Nonpsychoactive Cannabinoids: Their Chemistry and Role against Oxidative Stress, Inflammation, and Cancer. BioMed Res Int, 2018: 1691428.
- Pertwee RG. 2008. The diverse CB 1 and CB 2 receptor pharmacology of three plant cannabinoids: D 9 tetrahydrocannabinol, cannabidiol and D 9 tetrahydrocannabivarin. Br J Pharmacol, 153(2): 199-215.
- Rajesh M, Mukhopadhyay P, Btkai S, Patel V, Saito K, Matsumoto S. 2010. Cannabidiol attenuates cardiac dysfunction, oxidative stress, fibrosis, and inflammatory and cell death signaling pathways in diabetic cardiomyopathy. J Am Coll Cardiol, 56(25): 2115-2125.
- Salami SA, Luciano G, O'Grady MN, Biondi L, Newbold CJ, Kerry JP. 2019. Sustainability of feeding plant by-products: a review of the implications for ruminant meat production. Anim Feed Sci Technol, 251: 37-55. https://doi.org/10.1016/j.anifeedsci.
- Santos NAG, Martins NM, Sisti FM, Fernandes LS, Ferreira RS, Queiroz RHC. 2015. The neuroprotection of cannabidiol against MPP+-induced toxicity in PC12 cells involves trkA receptors, upregulation of axonal and synaptic proteins, neuritogenesis, and might be relevant to Parkinson's disease. Toxicol Vitr, 30(1): 231-240.
- Schultes RE, Klein WM, Plowman T, Lockwood TE. 1975. Cannabis: An example of taxonomic neglect. Cannabis Cult, 1975: 21-38.
- Siano F, Moccia S, Picariello G, Russo G, Sorrentino G, Di Stasio M, La Cara F, Volpe M. 2019. Comparative study of chemical, biochemical characteristic and ATR-FTIR analysis of seeds, oil and flour of the edible fedora cultivar Hemp (Cannabis sativa L.). Molecules, 24(1): 83.
- Silversides FG, Lefrancois MR. 2005. The effect of feeding hemp seed meal to laying hens. British Poult Sci, 46(2): 231-235.
- Small E, Cronquit A. 1976. A practical and natural taxonomy for cannabis. Taxon, 25: 405-435.
- Smirnov A, Tako E, Ferket PR, Uni Z. 2006. Effects of in ovo feeding of carbohydrates and beta-hydroxy-betamethylbutyrate on the development of chicken intestine. Poult Sci, 83(12): 2023-2028.
- Smith K. 2000. Hempseed oil: A smart start. The Hemp Rep, 2(14): 1488-3988.
- Wang Y, Mukhopadhyay P, Cao Z, Wang H, Feng D, Haskó G, et al. 2017. Cannabidiol attenuates alcohol-induced liver steatosis, metabolic dysregulation, inflammation and neutrophil-mediated injury. Sci Rep, 7(1): 1-12.
- Zuardi AW, Morais SL, Guimarães FS, Mechoulam R. 1995. Antipsychotic effect of cannabidiol. J Clin Psychiatry, 56(10): 485-486.

doi: 10.47115/bsagriculture.1518799

Open Access Journal e-ISSN: 2618 – 6578

Review

Volume 7 - Issue (ICABGEH-23): 464-476/ August 2024

NUTRITIONAL ASPECTS FOR MODULATION OF POULTRY IMMUNE STIMULATION

Merve GÜNDÜZ1*, Arda SÖZCÜ1, Üzeyir ÇAĞIRICI²

¹Bursa Uludağ University, Faculty of Agriculture, Department of Animal Science, 16059, Bursa, Türkiye ²Kaman Türk Telekom Osman Kulaksız Vocational and Technical Anatolian High School, 40300, Kırşehir, Türkiye

Abstract: The immune system's defense mechanism has been affected by the management and nutrition conditions in poultry production. In biological aspects, Bursa Fabricius and Thymus are primary lymphoid organs that defend the body against diseases. The spleen, cecal tonsils, immunoglobulins, and various forms of mucosal immunity, BALT (bronchus-associated lymphoid tissue), MALT (mucosa-associated lymphoid tissue), CALT (conjunctiva-associated lymphoid tissue), and GALT (gut-associated lymphoid tissue), are also vital to their overall health. The management program (environmental conditions, heating, lighting practices) and feed (feed ingredients, composition, and feed additives) significantly affect birds' immunity and performance. Dietary supplementation enhances immune system activation by improving the lymphoid tissues and beneficial immune modulators and responses. During the past fifteen years, alternative feed additives (amino acids, minerals, vitamins, probiotics, prebiotics, synbiotics, and organic acids) have impacted animal production performance and immunity. Besides, feed additives, such as probiotics, prebiotics, vitamins, minerals, and organic acids, are essential for sustaining hormonal, physiological, and immunological processes. These additives, probiotics, vitamins (A, C, D, E), mineral (selenium) supplementations, and organic acids boost the immune system's activity against microbial pathogens and physiological stress mechanisms. Finally, this paper aims to review the current improvement in the relationship between nutrition and immunity in poultry.

Keywords: Immune system, B and T cells, Vitamins, Minerals, Organic acids

1. Introduction

The immune system is characterized by the innate (maternal) and adaptive immunological developmental processes against with inadequate defence mechanism and health issues, predisposing individuals to microbial infections (Wynn and Levy, 2010; Alkie et al., 2019). To prevent the spread of viral, bacterial, fungal, and parasitic diseases in the body, the immune system plays a major role with its activity. The body produces defense molecules called antibodies, which recognize and deactivate pathogens (Sarıca et al., 2009). Besides, different pathogens potentiate the cellular and humoral response by altering the immune system's reaction (Shaji et al., 2023).

The immune system consists of various organs and tissues, including the Bursa Fabricius, thymus, bone marrow, spleen, the harderian gland, lymph nodes, circulating lymphocytes, and lymphoid tissue in the alimentary tract in poultry (Panda et al., 2015). Lymphoid tissue may be specialized into two types: primary and secondary. The thymus and Bursa Fabricius are considered primary organs, whereas the spleen is a secondary type of lymphoid tissue. The Bursa Fabricius belongs to birds and is absent from mammals due to

evolution. In both hens and mammals, primary lymphoid tissue reduces after puberty, with the effects of sex hormones (Junior et al., 2018).

Animals have a precise immune system mechanism that resists pathogens and adverse environmental conditions (Korver, 2023). Among the immunosuppressive components are management, feeding programs, and nutrition deficiencies, which cause health and welfare issues (Wlazlak et al., 2023). Also, immunosuppression decreases body weight gain and production performance and increases mortality and morbidity rates mainly due to secondary viral and bacterial infections (Fussell, 1998; Rehman et al., 2016). Immune system activation on metabolism has a significant impact on maintaining broiler health to optimize performance and cost-efficient production. Thus, acute activation of the immune system leads to inflammatory pathways with reduced feed intake. The hypothalamic-pituitary-adrenal axis is activated when the immune system detects a foreign substance and releases chemical messengers known as cytokines. These cytokines are responsible for initiating the production of acute phase proteins (APP) in the liver, transmitting feedback to the brain, resulting in common illness symptoms such as fever and decreased appetite

BSJ Agri / Merve GÜNDÜZ et al. 464

(Silverman et al., 2005; Rochell et al., 2016). It has been stated that immunity and production performance in poultry are positively correlated. Antibody production and immunity regulation depend on non-genetic factors, which include nutritional concentrations in the diet. At the same time, it has been proven that imbalance between nutrients or toxicity, disorders in the bird's physiology may occur due to immunosuppression. The immune system's reaction to harmful microorganisms penetrating the animal's body is related to its nutritional condition. Immune system suppression may result in inadequate body weight gain and feed conversion ratio, high mortality rate, uniformity issues, secondary bacterial infections, and lymphoid organ atrophy (Aviagen, 2009; Abo-Al-Ela et al., 2021). Currently, modern broilers have a rapid growth rate, high body weight, and efficient feed conversion, which makes them vulnerable to high disease rates and necessitates strong immune function. Because the nutrients in the feed are utilized for muscle growth, fast-growing modern broilers tend to have low disease resistance and are susceptible to diseases (Song et al., 2021).

Numerous factors, including diseases, environmental conditions, management practices, vaccination programs, physical type of feed material, and nutrition-related stress, weaken the immune system, potentially increasing susceptibility to infections (Abo-Al-Ela et al., 2021; Wlazlak et al., 2023). Nutritional deficiencies in the feed supplementation may result in deterioration in the growth and development mechanism of the immune system. Additionally, insufficient vitamin levels may compromise the immune system, enhancing susceptibility to infection and inflammation. During metabolic processes, vitamins of A, D, E, and C have been shown to significantly impact immune system function (Khan et al., 2023). For this reason, the review focused on the relationship between nutrition and immune system modulation of poultry.

2. Primary Immune Organs

The formation of the immune system organs begins in the embryonic stage and entirely functions with advancing age. The immune system organs generate functional lymphocytes and release them into the peripheral immune system to actively participate in immune responses (Naukkarinen and Hippelãinen, 1989; Gordon and Manley, 2011). In the avian immune system, there are two primary organs. The first one is Bursa Fabricius, associated with B-cells, and the second is the Thymus, associated with T-cells. Both organs produce B and T cells to improve poultry immunity. The thymus is located in the neck's ventral region and under the thyroid gland in front of the chest cavity. This gland is surrounded by a thin capsule composed of connective tissue. The compartments of the thymus gland contain reticular cells and lymphocytes. The thymus gland's primary function is to protect the body from infection by activating lymphocytes before and immediately after

birth. Autoreactive T cells are responsible for eliminating killing cells and synthesizing thymic hormones during the maturation of T lymphocytes in poultry (Sarıca et al., 2009). T and B cells progress to the secondary lymphoid tissue to provide a defense, which includes the spleen and mucosa-associated lymphoid tissue (MALT). The primary lymphoid tissue is crucial in selecting lymphocytes (including T-cells and B-cells) for a robust immune response while preventing autoimmunity (Yasuda et al., 2003).

The lymphoid stem cells in the bone marrow generate the T and B cell precursors (Ratcliffe and Jacobsen, 1994; Ratcliffe, 2006; Garcia et al., 2021). Bursa Fabricius is placed in the dorsal of the cloaca and is responsible for the maturation of B lymphocytes, the killing of autoreactive B cells, and the synthesis of hormones. The Bursa Fabricius produces B-cells, which are responsible for synthesizing antibodies. Moreover, bone marrowderived stem cells that resemble lymphoid cells mature in the thymus and Bursa Fabricius before proceeding via the bloodstream and lymphatic circulation to peripheral immune organs (Gordon and Manley, 2011; Davison, 2022). During the exposure to foreign antigens, the immune cells exhibit proliferation and differentiation. Additionally, in peripheral blood lymphocytes, the proliferative activity indicates a strengthening functional activity of T and B cells, reflecting an elevated cellular immune function (Dekruyff et al., 1980; Naukkarinen and Hippelãinen, 1989).

3. Secondary Lymphoid Organs

It has been stated that spleen, bone marrow, harderian gland (in orbit), pineal gland (in the brain), mucosaassociated lymphoid tissues (MALT), bronchusassociated lymphoid tissue (BALT), gut-associated lymphoid tissue (GALT), conjunctiva-associated lymphoid tissue (CALT) are secondary lymphoid organs in birds. MALT, BALT, GALT, and CALT lymphoid tissues comprise approximately 50% of the cells in the spleen. These lymphoid tissues accumulate on the skin or mucosal surface, where foreign antigens enter the body (Gurjar, 2013). The spleen is the largest of the secondary lymphoid organs and is surrounded by a capsule composed of connective tissue. It has several functions for the aging erythrocytes, erythrocytes production in fetal life, and granulocytes in postnatal life, to contribute to the formation of antibodies through B lymphocytes, phagocytosis through macrophage, and for accumulation of red blood cells (Júnior et al., 2018). On the other hand, spleen size increases through hyperplasia tissue, known as splenomegaly, after exposure to pathogens. Lymphocyte hyperplasia has also been observed in MALT. The germinal center is where avian antibodies are effectively produced (Junior et al., 2018; Lewis et al., 2019). Additionally, chickens have three classes of immunoglobulins similar to mammals: IgA, IgM, and IgY (equivalent to IgG). Although it has been suggested that chickens also have antibodies similar to mammalian IgE

and IgD, this has not been proven. Chicken IgY and mammalian IgG characterize immunological similarities, and the DNA sequence of chicken IgY is similar to human IgE (Shimizu et al., 1992; Aizenshtein et al., 2016; Pereira et al., 2019).

4. Nutrition and Immunity

Birds' immune systems benefit considerably from a wellbalanced nutrition program. The function of the immune system is to struggle with the negative impacts of stress and protect against pathogenic diseases. The regulation of the immune system in poultry involves the progress of anatomy in lymphoid tissues, mucus production, immunologically active substances synthesis, cellular proliferation, activation, movement, intracellular killing of pathogens, and modulation and regulation of the immune process. The nutritional intake may influence chickens' immune reactions, substantially benefiting their well-being and health. Therefore, a balanced diet could enhance chickens' overall health by preventing stress-related immune suppression (Butcher and Miles, 2002). The stimulation of immune system reactions in poultry nutrition has been revealed (Montout et al., 2021; Selim et al., 2021; Yu et al., 2021).

Poultry production has been focused on improving feed efficiency, decreasing disease risk, and increasing performance (Korver, 2023). Feed additives in poultry nutrition have been used as an alternative to antibiotics due to their potential positive effects on the growth rate, feed conversion, and resistance against pathogenic diseases (Fonseca et al., 2010; Ganan et al., 2012). It is widely known that antibiotics have extensively impacted poultry production in terms of maintaining gut microorganism balance and promotion (Selaledi et al., 2020). Due to food safety concerns and the public demand for antibiotic-free farm animal products, many countries have banned of antibiotics in poultry nutrition as a routine of growth promotion (Neveling and Dicks, 2021). So that, alternatives to antibiotics are needed to sustain poultry productivity and promote their performance and immunologic response (Wickramasuriya et al., 2024).

Nutraceuticals, including essential minerals and vitamins, are fundamental to animal nutrition. Nutritional components significantly benefit by preventing diseases, exhibiting immunomodulatory properties, promoting overall health, and enhancing productivity, regulating gut microbiota (Alagawany et al., 2021; Shehata et al., 2022). Feed additives essentially comprise amino acids, minerals, vitamins, fatty acids, enzymes, prebiotics, probiotics, synbiotics, pigments, medicinal herbs, herbal extracts, antioxidants, organic acids, and flavoring agents (Alagawany et al., 2018; Mohammed et al., 2019; Elgeddawy et al., 2020; Adhikari et al., 2020; Rehman et al., 2020; Shehata et al., 2022). Synthetic supplements include essential amino acids (such as lysine, methionine, threonine, and tryptophan), vitamins, and minerals. Besides, nutraceutical dietary elements may enhance digestion, absorption, utilization, metabolism, and positive health status (Ravindran, 2010; Haq at al., 2016; Alagawany et al., 2021) (Table 1).

The nutrients have been known to potentially affect embryonic development and hatchability with the prevention of musculoskeletal, immunological, and circulatory issues (Uni et al., 2012). During embryogenesis, nutrients are mainly acquired from yolk, which mainly contains lipids and low levels of carbohydrates (Santos et al., 2010). Supplementing with bioactive substances such as amino acids, polyphenols, and prebiotics will enhance the immune system, reduce osteoporosis, and lower the risk of heart diseases (Chalamaiah et al., 2018). The application of in-ovo injection for various biological agents has been recognized for its efficacy in poultry production. The application of in-ovo entails using nutritional supplements such as carbohydrates, proteins, amino acids, vitamins, and minerals (Ohta et al., 1999; Bhanja et al., 2005; Zhai et al., 2011; Bakyaraj et al., 2012). L-Arginine (100 μg/μL/egg) in-ovo injection on the 14th day of the embryonic stage enhanced survival rates, body weight, and immune response (IgM) in Subramaniyan et al. (2019) research.

After hatching, the early feeding program has gained immense importance for the developmental process of immunity in birds. Firstly, innate immunity is considered the defense mechanism, and adaptive immunity is for the protection against specific pathogens by the functions of lymphoid cells and antibodies. It has been stated that innate immunity is essential to secure all living organisms' cellular integrity, homeostasis, and livability (Buchmann, 2014; Romo et al., 2016).

The formation of birds' defense mechanisms begins in embryogenesis and continues to function for several days after hatching (Yasuda et al., 2003). Maternal antibodies in the yolk protect embryos and chicks (approximately one month) against microbial infection during egg incubation and after hatching. The secondary lymphoid tissues complete their development one week after hatching, facilitating an effective immune response and specific antibody production in chick vaccination (Yasuda et al., 2003; Hamal et al., 2006). Additionally, the posthatching period is significant due to a lack of maternal immunity (approximately one week) (O'Neal and Ketterson, 2012).

Table 1. Dietary supplementation in poultry immune stimulation

5. Nutritional Aspects in Poultry Immunity

5.1. Amino Acids

Dietary supplementation has been essential in optimizing poultry production due to obtaining better performance and health status with lower feed efficiency. The immune system consistently functions with sufficient levels of protein, energy, and micronutrients, vitamins, and minerals in nutrition (Butcher and Miles, 2002; Alagawany et al., 2021). Amino acids are the fundamental components of protein, the primary substances contributing to the growth of chickens and the production of eggs (Baker, 2009). The mechanism of lymphocyte activation, natural killer cells, and macrophages regulating intracellular redox balance, gene expression, and cytokine production related to amino acid supplementation. Therefore, appropriate dietary amino acids are critical for inhibiting infectious disorders, particularly viral infections (Chandra ve Kumari, 1994; Calder, 2013; Montout et al., 2021).

Several amino acids, such as methionine, threonine, and arginine, were studied to determine their contribution to a beneficial host immunological response (Montout et al., 2021). It has been indicated that different methionine supplementation levels of 0.60% and 0.90% in turkeys and broilers respectively resulted in increased methionine levels in the peripheral blood (Mirzaaghatabar et al., 2011; Kubinska et al., 2014). Also, in broilers, the dietary supplementation of %0.65 methionine and %0.13 choline may result in higher antibody levels (Swain and Johri, 2000).

Arginine has been characterized as a regulator of both innate and adaptive immunity (Lindez and Reith, 2021). Dietary arginine supplementation in metabolic processes may affect birds' immunological responses to physiological hemostasis under stressful conditions. A group of enzymes known as nitric oxide synthases are responsible for converting arginine into citrulline and nitric oxide as the sole route of nitric oxide synthesis (Förstermann et al., 1991; Fernandes et al., 2009). Nitric oxide exhibits cytotoxic effects on immuno-activated cells and regulates the immune system (Hibbs et al., 1988). Furthermore, arginine enhances interleukin production and T lymphocyte proliferation in broiler chickens (Rodriguez et al., 2017; Montout et al., 2021). Increasing dietary arginine over 0.45% may affect macrophage phagocytosis by stimulating the production of key cytokines, particularly IL-1, IL-2, INF, and TNF-a in broiler chickens (Emadi et al., 2010; Xu et al., 2018).

Supplementing lysine and methionine enhanced antibody production in broiler chickens against Newcastle and Gumboro infections (Bouyeh, 2012). A deficiency in dietary lysine inhibits protein synthesis, including cytokines, and lymphocyte proliferation, impairing immune responses in chickens and leading to morbidity and mortality in response to infection (Kidd, 2004). Insufficient dietary lysine levels may result in hen antibody responses and cell-mediated immunity (Chen et al., 2003).

Several essential amino acids (leucine, isoleucine, and valine) have been used in poultry nutrition. These amino acids contribute to protein synthesis, immune response modulation, intestinal tract health, and villi development (Kim et al., 2022; Liu et al., 2023). Leucine, isoleucine, and valine amino acids provide the amino groups for the synthesis of glutamine and recreate a crucial role in maintaining the gut barrier functions for epithelial cells. Furthermore, the immune system considerably depends on protein synthesis to produce cytokines, immunoglobulins, immune cells, and other essential immune molecules (Morgan, 2021; Montout et al., 2021). According to Liu et al. (2023), there were no significant differences between the effects of reduced-protein diets supplemented with arginine or branched-chain amino acids on the short-chain fatty acid profile in the cecal content of broiler chickens challenged with Eimeria spp on day 21.

L-carnitine is present in high lymphocyte concentrations for biological functions. Two essential amino acids (lysine and methionine), three vitamins (ascorbate, niacin in the form of nicotin amide adenine dinucleotide and vitamin B6), and reduced iron (Fe2+) are required as cofactors for the enzymes involved in the metabolic pathway of L-carnitine synthesis (Borum, 1983; Rebouche, 1992; Leibetseder, 1995; Adabi et al., 2011). These amino acids inhibit the apoptosis of immune cells and enhance their proliferative response to mitogens (Adabi et al., 2019). It has been reported that supplementing the diet of broiler chickens with 100 mg/kg of L-carnitine enhances the innate immune response by boosting acute phase protein production, and L-carnitine reveals glucocorticoid effects (Buyse et al., 2007). The supplementation of L-carnitine considerably enhanced total IgG and IgA responses but not IgM in broilers (Mast et al., 2000). In a study, the addition of 100 mg/kg L-carnitine supplementation indicated significantly effect on higher antibody response and Newcastle disease virus in broiler. Besides, 100 mg/kg L-carnitine supplementation group showed the highest Bursa Fabricius, spleen and thymus weight compared to 0, 25 and 75 mg/kg L-carnitine group (Golzar et al., 2007).

5.2. Probiotics, Prebiotics and Synbiotics

Bacterial population in the intestinal tract is necessary for appropriate immune function and obtaining better feed efficiency with probiotics in poultry. Primary, probiotics consist of live beneficial microorganisms for the host animal's health and welfare due to stimulating effects for intestinal microbial balance (Moreno et al., 2006). Probiotic species consist of Lactobacillus bulgaricus, Lactobacillus plantarum, Streptococcus thermophils, Bifidobacterim bifidum, Aspergillus oryzae (Khaksefidi and Rahimi, 2005; Ashayerizadeh et al., 2009; Kabir, 2009). Bacteria have a mutually beneficial relationship with the intestinal cells and microorganisms in the gastrointestinal tract. Probiotics significantly contribute to changes in the intestinal structure and digestive processes by stimulating the immune system. They also inhibit the growth of pathogenic bacteria, leading to an improvement in overall performance (Awad et al., 2009; El-Shenway and Soltan, 2015). Probiotics have importance for the immune system with including boosting the activity of macrophages, lymphocytes, and NK cells. They also enhance the oxidative burst of heterophils, resulting in increased production of immunoglobulins (IgG, IgM, and IgA). Also, probiotics have the potential effects to regulate the gastrointestinal tract and maintain an optimal balance between antiinflammatory and pro-inflammatory cytokines. Besides, the population of lamina propria lymphocytes (LPL) and intestinal epithelial lymphocytes (IEL) has increased with the probiotics activity in the small intestine while restricting the proliferation of detrimental microorganisms (Yeşilyurt et al., 2021).

Bacillus subtilis-based probiotic supplement has been approved to improve immune and oxidative responses in shackled broiler chickens under pre-slaughter stress challenge. The levels of IgG, IgA, and IgM of the shackled broiler chickens exposed to preslaughter stress were investigated. In the result of this study, IgG and IgA levels (mg/dL) in serum concentration were found similar between control, 0.25 g/kg feed, and 0.5 g/kg feed supplementation groups. In contrast, the level of IgM (mg/dL) in serum concentration was higher in the control group compared to 0.25 g/kg and 0.5 g/kg treatment groups (Mohammed et al., 2024).

Prebiotics are non-digestible feed ingredients that benefit by promoting the growth or activity of some bacterial species in the colon to improve host health (Ganguly, 2013). Prebiotics are small carbohydrate fragments, possibly commercially used as galactose, fructose, and mannose oligosaccharides. There are some prebiotics, such as fructooligosaccharide (FOS), transgalactooligosaccharide (TOS), inulin, glucooligosaccharide, xylooligosaccharide, isomaltooligosaccharide, soybean oligosaccharide, polydextrose, and lactosucrose (Vulevic et al., 2004; Propulla, 2008; Ganguly, 2013). Furthermore, mannan oligosaccharide (MOS) may be derived from the outer cell wall of Saccharomyces spp. yeast promotes gut health with pathogenic bacteria in type-I fimbriae or agglutinating various bacterial strains and increasing villi length, uniformity, and integrity (Spring et al., 2000; Loddi et al., 2004). Oligosaccharides are prebiotics that boost chickens' intestinal morphology and stimulate the activation of the immune system (Sayed et al., 2023). Oligosaccharides may be more active on macrophages to regulate the production of inflammatory cytokines and cytokines by T-helper cells (Th) during the immunological processes (Csernus et al., 2020).

Sittiya and Nii (2024) investigated the effects of oligosaccharides on the immune reactions in laying hens challenged with dextran sodium sulfate. Consequently, BOS (oligosaccharide extract from bamboo shoots) also increased *IFN-γ* in the liver and Th-17 cytokines in the

intestine in laying hens. In contrast, there are no significant differences in intestinal or liver morphology. On the other hand, BOS (oligosaccharide extract from bamboo shoots) suppressed an increase in leukocyte accumulation in the liver under DSS (dextran sodium sulfate treatment). These results approved that BOS (oligosaccharide extract from bamboo shoots) may enhance egg quality and Th-1 and Th-17 immune function without causing tissue damage under normal conditions, and it may suppress the excessive inflammatory responses during inflammation.

Synbiotics are a combination of probiotics and prebiotics that act as effective substitutes for antibiotics in feed (Dong et al., 2016; Mohammed et al., 2018; Ren et al., 2019). The beneficial effect of the synbiotics has impacted the gastrointestinal tract and health status in animals. Moreover, different synbiotics comprise probiotic strains, Lactobacillus plantarum, Lactobacillus reuteri, Lactobacillus pentosus, and Saccharomyces cerevisiae (Śliżewska et al., 2020). Due to the regulation of physiological balance and modifying both innate and adaptive immune responses against stress, synbiotics have gained immense importance in poultry nutrition (Seifert et al., 2011). It has been demonstrated that different levels of the synbiotic supplementation significantly changed plasma levels of the antioxidant enzyme GPx in heat-stressed broiler chickens (Mohammed et al., 2019).

5.3. Minerals

Minerals are essential micronutrients for activating optimal physiological processes and health status. The inclusion of microelements in poultry diets is common in commercial practice due to their importance in reproduction and immune function (Saripinar-Aksu et al., 2012). An adequate mineral supply in the diet is crucial for optimal poultry production. The lack of minerals in the diet may lead to various health issues in chicks (Pal, 2017). In poultry nutrition, optimizing animal health may be associated with adding zinc, selenium, chromium and zeolite to the diet program for rearing broilers and laying hens (Park et al., 2004; Tayeb and Qader, 2012; Jain et al., 2021; Elsherbeni et al., 2024).

Dietary zinc decreases feed intake and stimulates avian antibody production with plasma corticosterone levels (Onbasilar and Erol, 2007). It has been shown that dietary zinc supplementation of up to 80 ppm level enhances humoral and cell-mediated immune responses in broilers by Sunder et al. (2008). On the other hand, dietary zinc (up to 110 ppm) and higher doses (up to 10,000 mg/kg) in the diet result in inhibiting the colonization of Salmonella enteritidis (SE) in laying hens (Kubena et al., 2001).

Selenium is essential for optimal immune response and involves innate and acquired immune systems. This essential trace element is a component of selenoproteins, which are involved in various animal physiological processes. Although plants and fungi do not need selenium, they can convert mineral forms of selenium

present in the soil into various organic forms, such as selenomethionine and methyl selenocysteine, as a strategy of adaptation. Selenoproteins influence immunity through multiple mechanisms, and nutrition could modulate the immune system to resist pathogens. Selenium regulates oxidative stress, redox, and other cellular processes in all tissues and cell types, including immune responses. Selenium interacts with vitamin E in tissues to protect biological membranes from oxidative damage (Allmang et al., 2009). On the otherside, in Jain et al. (2021) study, the plasma IgG concentrations (μg/mL) showed significant differences between groups at 21, 28, 35 day age of broilers with feed different concentrations of organic and inorganic trace minerals (zinc, selenium, and chromium) supplementation.

Zeolite is a silicate mineral that is considered highly effective and non-toxic. Dietary supplementation of zeolite has been used as a feed additive. Additionally, the use of zeolite directly impacts on production performance and meat quality (Zhou et al., 2014; Abd El-Hady, 2020; Dashtestani et al., 2021). Zeolite has been used in biological processes to reduce ammonia and smell emissions from poultry production by absorbing nitrogenous metabolites such as ammonium (NH₄+) and ammonia (NH3) (Schneider et al., 2017). Elsherbeni et al. (2024) investigated the effects of using zeolite on lymphoid organs (%) in 42-day-old broiler chickens. In this study, thymus and Bursa Fabricus weights (%) were found to be higher in the 10g/kg zeolite diet supplementation group. In another study by Abdelrahman et al. (2023), it was shown that dietary zeolite supplementation at levels of 0.5%, 1.0%, 1.5%, and 2.0% resulted in a statistically significant reduction in spleen weight of broiler chicks compared to the untreated group. However, the different zeolite treatments did not significantly affect the thymus and Bursa Fabricus weights.

5.4. Vitamins

The use of vitamins in nutrition is essential for maintaining a healthy immune system in poultry. It has been stated that vitamin level insufficiency has led to compromised immune function, increased susceptibility to infections, and inflammation. In poultry nutrition, vitamins (A, D, E, and C) significantly impact the immunologic process with various activation mechanisms. Firstly, vitamin A enhances mucosal immunity, reduces free radicals, and maintains epithelial cell integrity. Vitamin A comprises several brightly colored and fat-soluble molecules involving retinol, retinal, retinoic acid (RA), and pro-vitamin A carotenoids. Carotenoids have multiple functions: immune regulation and stimulation, antioxidant, antimutagenic, and anticarcinogenic properties. Avian and mammalian species may not synthesize carotenoids, unlike plants and microorganisms. Therefore, chickens must obtain carotenoids from their diet as the yolk initially provides vitamin A during embryonic development. However, after hatching, they require dietary sources of vitamin A

(Khan et al., 2023). Post-hatch immune system function improved with the addition of vitamin A to the diet (Shojadoost et al., 2021).

Vitamin D is a type of fat-soluble vitamin that is obtained through sunlight exposure or supplements. Vitamin D is absorbed by the small intestine and then taken up by the liver, where it is converted into 25-hydroxyvitamin D3 (25(OH)D3), the stored form of vitamin D in the body. Vitamin D3 (1,25(OH)2D3), the physiologically active form of the vitamin, is produced in the kidneys by the enzyme 1a-hydroxylase. This form is responsible for the biological actions of vitamin D, regulating bone and mineral metabolism and modulating immune responses (Fakhoury et al., 2020; Wei et al., 2024). It has been demonstrated that 25(OH)D3 increases the synthesis of nitric oxide, promotes the activity of innate immune cells, modifies adaptive immunity, and produces antimicrobial proteins and inflammatory cytokines in chickens (Shanmugasundaram et al., 2019; Fakhoury et al., 2020; Sharma et al., 2024).

Vitamin C, commonly known as L-ascorbic acid, is a water-soluble vitamin that the body converts from glucose. In the body, vitamin C does not accumulate, and excess levels are promptly excreted by the kidneys, resulting in decreased absorption (Johnston et al., 2006; Shojadoost et al., 2021; Hieu et al., 2022). Vitamin C is an antioxidant that protects cells from damage caused by free radicals generated by infection or toxins. Birds may produce vitamin C due to an enzyme called Lgulonolactone oxidase in their renal tissue, which converts l-gulono-g-lactone into ascorbic acid. However, vitamin C requirements increase when chickens experience stressful conditions such as beak trimming, vaccination, transportation, thermal stress, or infection. Therefore, supplementing vitamin C alleviates the adverse effects of these stressful conditions (Abidin and Khatoon, 2013).

Vitamin E is a fat-soluble antioxidant with four distinct functional forms. Among these forms, a-tocopherol is the most biologically active and naturally abundant. This vitamin is well-known for its potential to counteract the adverse effects of free radicals on cell integrity that can occur during normal cell metabolism and inflammation. In poultry, it is crucial to supplement the diet with vitamin E to maintain fertility and hatchability in parent stocks and to prevent nutritional encephalopathy and myopathies in chickens and turkeys. Vitamin E supplementation (or other antioxidants) becomes even more critical when oxidizable fats are included in the feed. The supplementation of vitamin E may prevent the oxidation of unsaturated fats. However, the level of active vitamin E and unsaturated fat used in poultry nutrition may be adequate in intestinal absorption, potentially decreasing the antioxidant status by increasing lipid peroxidation (Pompeua et al., 2018). Vitamin E boosts immune system activity by inhibiting the synthesis of prostaglandins, which may cause inflammation and hinder the immune response. Vitamin E also prevents

oxidation, which is responsible for the production of prostaglandins. Vitamin E is primarily known for its antioxidant properties, effectively reducing the damage caused by free radicals during metabolic stress and immunological disorders. Vitamin E regulates the production of free radicals and gene expression caused by free radical signaling gene expression (Packer and Suzuki, 1993). Deficiency of vitamin E and selenium may impair the immune response in poultry, as indicated in the research conducted by Lewis et al. (2019).

5.5. Organic Acids

Organic acids are a type of feed additives that may promote good health and stimulate immunity in poultry. These acids are categorized as organic chemical compounds that contain the carboxyl group COOH in their structure. It is also known that formic, acetic, propionic, malic, fumaric, and citric acids are commonly utilized in poultry nutrition (Hajati, 2018; Khan et al., 2022). Besides, organic acids upregulate harmful microorganisms' inhibition mechanism by reducing the environment's pH level. By penetrating the biological membranes of microorganisms and disrupting their cell function through electrolytic dissociation, organic acids may also restrict bacterial growth (Feye et al., 2021; Pope et al., 2022). It has been stated that formic acid supplementation has beneficially affected broiler chicken performance (body weight gain, feed intake, and feed conversion ratio). It may also enhance the growth of intestinal villi and increase the number of lymphocytes in the spleen and antibody levels for the prevention of disease (Tawfeeq and Al-Mashhdani, 2020).

Organic acids have been significantly effective in body growth rate, feed efficiency, nutrient digestibility, meat quality, immune response, and suppressing pathogenic bacteria (Yang et al., 2018; Adhikari et al., 2020). The enhancements may be completed by lowering the pH in the gastrointestinal tract, optimizing nutrient utilization in diets, inhibiting pathogen growth, and improving immune response in poultry (Alagawany et al., 2021). Recently, Liao et al. (2020) found a strong correlation between gut microbial communities and organic acids, contributing to the gut health status of broilers. The function of gut microbiota is known as an endocrine organ regulating the immune system through synthesizing interferon/cytokines and immunoglobulins (IgG, IgM, and IgA), activating white blood cells like lymphocytes and natural killer cells, and boosting the activity of macrophages and heterophils (Clarke et al., 2014; Abd El-Hack et al., 2020).

Moreover, acetic acid (0.30 g/kg of) may be responsible to prevent gastric apoptosis and stimulate mucin synthesis (Liu et al., 2017). Also, butyric acid plays a crucial role in promoting intestinal development and maintaining the integrity of intestinal epithelial cells (Sun and O'riordan, 2013). Numerous studies have shown that dietary organic acids can inhibit the growth and spread of harmful opportunistic pathogens such as Salmonella, E. Coli, and Clostridium while also promoting the growth

of beneficial microorganisms such as Lactobacillus, Bacillus, and Bifidobacterium in the intestine (Lee et al., 2015; Nguyen et al., 2018; Gao et al., 2019). Additionally, in organic acids treatment group, the relative weights of lymphoid organs, such as Bursa Fabricius, thymus, and spleen, were found to be higher compared to the control treatment group, which is strongly related to immunological responses in chickens (Ghazala et al., 2011; Yang et al., 2018). These organic acids stimulate specific and non-specific immune responses in broiler chickens by activating macrophages, increasing cytokine production, and IgA, IgG, and IgM levels. Besides, organic acids inhibit bacterial infection growth, proliferation, and intestinal mucosa inflammation (Abd El-Hack et al., 2022).

5. Conclusion

A balanced diet with essential feed supplements (amino acids, probiotics, prebiotics, synbiotics, minerals, vitamins, and organic acids) stimulates the immune system. Dietary deficiencies may cause several health issues and economic losses in profitability for production. Besides, nutrition has been associated with immunologic processes in commercial production systems under various management conditions. Because of that, understanding the patterns of these feed additives is necessary for optimizing strong immunity.

Author Contributions

The percentage of the author(s) contributions is presented below. All authors reviewed and approved the final version of the manuscript.

 $C=Concept, D= design, S= supervision, L= literature search, W=$ writing, CR= critical review, SR= submission and revision.

Conflict of Interest

The authors declared that there is no conflict of interest.

Acknowledgments

This article was presented as an oral presentation at the VII. International Congress on Domestic Animal Breeding, Genetics and Husbandry - 2023 (ICABGEH-23) Krakow, POLAND, September 18 - 20, 2023.

References

- Abd El-Hack ME, El-Saadony MT, Salem HM, El-Tahan AM, Soliman MM, Youssef GBA, Taha AE, Soliman SM, Ahmed AE, El-kott AF, Al Syaad KM, Swelum AA. 2022. Alternatives to antibiotics for organic poultry production: types, modes of action and impacts on bird's health and production. Poult Sci, 101(4): 101696.
- Abd El-Hack ME, El-Saadony MT, Shafi ME, Qattan SYA, Batiha Ge, Khafaga AF, Abdel-Moneim AE, Alagawany M. 2020. Probiotics in poultry feed: A comprehensive review. J Anim Physiol Anim Nutr, 104(6): 1835-1850.
- Abd El-Hady AM. 2020. Effect of incorporating natural zeolite with or without phytase enzyme into broilers diets on blood constituents and carcass traits. Egypt Poult Sci J, 40: 225-242.
- Abdelrahman MM, Al-Baadani HH, Qaid MM, Al-Garadi MA, Suliman GM, Alobre MM, Al-Mufarrej SI. 2023. Using natural zeolite as a feed additive in broilers' diets for enhancing growth performance, carcass characteristics, and meat quality traits. Life, 13: 1548.
- Abidin Z, Khatoon A. 2013. Heat stress in poultry and the beneficial effects of ascorbic acid (vitamin C) supplementation during periods of heat stress. World's Poult Sci J, 69: 135-152.
- Abo-Al-Ela HG, El-Kassas S, El-Naggar K, E. Abdo S, Jahejo AR, Wakeel RA. 2021. Stress and immunity in poultry: light management and nanotechnology as effective immune enhancers to fight stress. Cell Stress and Chaperones, 26: 457-472.
- Adabi SG, Ceylan N, Çiftci İ, Ceylan, A. 2019. Response of growing chicks to supplementation of low protein diets with leucine, valine and glycine-glutamic acid. S Afr J Anim, 49: 1047-1062.
- Adabi SHG, Cooper RG, Ceylan N, Corduk M. 2011. L-carnitine and its functional effects in poultry nutrition. J World's Poult Sci, 67(2): 277-296.
- Adhikari P, Yadav S, Cosby DE, Cox NA, Jendza JA, Kimy WK. 2020. Research Note: Effect of organic acid mixture on growth performance and Salmonella Typhimurium colonization in broiler chickens. Poult Sci, 99(5): 2145862.
- Aizenshtein R, Yosipovicha M, Kvint R, Shadmon S, Krispel E, Shuster E, Eliyahu D, Finger A, Banet-Noach C, Shahar E, Pitcovski J. 2016. Practical aspects in the use of passive immunization as an alternative to attenuated viral vaccines. Vaccine, 34(22): 2513-2518.
- Alagawany M, Abd El-Hack ME, Farag MR, Sachan S, Karthik K, Dhama K. 2018. The use of probiotics as eco-friendly alternatives for antibiotics in poultry nutrition. Environ Sci Pollut Res Int, 25(11): 10611-10618.
- Alagawany M, Elnesr SS, Farag MR, Tiwari R, Yatoo MI, Karthik K, Michalak I, Dhama K. 2021. Nutritional significance of amino acids, vitamins and minerals as nutraceuticals in poultry production and health - a comprehensive review. Vet Quart, 41(1): 1-29.
- Alkie TN, Yitbarek A, Hodgins DC, Kulkarni RR, Taha-Abdelaziz K, Sharif S. 2019. Development of innate immunity in chicken embryos and newly hatched chicks: a disease control perspective. Avian Pathol, 48(4): 288-310.
- Allmang C, Wurth L, Krol A. 2009. The selenium to selenoprotein pathway in eukaryotes: More molecular partners than anticipated. Biochim Biophys Acta, 1790(11): 1415-1423.
- Ashayerizadeh A, Dabiri N, Ashayerizadeh O, Mirzadeh KH, Roshanfekr H, Mamooee M. 2009. Effect of dietary antibiotic, probiotic and prebiotic as growth promoters, on growth performance, carcass characteristics and hematological

indices of broiler chickens. Pak J Biol Sci, 12: 52-57.

- Aviagen. 2009. Immunosuppression in broilers. Aviagen publishers, New York, USA, 0809-AVN-021, pp:254.
- Awad WA, Chareeb K, Abdel-Raheem S, Bohm J. 2009. Effects of dietary inclusion of probiotic and symbiotic on broiler chickens' growth performance, organ weight, and intestinal histomorphology. Poult Sci, 88: 49-56.
- Azzam MMM, Dong XY, Xie P, Wang C, Zou XT. 2011. The effect of supplemental l-threonine on laying performance, serum free amino acids, and immune function of laying hens under high-temperature and high-humidity environmental climates. J Appl Poult Res, 20: 361-370.
- Bai SP, Wu AM, Ding XM, Lei Y, Bai J, Zhang KY, Chio JS. 2013. Effects of probiotic-supplemented diets on growth performance and intestinal immune characteristics of broiler chickens. Poult Sci, 92: 663-670.
- Baker DH. 2009. Advances in protein-amino acid nutrition of poultry. Amino Acids, 37: 29-41.
- Bakyaraj S, Bhanja SK, Majumdar S, Dash B. 2012. Modulation of post-hatch growth and immunity through in ovo supplementated nutrients in broiler chickens. J Sci Food Agric, 92: 313-20.
- Bhanja SK, Mandal AB. 2005. Effect of in ovo injection of critical amino acids on pre- and post-hatch growth, immunocompetence, and development of digestive organs in broiler chickens. Asian Australas J Anim Sci, 18: 524-31.
- Borum PR. 1983. Carnitine. Annu Rev Nutr, 3: 233-259.
- Bouyeh, M. 2012. Effect of excess lysine and methionine on immune system and performance of broilers. Ann Biol Res, 3: 3218-3224.
- Buchmann K. 2014. Evolution of innate immunity: clues from invertebrates via fish to mammals. Front Immunol, 5: 459.
- Butcher GD, Miles D. 2002. Interrelationship between nutrition and immunity. UF/IFAS Extension. University of Florida.
- Buyse J, Swennen Q, Niewold TA, Klasing KC, Janssens GPJ, Baumgartner M, Goddeeris BM. 2007. Dietary L-carnitine supplementation enhances the lipopolysaccharide-induced acute phase protein response in broiler chickens. Vet Immunol Immunopathol, 118(1-2): 154-119.
- Calder PC. 2013. Feeding the immune system. Proc Nutr Soc, 72: 299-309.
- Chalamaiah M, Yu W, Wu J. 2018. Immunomodulatory and anticancer protein hydrolysates (peptides) from food proteins: A review. Food Chem, 245: 205-222.
- Chandra RK, Kumari S. 1994. Nutrition and immunity: An overview. J Nutr, 124: 1433-1435.
- Chen C, Sander J, Dale NM. 2003. The effect of dietary lysine deficiency on the immune response to Newcastle disease vaccination in chickens. Avian Diseases, 47(4): 1346-1351.
- Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG. 2014. Minireview: gut microbiota: the neglected endocrine organ. Mol Endocrinol, 28: 1221-1238.
- Csernus B, Biró S, Babinszky L, Komlósi I, Jávor A, Stündl L, Remenyik J, Bai P, Oláh J, Pesti-Asbóth G, Czeglédi L. 2020. Effect of carotenoids, oligosaccharides and anthocyanins on growth performance, immunological parameters and intestinal morphology in broiler chickens challenged with escherichia coli lipopolysaccharide. Animals (Basel), 10(2): 347.
- Dashtestani F, Ma'mani L, Jokar F, Maleki M, Fard EM, Salekdeh HG. 2021. Zeolite-based nanocomposite as a smart pHsensitive nanovehicle for release of xylanase as poultry feed supplement. Sci Rep, 11: 21386.
- Dashtestani SA, Kang DR, Park JR, Siddiqui SH, Ravichandiran P, Yoo DJ, Na CS, Shim KS. 2019. Effect of in ovo injection of l-

arginine in different chicken embryonic development stages on post-hatchability, immune response, and myo-d and myogenin proteins. Animals, 9: 357.

- Davison F. 2022. The importance of the avian immune system and its unique features. Elsevier, London, UK, pp: 1-9.
- Dekruyff R, Kim YT, Siskind GW, Weksler ME. 1980. Age related changes in the in vitro immune response: increased suppressor cell activity in immature and aged mice. J Immunol, 125: 142.
- Dong ZL, Wang YW, Song D, Hou YJ, Wang WW, Qi WT, Yun TT, Li AK. 2016. The effects of dietary supplementation of microencapsulated enterococcus fecalis and the extract of camellia oleifera seed on growth performance, intestinal morphology, and intestinal mucosal immune functions in broiler chickens. Anim Feed Sci Technol, 212: 42-51.
- Elgeddawy SA, Shaheen HM, El-Sayed YS, Abd Elaziz M, Darwish A, Samak D, Alagawany M. 2020. Effects of the dietary inclusion of a probiotic or prebiotic on florfenicol pharmacokinetic profile in broiler chicken. J Anim Physiol Anim Nutr, 104: 549-557.
- El-Shenway MA, Soltan AM. 2015. Effect of dietary probiotic and/or prebiotic supplementation on growth performance, carcass traits and some serum biochemical alterations in broiler chicken. J Anim Sci Adv, 5: 1480-1492.
- Elsherbeni A, Youssef I, Kamal M, Youssif MAM. 2024. Impact of adding zeolite to broilers' diet and litter on growth, blood parameters, immunity, and ammonia emission. Poult Sci, 103(7): 103981.
- Emadi M, Jahanshiri F, Azizi JF, Kaveh K, Bejo MH, Ideris A, Assumaidaee AA, Alimon RA. 2010. Immunostimulatory effects of arginine in broiler chickens challenged with vaccine strain of infectious Bursal Disease virus. J Anim Vet Adv, 9: 594-600.
- Fakhoury HMA, Kvietys PR, AlKattan W, Anouti FA, Elahi MA, Karras SN, Grant WB. 2020. Vitamin D and intestinal homeostasis: Barrier, microbiota, and immune modulation. J Steroid Biochem Mol Biol, 200: 105663.
- Fernandes JI, Murakami AE, Martins EN, Sakamoto MI, Garcia ER. 2009. Effect of arginine on the development of the pectoralis muscle and the diameter and the protein: deoxyribonucleic acid rate of its skeletal myofibers in broilers. Poultr Sci, 88: 1399-406.
- Feye KM, Dittoe DK, Jendza JA, Caldas-Cueva yJP, Mallmann zBA, Booher zB, Tellez-Isaias G, Owens zCM, Kidd, zMT, Rickex SC. 2021. A comparison of formic acid or monoglycerides to formaldehyde on production efficiency, nutrient absorption, and meat yield and quality of Cobb 700 broilers. Poult Sci, 100: 101476.
- Fonseca BB, Beletti ME, Da Silva MS, Da Silva PL, Duarte IN, Rossi DA. 2010. Microbiota of the cecum, ileum morphology, pH of the crop and performance of broiler chickens supplemented with probiotics. Rev Bras Zootec, 39: 1756- 1760.
- Förstermann U, Pollock JS, Schmidt HH, Heller M, Murad F. 1991. Calmodulin-dependent endothelium-derived relaxing factor/nitric oxide synthase activity is present in the particulate and cytosolic fractions of bovine aortic endothelial cells. Proc Natl Acad Sci, 88: 1788-1792.
- Fussell LW. 1998. Poultry industry strategies for control of immunosuppressive diseases. Poult Sci, 77: 1193-1196.
- Ganan M, Silván JM, Carrascosa AV, Martínez-Rodríguez AJ. 2012. Alternative strategies to use antibiotics or chemical products for controlling Campylobacter in the food chain. Food Contr, 24: 6-14.

Ganguly S. 2013. Supplementation of prebiotics, probiotics and

acids on immunity in poultry feed: a brief review. World's Poult Sci J, 69(3): 639-648.

- Gao YY, Zhang X, Xu L, Peng H, Wang C, Bi Y. 2019. Encapsulated blends of essential oils and organic acids improved performance, intestinal morphology, cecal microflora, and jejunal enzyme activity of broilers. Czech J Anim Sci, 64(5): 189-198.
- Garcia P, Wang Y, Viallet J, Jilkova ZM. 2021. The chicken embryo model: a novel and relevant model for immune-based studies. Front Immunol, 12: 791081.
- Ghazala AA, Atta AM, Elkloub K, Mustafa MEL, Shata RFH. 2011. Effect of dietary supplementation of organic acids on performance, nutrients digestibility and health of broiler chicks. Int J Poultry Sci, 10(3): 176-184.
- Golzar SHA, Rahimi SH, Kamali MA, Torshizi KMA. 2007. The effects of two dietary levels of L-carnitine and vegetable fat powder on quality of cockerels sperm, and fertility and hatchability in broiler breeders. J Vet Res, 62: 107-114.
- Gordon J, Manley NR. 2011. Mechanisms of thymus organogenesis and morphogenesis. Development, 138: 3865- 78.
- Gottardo ET, Junior AMB, Lemke BV, Silva AM, Pasa CLB, Fernandes JIM. 2017. Immune response in Eimeria sp. and E. coli challenged broilers supplemented with amino acids. Austral J Vet Sci, 49: 175-184.
- Gurjar RS. 2013. Cell-mediated immunity after ocular Arktype infectious bronchitis virus vaccination. MSC thesis, Auburn University.
- Gül M, Tunç MA, Cengiz S, Yildiz A. 2014. Effect of organic acids in diet on laying hens' performance, egg quality indices, intestinal microflora, and small intestinal villi height. Europ Poult Sci, 78: 1612-9199.
- Hajati H. 2018. Application of organic acids in poultry nutrition. Int J Avian & Wildlife Biol, 3(4): 324-329.
- Hamal KR, Burgess SC, Pevzner IY, Erf GF. 2006. Maternal antibody transfer from dams to their egg yolks, egg whites, and chicks in meat lines of chickens. Poult Sci, 85(8): 1364- 72.
- Haq Z, Jain RK, Khan N, Dar MY, Ali S, Gupta M, Varun TK. 2016. Recent advances in role of chromium and its antioxidant combinations in poultry nutrition: a review. Vet World, 9(12): 1392-1399.
- Heak C, Sukon P, Sornplang P. 2018. Effect of direct-fed microbials on culturable gut microbiotas in broiler chickens: A meta-analysis of controlled trials. Asian Australas J Anim Sci, 31: 1781-1794.
- Hibbs JR, Taintor RR, Vavrin Z, Rachlin EM. 1988. Nitric oxide: A cytotoxic activated macrophage effector molecule. Biochem Biophys Res Commun, 157: 87-94.
- Hieu TV, Guntoro B, Qui NH, Quyen NTK, Hafiz FAA. 2022. The application of ascorbic acid as a therapeutic feed additive to boost immunity and antioxidant activity of poultry in heat stress environment. Vet World, 15(3): 685-693.
- Jain AK, Mishra A, Singh AP, Patel P, Sheikh AA, Chandraker TR, Vandre R. 2021. Effects of different concentration of organic and inorganic trace minerals (zinc, selenium, and chromium) supplementation on expression of chTLR4 gene and humoral immune response in broilers. Vet World, 14(5): 1093-1101.
- Johnson CS, Corte C, Swan PD. 2006. Marginal vitamin C status is associated with reduced fat oxidation during submaximal exercise in young adults. Nutr Metab, 3: 35.
- Junior AF, dos Santos JP, Sousa IdO, Martin I, Alves EGL, Rosado IR. 2018. Gallus gallus domesticus: immune system and its potential for generation of immunobiologics. Ciênc Rural, 48(08): e20180250.

Kabir SML. 2009. The role of probiotics in the poultry industry. Int J Mol Sci, 10: 3531-3546.

Karachaliou CE, Vassilakopoulou V, Livaniou E. 2021. IgY technology: Methods for developing and evaluating avian immunoglobulins for the in vitro detection of biomolecules. World J Methodol, 11(5): 243-262.

Khaksefidi A, Rahimi SH. 2005. Effect of probiotic inclusion in the diet of broiler chickens on performance, feed efficiency and carcass quality. Asian-Aust J Anim Sci, 18: 1153-1156.

Khan RU, Khan A, Naz S, Ullah Q, Puvaˇca N, Laudadio V, Mazzei D, Seidavi A, Ayasan T, Tufarelli V. 2023. Pros and cons of dietary Vitamin A and its precursors in poultry health and production: A comprehensive review antioxidants. Antioxidants, 12: 1131.

- Khan RU, Naz S, Raziq F, Qudratullah Q, Khan NA, Laudadio V, Tufarelli V, Ragni M. 2022. Prospects of organic acids as safe alternative to antibiotics in broiler chickens diet. Environ Sci Pollut R, 29: 32594-32604.
- Kidd MT. 2004. Nutritional modulation of immune function in broilers. Poult Sci, 83: 650-657.
- Kim WK, Singh AK, Wang J, Applegate T. 2022. Functional role of branched chain aminoacids in poultry: a review. Poult Sci, 101: 101715.
- Korver DR. 2023. Review: Current challenges in poultry nutrition, health, and welfare. Anim, 17: 100755.
- Kubena LF, Kwon YM, Byrd JA, Woodward CL, Moore RW, Ziprin RL, Anderson RC, Nisbet DJ, Ricke SC. 2001. Drinking water treatment and dietary treatment effects on Salmonella enteritidis in leghorn hens during forced molt. Poult Sci, 80: 88.
- Kubinska M, Tykałowski B, Jankowski J, Koncicki A. 2014. Immunological and biochemical indicators in turkeys fed diets with a different Methionine content. Pol J Vet Sci, 17: 687-695.

Lee SI, Kim HS, Kim I. 2015. Microencapsulated organic acid blend with MCFAs can be used as an alternative to antibiotics for laying hens. Turk J of Vet Anim Sci, 39: 520-527.

- Leibetseder J. 1995. Studies on the effects of L-carnitine in poultry. Arch Anim Nutr, 48: 97-108.
- Lewis ED, Meydani SN, Wu D. 2019. Regulatory role of vitamin E in the immune system and inflammation. HHS Public Access, 71(4): 487-494.
- Li L, Liu Z, Fang B, Xu J, Dong X, Yang L, Zhang Z, Guo S, Ding B. 2022. Effects of Vitamin A and K3 on immune function and intestinal antioxidant capacity of aged laying hens. Braz J Poult Sci, 24(4): 1-10.
- Liao X, Shao Y, Sun G, Yang Y, Zhang L, Guo Y, Luo X, Lu L. 2020. The relationship among gut microbiota, short-chain fatty acids, and intestinal morphology of growing and healthy broilers. Poult Sci, 99(11): 5883-5895.
- Líndez AMI, Reith W. 2021. Arginine-dependent immune responses. Cell Mol Life Sci, 78(13): 5303-5324.
- Liu G, Ajao AM, Shanmugasundaram R, Taylor J, Ball E, Applegate TJ, Selvaraj R, Kyriazakis I, Olukosi OA, Kim WK. 2023. The effects of arginine and branched-chain amino acid supplementation to reduced-protein diet on intestinal health, cecal short-chain fatty acid profiles, and immune response in broiler chickens challenged with Eimeria spp: Poult Sci, 102(7): 102773.
- Liu Y, Yang X, Xin H, Chen S, Yang C, Duan Y, Yang X. 2017. Effects of a protected inclusion of organic acids and essential oils as antibiotic growth promoter alternative on growth performance, intestinal morphology and gut microflora in broilers. Anim Sci J, 88: 1414-1424.

Loddi MM, Sato RN, Ariki J, Pedroso AA, Moraes VM, Kishibe R.

2004. Ação isolada ou combinada de antibiótico ou probiótico como promotores de crescimento em rações iniciais de frangos de corte. In: Reunião Anual da Sociedade Brasileira de Zootecnia, pp: 254.

- Manangi MK, Vazques-A˜non M, Richards JD, Carter S, Knight CD. 2015. The impact of feeding supplemental chelated trace minerals on shell quality, tibia breaking strength, and immune response in laying hens. J Appl Poult Res, 24: 316- 326.
- Mast J., Buyse J, Goddeeris B.M. 2000. Dietary L-carnitine supplementation increases antigen specific Ig G production in broiler chickens. B J Nutr, 83: 161-166.
- Mirzaaghatabar F, Saki AA, Zamani P, Aliarabi H, Matin HRH. 2011. Effect of different levels of diet methionine and metabolisable energy on broiler performance and immune system. Food Agric Immunol, 22: 93-103.
- Mohammadi FF, Seidavi A, Bouyeh M. 2023. The effect of the chelated form of trace elements in diet on weight gain, production traits, egg specific gravity, immune system, blood parameters, liver enzymes, and progesterone hormone in Ross 308 broiler breeder chickens. Ital J Anim Sci, 22(1): 524- 536.
- Mohammed AA, Jacobs JA, Murugesan GR, Cheng HW. 2018. Effect of dietary synbiotic supplement on behavioral patterns and growth performance of broiler chickens reared under heat stress. Poult Sci, 97: 1101-1108.
- Mohammed AA, Jiang S, Jacobs JA, Cheng HW. 2019. Effect of a synbiotic supplement on cecal microbial ecology, antioxidant status, and immune response of broiler chickens reared under heat stress. Poult Sci, 98: 4408-4415.
- Mohammed AA, Mahmoud MA, Zaki RS, Cheng HW. 2024. Effect of a probiotic supplement (Bacillus subtilis) on struggling behavior, immune response, and meat quality of shackled broiler chickens exposed to pre-slaughter stress. Poult Sci, 2024: 104051.
- Montout L, Poullet N, Bambou J. 2021. Systematic review of the interaction between nutrition and immunity in livestock: effect of dietary supplementation with synthetic amino acids. Animals, 11: 2813.
- Moreno FMR, Sarantinopoulos P, Tsakalidou R, Vuyst LD. 2006. The role and application of enterococci in food and health. Int J Food Microbiol, 106: 1-24.
- Morgan PM. 2021. Immune response in mammals and chickens. IgY-technology: production and application of egg yolk antibodies. Springer, Heidelberg, Germany, pp: 31-47.
- Naukkarinen A, Hippelãinen M. 1989. Development of the peripheral immune function in the chicken: A study on the bursa of Fabricius isolated from the rest of the gut-associated lymphoid tissue (GALT). Apmis Acta Pathol Microbiol Immunol Scand, 97: 787-92.
- Neveling DP, Dicks LMT. 2021. Probiotics: An Antibiotic replacement strategy for healthy broilers and productive rearing. Probiotics Antimicrob Proteins, 13: 1-11.
- Nguyen DH, Lee KY, Mohammadigheisar M, Kim IH. 2018. Evaluation of the blend of organic acids and medium-chain fatty acids in matrix coating as antibiotic growth promoter alternative on growth performance, nutrient digestibility, blood profiles, excreta microflora, and carcass quality in broilers. Poult Sci, 97(12): 4351-4358.
- O'Neal DM, Ketterson ED. 2012. Life-history evolution, hormones, and avian immune function. In: Demas GE, Nelson RJ (eds) Ecoimmunology. Oxford University Press, Inc., New York, NY, USA, pp: 142.
- Ohta Y, Tsushima N, Koide K, Kidd MT, Ishibashi T. 1999. Effect of amino acid injection in broiler breeder eggs on embryonic

growth and hatchability of chicks. Poult Sci, 78: 1493-1498.

- Onbaşılar EE, Erol H. 2007. Effects of different forced molting methods on postmolt production, corticosterone level, and immune response to sheep red blood cells in laying hens. J Appl Poult Res, 16(4): 529-536.
- Packer L, Suzuki Y. 1993. Vitamin E and alpha-lipoate: Role in antioxidant recycling and activation of the NK-B transcription factor. Mol Asp Med, 14: 229-239.
- Pal M. 2017. The role of minerals and vitamins in poultry production. Agri World, 2017: 68-71.
- Panda AK, Bhanja SK, Sunder GS. 2015. Early post-hatch nutrition on immune system development and function in broiler chickens. World's Poult Sci J, 71(2): 285-296.
- Park SY, Birkhold SG, Kubena LF, Nisbet DJ, Ricke SC. 2004. Review on the role of dietary zinc in poultry nutrition, immunity, and reproduction. Biol Trace Elem Res, 101: 147- 163.
- Pereira EPV, Tilburg MFV, Florean EOPT, Guedes MIF. 2019. Egg yolk antibodies (IgY) and their applications in human and veterinary health: A review. Int Immunopharmacol, 73: 293- 303.
- Pompeua MA, Cavalcantib LFL, Torala FLB. 2018. Effect of vitamin E supplementation on growth performance, meat quality, and immune response of male broiler chickens: A meta-analysis. Livest Sci, 208: 5-13.
- Pope JT, Walker GK, Rubio AA, Brake J, Jendza, yJA, Fahrenholzy AC. 2022. Effects of corn particle size distributions and formic acid on productive and processing performance of broilers. J Appl Poult Res, 31: 100288.
- Propulla SG. 2008. Prebiotic: an emerging functional food. pp: 47-53. In: harnessing microbial diversity for use in animal nutrition. Natl Inst Anim Nutr Physiol, Adugodi, Bangalore, India, pp: 412.
- Ratcliffe MJH, Jacobsen KA. 1994. Rearrangement of immunoglobulin genes in chicken B cell development. Semin Immunol, 6: 175-84.
- Ratcliffe MJH. 2006. Antibodies, immunoglobulin genes and the bursa of Fabricius in chicken B cell development. Dev Comp Immunol, 30(1-2): 101-118.
- Ravindran V. 2010. Poultry feed availability and nutrition in developing countries. Poultry Development Review. Food and Agriculture Organization of the United Nations.
- Rebouche CJ. 1992. Carnitine function and requirements during the life cycle. Faseb J, 6: 3379-3386.
- Rehman A, Arif M, Sajjad N, Al-Ghadi MQ, Alagawany M, Abd El-Hack ME, Alhimaidi AR, Elnesr SS, Almutairi BO, Amran RA, Hussein EOS, Swelum AA. 2020. Dietary effect of probiotics and prebiotics on broiler performance, carcass, and immunity. Poult Sci, 99: 6946-6953.
- Rehman ZU, Meng C, Umar S, Munir M, Ding C. 2016. Interaction of infectious bursal disease virus with the immune system of poultry. World Poult Sci J, 72: 805-820.
- Ren H, Vahjen W, Dadi T, Saliu EM, Zentek J. 2019. Synergistic effects of probiotics and phytobiotics on the intestinal microbiota in young broiler chicken. Microorganisms, 7: 684.
- Rochell SA, Parsons HC, Dilger R. 2016. Influence of dietary amino acids reductions and Eimeria acervulina infection on growth performance and intestinal cytokine response of broilers fed low crude protein diets. Poult Sci, 95: 2602-2614.
- Rodriguez PC, Ochoa AC, Al-Khami AA. 2017. Arginine metabolism in myeloid cells shapes innate and adaptive immunity. Front Immunol, 8: 93.
- Romo MR, Perez-Martinez D, Ferrer CC. 2016. Innate immunity in vertebrates: an overview. Immunol, 148(2): 125-39.

Sadiq RK, Abrahimkhil MA, Rahimi N, Banuree SZ, Banuree SAH.

2023. Effects of dietary supplementation of Vitamin E on growth performance and immune system of broiler chickens. J World Poult Res, 13(1): 120-126.

- Sahin K, Sahin N, Kucuk O. 2003. Effects of chromium and ascorbic acid supplementation on growth, carcass traits, serum metabolites, and antioxidant status of broiler chickens reared at a high ambient temperature (32°C). Nutr Res, 23(2): 225-238.
- Santos DTT, Corzo A, Kidd MT, McDaniel CD, Torres FRA, Araujo LF. 2010. Influence of in ovo inoculation with various nutrients and egg size on broiler performance. J Appl Poult Res, 19: 1-12.
- Sarıca Ş, Karataş Ü, Gözalan R. 2009. Kanatlılarda bağışıklık sistemi ve bağışıklık sistemini etkileyen besinsel faktörler. GOÜ Ziraat Fakültesi Dergisi, 26(2): 81-86.
- Saripinar-Aksu D, Aksu T, Onel SE. 2012. Does inclusion at low levels of organically complexed minerals versus inorganic forms create a weakness in performance or antioxidant defense system in broiler diets? Int J Poult Sci, 11(10): 666- 672.
- Sayed Y, Hassan M, Salem HB, Al‑Amry K, E. Eid G. 2023. Prophylactic influences of prebiotics on gut microbiome and immune response of heat-stressed broiler chickens. Nature Scientific Reports, 13: 13991.
- Schneider AF, Zimmermann OF, Gewehr CE. 2017. Zeolites in poultry and swine production. Ciencia Rural, 47: 1-8.
- Seifert C, Fritz C, Carlini N, Barth SW, Franz CMAP, Watzl B. 2011. Modulation of innate and adaptive immunity by the probiotic Bifidobacterium longum PCB133 in turkeys. Poult Sci, 90: 2275-2280.
- Selaledi LA, Hassan ZM, Manyelo TG, Mabelebele M. 2020. The current status of the alternative use to antibiotics in poultry production: an african perspective. Antibiotics, 9: 594.
- Selim S, Abdel-Megeid NS, Abou-Elnaga MK, Mahmoud SF. 2021. Early nutrition with different diets composition versus fasting on immunity-related gene expression and histomorphology of digestive and lymphoid organs of layertype chicks. Animals, 11: 1568.
- Shaji S, Selvaraj RK, Shanmugasundaram R. 2023. Salmonella infection in poultry: a review on the pathogen and control strategies. Microorganisms, 11: 2814.
- Shanmugasundaram, R., Morris, A., Selvaraj, R., 2019. Effect of 25-hydroxycholecalciferol supplementation on turkey performance and immune cell parameters in a coccidial infection model. Poult Sci, 98: 1127-1133.
- Sharma MK, Lee J, Shi H, Ko H, Goo D, Paneru D, Holladay SD, Gogal RM, Kim WK. 2024. Effect of dietary inclusion of 25 hydroxyvitamin D and vitamin E on performance, gut health, oxidative status, and immune response in laying hens infected with coccidiosis. Poult Sci, 103: 104033.
- Shehata AA, Yalçın S, Latorre JD, Basiouni S, Attia YA, El-Wahab AA, Visscher C, El-Seedi HR, Huber C, Hafez HM, Eisenreich W, Tellez-Isaias G. 2022. Probiotics, prebiotics, and phytogenic substances for optimizing gut health in poultry. Microorganisms, 10: 395.
- Shimizu M, Nagashima H, Sano K, Hashimoto K, Ozeki M, Tsuda K, Hatta H. 1992. Molecular stability of chicken and rabbit immunoglobulin G. Biosci Biotech Biochem, 56(2): 270-274.
- Shojadoost B, Yitbarek A, Alizadeh M, Kulkarni RR, Astill J, Boodhoo N, Sharif S. 2021. Centennial review: effects of vitamins A, D, E, and C on the chicken immune system. Poult Sci, 100(4): 100930.
- Silverman MN, Pearce BD, Biron CA, Miller AH. 2005. Immune modulation of the hypothalamic-pituitary-adrenal (hpa) axis during viral infection. Viral Immunol, 18(1): 41-78.
- Sittiya J, Nii T. 2024. Effects of oligosaccharides on performance, intestinal morphology, microbiota and immune reactions in laying hens challenged with dextran sodium sulfate. Poult Sci, 2024: 104062.
- Śliżewska K, Markowiak-Kopeć P, Żbikowski A, Szeleszczuk P. 2020. The effect of synbiotic preparations on the intestinal microbiota and her metabolism in broiler chickens. Sci Rep, 10: 4281.
- Song B, Tang D, Yan S, Fan H, Li G, Shahid MS, Mahmood T and Guo Y. 2021. Effects of age on immune function in broiler chickens. J Anim Sci Biotechnol, 12: 42.
- Spring P, Wenk C, Dawson KA, Newman KE. 2000. The effects of dietary mannan-oligosaccharides on cecal parameters and the concentrations of enteric bacteria in the ceca of Salmonella challenged broiler chicks. Poult Sci, 79: 205-211.
- Subramaniyan SA, Kang DA, Park JR, Siddiqui SH, Ravichandiran P, Jing D, Sam Na C, Shim KS. 2019. Effect of in ovo injection of l-arginine in different chicken embryonic development stages on post-hatchability, immune response, and myo-d and myogenin proteins. Animals, 9(6): 357.
- Sun Y, O'Riordan MX. 2013. Regulation of bacterial pathogenesis by intestinal short-chain fatty acids. Adv Appl Microbiol, 85: 93-118.
- Sunder GS, Panda AK, Gopinath NCS, Rao SVR, Raju MVLN, Reddy MR, Kumar CV. 2008. Effects of higher levels of zinc supplementation on performance, mineral availability, and immune competence in broiler chickens. J Appl Poult Res, 17: 79-86.
- Swain BK, Johri TS. 2000. Effect of supplemental methionine, choline and their combinations on the performance and immune response of broilers. Br Poult Sci, 41: 83-88.
- Tawfeeq WS, Al-Mashhdani HE. 2020. Effect of adding propionic acid, formic acid, and antibiotics to broiler diet on the production performance, some histological traits, and microbial characteristics. Plant Arch, 20: 468-472.
- Tayeb IH, Qader GK. 2012. Effect of feed supplementation of selenium and Vitamin E on production performance and some hematological parameters of broiler. KSU Doğa Bil Derg, 15(3): 46-56.
- Uni Z, Yadgary L, Yair R. 2012. Nutritional limitations during poultry embryonic development. J Appl Poult Res, 21: 175- 184.
- Vulevic J, Rastall RA, Gibson GR. 2004. Developing a quantitative approach for determining the in vitro prebiotic potential of dietary oligosaccharides. FEMS Microbiol Lett, 236: 153-159.
- Wei J, Li L, Peng Y, Luo J, Chen T, Xi Q, Zhang Y, Sun J. 2024. The effects of optimal dietary vitamin D3 on growth and carcass performance, tibia traits, meat quality, and ıntestinal morphology of chinese yellow-feathered broiler chickens.

Animals, 14: 920.

- Wickramasuriya SS, Ault J, Ritchie S, Gay CG, Lillehoj HS. 2024. Alternatives to antibiotic growth promoters for poultry: a bibliometric analysis of the research journals. Poult Sci, 103(9): 103987.
- Wlazlak S, Pietrzak E, Biesek J, Dunislawska A. 2023. Modulation of the immune system of chickens a key factor in maintaining poultry production - a review. Poult Sci, 102: 102785.
- Wynn JL, Levy O. 2010. Role of Innate Host Defenses in Susceptibility to Early Onset Neonatal Sepsis. Clin Perinatol, 37(2): 307-337.
- Xu H, Lu Y, Li D, Yan C, Jiang Y, Hu Z, Zhang Z, Du R, Zhao X, Zhang Y, Tian Y, Zhu Q, Liu Y, Wang Y. 2023. Probiotic mediated intestinal microbiota and improved performance, egg quality and ovarian immune function of laying hens at different laying stage. Front Microbiol, 14: 1041072.
- Xu YQ, Guo YW, Shi BL, Yan SM, Guo XY. 2018. Dietary arginine supplementation enhances the growth performance and immune status of broiler chickens. Livest Sci, 209: 8-13.
- Yang X, Xin H, Yang C, Yang X. 2018. Impact of essential oils and organic acids on the growth performance, digestive functions and immunity of broiler chickens. Anim Nutr, 4: 388-393.
- Yasuda M, Kajiwara E, Ekino S, Taura Y, Hirota Y, Horiuchi H, Matsuda H, Furusawa S. 2003. Immunobiology of chicken germinal center: I. Changes in surface Ig class expression in the chicken splenic germinal center after antigenic stimulation. Dev Comp Immunol, 27(2): 159-166.
- Yeşilyurt N, Yılmaz B, Ağagündüz D, Capasso R. 2021. Involvement of probiotics and postbiotics in the immune system modulation. Biologics, 1: 89-110.
- Yu J, Dong B, Zhao M, Liu L, Geng T, Gong D, Wang J. 2021. Dietary clostridium butyricum and bacillus subtilis promote goose growth by improving intestinal structure and function, antioxidative capacity and microbial composition. Animals, 11: 3174.
- Yu Y, Li Q, Zeng X, Xu Y, Jin K, Liu J, Cao G. 2022. Effects of probiotics on the growth performance, antioxidant functions, immune responses, and caecal microbiota of broilers challenged by Lipopolysaccharide. Front Vet Sci, 9: 846649.
- Zhai W, Gerard PD, Pulikanti R, Peebles ED. 2011. Effects of in ovo injection of carbohydrates on embryonic metabolism, hatchability, and subsequent somatic characteristics of broiler hatchlings. Poult Sci, 90: 2134-43.
- Zhou P, Tan YQ, Zhang L, Zhou YM, Gao F, Zhou GH. 2014. Effects of dietary supplementation with the combination of zeolite and attapulgite on growth performance, nutrient digestibility, secretion of digestive enzymes and intestinal health in broiler chickens. Asian-Australas J Anim Sci, 27: 1311.